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**Permian *Circulipuncturites discinispuris* Labandeira, Wang, Zhang, Bek et
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Ichnotaxon of a Punch-and-Sucking Insect on Noeggerathialean Spores**

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28
29**Abstract**

30 The generic name *Discinispora* Wang, Zhang, Bek et Pfefferkorn was originally created for
31 spores with an operculum-like structure that were found in a permineralized Noeggeranthialean
32 cone. Subsequently it was observed that up to three round and smooth openings can occur in
33 different positions on the surface of a single spore. In light of the new observations, the previous
34 interpretation as an operculum cannot be sustained. An interpretation implicating insect punch-
35 and-sucking activity was suggested for these round structures. This new interpretation makes it
36 necessary to withdraw the original diagnosis and the taxon. The insect-inflicted damage now is
37 assigned to the ichnotaxon *Circulipuncturites discinisporeis* Labandeira, Wang, Zhang, Bek et
38 Pfefferkorn under the rules of the ICZN, rather than those of the ICBN that typified the insect-
39 damaged host-plant spore.

40
41 Key words: *Discinispora*, ichnotaxon, *Circulipuncturites*, insect damage, Thysanoptera

42
43**Introduction**

44 During an investigation of the morphology of the spores from the cone *Discinites sinensis*
45 Wang, which is the sole petrified species of the Noeggerathiales and bears significant
46 information on the systematic position of the order, a circular opening was recognized in a
47 number of the microspores. Based on the consistent dimensions, morphology, and position of
48 these circular apertures, it was interpreted as an original reproductive structure of the spore and
49 this palynomorph was published as the new genus and species *Discinispora sinensis* Wang,
50 Zhang, Bek et Pfefferkorn (Wang et al., 2004). However, during further study on the petrified
51 noeggeranthialean strobilus, it came to our attention that this structure might be the result of
52 animal activity. It was not until recently that subsequent critical examination of in situ spores of

53 *Discinites sinensis* Wang provided evidence that the opening could not be an operculum, that is,
54 an integral part of the spore. Therefore, given this new interpretation, it is necessary to establish
55 a new generic name for this insect-mediated damage according to the International Code of
56 Zoological Nomenclature under Articles 2.1 and 10.5 (Ride et al., 1999). The name *Discinispora*
57 *sinensis* Wang, Zhang, Bek et Pfefferkorn has to be abandoned as an empty name because spores
58 of this cone that lack the circular damage described herein could be called *Deltoidospora* Miner,
59 *Calamospora* Schopf, or *Punctatisporites* Ibrahim.

60 Our critical examination of the in situ spores of *Discinites sinensis* Wang is based on 1000
61 spores from two sporangia of the holotype (Wang, 2000, 2005). Methods have been described in
62 detail by Wang (2000). For the current study, isolated sporangia were macerated with HF (36
63 percent) for about 24 hours and washed with distilled water. Spores were etched with ethanol and
64 coated with gold for observation in a LEO1530BP Scanning Electron Microscope (SEM). The
65 sporangia of *Discinites sinensis* Wang have been fragmental or broken to some degree due to
66 weathering. There is not a certainly complete sporangium available for a critical investigation of
67 the insect damage on sporangial walls.

68 **Systematic Paleontology**

69
70 According to Article 10.3 of the International Code of Zoological Nomenclature (Ride et al.,
71 1999), a name proposed for an ichnotaxon is a family group name, a genus-group name, or a
72 species-group name, based on the way in which it is first established. We propose a genus-group
73 name.

74
75 GENUS *Circulipuncturites discinisporeis* Labandeira, Wang, Zhang, Bek et Pfefferkorn

76 **Diagnosis.** A circular orifice, 40–60 μm in diameter, representing severance of the spore wall
77 and from one to three occurrences per spore that are variably positioned on the exposed surface.

78 **Host plant.** This distinctive damage occurs on the spores of *Discinites sinensis* Wang that
79 belongs to the late Paleozoic Noeggerathiales.

