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LIST OF RECENT LAND MAMMALS OF MEXICO, 2014



*JOSÉ RAMÍREZ-PULIDO, NOÉ GONZÁLEZ-RUIZ, ALFRED L. GARDNER, AND
JOAQUÍN ARROYO-CABRALES*

Front cover: Image of the cover of *Nova Plantarvm, Animalivm et Mineralivm Mexicanorvm Historia*, by Francisci Hernández et al. (1651), which included the first list of the mammals found in Mexico. Cover image courtesy of the John Carter Brown Library at Brown University.

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José Ramírez-Pulido, Noé González-Ruiz, Alfred L. Gardner, and Joaquín Arroyo-Cabrales

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Museum of Texas Tech University
Lubbock, TX 79409-3191 USA
(806)742-2442

LIST OF RECENT LAND MAMMALS OF MEXICO, 2014

JOSÉ RAMÍREZ-PULIDO, NOÉ GONZÁLEZ-RUIZ, ALFRED L. GARDNER, AND JOAQUÍN ARROYO-CABRALES

ABSTRACT

We provide an updated list of the Recent land mammals of Mexico and include information on the taxonomy of certain species, and where appropriate, the endemic and threatened status of all species listed. Several taxonomic and nomenclatural changes have been made since publication of the last list of the Mexican terrestrial mammalian fauna. Within the period from 2005 to present, there have been at least 209 changes concerning the nomenclature of this fauna; these we evaluated in this paper. The land mammals of Mexico comprise 168 genera, 496 species, and 881 subspecies.

Key words: nomenclature, mammals, Mexico, species checklist

RESUMEN

Se presenta una lista actualizada de los mamíferos terrestres nativos de México, incluyendo información acerca de la situación taxonómica de varias de las especies, así como las especies endémicas y amenazadas. Desde la última publicación de los mamíferos terrestres de México, se han realizado importantes cambios taxonómicos y nomenclaturales. En el período que abarca desde el año 2005, se han registrado al menos 209 cambios nomenclatoriales que se discuten en este trabajo. Los mamíferos terrestres de México se componen de 168 géneros, 496 especies y 881 subespecies.

Palabras clave: nomenclatura, mamíferos, México, lista de especies

INTRODUCTION AND METHODS

It has been 32 years since the first list of Mexican land mammals appeared (Ramírez–Pulido et al. 1982). This list has been periodically updated to incorporate additions and taxonomic changes to the mammal fauna. In that period, five lists have been provided (Cervantes et al. 1994; Ramírez–Pulido et al. 1982, 1983, 1996, 2005), each of which has proven to be a useful synthesis of the taxonomic changes since the previous list and facilitating rapid access to the current knowledge on the diversity of the nonmarine Mexican mammal fauna.

The past 9 years have seen many changes in the number of species. For example, Ramírez–Pulido et al. (2005) included 475 species, whereas in this list we recognize 496, an increase of 21 species in just 9 years of taxonomic study of the Mexican mammal fauna. The increase in the number of species is due not only to the recent discovery of new species, but also to the

use of new research techniques, including molecular genetics, that have resulted in a better understanding of taxonomic relationships. Only eight new species have been described in those 9 years and of those, only two (*Thomomys navarensis* and *Habromys schmidlyi*) were based on specimens previously unavailable to science. Other increases were based on specimens previously misidentified as other species, but when restudied using new techniques and different paradigms, it became clear that they did not represent the species as originally assigned. Some of these specimens represented named species previously considered synonyms of valid species; others were undescribed. There are mainly two scientific approaches that have led to the recent increase in number of species. First, molecular techniques are evolving and becoming cheaper, faster, and more commonly used in taxonomic research. Second, recent field work has provided critically needed additional

specimens, thus increasing the sample sizes necessary for morphologic and morphometric analyses, which have resulted in higher confidence levels in the analyses of variation and species limits. Nothing has been published since the last list (Ramírez–Pulido et al. 2005) that brings together in one place all of the taxonomic and nomenclatorial changes that have occurred at the generic, species, and subspecies levels. Several papers have chronicled recent taxonomic changes (Ceballos and Arroyo–Cabral 2013; Gardner 2008a; Godínez et al. 2011; Wilson and Reeder 2005) and, in fact those reports have generated the need for the discussions of taxonomic decisions we provide in this list.

We recognize the need and utility for a comprehensive list that makes any literature search easier; therefore, we compiled this list with two main objectives. First, to produce a current list of species and subspecies of Mexican land mammals, and second, a review, analysis, and discussion of the nomenclatorial and taxonomic changes from 2005 up to early 2014.

We have found that taxonomic proposals often are accepted or rejected without careful scrutiny by the scientific community as soon as these proposals are published. Taxonomy is a dynamic endeavor and all taxonomic judgments are hypotheses worthy of evaluation. Our list is not simply a collection of recent literature. We have based our evaluations of the taxonomic proposals discussed in this paper on a variety of criteria: scientific authority, proposal complexity, analysis type, and on information available to us on taxonomy, biogeography, ecology, and evolution. We emphasize that most of the information presented is not our own and acknowledge that our evaluations of these taxonomic hypotheses are based on our combined experience and involve a certain level of subjectivity.

Taxonomic ranks above the level of genus follow McKenna and Bell (1997); ranks at the generic level and below primarily follow the third edition of *Mammal Species of the World* (Wilson and Reeder 2005), including some changes proposed in *Mammals of South America* (Gardner 2008a). Within each suprageneric

level, genera are arranged alphabetically, and within each genus, species also are arranged alphabetically.

The valid name for each taxon is followed by the author's name and year of publication. The reader will notice that some of the authors and dates differ from those provided in previous lists and in other published sources (e.g., Hall 1981). These changes are the result of our nomenclatorial diligence in determining the correct author(s) of a name and the actual date of publication, which may differ from that printed.

All taxonomic and nomenclatorial changes covered in this report are those published from 2005 (after the closing date for Ramírez–Pulido et al. 2005) to March 2014. For information concerning taxonomic changes prior to 2005, we suggest reviewing the compilations by Cervantes et al. (1994), and Ramírez–Pulido et al. (1982, 1983, 1996, and 2005). Since 2005, there have been at least 209 nomenclatorial acts, which we evaluate in the next section.

We do not include recently introduced species for the following reasons: 1) we lack objective records verifying the success of the introduction (e.g., *Mustela nigripes*); 2) we lack evidence of their current presence in Mexico (*Sciurus carolinensis* and *Cervus elaphus*); and 3) the species were reintroduced into areas within their known previous distributions (e.g., *Canis lupus*, *Ovis canadensis*, *Bison bison*, and *Antilocapra americana*) and these we treat as native.

To provide some helpful data on conservation status, we identify species that are endemic (En) to Mexico, as well as the conservation status according to the Mexican Ministry of Environment and Natural Resources (NOM–059–SEMARNAT–2010; see Norma Oficial Mexicana 2010), as follows: species probably extinct in the wild (E), endangered species (P), threatened species (A), and special protection species (Pr). For geographic distribution, we distinguish between island (In) and continental (Co); for taxonomic diversity, species are listed as either monotypic (Mo) or polytypic (Po).

TAXONOMIC LIST AND CONSERVATION STATUS OF THE LAND MAMMALS OF MEXICO

	Conservation Status	Endemism	Taxonomic Diversity	Geographic Distribution
CLASS MAMMALIA Linnaeus, 1758				
COHORT MARSUPIALIA Illiger, 1811				
MAGNORDER AMERIDELPHIA Szalay in: Archer, 1982				
ORDER DIDELPHIMORPHIA Gill, 1872				
FAMILY DIDELPHIDAE Gray, 1821				
SUBFAMILY CALUROMYINAE Kirsch, 1977				
<i>Caluromys</i> J. A. Allen, 1900				
<i>Caluromys derbianus</i> (Waterhouse, 1841)	A		Po	Co
<i>Caluromys derbianus aztecus</i> (Thomas, 1913)				
<i>Caluromys derbianus fervidus</i> (Thomas, 1913)				
SUBFAMILY DIDELPHINAE Gray, 1821				
TRIBE DIDELPHINI Gray, 1821				
<i>Chironectes</i> Illiger, 1811				
<i>Chironectes minimus</i> (Zimmermann, 1780)	P		Po	Co
<i>Chironectes minimus argyrodytes</i> Dickey, 1928				
<i>Didelphis</i> Linnaeus, 1758				
<i>Didelphis marsupialis</i> Linnaeus, 1758			Po	Co–In
<i>Didelphis marsupialis cauae</i> J. A. Allen, 1900				
<i>Didelphis virginiana</i> Kerr, 1792			Po	Co–In
<i>Didelphis virginiana californica</i> Bennett, 1833				
<i>Didelphis virginiana yucatanensis</i> J. A. Allen, 1901				
<i>Philander</i> Brisson, 1762				
<i>Philander opossum</i> (Linnaeus, 1758)			Po	Co
<i>Philander opossum fuscogriseus</i> (J. A. Allen, 1900)				
TRIBE MARMOSINI Hershkovitz, 1992				
<i>Marmosa</i> Gray, 1821				
<i>Marmosa mayensis</i> Osgood, 1913		En	Mo	Co
<i>Marmosa mexicana</i> Merriam, 1897			Po	Co
<i>Marmosa mexicana mexicana</i> Merriam, 1897				
<i>Tlacuatzin</i> Voss and Jansa, 2003				
<i>Tlacuatzin canescens</i> (J. A. Allen, 1893)		En	Po	Co–In
<i>Tlacuatzin canescens canescens</i> (J. A. Allen, 1893)				
<i>Tlacuatzin canescens gaumeri</i> (Osgood, 1913)				
<i>Tlacuatzin canescens insularis</i> (Merriam, 1898)				

TRIBE METACHIRINI Reig, Kirsch, and Marshall, 1985				
<i>Metachirus</i> Burmeister, 1854				
	<i>Metachirus nudicaudatus</i> (É. Geoffroy Saint-Hilaire, 1803)	A	Po	Co
	<i>Metachirus nudicaudatus colombianus</i> J. A. Allen, 1900			
COHORT PLACENTALIA Owen, 1837				
MAGNORDER XENARTHRA Cope, 1889				
ORDER CINGULATA Illiger, 1811				
SUPERFAMILY DASYPODOIDEA Gray, 1821				
FAMILY DASYPODIDAE Gray, 1821				
SUBFAMILY DASYPODINAE Gray, 1821				
<i>Dasypus</i> Linnaeus, 1758				
	<i>Dasypus novemcinctus</i> Linnaeus, 1758		Po	Co-In
	<i>Dasypus novemcinctus mexicanus</i> Peters, 1864			
SUBFAMILY TOLYPEUTINAE Gray, 1865				
<i>Cabassous</i> McMurtrie, 1831				
	<i>Cabassous centralis</i> (Miller, 1899)	P	Mo	Co
ORDER PILOSA Flower, 1883				
SUBORDER VERMILINGUA Illiger, 1811				
FAMILY MYRMECOPHAGIDAE Gray, 1825				
<i>Tamandua</i> Gray, 1825				
	<i>Tamandua mexicana</i> (de Saussure, 1860)		Po	Co
	<i>Tamandua mexicana mexicana</i> (de Saussure, 1860)	P		
FAMILY CYCLOPEDIDAE Pocock, 1924				
<i>Cyclopes</i> Gray, 1821				
	<i>Cyclopes didactylus</i> (Linnaeus, 1758)	P	Po	Co
	<i>Cyclopes didactylus mexicanus</i> Hollister, 1914			
ORDER SORICOMORPHA Gregory, 1910				
SUPERFAMILY SORICOIDEA G. Fischer, 1814				
FAMILY SORICIDAE G. Fischer, 1814				
SUBFAMILY SORICINAE G. Fischer, 1814				
TRIBE BLARININI G. Fischer, 1814				
<i>Cryptotis</i> Pomel, 1848				
	<i>Cryptotis alticola</i> (Merriam, 1895)	Pr	En	Mo
	<i>Cryptotis goldmani</i> (Merriam, 1895)	Pr	En	Po
	<i>Cryptotis goldmani goldmani</i> (Merriam, 1895)			
	<i>Cryptotis goldmani machetes</i> (Merriam, 1895)			
	<i>Cryptotis goodwini</i> Jackson, 1933		Mo	Co

<i>Cryptotis griseiventris</i> Jackson, 1933		En	Mo	Co
<i>Cryptotis magnus</i> (Merriam, 1895)	Pr	En	Mo	Co
<i>Cryptotis mayensis</i> (Merriam, 1901)	Pr		Mo	Co
<i>Cryptotis merriami</i> Choate, 1970			Mo	Co
<i>Cryptotis mexicanus</i> (Coues, 1877)		En	Mo	Co
<i>Cryptotis nelsoni</i> (Merriam, 1895)	Pr	En	Mo	Co
<i>Cryptotis obscurus</i> (Merriam, 1895)	Pr	En	Mo	Co
<i>Cryptotis parvus</i> (Say, 1822)			Po	Co
<i>Cryptotis parvus berlandieri</i> (Baird, 1857)				
<i>Cryptotis parvus pueblensis</i> Jackson, 1933				
<i>Cryptotis parvus soricinus</i> (Merriam, 1895)	Pr			
<i>Cryptotis peregrina</i> (Merriam, 1895)	Pr	En	Mo	Co
<i>Cryptotis phillipsii</i> (Schaldach, 1966)		En	Mo	Co
<i>Cryptotis tropicalis</i> (Merriam, 1895)	Pr		Mo	Co
TRIBE NOTIOSORICINI Reumer, 1984				
<i>Megasorex</i> Hibbard, 1950		En		
<i>Megasorex gigas</i> (Merriam, 1897)	A	En	Mo	Co
<i>Notiosorex</i> Coues, 1877				
<i>Notiosorex cockrumi</i> R. J. Baker, O'Neill, and McAliley, 2003			Mo	Co
<i>Notiosorex crawfordi</i> (Coues, 1877)	A		Mo	Co
<i>Notiosorex evotis</i> (Coues, 1877)	A	En	Mo	Co
<i>Notiosorex villai</i> Carraway and Timm, 2000	A	En	Mo	Co
TRIBE SORICINI G. Fischer, 1814				
<i>Sorex</i> Linnaeus, 1758				
<i>Sorex arizonae</i> Diersing and Hoffmeister, 1977	P		Mo	Co
<i>Sorex emarginatus</i> Jackson, 1925		En	Mo	Co
<i>Sorex ixtlanensis</i> Carraway, 2007		En	Mo	Co
<i>Sorex macrodon</i> Merriam, 1895	A	En	Mo	Co
<i>Sorex mediopua</i> Carraway, 2007		En	Mo	Co
<i>Sorex milleri</i> Jackson, 1947	Pr	En	Mo	Co
<i>Sorex monticola</i> Merriam, 1890	A		Po	Co
<i>Sorex monticola monticola</i> Merriam, 1890				
<i>Sorex oreopolus</i> Merriam, 1892		En	Mo	Co
<i>Sorex orizabae</i> Merriam, 1895		En	Mo	Co
<i>Sorex ornatus</i> Merriam, 1895			Po	Co
<i>Sorex ornatus juncensis</i> Nelson and Goldman, 1909	A			
<i>Sorex ornatus lagunae</i> Nelson and Goldman, 1909				
<i>Sorex ornatus ornatus</i> Merriam, 1895	A			
<i>Sorex saussurei</i> Merriam, 1892		En	Mo	Co
<i>Sorex sclateri</i> Merriam, 1897	A	En	Mo	Co

<i>Sorex stizodon</i> Merriam, 1895	A	En	Mo	Co
<i>Sorex ventralis</i> Merriam, 1895		En	Mo	Co
<i>Sorex salvini</i> Merriam, 1897		En	Po	Co
<i>Sorex salvini altoensis</i> Carraway, 2007				
<i>Sorex salvini cristobalensis</i> Jackson, 1925	A			
<i>Sorex salvini oaxacae</i> Jackson, 1925	A			
<i>Sorex salvini veraecrucis</i> Jackson, 1925	Pr			
<i>Sorex veraepacis</i> Alston, 1877		En	Po	Co
<i>Sorex veraepacis chiapensis</i> Jackson, 1925	A			
<i>Sorex veraepacis mutabilis</i> Merriam, 1898	A			
<i>Sorex veraepacis veraepacis</i> Alston, 1877				
SUPERFAMILY TALPOIDEA G. Fischer, 1814				
FAMILY TALPIDAE G. Fischer, 1814				
SUBFAMILY SCALOPINAE Gill, 1875				
TRIBE SCALOPINI Trouessart, 1879				
<i>Scalopus</i> È. Geoffroy Saint-Hilaire, 1803				
<i>Scalopus aquaticus</i> (Linnaeus, 1758)	P		Po	Co
<i>Scalopus aquaticus inflatus</i> Jackson, 1914				
<i>Scalopus aquaticus montanus</i> R. H. Baker, 1951				
<i>Scapanus</i> Pomel, 1848				
<i>Scapanus anthonyi</i> J. A. Allen, 1893	P	En	Mo	Co
<i>Scapanus latimanus</i> (Bachman, 1842)	A		Po	Co
<i>Scapanus latimanus occultus</i> Grinnell and Swarth, 1912				
ORDER CHIROPTERA Blumenbach, 1779				
SUBORDER MICROCHIROPTERA Dobson, 1875				
INFRAORDER incertae sedis Simmons, 1998				
SUPERFAMILY EMBALLONUROIDEA Weber, 1928				
FAMILY EMBALLONURIDAE Gervais, 1856				
SUBFAMILY EMBALLONURINAE Gervais, 1856				
<i>Balantiopteryx</i> Peters, 1867				
<i>Balantiopteryx io</i> Thomas, 1904			Mo	Co
<i>Balantiopteryx plicata</i> Peters, 1867			Po	Co-In
<i>Balantiopteryx plicata pallida</i> Burt, 1948				
<i>Balantiopteryx plicata plicata</i> Peters, 1867				
<i>Centronycteris</i> Gray, 1838				
<i>Centronycteris centralis</i> Thomas, 1912	Pr		Mo	Co
<i>Diclidurus</i> Wied-Neuwied, 1820				
<i>Diclidurus albus</i> Wied-Neuwied, 1820			Po	Co
<i>Diclidurus albus virgo</i> Thomas, 1903				
<i>Peropteryx</i> Peters, 1867				

<i>Peropteryx kappleri</i> Peters, 1867			Po	Co
<i>Peropteryx kappleri kappleri</i> Peters, 1867	Pr			
<i>Peropteryx macrotis</i> (J. A. Wagner, 1843)			Mo	Co
<i>Rhynchonycteris</i> Peters, 1867				
<i>Rhynchonycteris naso</i> (Wied–Neuwied, 1820)	Pr		Mo	Co
<i>Saccopteryx</i> Illiger, 1811				
<i>Saccopteryx bilineata</i> (Temminck, 1838)			Mo	Co
<i>Saccopteryx leptura</i> (Schreber, 1774)	Pr		Mo	Co
INFRAORDER YANGOCHIROPTERA Koopman, 1984				
SUPERFAMILY MOLOSSOIDEA Gervais, 1856				
FAMILY MOLOSSIDAE Gervais, 1856				
SUBFAMILY MOLOSSINAE Gervais, 1856				
<i>Cynomops</i> Thomas, 1920				
<i>Cynomops mexicanus</i> (J. K. Jones and Genoways, 1967)	Pr	En	Mo	Co
<i>Eumops</i> Miller, 1906				
<i>Eumops auripendulus</i> (Shaw, 1800)			Po	Co
<i>Eumops auripendulus auripendulus</i> (Shaw, 1800)				
<i>Eumops ferox</i> (Gundlach, 1861)			Mo	Co
<i>Eumops hansae</i> Sanborn, 1932			Mo	Co
<i>Eumops nanus</i> (Miller, 1900)	Pr		Mo	Co
<i>Eumops perotis</i> (Schinz, 1821)			Po	Co
<i>Eumops perotis californicus</i> (Merriam, 1890)				
<i>Eumops underwoodi</i> Goodwin, 1940			Po	Co
<i>Eumops underwoodi sonoriensis</i> Benson, 1947				
<i>Eumops underwoodi underwoodi</i> Goodwin, 1940				
<i>Molossus</i> È. Geoffroy Saint–Hilaire, 1805				
<i>Molossus alvarezii</i> González–Ruiz, Ramírez–Pulido, and Arroyo–Cabrales 2011			Mo	Co
<i>Molossus aztecus</i> de Saussure, 1860			Mo	Co
<i>Molossus coibensis</i> J. A. Allen, 1904			Mo	Co
<i>Molossus molossus</i> (Pallas, 1766)			Mo	Co
<i>Molossus rufus</i> È. Geoffroy Saint–Hilaire, 1805			Po	Co
<i>Molossus rufus nigricans</i> Miller, 1902				
<i>Molossus sinaloae</i> J. A. Allen, 1906			Po	Co
<i>Molossus sinaloae sinaloae</i> J. A. Allen, 1906				
<i>Nyctinomops</i> Miller, 1902				
<i>Nyctinomops aurispinosus</i> (Peale, 1848)			Mo	Co
<i>Nyctinomops femorosaccus</i> (Merriam, 1889)			Mo	Co
<i>Nyctinomops laticaudatus</i> (È. Geoffroy Saint–Hilaire, 1805)			Po	Co
<i>Nyctinomops laticaudatus ferrugineus</i> (Goodwin, 1954)				
<i>Nyctinomops laticaudatus yucatanicus</i> Miller, 1902				

<i>Nyctinomops macrotis</i> (Gray, 1839)		Mo	Co
<i>Promops</i> Gervais, 1856			
<i>Promops centralis</i> Thomas, 1915		Po	Co
<i>Promops centralis centralis</i> Thomas, 1915			
<i>Tadarida</i> Rafinesque, 1814			
<i>Tadarida brasiliensis</i> (I. Geoffroy Saint-Hilaire, 1824)		Po	Co
<i>Tadarida brasiliensis intermedia</i> Shamel, 1931			
<i>Tadarida brasiliensis mexicana</i> (de Saussure, 1860)			
SUPERFAMILY NATALOIDEA Gray, 1866			
FAMILY NATALIDAE Gray, 1866			
<i>Natalus</i> Gray, 1838			
<i>Natalus lanatus</i> Tejedor, 2005	En	Mo	Co
<i>Natalus mexicanus</i> Miller, 1902		Mo	Co-In
FAMILY THYROPTERIDAE Miller, 1907			
<i>Thyroptera</i> Spix, 1823			
<i>Thyroptera tricolor</i> Spix, 1823		Po	Co
<i>Thyroptera tricolor albiventer</i> (Tomes, 1856)	Pr		
SUPERFAMILY NOCTILIONOIDEA Gray, 1821			
FAMILY MORMOOPIDAE de Saussure, 1860			
<i>Mormoops</i> Leach, 1821			
<i>Mormoops megalophylla</i> (Peters, 1864)		Po	Co-In
<i>Mormoops megalophylla megalophylla</i> (Peters, 1864)			
<i>Pteronotus</i> Gray, 1838			
<i>Pteronotus davyi</i> Gray, 1838		Po	Co-In
<i>Pteronotus davyi fulvus</i> (Thomas, 1892)			
<i>Pteronotus gymnotus</i> (J. A. Wagner, 1843)	A	Mo	Co
<i>Pteronotus parnellii</i> (Gray, 1843)		Po	Co-In
<i>Pteronotus parnellii mesoamericanus</i> Smith, 1972			
<i>Pteronotus parnellii mexicanus</i> (Miller, 1902)			
<i>Pteronotus personatus</i> (J. A. Wagner, 1843)		Po	Co-In
<i>Pteronotus personatus psilotis</i> (Dobson, 1878)			
FAMILY NOCTILIONIDAE Gray, 1821			
<i>Noctilio</i> Linnaeus, 1766			
<i>Noctilio albiventris</i> Desmarest, 1818		Po	Co
<i>Noctilio albiventris minor</i> Osgood, 1910	Pr		
<i>Noctilio leporinus</i> (Linnaeus, 1758)		Po	Co
<i>Noctilio leporinus mastivus</i> (Vahl, 1797)			