80 **Holotype.** The spores of *Discinites sinensis* Wang figured by Wang, Zhang, Bek and
81 Pfefferkorn (2004, Plate 1, fig. 4), here refigured in Plate 1, fig. 1.

82 **Type locality.** Shitanjing, Shizuishan, Ningxia, China.

83 **Stratigraphic horizon.** The Lower Shihhotse Formation; Early Permian (Late Artinskian to
84 Kungurian) in age.

85 **Etymology.** From the Latin (masculine): *circulus*, for “small circle,” and *punctura*, for
86 “hole.”

87 **Description.** For a critical examination on the presence or absence of these orifices, two
88 complete sporangia were isolated from the cone *Discinites sinensis* Wang, resulting in about
89 1000 spores after maceration. After checking our SEM observations, one to three circular
90 openings per spore were found on more than 50 spores, of which six were selected and are
91 shown in Plate 1, figs. 2–8. These orifices are usually circular (Plate 1, figs. 1–5, 6 for lower and
92 left orifices, 7), although occasionally some are slightly irregular (Plate 1, fig. 6 for the upper-
93 right orifice). The diameter typically ranges between 40 and 60 μm but the total range is between
94 10 and 80 μm . The apertures usually occur on the lateral and distal walls (Plate 1, figs. 1–6) but
95 also on the proximal spore wall (Plate 1, figs. 7, 8). They always traverse all three layers of the
96 spore wall (Wang et al., 2004). Occasionally spores have been incompletely penetrated,
97 preserving a disk preserved in place prior to breakage (Plate 1, fig. 2) whereas other examples
98 have the dislocated and fragmented disc occurring within the spore itself (Plate 1, fig. 4).

99
100**Remarks**

101 The fact that these highly stereotyped, recurring, circular openings can occur at any
102 orientation on the spore surface excludes the possibility that they are a functional part of the
103 spore, such as an operculum. Thus, an alternative explanation has been sought. The highly
104 regular, circular shape of the opening suggests a biological explanation, possibly by the action of
105 an organism such as a fungus or an insect. Although fungal predation on spores is known
106 (Sinclair et al., 1987), no known extant fungus produces the *Circulipuncturites discinisporis* type
107 of damage.

108 Sporophagy is a common feeding habit among a variety of extant microarthropods, including
109 springtails (Collembola), thrips (Thysanoptera), and major clades of holometabolous insects,
110 particularly beetles (Coleoptera), sawflies (Hymenoptera) and moths (Lepidoptera). Much of the
111 consumption of spores, including sporangial and synangial tissues, is accomplished by relatively
112 large, mandibulate larval and adult external feeders, particularly beetles (Leschen and Lawrence,
113 1991). However, taxa with comparatively small adult and larval body sizes are known to
114 consume spores within enclosed sporangia, including the curculionid weevils *Endalus celatus*
115 and *Echinochneumus* sp. that bore small apertures for entry into sporocarps of *Marsilea* sp. (Board
116 and Burke, 1971; Loyal and Kumar, 1977); the tenthredinid sawfly *Teichobia filicivora* that
117 feeds in the sori of *Dryopteris borreri* and *Pteridium aquilinum* (Ottosson and Anderson, 1983);
118 and the primitive oecophorid moth *Erineda aenea* on *Asplenium angustifolium*, which produces a
119 hollowed indusium after feeding (Braun, 1918). Examples also are known from the fossil record,
120 prominently the presence of pollen-laden coprolites within the pollen sacs of a pollen cone from
121 the Middle Triassic of Antarctica (Klavins et al., 2005). In the present case, we have not seen any
122 evidence that the sporangial wall was ruptured, but it is highly possible that certain apertures for

123 entry into the sporangia are too small to recognize. No coprolites are detected. They might have
124 been lost during maceration. The spores are considered to be nearly but not fully mature, since
125 they are in situ and completely mature spores would have been dispersed (Wang, 2005).