FAMILY PHYLLOSTOMIDAE Gray, 1825			
SUBFAMILY CAROLLIINAE Miller, 1924			
<i>Carollia</i> Gray, 1838			
<i>Carollia perspicillata</i> (Linnaeus, 1758)		Po	Co
<i>Carollia perspicillata azteca</i> de Saussure, 1860			
<i>Carollia sowelli</i> R. J. Baker, Solari, and Hoffmann, 2002		Mo	Co
<i>Carollia subrufa</i> (Hahn, 1905)		Mo	Co
SUBFAMILY DESMODONTINAE J. A. Wagner, 1840			
TRIBE DESMODONTINI J. A. Wagner, 1840			
<i>Desmodus</i> Wied–Neuwied, 1826			
<i>Desmodus rotundus</i> (É. Geoffroy Saint–Hilaire, 1810)		Po	Co
<i>Desmodus rotundus murinus</i> J. A. Wagner, 1840			
<i>Diaemus</i> Miller, 1906			
<i>Diaemus youngii</i> (Jentink, 1893)	Pr	Mo	Co
TRIBE DIPHYLLINI R. J. Baker et al., 2003			
<i>Diphylla</i> Spix, 1823			
<i>Diphylla ecaudata</i> Spix, 1823		Mo	Co
SUBFAMILY GLOSSOPHAGINAE Bonaparte, 1845			
TRIBE CHOERONYCTERINI R. J. Baker et al., 2003			
SUBTRIBE ANOURINA R. J. Baker et al., 2003			
<i>Anoura</i> Gray, 1838			
<i>Anoura geoffroyi</i> Gray, 1838		Po	Co
<i>Anoura geoffroyi lasiopyga</i> (Peters, 1868)			
SUBTRIBE CHOERONYCTERINA R. J. Baker et al., 2003			
<i>Choeroniscus</i> Thomas, 1928			
<i>Choeroniscus godmani</i> (Thomas, 1903)		Mo	Co
<i>Choeronycteris</i> Tschudi, 1844			
<i>Choeronycteris mexicana</i> Tschudi, 1844	A	Mo	Co
<i>Hylonycteris</i> Thomas, 1903			
<i>Hylonycteris underwoodi</i> Thomas, 1903		Po	Co
<i>Hylonycteris underwoodi minor</i> Phillips and Jones, 1971			
<i>Hylonycteris underwoodi underwoodi</i> Thomas, 1903			
<i>Lichonycteris</i> Thomas, 1895			
<i>Lichonycteris obscura</i> Thomas, 1895		Mo	Co
<i>Musonycteris</i> Schaldach and McLaughlin, 1960			
<i>Musonycteris harrisoni</i> Schaldach and McLaughlin, 1960	P	En	Mo
TRIBE GLOSSOPHAGINI Bonaparte, 1845			

<i>Glossophaga</i> È. Geoffroy Saint-Hilaire, 1818			
<i>Glossophaga commissarisi</i> Gardner, 1962		Po	Co
<i>Glossophaga commissarisi commissarisi</i> Gardner, 1962			
<i>Glossophaga commissarisi hespera</i> Webster and Jones, 1982			
<i>Glossophaga leachii</i> (Gray, 1844)		Mo	Co
<i>Glossophaga morenoi</i> Martínez and Villa, 1938	En	Po	Co
<i>Glossophaga morenoi mexicana</i> Webster and Jones, 1980			
<i>Glossophaga morenoi morenoi</i> Martínez and Villa, 1938			
<i>Glossophaga soricina</i> (Pallas, 1766)		Po	Co-In
<i>Glossophaga soricina handleyi</i> Webster and Jones, 1980			
<i>Glossophaga soricina mutica</i> Merriam, 1898			
<i>Leptoncyteris</i> Lydekker, 1891			
<i>Leptoncyteris nivalis</i> (de Saussure, 1860)	A	Mo	Co
<i>Leptoncyteris yerbabuenae</i> Martínez and Villa, 1940	A	Mo	Co-In
SUBFAMILY GLYPHONYCTERINAE R. J. Baker et al., 2003			
<i>Glyphonycteris</i> Thomas, 1896			
<i>Glyphonycteris sylvestris</i> Thomas, 1896		Mo	Co
SUBFAMILY LONCHORHININAE Gray, 1866			
<i>Lonchorhina</i> Tomes, 1863			
<i>Lonchorhina aurita</i> Tomes, 1863	A	Po	Co
<i>Lonchorhina aurita aurita</i> Tomes, 1863			
SUBFAMILY MACROTINAE R. J. Baker et al., 1989			
<i>Macrotus</i> Gray, 1843			
<i>Macrotus californicus</i> Baird, 1858		Mo	Co
<i>Macrotus waterhousii</i> Gray, 1843		Po	Co-In
<i>Macrotus waterhousii bulleri</i> H. Allen, 1890			
<i>Macrotus waterhousii mexicanus</i> de Saussure, 1860			
SUBFAMILY MICRONYCTERINAE R. J. Baker et al., 1989			
<i>Lamproncyteris</i> Sanborn, 1949			
<i>Lamproncyteris brachyotis</i> (Dobson, 1879)	A	Mo	Co
<i>Microncyteris</i> Gray, 1866			
<i>Microncyteris microtis</i> Miller, 1898		Po	Co-In
<i>Microncyteris microtis mexicana</i> Miller, 1898			
<i>Microncyteris schmidtorum</i> Sanborn, 1935	A	Mo	Co
<i>Trinycteris</i> Sanborn, 1949			
<i>Trinycteris nicefori</i> (Sanborn, 1949)		Mo	Co
SUBFAMILY PHYLLOSTOMINAE Gray, 1852			
TRIBE MACROPHYLLINI R. J. Baker et al., 2003			

<i>Macrophyllum</i> Gray, 1838			
<i>Macrophyllum macrophyllum</i> (Schinz, 1821)	A	Mo	Co
<i>Trachops</i> Gray, 1847			
<i>Trachops cirrhosus</i> (Spix, 1823)	A	Po	Co
<i>Trachops cirrhosus coffini</i> Goldman, 1925			
TRIBE PHYLLOSTOMINI Gray, 1825			
<i>Lophostoma</i> d'Orbigny, 1836			
<i>Lophostoma brasiliense</i> Peters, 1867	A	Mo	Co
<i>Lophostoma evotis</i> (W. B. Davis and Carter, 1978)	A	Mo	Co
<i>Mimon</i> Gray, 1847			
<i>Mimon cozumelae</i> Goldman, 1914		Mo	Co
<i>Mimon crenulatum</i> (É. Geoffroy Saint-Hilaire, 1803)	A	Po	Co
<i>Mimon crenulatum keenani</i> Handley, 1960			
<i>Phylloderma</i> Peters, 1865			
<i>Phylloderma stenops</i> Peters, 1865	A	Po	Co
<i>Phylloderma stenops septentrionalis</i> Goodwin, 1940			
<i>Phyllostomus</i> Lacépède, 1799			
<i>Phyllostomus discolor</i> (J. A. Wagner, 1843)		Po	Co
<i>Phyllostomus discolor verrucosus</i> (Elliot, 1905)			
<i>Tonatia</i> Gray, 1827			
<i>Tonatia saurophila</i> Koopman and Williams, 1951	A	Po	Co-In
<i>Tonatia saurophila bakeri</i> Williams, Willig, and Reid, 1995			
TRIBE VAMPYRINI Bonaparte, 1838			
<i>Chrotopterus</i> Peters, 1865			
<i>Chrotopterus auritus</i> (Peters, 1856)	A	Po	Co
<i>Chrotopterus auritus auritus</i> (Peters, 1856)			
<i>Vampyrum</i> Rafinesque, 1815			
<i>Vampyrum spectrum</i> (Linnaeus, 1758)	P	Mo	Co
SUBFAMILY STENODERMATINAE Gervais, 1856			
TRIBE MESOSTENODERMATINI R. J. Baker et al., 2003			
SUBTRIBE ENCHISTHENINA R. J. Baker et al., 2003			
<i>Artibeus</i> Leach, 1821			
<i>Artibeus hirsutus</i> Andersen, 1906		En	Mo
<i>Artibeus jamaicensis</i> Leach, 1821			Po
<i>Artibeus jamaicensis paulus</i> W. B. Davis, 1970			Co-In
<i>Artibeus jamaicensis richardsoni</i> J. A. Allen, 1908			
<i>Artibeus jamaicensis triomylus</i> Handley, 1966			
<i>Artibeus jamaicensis yucatanicus</i> J. A. Allen, 1904			
<i>Artibeus lituratus</i> (Olfers, 1818)		Po	Co-In

<i>Artibeus lituratus koopmani</i> Wilson, 1991			
<i>Artibeus lituratus palmarum</i> J. A. Allen and Chapman, 1897			
<i>Dermanura</i> Gervais, 1856			
<i>Dermanura azteca</i> (Andersen, 1906)		Po	Co
<i>Dermanura azteca azteca</i> (Andersen, 1906)			
<i>Dermanura azteca minor</i> (W. B. Davis, 1969)			
<i>Dermanura phaeotis</i> Miller, 1902		Po	Co-In
<i>Dermanura phaeotis nana</i> (Andersen, 1906)			
<i>Dermanura phaeotis palatina</i> (W. B. Davis, 1970)			
<i>Dermanura phaeotis phaeotis</i> Miller, 1902			
<i>Dermanura tolteca</i> (de Saussure, 1860)		Po	Co
<i>Dermanura tolteca hespera</i> (W. B. Davis, 1969)			
<i>Dermanura tolteca tolteca</i> (de Saussure, 1860)			
<i>Dermanura watsoni</i> (Thomas, 1901)	Pr	Mo	Co
<i>Enchisthenes</i> Andersen, 1906			
<i>Enchisthenes hartii</i> (Thomas, 1892)	Pr	Mo	Co
TRIBE STENODERMATINI Gervais, 1856			
SUBTRIBE STENODERMATINA Gervais, 1856			
<i>Centurio</i> Gray, 1842			
<i>Centurio senex</i> Gray, 1842		Po	Co
<i>Centurio senex senex</i> Gray, 1842			
SUBTRIBE VAMPYRESSINA R. J. Baker et al., 2003			
<i>Chiroderma</i> Peters, 1860			
<i>Chiroderma salvini</i> Dobson, 1878		Po	Co
<i>Chiroderma salvini salvini</i> Dobson, 1878			
<i>Chiroderma salvini scopaeum</i> Handley, 1966			
<i>Chiroderma villosum</i> Peters, 1860		Po	Co
<i>Chiroderma villosum jesupi</i> J. A. Allen, 1900			
<i>Platyrrhinus</i> de Saussure, 1860			
<i>Platyrrhinus helleri</i> (Peters, 1866)		Mo	Co
<i>Uroderma</i> Peters, 1865			
<i>Uroderma bilobatum</i> Peters, 1866		Po	Co
<i>Uroderma bilobatum convexum</i> Lyon, 1902			
<i>Uroderma bilobatum davisi</i> R. J. Baker and McDaniel, 1972			
<i>Uroderma magnirostrum</i> W. B. Davis, 1968		Mo	Co
<i>Vampyressa</i> Thomas, 1900			
<i>Vampyressa thyone</i> Thomas, 1909		Mo	Co
<i>Vampyrodes</i> Thomas, 1900			
<i>Vampyrodes major</i> G. M. Allen, 1908		Mo	Co

TRIBE STURNIRINI Miller, 1907			
<i>Sturnira</i> Gray, 1842			
<i>Sturnira hondurensis</i> Goodwin, 1940	Po	Co	
<i>Sturnira hondurensis hondurensis</i> Goodwin, 1940			
<i>Sturnira hondurensis occidentalis</i> J. K. Jones and Phillips, 1964			
<i>Sturnira parvidens</i> Goldman, 1917	Po	Co	
SUPERFAMILY VESPERTILIONOIDEA Gray, 1821			
FAMILY VESPERTILIONIDAE Gray, 1821			
SUBFAMILY ANTROZOINAE Miller, 1897			
<i>Antrozous</i> H. Allen, 1862			
<i>Antrozous pallidus</i> (J. Le Conte, 1855)	Po	Co-In	
<i>Antrozous pallidus minor</i> Miller, 1902			
<i>Antrozous pallidus packardi</i> Martin and Schmidly, 1982			
<i>Antrozous pallidus pallidus</i> (J. Le Conte, 1855)			
<i>Bauerus</i> Van Gelder, 1959			
<i>Bauerus dubiaquercus</i> (Van Gelder, 1959)	Mo	Co-In	
SUBFAMILY MYOTINAE Tate, 1942			
<i>Lasionycteris</i> Peters, 1866			
<i>Lasionycteris noctivagans</i> (J. Le Conte, 1831)	Pr	Mo	Co
<i>Myotis</i> Kaup, 1829			
<i>Myotis albescens</i> (È. Geoffroy Saint-Hilaire, 1806)	Pr	Mo	Co
<i>Myotis auriculus</i> R. H. Baker and Stains, 1955		Po	Co
<i>Myotis auriculus apache</i> Hoffmeister and Krutzch, 1955			
<i>Myotis auriculus auriculus</i> R. H. Baker and Stains, 1955			
<i>Myotis californicus</i> (Audubon and Bachman, 1842)		Po	Co
<i>Myotis californicus californicus</i> (Audubon and Bachman, 1842)			
<i>Myotis californicus mexicanus</i> (de Saussure, 1860)			
<i>Myotis californicus stephensi</i> Dalquest, 1946			
<i>Myotis elegans</i> Hall, 1962		Mo	Co
<i>Myotis evotis</i> (H. Allen, 1864)	Pr	Po	Co
<i>Myotis evotis micronyx</i> Nelson and Goldman, 1909			
<i>Myotis evotis milleri</i> Elliot, 1903			
<i>Myotis findleyi</i> Bogan, 1978	En	Mo	In
<i>Myotis fortidens</i> Miller and G. M. Allen, 1928	En	Po	Co
<i>Myotis fortidens fortidens</i> Miller and G. M. Allen, 1928			
<i>Myotis fortidens sonoriensis</i> Findley and Jones, 1967			
<i>Myotis keaysi</i> J. A. Allen, 1914		Po	Co
<i>Myotis keaysi pilosatibialis</i> LaVal, 1973			

<i>Myotis melanorhinus</i> (Merriam, 1890)			Mo	Co
<i>Myotis nigricans</i> (Schinz, 1821)			Po	Co
<i>Myotis nigricans carteri</i> LaVal, 1973	Pr			
<i>Myotis nigricans extremus</i> Miller and G. M. Allen, 1928				
<i>Myotis nigricans nigricans</i> (Schinz, 1821)				
<i>Myotis occultus</i> Hollister, 1909			Mo	Co
<i>Myotis peninsularis</i> Miller, 1898		En	Mo	Co
<i>Myotis planiceps</i> R. H. Baker, 1955	P	En	Mo	Co
<i>Myotis thysanodes</i> Miller, 1897			Po	Co
<i>Myotis thysanodes aztecus</i> Miller and G. M. Allen, 1928				
<i>Myotis thysanodes thysanodes</i> Miller, 1897				
<i>Myotis velifer</i> (J. A. Allen, 1890)			Po	Co
<i>Myotis velifer brevis</i> Vaughan, 1954				
<i>Myotis velifer incautus</i> (J. A. Allen, 1896)				
<i>Myotis velifer velifer</i> (J. A. Allen, 1890)				
<i>Myotis vivesi</i> Menegaux, 1901	P	En	Mo	Co-In
<i>Myotis volans</i> (H. Allen, 1866)			Po	Co
<i>Myotis volans amotus</i> Miller, 1914				
<i>Myotis volans interior</i> Miller, 1914				
<i>Myotis volans volans</i> (H. Allen, 1866)				
<i>Myotis yumanensis</i> (H. Allen, 1864)			Po	Co
<i>Myotis yumanensis lambi</i> Benson, 1947				
<i>Myotis yumanensis lutosus</i> Miller and G. M. Allen, 1928				
<i>Myotis yumanensis yumanensis</i> (H. Allen, 1864)				
SUBFAMILY VESPERTILIONINAE Miller, 1897				
TRIBE incertae sedis (Hooper et al., 2003)				
<i>Parastrellus</i> Hooper, Van Den Bussche, and Horáček, 2006				
<i>Parastrellus hesperus</i> (H. Allen, 1864)			Po	Co-In
<i>Parastrellus hesperus hesperus</i> (H. Allen, 1864)				
<i>Parastrellus hesperus maximus</i> (Hatfield, 1936)				
<i>Perimyotis</i> Menu, 1984				
<i>Perimyotis subflavus</i> (F. Cuvier, 1832)			Po	Co
<i>Perimyotis subflavus clarus</i> (R. H. Baker, 1954)				
<i>Perimyotis subflavus subflavus</i> (F. Cuvier, 1832)				
<i>Perimyotis subflavus veraecrucis</i> (Ward, 1891)				
TRIBE EPTESICINI Volleth and Heller, 1994				
<i>Eptesicus</i> Rafinesque, 1820				
<i>Eptesicus brasiliensis</i> (Desmarest, 1819)			Po	Co
<i>Eptesicus brasiliensis brasiliensis</i> (Desmarest, 1819)				
<i>Eptesicus furinalis</i> (d'Orbigny and Gervais, 1847)			Po	Co
<i>Eptesicus furinalis gaumeri</i> (J. A. Allen, 1897)				

<i>Eptesicus fuscus</i> (Palisot de Beauvois, 1796)		Po	Co	
<i>Eptesicus fuscus fuscus</i> (Palisot de Beauvois, 1796)				
<i>Eptesicus fuscus miradorensis</i> (H. Allen, 1866)				
<i>Eptesicus fuscus pallidus</i> Young, 1908				
<i>Eptesicus fuscus peninsulae</i> (Thomas, 1898)				
TRIBE LASIURINI Tate, 1942				
<i>Lasiurus</i> Gray, 1831				
<i>Lasiurus blossevillii</i> (Lesson, 1826)		Po	Co–In	
<i>Lasiurus blossevillii frantzii</i> (Peters, 1870)				
<i>Lasiurus blossevillii teliotis</i> (H. Allen, 1891)				
<i>Lasiurus borealis</i> (Müller, 1776)		Mo	Co	
<i>Lasiurus cinereus</i> (Palisot de Beauvois, 1796)		Po	Co	
<i>Lasiurus cinereus cinereus</i> (Palisot de Beauvois, 1796)				
<i>Lasiurus ega</i> (Gervais, 1856)		Po	Co	
<i>Lasiurus ega panamensis</i> (Thomas, 1901)				
<i>Lasiurus intermedius</i> H. Allen, 1862		Po	Co	
<i>Lasiurus intermedius intermedius</i> H. Allen, 1862				
<i>Lasiurus seminolus</i> (Rhoads, 1895)		Mo	Co	
<i>Lasiurus xanthinus</i> (Thomas, 1897)		Mo	Co	
TRIBE NYCTICEINI Gervais, 1856				
<i>Nycticeius</i> Rafinesque, 1818				
<i>Nycticeius humeralis</i> (Rafinesque, 1818)		Po	Co	
<i>Nycticeius humeralis mexicanus</i> W. B. Davis, 1944				
<i>Rhogeessa</i> H. Allen, 1866				
<i>Rhogeessa aeneus</i> Goodwin, 1958	En	Mo	Co	
<i>Rhogeessa alleni</i> Thomas, 1892	En	Mo	Co	
<i>Rhogeessa bickhami</i> Baird, Marchán-Rivadeneira, Pérez, and R. J. Baker, 2012	En	Mo	Co	
<i>Rhogeessa genowaysi</i> R. J. Baker, 1984	A	En	Mo	Co
<i>Rhogeessa gracilis</i> (Miller, 1897)		En	Mo	Co
<i>Rhogeessa mira</i> LaVal, 1973	Pr	En	Mo	Co
<i>Rhogeessa parvula</i> H. Allen, 1866		En	Po	Co–In
<i>Rhogeessa parvula major</i> Goodwin, 1958				
<i>Rhogeessa parvula parvula</i> H. Allen, 1866				
<i>Rhogeessa tumida</i> H. Allen, 1866		Mo	Co	
TRIBE PLECOTINI Gray, 1866				
<i>Corynorhinus</i> H. Allen, 1865				
<i>Corynorhinus mexicanus</i> G. M. Allen, 1916		En	Mo	Co
<i>Corynorhinus townsendii</i> (Cooper, 1837)			Po	Co–In
<i>Corynorhinus townsendii australis</i> Handley, 1955				

<i>Corynorhinus townsendii townsendii</i> (Cooper, 1837)			
<i>Euderma</i> H. Allen, 1892			
<i>Euderma maculatum</i> (J. A. Allen, 1891)	Pr	Mo	Co
<i>Idionycteris</i> Anthony, 1923			
<i>Idionycteris phyllotis</i> (G. M. Allen, 1916)		Po	Co
<i>Idionycteris phyllotis phyllotis</i> (G. M. Allen, 1916)			
ORDER PRIMATES Linnaeus, 1758			
SUBORDER EUPRIMATES Hoffstetter, 1978			
FAMILY ATELIDAE Gray, 1825			
SUBFAMILY ATELINAE Gray, 1825			
<i>Ateles</i> È. Geoffroy Saint-Hilaire, 1806			
<i>Ateles geoffroyi</i> Kuhl, 1820	P	Po	Co
<i>Ateles geoffroyi vellerosus</i> Gray, 1866			
<i>Ateles geoffroyi yucatanensis</i> Kellogg and Goldman, 1944			
SUBFAMILY ALOUATTINAE Trouessart, 1897			
<i>Alouatta</i> Lacépède, 1799			
<i>Alouatta palliata</i> (Gray, 1849)	P	Po	Co
<i>Alouatta palliata mexicana</i> Merriam, 1902			
<i>Alouatta villosa</i> (Gray, 1845)	P	Mo	Co
MAGNORDER EPITHERIA McKenna, 1975			
ORDER LAGOMORPHA Brandt, 1855			
FAMILY LEPORIDAE G. Fischer, 1817			
<i>Lepus</i> Linnaeus, 1758			
<i>Lepus alleni</i> Mearns, 1890		Po	Co-In
<i>Lepus alleni alleni</i> Mearns, 1890			
<i>Lepus alleni tiburonensis</i> Townsend, 1912	Pr		
<i>Lepus californicus</i> Gray, 1837		Po	Co-In
<i>Lepus californicus californicus</i> Gray, 1837			
<i>Lepus californicus deserticola</i> Mearns, 1896			
<i>Lepus californicus insularis</i> Bryant, 1891	Pr		
<i>Lepus californicus magdalenae</i> Nelson, 1907	Pr		
<i>Lepus californicus melanotis</i> Mearns, 1890			
<i>Lepus californicus texianus</i> Waterhouse, 1848			
<i>Lepus californicus xanti</i> Thomas, 1898			
<i>Lepus callotis</i> J. A. Wagler, 1830		En	Po
<i>Lepus callotis callotis</i> J. A. Wagler, 1830			
<i>Lepus callotis gaillardi</i> Mearns, 1896			
<i>Lepus flavigularis</i> J. A. Wagner, 1844	P	En	Mo
<i>Sylvilagus</i> Gray, 1867			