126 After a review of the modern literature on phytophagous insects, we noted similarities
127 between the circular openings of *Circulipuncturites discinisporis* on Permian spores of *Discinites*
128 *sinensis* Wang and similar damage inflicted on a variety of modern angiosperm pollen by thrips
129 (Grinfel'd, 1960; Kirk, 1984, 1985, 1987, 1997; Plate 2, fig. 1). Thrips are a moderately speciose
130 group of hemipteroid, piercing-and-sucking insects constituting the order Thysanoptera, and are
131 comprised of approximately 4500 described species that are rarely longer than 3 mm in body
132 length (Lewis, 1973, Ananthkrishnan, 1984). Thysanoptera are associated predominantly with
133 plants, and are common associates of flowering plants (Kirk, 1997, Childers, 1997) and cycads
134 (Terry, 2001). Their economic importance is considerable because of their damage to surface
135 tissues, frequently as circular damaged areas in epidermis (Plate 2, fig. 6), mesophyll,
136 parenchyma, and ovular surfaces (Plate 2, fig. 7) in plants of natural and managed ecosystems
137 (Kirk, 1997; Childers, 1997; Kindt, 2004).

138 The distinctive ensemble of downwardly directed, asymmetric, fluid feeding mouthparts in
139 the Thysanoptera consists of a central, tapering mouthcone that houses a single, stout mandibular
140 stylet and two, more gracile maxillary stylets that are used to channel imbibed fluids into the
141 esophagus (Risler, 1957; Mickoleit, 1963; Heming, 1993). Feeding on plants typically targets the
142 protoplasts of individual cells in surface tissues, but also pollen. For pollen, thysanopterans use
143 the punch-and-suck mechanism (Kirk, 1984), activated by powerful cibarial musculature within
144 the head capsule, and characterized by an initial probe along the outer wall by the mandibular
145 stylet (Heming, 1993). The pollen grain is secured by the mouthcone, maxillary and labial palps

146 (Plate 2, fig. 1) and is buttressed by robust prothoracic legs from the right and left sides
147 (Grinfel'd, 1960; Kirk, 1984). After penetration and subsequent insertion of channeled maxillary
148 stylets, the protoplast is imbibed (Chisholm and Lewis, 1984), often as the pollen grain is held in
149 midair attached to the mouthcone opening. Thrips also invade the confined interstices of
150 fructifications, gaining purchase and feeding on individual pollen grains anchored to a solid
151 substrate surface by prothoracic legs, palps, and an adpressing mouthcone (Kirk, 1996). After
152 stylet withdrawal, a circular region of surface damage is accentuated, accompanied by pollen
153 wall collapse within a circular area (Childers and Achor, 1984; Plate 2, figs. 4, 5). Frequently
154 there is evidence of initial stylet penetration, in the form of a figure-8 hole representing the dual
155 puncture events initially by the mandibular stylet and thereafter by the paired maxillary stylets
156 (Chisholm and Lewis, 1984; Kirk, 1997). This distinctive type of microscale piercing-and-
157 sucking damage occurs at the cellular level and has been difficult to document unambiguously in
158 the plant fossil record.

159 The occurrence of multiple holes of *Circulipuncturites discinisporis* for particular *Discinites*
160 *sinensis* spores may indicate a hard-wired feeding behavior. Repetitive feeding routines are
161 generally common among insects (Chapman, 1982), and have been specifically documented
162 from three-dimensional sections in piercing-and-sucking taxa such as the aphid *Aphis fabae*
163 (Hemiptera), which produce radiating multiple probes from the same localized stylet insertion
164 point for targeting individual cell protoplasts (Tjallingii and Hogen Esch, 1993). This is a
165 stereotyped feeding pattern that has been found in Early Devonian land plants as well, attributed
166 to an unknown microarthropod (Labandeira, 2002, fig. 2.8d). A parallel example occurs for
167 modern thrips, which produce linear tracks of stylate puncture marks on leaf surfaces, indicating
168 a repetitive puncturing of epidermal tissue along particular ambulatory trajectories, such as

169 *Selenothrips rubrocinctus* on the cashew *Anacardium occidentale* (Fennah, 1963). This repeated
170 pattern of linear punctures extends to the Late Permian of South Africa, expressed by Damage
171 Type 138 (Labandeira et al., 2007, p. 10), which occurs along the intersection of midrib and
172 blade for particular glossopterid foliage. An alternative explanation for the occurrence of these
173 multiple probes on Permian spores is related to ingestion limitations based on spore protoplast
174 volumes that would exceed a single feeding event. Such a hypothesis remains untested because
175 the body size and digestive tract volume of the hypothetical feeding insect remains unknown.

176 Punch and sucking has originated independently multiple times among small
177 microarthropods. In addition to Thysanoptera, punch-and-suckers presently include members of
178 forcipine biting midges (Diptera: Ceratopogonidae) that similarly have robust piercing
179 mouthparts (Downes, 1955; Labandeira, 2005). As well, piercing-and-sucking may occur among
180 members of primitively entognathan insects such as the Protura and some springtails (Schaller
181 and Wolter, 1963; François, 1969), and in certain small beetles such as the Leiodidae,
182 Eucinetidae and Cerylonidae (Besuchet, 1972; Vit, 1981; Pakaluk, 1987). These assemblages of
183 taxa have developed gracile, multistylate, piercing-and-sucking mouthparts for consumption of
184 fungal hyphae, algal filaments and plant cells, and are known to ingest intact spores and pollen
185 grains (Labandeira, 1997), but have not been suspected of consuming pollen protoplasts through
186 punch-and-sucking. Primitively entognathous insects are both known from the terrestrial
187 Paleozoic (Riek, 1976; Greenslade, 1986), and Protura and probably piercing-and-sucking
188 springtails were present during the Permian; however, it is not clear that the mouthparts of any
189 members of these groups were capable of punch-and-sucking. In addition, the presence of a
190 circular hole, with beveled edges on the surface of the lycopod megaspore *Setisisporites* sp.
191 (Scott et al., 1992), has been recorded during the Middle Pennsylvanian, preceding the earliest

192 occurrence of the basal thysanopteran clade, Lophioneuridae, by ~ 30 m.y., suggesting damage
193 inflicted by another insect group, probably a piercing-and-sucking paleodictyopteroid with
194 truncate stylate mouthparts. By contrast, from what is known of beetle phylogeny and their
195 Permian fossil record, the occurrence of *Circulipuncturites discinispuris* damage antedates
196 considerably the inferred earliest occurrences of beetle culprits that could have produced similar
197 spore damage.

198 The most likely culprit is the Lophioneuridae, an immediate, ancestral clade of the
199 Thysanoptera, or possibly a closely related stem-group or early paraphyletic lineage (Zherikhin,
200 2000; Grimaldi et al., 2004; Grimaldi and Engel, 2005; Labandeira, 2006). The Lophioneuridae
201 bore a mouthcone housing relatively symmetrical mandibular and maxillary stylets and bearing
202 typical maxillary and labial palps (Vishniakova, 1981); judging from mouthpart structure, punch-
203 and-suck feeding was likely. The earliest lophioneurid occurrence is *Cyphoneurodes patriciae*
204 Beckemeyer from the Early Permian (Artinskian) Wellington Formation of Elmo, Kansas
205 (Beckemeyer, 2004), after which they are variably represented elements in later Permian and
206 Triassic deposits (Plate 2, figs. 2, 3; Becker-Migdisova, 1991; Grimaldi et al., 2004), becoming
207 rare thereafter, and achieving relict status into the mid Cretaceous. The last known, surviving
208 taxon is *Burmacypha longicornis* Zherikhin from the Early Cretaceous (late Albian) from
209 Burmese amber in northern Myanmar (Zherikhin, 2000).