<i>Sylvilagus audubonii</i> (Baird, 1857)			Po	Co
<i>Sylvilagus audubonii arizonae</i> (J. A. Allen, 1877)				
<i>Sylvilagus audubonii confinis</i> (J. A. Allen, 1898)				
<i>Sylvilagus audubonii goldmani</i> (Nelson, 1904)				
<i>Sylvilagus audubonii minor</i> (Mearns, 1896)				
<i>Sylvilagus bachmani</i> (Waterhouse, 1839)			Po	Co-In
<i>Sylvilagus bachmani cerrosensis</i> (J. A. Allen, 1898)	P			
<i>Sylvilagus bachmani cinerascens</i> (J. A. Allen, 1890)				
<i>Sylvilagus bachmani exiguus</i> Nelson, 1907				
<i>Sylvilagus bachmani howelli</i> Huey, 1927				
<i>Sylvilagus bachmani peninsularis</i> (J. A. Allen, 1898)				
<i>Sylvilagus brasiliensis</i> (Linnaeus, 1758)			Po	Co
<i>Sylvilagus brasiliensis truei</i> (J. A. Allen, 1890)				
<i>Sylvilagus cunicularius</i> (Waterhouse, 1848)		En	Po	Co
<i>Sylvilagus cunicularius cunicularius</i> (Waterhouse, 1848)				
<i>Sylvilagus cunicularius insolitus</i> (J. A. Allen, 1890)				
<i>Sylvilagus floridanus</i> (J. A. Allen, 1890)			Po	Co
<i>Sylvilagus floridanus aztecus</i> (J. A. Allen, 1890)				
<i>Sylvilagus floridanus chapmani</i> (J. A. Allen, 1899)				
<i>Sylvilagus floridanus holzneri</i> (Mearns, 1896)				
<i>Sylvilagus floridanus macrocorpus</i> Diersing and Wilson, 1980				
<i>Sylvilagus floridanus orizabae</i> (Merriam, 1893)				
<i>Sylvilagus floridanus russatus</i> (J. A. Allen, 1904)				
<i>Sylvilagus floridanus yucatanicus</i> (Miller, 1899)				
<i>Sylvilagus graysoni</i> (J. A. Allen, 1877)	P	En	Mo	In
<i>Sylvilagus insonus</i> (Nelson, 1904)	P	En	Mo	Co
<i>Sylvilagus mansuetus</i> Nelson, 1907	P	En	Mo	In
<i>Sylvilagus robustus</i> (Bailey, 1905)			Mo	Co
<i>Romerolagus</i> Merriam, 1896		En		
<i>Romerolagus diazi</i> (Ferrari-Pérez, 1893)	P	En	Mo	Co
ORDER RODENTIA Bowdich, 1821				
SUBORDER SCIUROMORPHA Brandt, 1855				
INFRAORDER SCIURIDA Carus, 1868				
FAMILY SCIURIDAE G. Fischer, 1817				
SUBFAMILY PTEROMYINAE Brandt, 1855				
<i>Glaucomys</i> Thomas, 1908				
<i>Glaucomys volans</i> (Linnaeus, 1758)	A		Po	Co
<i>Glaucomys volans goldmani</i> (Nelson, 1904)				
<i>Glaucomys volans guerreroensis</i> Diersing, 1980				
<i>Glaucomys volans madrensis</i> Goldman, 1936				
<i>Glaucomys volans oaxacensis</i> Goodwin, 1961				

SUBFAMILY SCIURINAE G. Fischer, 1817				
TRIBE MARMOTINI Pocock, 1923				
<i>Ammospermophilus</i> Merriam, 1892				
<i>Ammospermophilus harrisi</i> (Audubon and Bachman, 1854)			Po	Co
<i>Ammospermophilus harrisi harrisi</i> (Audubon and Bachman, 1854)				
<i>Ammospermophilus harrisi saxicola</i> (Mearns, 1896)				
<i>Ammospermophilus interpres</i> (Merriam, 1890)			Mo	Co
<i>Ammospermophilus leucurus</i> (Merriam, 1889)			Po	Co
<i>Ammospermophilus leucurus canfieldae</i> Huey, 1929				
<i>Ammospermophilus leucurus extimus</i> Nelson and Goldman, 1929				
<i>Ammospermophilus leucurus insularis</i> Nelson and Goldman, 1909	A			
<i>Ammospermophilus leucurus leucurus</i> (Merriam, 1889)				
<i>Ammospermophilus leucurus peninsulae</i> (J. A. Allen, 1893)				
<i>Callospermophilus</i> Merriam, 1897				
<i>Callospermophilus madrensis</i> Merriam, 1901	Pr	En	Mo	Co
<i>Cynomys</i> Rafinesque, 1817				
<i>Cynomys ludovicianus</i> (Ord, 1815)	A		Po	Co
<i>Cynomys ludovicianus arizonensis</i> Mearns, 1890				
<i>Cynomys mexicanus</i> Merriam, 1892	P	En	Mo	Co
<i>Ictidomys</i> J. A. Allen, 1877				
<i>Ictidomys mexicanus</i> (Erxleben, 1777)		En	Mo	Co
<i>Ictidomys parvidens</i> (Mearns, 1896)			Mo	Co
<i>Notocitellus</i> A. H. Howell, 1938				
<i>Notocitellus adocetus</i> (Merriam, 1903)		En	Po	Co
<i>Notocitellus adocetus adocetus</i> (Merriam, 1903)				
<i>Notocitellus adocetus infernatus</i> (Álvarez and Ramírez-Pulido, 1968)				
<i>Notocitellus annulatus</i> (Audubon and Bachman, 1842)		En	Po	Co
<i>Notocitellus annulatus annulatus</i> (Audubon and Bachman, 1842)				
<i>Notocitellus annulatus goldmani</i> (Merriam, 1902)				
<i>Otospermophilus</i> Brandt, 1844				
<i>Otospermophilus atricapillus</i> (Bryant, 1889)		En	Mo	Co
<i>Otospermophilus beecheyi</i> (Richardson, 1829)			Po	Co
<i>Otospermophilus beecheyi nudipes</i> (Huey, 1931)				
<i>Otospermophilus beecheyi rupinarum</i> (Huey, 1931)				
<i>Otospermophilus variegatus</i> (Erxleben, 1777)			Po	Co-In
<i>Otospermophilus variegatus couchii</i> (Baird, 1855)				
<i>Otospermophilus variegatus grammurus</i> (Say, 1822)				
<i>Otospermophilus variegatus rupestris</i> (J. A. Allen, 1903)				

<i>Otospermophilus variegatus variegatus</i> (Erxleben, 1777)				
<i>Xerospermophilus</i> Merriam, 1892				
<i>Xerospermophilus spilosoma</i> (Bennett, 1833)		Po		Co
<i>Xerospermophilus spilosoma altiplanensis</i> (Anderson, 1972)				
<i>Xerospermophilus spilosoma ammophilus</i> (Hoffmeister, 1959)				
<i>Xerospermophilus spilosoma bavicorensis</i> (Anderson, 1972)				
<i>Xerospermophilus spilosoma cabrerai</i> (Dalquest, 1951)				
<i>Xerospermophilus spilosoma canescens</i> (Merriam, 1890)				
<i>Xerospermophilus spilosoma marginatus</i> (Bailey, 1902)				
<i>Xerospermophilus spilosoma oricola</i> (Álvarez, 1962)				
<i>Xerospermophilus spilosoma pallescens</i> (A. H. Howell, 1928)				
<i>Xerospermophilus spilosoma perotensis</i> (Merriam, 1893)	A			
<i>Xerospermophilus spilosoma spilosoma</i> (Bennett, 1833)				
<i>Xerospermophilus tereticaudus</i> (Baird, 1857)		Po		Co-In
<i>Xerospermophilus tereticaudus apricus</i> (Huey, 1927)				
<i>Xerospermophilus tereticaudus neglectus</i> (Merriam, 1889)				
<i>Xerospermophilus tereticaudus tereticaudus</i> (Baird, 1857)				
TRIBE SCIURINI G. Fischer, 1817				
<i>Sciurus</i> Linnaeus, 1758				
<i>Sciurus aberti</i> Woodhouse, 1853		Po		Co
<i>Sciurus aberti barberi</i> J. A. Allen, 1904	Pr			
<i>Sciurus aberti durangi</i> Thomas, 1893	Pr			
<i>Sciurus alleni</i> Nelson, 1898		En	Mo	Co
<i>Sciurus arizonensis</i> Coues, 1867	A		Po	Co
<i>Sciurus arizonensis huachuca</i> J. A. Allen, 1894				
<i>Sciurus aureogaster</i> F. Cuvier, 1829			Po	Co
<i>Sciurus aureogaster aureogaster</i> F. Cuvier, 1829				
<i>Sciurus aureogaster nigrescens</i> Bennett, 1833				
<i>Sciurus colliae</i> Richardson, 1839		En	Po	Co
<i>Sciurus colliae colliae</i> Richardson, 1839				
<i>Sciurus colliae nuchalis</i> Nelson, 1899				
<i>Sciurus colliae sinaloensis</i> Nelson, 1899				
<i>Sciurus colliae truei</i> Nelson, 1899				
<i>Sciurus deppei</i> Peters, 1864			Po	Co
<i>Sciurus deppei deppei</i> Peters, 1864				
<i>Sciurus deppei negligens</i> Nelson, 1898				
<i>Sciurus deppei vivax</i> Nelson, 1901				
<i>Sciurus griseus</i> Ord, 1818	A		Po	Co
<i>Sciurus griseus anthonyi</i> Mearns, 1897				
<i>Sciurus nayaritensis</i> J. A. Allen, 1890		En	Po	Co
<i>Sciurus nayaritensis apache</i> J. A. Allen, 1893				

<i>Sciurus nayaritensis nayaritensis</i> J. A. Allen, 1890				
<i>Sciurus niger</i> Linnaeus, 1758			Po	Co
<i>Sciurus niger limitis</i> Baird, 1855				
<i>Sciurus oculus</i> Peters, 1863	Pr	En	Po	Co
<i>Sciurus oculus oculus</i> Peters, 1863				
<i>Sciurus oculus shawi</i> Dalquest, 1950				
<i>Sciurus oculus tolucae</i> Nelson, 1898				
<i>Sciurus variegatoides</i> Ogilby, 1839	Pr		Po	Co
<i>Sciurus variegatoides goldmani</i> Nelson, 1898				
<i>Sciurus yucatanensis</i> J. A. Allen, 1877			Po	Co
<i>Sciurus yucatanensis baliolus</i> Nelson, 1901				
<i>Sciurus yucatanensis phaeopus</i> Goodwin, 1932				
<i>Sciurus yucatanensis yucatanensis</i> J. A. Allen, 1877				
TRIBE TAMIINI Weber, 1928				
<i>Neotamias</i> A. H. Howell, 1929				
<i>Neotamias bulleri</i> (J. A. Allen, 1889)			Mo	Co
<i>Neotamias dorsalis</i> (Baird, 1855)			Po	Co
<i>Neotamias dorsalis carminis</i> (Goldman, 1938)				
<i>Neotamias dorsalis dorsalis</i> (Baird, 1855)				
<i>Neotamias dorsalis nidoensis</i> (Lidicker, 1960)				
<i>Neotamias dorsalis sonoriensis</i> (Callahan and Davis, 1977)				
<i>Neotamias durangae</i> J. A. Allen, 1903		En	Mo	Co
<i>Neotamias merriami</i> (J. A. Allen, 1889)	Pr		Po	Co
<i>Neotamias merriami merriami</i> (J. A. Allen, 1889)				
<i>Neotamias obscurus</i> (J. A. Allen, 1890)			Po	Co
<i>Neotamias obscurus meridionalis</i> (Nelson and Goldman, 1909)				
<i>Neotamias obscurus obscurus</i> (J. A. Allen, 1890)				
<i>Neotamias solivagus</i> (A. H. Howell, 1922)		En	Mo	Co
TRIBE TAMIASCIURINI Pocock, 1923				
<i>Tamiasciurus</i> Trouessart, 1880				
<i>Tamiasciurus mearnsi</i> (Townsend, 1897)	A	En	Mo	Co
INFRAORDER GEOMORPHA Thaler, 1966				
SUPERFAMILY GEOMYOIDEA Bonaparte, 1845				
FAMILY GEOMYIDAE Bonaparte, 1845				
SUBFAMILY GEOMYINAE Bonaparte, 1845				
TRIBE GEOMYINI Bonaparte, 1845				
<i>Cratogeomys</i> Merriam, 1895				
<i>Cratogeomys castanops</i> (Baird, 1852)			Po	Co