210
211

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Plates

Plate 1

342 Eight figures below shows from one to three circular punctures of *Circulipunctarites*
343 *discinisporis* Labandeira, Wang, Zhang, Bek and Pfefferkorn occurring on spores of *Discinites*
344 *sinensis* Wang.

- 345 1. Holotype of *Circulipunctarites discinisporis*, figured in Wang et al. (2004; Plate 1, fig. 4) as
346 *Discinispora sinensis* Wang, Zhang, Bek and Pfefferkorn. Scale bar = 20 µm.
- 347 2. An initially cut disk that did not separate from the spore wall. Scale bar = 20 µm.
- 348 3. An orifice on the distal wall, similar to the holotype. Scale bar = 30 µm.
- 349 4. An orifice with a diameter larger than half of the spore diameter. Scale bar = 30 µm.

- 350 5. Two orifices of different sizes on the lateral (left and smaller) and distal (right and larger)
351 spore walls, respectively. Scale bar = 30 μm .
- 352 6. Three orifices of different sizes on the lateral distal wall, of which the upper-right one has a
353 slightly irregular circular margin. Scale bar = 30 μm .
- 354 7. An orifice on the proximal spore wall. Scale = 30 μm .
- 355 8. Enlargement of the orifice in (7) at left, showing the aperture margin. Scale = 10 μm .

356
357 **Plate 2**

358 At left are modern (1) and Permian (2, 3) specimens of Thysanoptera; at right are examples of
359 thysanopteran damage to angiosperm pollen (4, 5), a leaf (6) and an ovule (7).

360 1. The punch-and-suck feeding technique in *Thrips vulgatissimus* Haliday (Thripidae) above
361 and *Aeolothrips intermedius* Bagnall (Aeolothripidae) below, showing the head profile and
362 position of the mouthcone and associated mouthparts during puncture of a pollen grain. From
363 Kirk (1997), reproduced with permission by the Commonwealth Agricultural Board
364 International, London, U.K. Scale bars = 100 μm .

365 2. An early Middle Permian (Roadian) lophioneurid, *Lophiocypha incompleta* Vishniakova,
366 from Soyana, Ural Region, Russia. The arrow points to mouthcone and stylet mouthparts.
367 From Vishniakova (1981), reproduced with permission by the Russian Academy of Sciences.
368 Scale bar = 1 mm.

369 3. A dorsal view of the head, mouthparts and antennae of a different specimen of *Lophiocypha*
370 *incompleta* Vishniakova but from the same locality than shown in (2) above. From Vishniakova
371 (1981), reproduced with permission by the Russian Academy of Sciences. Scale bar = 100 μm .

- 372 4. A SEM micrograph of a pollen grain of *Malva sylvestris* L. (Malvaceae), exhibiting a central
373 collapsed area following a feeding puncture. From Kirk (1997), used with permission by the
374 Commonwealth Agricultural Board International, London, U.K. Scale bar = 20 μm .
- 375 5. A drawing of the specimen in (4) above, showing the envelope of the circular collapsed area.
376 From Kirk (1996), used with permission from the Company of Biologists, Ltd., U.K. Scale bar =
377 20 mm.
- 378 6. A SEM micrograph showing a circular damaged area and a central figure-8 puncture by
379 *Limothrips cerealium* Haliday (Thripidae) on a *Triticum aestivium* L. (Poaceae) leaf. From Kirk
380 (1997), used with permission by Commonwealth Agricultural Board International, London, U.K.
381 Scale bar = 5 μm
- 382 7. SEM micrograph of a feeding puncture of *Frankliniella bispinosa* (Morgan) (Thripidae) on a
383 *Citrus sinensis* Pers. (Rutaceae) ovary, showing a circular damaged area with (left arrow) and
384 without (right arrow) a saliva plug. From Childers and Achor (1984), reproduced with
385 permission by the Entomological Society of America. Scale bar = 10 μm .