<i>Cratogeomys castanops castanops</i> (Baird, 1852)				
<i>Cratogeomys castanops consitus</i> Nelson and Goldman, 1934				
<i>Cratogeomys fulvescens</i> Merriam, 1895		En	Mo	Co
<i>Cratogeomys fumosus</i> (Merriam, 1892)	A	En	Po	Co
<i>Cratogeomys fumosus angustirostris</i> (Merriam 1903)				
<i>Cratogeomys fumosus fumosus</i> (Merriam, 1892)				
<i>Cratogeomys fumosus imparilis</i> (Goldman, 1939)				
<i>Cratogeomys fumosus tylorhinus</i> (Merriam, 1895)				
<i>Cratogeomys goldmani</i> Merriam, 1895		En	Po	Co
<i>Cratogeomys goldmani goldmani</i> Merriam, 1895				
<i>Cratogeomys goldmani subnubilus</i> Nelson and Goldman, 1934				
<i>Cratogeomys merriami</i> (Thomas, 1893)		En	Mo	Co
<i>Cratogeomys perotensis</i> Merriam, 1895		En	Mo	Co
<i>Cratogeomys planiceps</i> (Merriam, 1895)		En	Mo	Co
<i>Geomys Rafinesque, 1817</i>				
<i>Geomys arenarius</i> Merriam, 1895			Po	Co
<i>Geomys arenarius arenarius</i> Merriam, 1895				
<i>Geomys personatus</i> True, 1889	A		Po	Co
<i>Geomys personatus megapotamus</i> W. B. Davis, 1940				
<i>Geomys tropicalis</i> Goldman, 1915	A	En	Mo	Co
<i>Orthogeomys</i> Merriam, 1895				
<i>Orthogeomys cuniculus</i> Elliot, 1905	A	En	Mo	Co
<i>Orthogeomys grandis</i> (Thomas, 1893)			Po	Co
<i>Orthogeomys grandis alleni</i> Nelson and Goldman, 1930				
<i>Orthogeomys grandis alvarezi</i> Schaldach, 1966				
<i>Orthogeomys grandis annexus</i> Nelson and Goldman, 1933				
<i>Orthogeomys grandis carbo</i> Goodwin, 1956				
<i>Orthogeomys grandis felipensis</i> Nelson and Goldman, 1930				
<i>Orthogeomys grandis guerrierensis</i> Nelson and Goldman, 1930				
<i>Orthogeomys grandis huixtlae</i> Villa R., 1944				
<i>Orthogeomys grandis nelsoni</i> Merriam, 1895				
<i>Orthogeomys grandis scalops</i> (Thomas, 1894)				
<i>Orthogeomys grandis soconuscensis</i> Villa R., 1949				
<i>Orthogeomys hispidus</i> (J. L. Le Conte, 1852)			Po	Co
<i>Orthogeomys hispidus chiapensis</i> (Nelson and Goldman, 1929)				
<i>Orthogeomys hispidus concavus</i> (Nelson and Goldman, 1929)				
<i>Orthogeomys hispidus hispidus</i> (J. L. Le Conte, 1852)				
<i>Orthogeomys hispidus isthmicus</i> (Nelson and Goldman, 1929)				
<i>Orthogeomys hispidus latirostris</i> (Hall and Álvarez, 1961)				
<i>Orthogeomys hispidus negatus</i> (Goodwin, 1953)				
<i>Orthogeomys hispidus teapensis</i> (Goldman, 1939)				

<i>Orthogeomys hispidus tehuantepecus</i> (Goldman, 1939)				
<i>Orthogeomys hispidus torridus</i> (Merriam, 1895)				
<i>Orthogeomys hispidus yucatanensis</i> (Nelson and Goldman, 1929)				
<i>Orthogeomys lanius</i> (Elliot, 1905)	A	En	Mo	Co
<i>Pappogeomys</i> Merriam, 1895		En		
<i>Pappogeomys bulleri</i> (Thomas, 1892)		En	Po	Co
<i>Pappogeomys bulleri albinasus</i> Merriam, 1895				
<i>Pappogeomys bulleri alcorni</i> Russell, 1957	Pr			
<i>Pappogeomys bulleri bulleri</i> (Thomas, 1892)				
<i>Pappogeomys bulleri burti</i> Goldman, 1939				
<i>Pappogeomys bulleri nayaritensis</i> Goldman, 1939				
<i>Zygozemys</i> Merriam, 1895		En		
<i>Zygozemys trichopus</i> Merriam, 1895	P	En	Po	Co
<i>Zygozemys trichopus tarascensis</i> Goldman, 1938				
<i>Zygozemys trichopus trichopus</i> Merriam, 1895				
TRIBE THOMOMYINI Russell, 1968				
<i>Thomomys</i> Wied–Neuwied, 1839				
<i>Thomomys atrovarius</i> J. A. Allen, 1898		En	Po	Co
<i>Thomomys atrovarius atrovarius</i> J. A. Allen, 1898				
<i>Thomomys atrovarius parviceps</i> Nelson and Goldman, 1934				
<i>Thomomys bottae</i> (Eydoux and Gervais, 1836)			Po	Co–In
<i>Thomomys bottae abbotti</i> Huey, 1928				
<i>Thomomys bottae albatu</i> s Grinnell, 1912				
<i>Thomomys bottae analogus</i> Goldman, 1938				
<i>Thomomys bottae angustidens</i> R. H. Baker, 1953				
<i>Thomomys bottae anitae</i> J. A. Allen, 1898				
<i>Thomomys bottae ahrastus</i> Elliot, 1903				
<i>Thomomys bottae basilicae</i> Benson and Tillotson, 1939				
<i>Thomomys bottae borjasensis</i> Huey, 1945				
<i>Thomomys bottae bottae</i> (Eydoux and Gervais, 1836)				
<i>Thomomys bottae brazierhowelli</i> Huey, 1960				
<i>Thomomys bottae cactophilus</i> Huey, 1929				
<i>Thomomys bottae camoae</i> Burt, 1937				
<i>Thomomys bottae catavinensis</i> Huey, 1931				
<i>Thomomys bottae convergens</i> Nelson and Goldman, 1934				
<i>Thomomys bottae cunicularis</i> Huey, 1945				
<i>Thomomys bottae divergens</i> Nelson and Goldman, 1934				
<i>Thomomys bottae estanciae</i> Benson and Tillotson, 1939				
<i>Thomomys bottae homorus</i> Huey, 1949				
<i>Thomomys bottae humilis</i> R. H. Baker, 1953				

<i>Thomomys bottae jobobae</i> Huey, 1945			
<i>Thomomys bottae juarezensis</i> Huey, 1945			
<i>Thomomys bottae lucidus</i> Hall, 1932			
<i>Thomomys bottae martirensis</i> J. A. Allen, 1898			
<i>Thomomys bottae mearnsi</i> Bailey, 1914			
<i>Thomomys bottae modicus</i> Goldman, 1931			
<i>Thomomys bottae nigricans</i> Rhoads, 1895			
<i>Thomomys bottae perditus</i> Merriam, 1901			
<i>Thomomys bottae proximarinus</i> Huey, 1945			
<i>Thomomys bottae pusillus</i> Goldman, 1931			
<i>Thomomys bottae retractus</i> R. H. Baker, 1953			
<i>Thomomys bottae rhizophagus</i> Huey, 1949			
<i>Thomomys bottae ruricola</i> Huey, 1949			
<i>Thomomys bottae russeolus</i> Nelson and Goldman, 1909			
<i>Thomomys bottae siccovallis</i> Huey, 1945			
<i>Thomomys bottae simulus</i> Nelson and Goldman, 1934			
<i>Thomomys bottae sinaloae</i> Merriam, 1901			
<i>Thomomys bottae sturgisi</i> Goldman, 1938			
<i>Thomomys bottae toltecus</i> J. A. Allen, 1893			
<i>Thomomys bottae vanrossemi</i> Huey, 1934			
<i>Thomomys bottae varus</i> Hall and Long, 1960			
<i>Thomomys bottae villai</i> R. H. Baker, 1953			
<i>Thomomys bottae winthropi</i> Nelson and Goldman, 1934			
<i>Thomomys bottae xerophilus</i> Huey, 1945			
<i>Thomomys nayarensis</i> Mathis, Hafner, Hafner, and Demastes, 2013	En	Mo	Co
<i>Thomomys sheldoni</i> Bailey, 1915	En	Po	Co
<i>Thomomys sheldoni chihuahuae</i> Nelson and Goldman, 1934			
<i>Thomomys sheldoni sheldoni</i> Bailey, 1915			
<i>Thomomys umbrinus</i> (Richardson, 1829)	En	Po	Co
<i>Thomomys umbrinus arriagensis</i> Dalquest, 1951			
<i>Thomomys umbrinus atrodorsalis</i> Nelson and Goldman, 1934			
<i>Thomomys umbrinus camargensis</i> Anderson, 1972			
<i>Thomomys umbrinus crassidens</i> Nelson and Goldman, 1934			
<i>Thomomys umbrinus durangi</i> Nelson and Goldman, 1934			
<i>Thomomys umbrinus enixus</i> Nelson and Goldman, 1934			
<i>Thomomys umbrinus eximius</i> Nelson and Goldman, 1934			
<i>Thomomys umbrinus goldmani</i> Merriam, 1901			
<i>Thomomys umbrinus juntae</i> Anderson, 1972			
<i>Thomomys umbrinus madrensis</i> Nelson and Goldman, 1934			
<i>Thomomys umbrinus nelsoni</i> Merriam, 1901			
<i>Thomomys umbrinus newmani</i> Dalquest, 1951			

Thomomys umbrinus potosinus Nelson and Goldman, 1934
Thomomys umbrinus pullus Hall and Villa R., 1948
Thomomys umbrinus sonoriensis Nelson and Goldman, 1934
Thomomys umbrinus supernus Nelson and Goldman, 1934
Thomomys umbrinus umbrinus (Richardson, 1829)
Thomomys umbrinus zacatecae Nelson and Goldman, 1934

FAMILY HETEROMYIDAE Gray, 1868

SUBFAMILY DIPODOMYINAE GERVAIS, 1853

Dipodomys Gray, 1841***Dipodomys compactus* True, 1889**

Po Co

Dipodomys compactus compactus True, 1889***Dipodomys deserti* Stephens, 1887**

Po Co

Dipodomys deserti deserti Stephens, 1887*Dipodomys deserti sonoriensis* Goldman, 1923***Dipodomys gravipes* Huey, 1925**

E En Mo Co

***Dipodomys merriami* Mearns, 1890**

Po Co-In

Dipodomys merriami ambiguus Merriam, 1890*Dipodomys merriami annulus* Huey, 1951*Dipodomys merriami arenivagus* Elliot, 1903*Dipodomys merriami atronasus* Merriam, 1894*Dipodomys merriami brunensis* Huey, 1951*Dipodomys merriami insularis* Merriam, 1907 P*Dipodomys merriami margaritae* Merriam, 1907 P*Dipodomys merriami mayensis* Goldman, 1928*Dipodomys merriami melanurus* Merriam, 1893*Dipodomys merriami merriami* Mearns, 1890*Dipodomys merriami mitchelli* Mearns, 1897 A*Dipodomys merriami olivaceus* Swarth, 1929*Dipodomys merriami platycephalus* Merriam, 1907*Dipodomys merriami quintinensis* Huey, 1951*Dipodomys merriami trinidadensis* Huey, 1951***Dipodomys nelsoni* Merriam, 1907**

En Mo Co

***Dipodomys ordii* Woodhouse, 1853**

Po Co

Dipodomys ordii durranti Setzer, 1952*Dipodomys ordii extractus* Setzer, 1949*Dipodomys ordii obscurus* (J. A. Allen, 1903)*Dipodomys ordii ordii* Woodhouse, 1853*Dipodomys ordii palmeri* (J. A. Allen, 1891)*Dipodomys ordii pullus* Anderson, 1972***Dipodomys ornatus* Merriam, 1894**

En Mo Co

***Dipodomys phillipsii* Gray, 1841**

Pr En Po Co

<i>Dipodomys phillipsii oaxacae</i> Hooper, 1947	A			
<i>Dipodomys phillipsii perotensis</i> Merriam, 1894	A			
<i>Dipodomys phillipsii phillipsii</i> Gray, 1841	A			
<i>Dipodomys simulans</i> (Merriam, 1904)			Po	Co
<i>Dipodomys simulans peninsularis</i> (Merriam, 1907)				
<i>Dipodomys simulans simulans</i> (Merriam, 1904)				
<i>Dipodomys spectabilis</i> Merriam, 1890			Po	Co
<i>Dipodomys spectabilis cratodon</i> Merriam, 1907				
<i>Dipodomys spectabilis intermedius</i> Nader, 1965				
<i>Dipodomys spectabilis perblandus</i> Goldman, 1933				
<i>Dipodomys spectabilis spectabilis</i> Merriam, 1890				
<i>Dipodomys spectabilis zygomaticus</i> Goldman, 1923				
SUBFAMILY HETEROMYINAE Gray, 1868				
<i>Heteromys</i> Desmarest, 1817				
<i>Heteromys desmarestianus</i> Gray, 1868			Po	Co
<i>Heteromys desmarestianus desmarestianus</i> Gray, 1868				
<i>Heteromys gaumeri</i> J. A. Allen and Chapman, 1897	En		Mo	Co
<i>Heteromys goldmani</i> Merriam, 1902			Mo	Co
<i>Heteromys irroratus</i> Gray, 1868			Po	Co
<i>Heteromys irroratus alleni</i> Coues, 1881				
<i>Heteromys irroratus bulleri</i> Thomas, 1893				
<i>Heteromys irroratus guerrensis</i> (Goldman, 1911)				
<i>Heteromys irroratus irroratus</i> Gray, 1868				
<i>Heteromys irroratus jaliscensis</i> J. A. Allen, 1906				
<i>Heteromys irroratus texensis</i> (Merriam, 1902)				
<i>Heteromys irroratus torridus</i> (Merriam, 1902)				
<i>Heteromys nelsoni</i> Merriam, 1902	Pr	En	Mo	Co
<i>Heteromys pictus</i> Thomas, 1893			Po	Co
<i>Heteromys pictus annectens</i> Merriam, 1902				
<i>Heteromys pictus hispidus</i> J. A. Allen, 1897				
<i>Heteromys pictus pictus</i> Thomas, 1893				
<i>Heteromys pictus plantinarenis</i> (Merriam, 1902)				
<i>Heteromys salvini</i> Thomas, 1893			Po	Co
<i>Heteromys salvini crispus</i> (Merriam, 1902)				
<i>Heteromys spectabilis</i> (Genoways, 1971)	Pr	En	Mo	Co
<i>Heteromys temporalis</i> Goldman, 1911			Mo	Co
SUBFAMILY PEROGNATHINAE Coues, 1875				
<i>Chaetodipus</i> Merriam, 1889				
<i>Chaetodipus ammophilus</i> (Osgood, 1907)	A	En	Po	Co-In
<i>Chaetodipus ammophilus ammophilus</i> (Osgood, 1907)				

<i>Chaetodipus ammophilus dalquesti</i> (Roth, 1976)	Pr		
<i>Chaetodipus ammophilus sublucidus</i> (Nelson and Goldman, 1929)			
<i>Chaetodipus arenarius</i> (Merriam, 1894)		En	Po Co
<i>Chaetodipus arenarius albescens</i> (Huey, 1926)			
<i>Chaetodipus arenarius albulus</i> (Nelson and Goldman, 1923)	A		
<i>Chaetodipus arenarius ambiguus</i> (Nelson and Goldman, 1929)			
<i>Chaetodipus arenarius arenarius</i> (Merriam, 1894)			
<i>Chaetodipus arenarius helleri</i> (Elliot, 1903)			
<i>Chaetodipus arenarius mexicalis</i> (Huey, 1939)			
<i>Chaetodipus arenarius paralios</i> (Huey, 1964)			
<i>Chaetodipus arenarius ramirezpulidoi</i> Álvarez–Castañeda and Cortés–Calva, 2004			
<i>Chaetodipus arenarius sabulosus</i> (Huey, 1964)			
<i>Chaetodipus artus</i> (Osgood, 1900)		En	Mo Co
<i>Chaetodipus baileyi</i> (Merriam, 1894)			Po Co–In
<i>Chaetodipus baileyi baileyi</i> (Merriam, 1894)			
<i>Chaetodipus baileyi insularis</i> (Townsend, 1912)	P		
<i>Chaetodipus californicus</i> (Merriam, 1889)			Po Co
<i>Chaetodipus californicus femoralis</i> (J. A. Allen, 1891)			
<i>Chaetodipus californicus mesopolius</i> (Elliot, 1903)			
<i>Chaetodipus eremicus</i> (Mearns, 1898)			Po Co
<i>Chaetodipus eremicus atrodorsalis</i> (Dalquest, 1951)			
<i>Chaetodipus eremicus eremicus</i> (Mearns, 1898)			
<i>Chaetodipus fallax</i> (Merriam, 1889)			Po Co–In
<i>Chaetodipus fallax anthonyi</i> (Osgood, 1900)	A		
<i>Chaetodipus fallax fallax</i> (Merriam, 1889)			
<i>Chaetodipus fallax inopinus</i> (Nelson and Goldman, 1929)			
<i>Chaetodipus fallax majusculus</i> (Huey, 1960)			
<i>Chaetodipus fallax xerotrophicus</i> (Huey, 1960)			
<i>Chaetodipus formosus</i> (Merriam, 1889)			Po Co
<i>Chaetodipus formosus cinerascens</i> (Nelson and Goldman, 1929)			
<i>Chaetodipus formosus infolatus</i> (Huey, 1954)			
<i>Chaetodipus formosus mesembrinus</i> (Elliot, 1904)			
<i>Chaetodipus goldmani</i> (Osgood, 1900)		En	Mo Co
<i>Chaetodipus hispidus</i> (Baird, 1857)			Po Co
<i>Chaetodipus hispidus conditi</i> (J. A. Allen, 1894)			
<i>Chaetodipus hispidus hispidus</i> (Baird, 1857)			
<i>Chaetodipus intermedius</i> (Merriam, 1889)			Po Co–In
<i>Chaetodipus intermedius intermedius</i> (Merriam, 1889)			
<i>Chaetodipus intermedius lithophilus</i> (Huey, 1937)			

<i>Chaetodipus intermedius minimus</i> (Burt, 1932)	A			
<i>Chaetodipus intermedius phasma</i> (Goldman, 1918)				
<i>Chaetodipus lineatus</i> (Dalquest, 1951)		En	Mo	Co
<i>Chaetodipus nelsoni</i> (Merriam, 1894)		En	Po	Co
<i>Chaetodipus nelsoni canescens</i> (Merriam, 1894)				
<i>Chaetodipus nelsoni nelsoni</i> (Merriam, 1894)				
<i>Chaetodipus penicillatus</i> (Woodhouse, 1852)			Po	Co–In
<i>Chaetodipus penicillatus angustirostris</i> (Osgood, 1900)				
<i>Chaetodipus penicillatus pricei</i> (J. A. Allen, 1894)				
<i>Chaetodipus penicillatus seri</i> (Nelson, 1912)	A			
<i>Chaetodipus pernix</i> (J. A. Allen, 1898)		En	Po	Co
<i>Chaetodipus pernix pernix</i> (J. A. Allen, 1898)				
<i>Chaetodipus pernix rostratus</i> (Osgood, 1900)				
<i>Chaetodipus rudinoris</i> (Elliot, 1903)			Po	Co–In
<i>Chaetodipus rudinoris extimus</i> (Nelson and Goldman, 1930)				
<i>Chaetodipus rudinoris fornicatus</i> (Burt, 1932)				
<i>Chaetodipus rudinoris hueyi</i> (Nelson and Goldman, 1929)				
<i>Chaetodipus rudinoris mesidios</i> (Huey, 1964)				
<i>Chaetodipus rudinoris rudinoris</i> (Elliot, 1903)				
<i>Chaetodipus siccus</i> (Osgood, 1907)	A	En	Mo	Co–In
<i>Chaetodipus spinatus</i> (Merriam, 1889)			Po	Co–In
<i>Chaetodipus spinatus broccus</i> (Huey, 1960)				
<i>Chaetodipus spinatus bryanti</i> (Merriam, 1894)	P			
<i>Chaetodipus spinatus evermanni</i> (Nelson and Goldman, 1929)	E			
<i>Chaetodipus spinatus guardiaie</i> (Burt, 1932)	A			
<i>Chaetodipus spinatus lambi</i> (Benson, 1930)	A			
<i>Chaetodipus spinatus latijugularis</i> (Burt, 1932)	P			
<i>Chaetodipus spinatus lorenzi</i> (Banks, 1967)	A			
<i>Chaetodipus spinatus magdalenae</i> (Osgood, 1907)				
<i>Chaetodipus spinatus marcosensis</i> (Burt, 1932)	A			
<i>Chaetodipus spinatus margaritae</i> (Merriam, 1894)	A			
<i>Chaetodipus spinatus occultus</i> (Nelson, 1912)	A			
<i>Chaetodipus spinatus oribates</i> (Huey, 1960)				
<i>Chaetodipus spinatus peninsulae</i> (Merriam, 1894)				
<i>Chaetodipus spinatus prietae</i> (Huey, 1930)				
<i>Chaetodipus spinatus pullus</i> (Burt, 1932)	A			
<i>Chaetodipus spinatus seorsus</i> (Burt, 1932)	A			
<i>Chaetodipus spinatus spinatus</i> (Merriam, 1889)				
<i>Perognathus</i> Wied–Neuwied, 1839				
<i>Perognathus amplus</i> Osgood, 1900			Po	Co
<i>Perognathus amplus amplus</i> Osgood, 1900	Pr			
<i>Perognathus amplus taylori</i> Goldman, 1932				

<i>Perognathus flavescens</i> Merriam, 1889		Po	Co
<i>Perognathus flavescens melanotis</i> Osgood, 1900			
<i>Perognathus flavus</i> Baird, 1855		Po	Co
<i>Perognathus flavus flavus</i> Baird, 1855			
<i>Perognathus flavus fuscus</i> Anderson, 1972			
<i>Perognathus flavus medius</i> R. H. Baker, 1954			
<i>Perognathus flavus mexicanus</i> Merriam, 1894			
<i>Perognathus flavus pallescens</i> R. H. Baker, 1954			
<i>Perognathus flavus parviceps</i> R. H. Baker, 1954			
<i>Perognathus flavus sonoriensis</i> Nelson and Goldman, 1934			
<i>Perognathus longimembris</i> (Coues, 1875)		Po	Co
<i>Perognathus longimembris aestivus</i> Huey, 1928			
<i>Perognathus longimembris bombycinus</i> Osgood, 1907			
<i>Perognathus longimembris internationalis</i> Huey, 1939			
<i>Perognathus longimembris kinoensis</i> Huey, 1935			
<i>Perognathus longimembris venustus</i> Huey, 1930			
<i>Perognathus merriami</i> J. A. Allen, 1892		Po	Co
<i>Perognathus merriami gilvus</i> Osgood, 1900			
<i>Perognathus merriami merriami</i> J. A. Allen, 1892			
SUBORDER HYSTRICOGNATHA Woods, 1976			
INFRAORDER HYSTRICOGNATHI Tullberg, 1899			
FAMILY ERETHIZONTIDAE Bonaparte, 1845			
SUBFAMILY ERETHIZONTINAE Bonaparte, 1845			
<i>Erethizon</i> F. Cuvier, 1823			
<i>Erethizon dorsatum</i> (Linnaeus, 1758)	P	Po	Co
<i>Erethizon dorsatum epixanthum</i> Brandt, 1835			
<i>Coendou</i> Lacépède, 1799			
<i>Coendou mexicanus</i> (Kerr, 1792)	A	Po	Co
<i>Coendou mexicanus mexicanus</i> (Kerr, 1792)			
<i>Coendou mexicanus yucataniae</i> (Thomas, 1902)			
PARVORDER CAVIIDA Bryant and McKenna, 1995			
SUPERFAMILY CAVIOIDEA G. Fischer, 1817			
FAMILY AGOUTIDAE Gray, 1821			
SUBFAMILY DASYPROCTINAE Gray, 1825			
<i>Dasyprocta</i> Illiger, 1811			
<i>Dasyprocta mexicana</i> de Saussure, 1860	En	Mo	Co
<i>Dasyprocta punctata</i> Gray, 1842		Po	Co-In
<i>Dasyprocta punctata chiapensis</i> Goldman, 1913			
<i>Dasyprocta punctata yucatanica</i> Goldman, 1913			

FAMILY CUNICULIDAE Miller and Gidley, 1918				
<i>Cuniculus</i> Brisson, 1762				
<i>Cuniculus paca</i> (Linnaeus, 1766)			Po	Co–In
<i>Cuniculus paca nelsoni</i> (Goldman, 1913)				
INFRAORDER CASTORIMORPHA Wood, 1955				
FAMILY CASTORIDAE Hemprich, 1820				
SUBFAMILY CASTORINAE Hemprich, 1820				
TRIBE CASTORINI Hemprich, 1820				
SUBTRIBE CASTORINA Hemprich, 1820				
<i>Castor</i> Linnaeus, 1758				
<i>Castor canadensis</i> Kuhl, 1820	P		Po	Co
<i>Castor canadensis frondator</i> Mearns, 1897				
<i>Castor canadensis mexicanus</i> Bailey, 1913				
SUBORDER MYOMORPHA Brandt, 1855				
INFRAORDER MYODONTA Schaub in: Grassé and Dekeyser, 1955				
SUPERFAMILY MUROIDEA Illiger, 1811				
FAMILY CRICETIDAE G. Fischer, 1817				
SUBFAMILY ARVICOLINAE Gray, 1821				
TRIBE ARVICOLINI Gray, 1821				
<i>Microtus</i> Schrank, 1798				
<i>Microtus californicus</i> (Peale, 1848)	P		Po	Co
<i>Microtus californicus aequivocatus</i> Osgood, 1928				
<i>Microtus californicus grinnelli</i> Huey, 1931				
<i>Microtus californicus hyperythrus</i> Elliot, 1903				
<i>Microtus guatemalensis</i> Merriam, 1898	A		Mo	Co
<i>Microtus mexicanus</i> (de Saussure, 1861)			Po	Co
<i>Microtus mexicanus fulviventer</i> Merriam, 1898				
<i>Microtus mexicanus fundatus</i> Hall, 1948				
<i>Microtus mexicanus madrensis</i> Goldman, 1938				
<i>Microtus mexicanus mexicanus</i> (de Saussure, 1861)				
<i>Microtus mexicanus neveriae</i> Hooper, 1955				
<i>Microtus mexicanus ocotensis</i> Álvarez and Hernández–Chávez, 1993				
<i>Microtus mexicanus phaeus</i> (Merriam, 1892)				
<i>Microtus mexicanus salvus</i> Hall, 1948				
<i>Microtus mexicanus subsimus</i> Goldman, 1938				
<i>Microtus oaxacensis</i> Goodwin, 1966	A	En	Mo	Co
<i>Microtus pennsylvanicus</i> (Ord, 1815)	P		Po	Co
<i>Microtus pennsylvanicus chihuahuensis</i> W. G. Bradley and Cockrum, 1968				

<i>Microtus quasiater</i> (Coues, 1874)	Pr	En	Mo	Co
<i>Microtus umbrosus</i> Merriam, 1898	Pr	En	Mo	Co
TRIBE ONDATRINI Gray, 1825				
<i>Ondatra</i> Link, 1795				
<i>Ondatra zibethicus</i> (Linnaeus, 1766)	A		Po	Co
<i>Ondatra zibethicus pallidus</i> (Mearns, 1890)				
<i>Ondatra zibethicus ripensis</i> (Bailey, 1902)				
SUBFAMILY NEOTOMINAE Merriam, 1894				
TRIBE BAIOMYINI Musser and Carleton, 2005				
<i>Baiomys</i> True, 1894				
<i>Baiomys musculus</i> (Merriam, 1892)			Po	Co
<i>Baiomys musculus brunneus</i> (J. A. Allen and Chapman, 1897)				
<i>Baiomys musculus infernalis</i> Hooper, 1952				
<i>Baiomys musculus musculus</i> (Merriam, 1892)				
<i>Baiomys musculus nigrescens</i> (Osgood, 1904)				
<i>Baiomys musculus pallidus</i> Russell, 1952				
<i>Baiomys taylori</i> (Thomas, 1887)			Po	Co
<i>Baiomys taylori allex</i> (Osgood, 1904)				
<i>Baiomys taylori analogus</i> (Osgood, 1909)				
<i>Baiomys taylori ater</i> Blossom and Burt, 1942				
<i>Baiomys taylori canutus</i> Packard, 1960				
<i>Baiomys taylori fuliginatus</i> Packard, 1960				
<i>Baiomys taylori paulus</i> (J. A. Allen, 1903)				
<i>Baiomys taylori taylori</i> (Thomas, 1887)				
<i>Scotinomys</i> Thomas, 1913				
<i>Scotinomys teguina</i> (Alston, 1877)			Po	Co
<i>Scotinomys teguina teguina</i> (Alston, 1877)				
TRIBE NEOTOMINI Vorontsov, 1959				
<i>Hodomys</i> Merriam, 1894				
<i>Hodomys alleni</i> (Merriam, 1892)		En	Po	Co
<i>Hodomys alleni alleni</i> (Merriam, 1892)				
<i>Hodomys alleni elatturus</i> Osgood, 1904				
<i>Hodomys alleni guerrerensis</i> Goldman, 1938				
<i>Hodomys alleni vetulus</i> Merriam, 1894				
<i>Nelsonia</i> Merriam, 1897				
<i>Nelsonia goldmani</i> Merriam, 1903	Pr	En	Po	Co
<i>Nelsonia goldmani cliftoni</i> Genoways and J. K. Jones, 1968				
<i>Nelsonia goldmani goldmani</i> Merriam, 1903				
<i>Nelsonia neotomodon</i> Merriam, 1897	Pr	En	Mo	Co

Neotoma Say and Ord, 1825

<i>Neotoma albigula</i> Hartley, 1894		Po	Co–In
<i>Neotoma albigula albigula</i> Hartley, 1894			
<i>Neotoma albigula melanura</i> Merriam, 1894			
<i>Neotoma albigula seri</i> Townsend, 1912	A		
<i>Neotoma albigula mearnsi</i> Goldman, 1915			
<i>Neotoma albigula varia</i> Burt, 1932	P		
<i>Neotoma albigula venusta</i> True, 1894			
<i>Neotoma angustapalata</i> R. H. Baker, 1951		En	Mo Co
<i>Neotoma bryanti</i> Merriam, 1887	A	En	Po Co–In
<i>Neotoma bryanti anthonyi</i> J. A. Allen, 1898	E		
<i>Neotoma bryanti bryanti</i> Merriam, 1887			
<i>Neotoma bryanti intermedia</i> Rhoads, 1894			
<i>Neotoma bryanti marcosensis</i> Burt, 1932	A		
<i>Neotoma bryanti martinensis</i> Goldman, 1905	P		
<i>Neotoma devia</i> Goldman, 1927			Mo Co
<i>Neotoma goldmani</i> Merriam, 1903		En	Mo Co
<i>Neotoma insularis</i> Townsend, 1912	A	En	Mo In
<i>Neotoma lepida</i> Thomas, 1893			Po Co–In
<i>Neotoma lepida lepida</i> Thomas, 1893			
<i>Neotoma leucodon</i> Merriam, 1894		En	Po Co
<i>Neotoma leucodon durangae</i> J. A. Allen, 1903			
<i>Neotoma leucodon latifrons</i> Merriam, 1894			
<i>Neotoma leucodon leucodon</i> Merriam, 1894			
<i>Neotoma leucodon subsolana</i> Álvarez, 1962			
<i>Neotoma macrotis</i> Thomas, 1893		En	Mo Co
<i>Neotoma mexicana</i> Baird, 1855			Po Co
<i>Neotoma mexicana distincta</i> Bangs, 1903			
<i>Neotoma mexicana eremita</i> Hall, 1955			
<i>Neotoma mexicana griseoventer</i> Dalquest, 1951			
<i>Neotoma mexicana inornata</i> Goldman, 1938			
<i>Neotoma mexicana isthmica</i> Goldman, 1904			
<i>Neotoma mexicana mexicana</i> Baird, 1855			
<i>Neotoma mexicana navus</i> Merriam, 1903			
<i>Neotoma mexicana ochracea</i> Goldman, 1905			
<i>Neotoma mexicana parvidens</i> Goldman, 1904			
<i>Neotoma mexicana picta</i> Goldman, 1904			
<i>Neotoma mexicana sinaloae</i> J. A. Allen, 1898			
<i>Neotoma mexicana tenuicauda</i> Merriam, 1892			
<i>Neotoma mexicana torquata</i> Ward, 1891			
<i>Neotoma mexicana tropicalis</i> Goldman, 1904			
<i>Neotoma mexicana vulcani</i> Sanborn, 1935			

<i>Neotoma micropus</i> Baird, 1855			Po	Co
<i>Neotoma micropus canescens</i> J. A. Allen, 1891				
<i>Neotoma micropus micropus</i> Baird, 1855				
<i>Neotoma micropus planiceps</i> Goldman, 1905				
<i>Neotoma nelsoni</i> Goldman, 1905		En	Mo	Co
<i>Neotoma palatina</i> Goldman, 1905		En	Mo	Co
<i>Neotoma phenax</i> (Merriam, 1903)	Pr	En	Mo	Co
<i>Xenomys</i> Merriam, 1892		En		
<i>Xenomys nelsoni</i> Merriam, 1892	A	En	Mo	Co
TRIBE REITHRODONTOMYINI Vorontsov, 1959				
<i>Habromys</i> Hooper and Musser, 1964				
<i>Habromys chinanteco</i> (Robertson and Musser, 1976)		En	Mo	Co
<i>Habromys delicatulus</i> Carleton, Sánchez, and Urbano Vidales, 2002		En	Mo	Co
<i>Habromys ixtlani</i> (Goodwin, 1964)		En	Mo	Co
<i>Habromys lepturus</i> (Merriam, 1898)		En	Mo	Co
<i>Habromys lophurus</i> (Osgood, 1904)			Mo	Co
<i>Habromys schmidlyi</i> Romo-Vázquez, León-Paniagua, and Sánchez, 2005		En	Mo	Co
<i>Habromys simulatus</i> (Osgood, 1904)	Pr	En	Mo	Co
<i>Megadontomys</i> Merriam, 1898		En		
<i>Megadontomys cryophilus</i> (Musser, 1964)	A	En	Mo	Co
<i>Megadontomys nelsoni</i> (Merriam, 1898)	A	En	Mo	Co
<i>Megadontomys thomasi</i> (Merriam, 1898)	Pr	En	Mo	Co
<i>Neotomodon</i> Merriam, 1898		En		
<i>Neotomodon alstoni</i> Merriam, 1898		En	Mo	Co
<i>Onychomys</i> Baird, 1857				
<i>Onychomys arenicola</i> Mearns, 1896			Po	Co
<i>Onychomys arenicola ater</i> Anderson, 1972				
<i>Onychomys arenicola canus</i> Merriam, 1904				
<i>Onychomys arenicola sarrufus</i> Hollister, 1914				
<i>Onychomys leucogaster</i> (Wied-Neuwied, 1841)			Po	Co
<i>Onychomys leucogaster albescens</i> Merriam, 1904				
<i>Onychomys leucogaster longipes</i> Merriam, 1889				
<i>Onychomys leucogaster ruidosae</i> Stone and Rehn, 1903				
<i>Onychomys torridus</i> (Coues, 1874)			Po	Co
<i>Onychomys torridus knoxjonesi</i> Hollander and Willig, 1992				
<i>Onychomys torridus macrotis</i> Elliot, 1903				
<i>Onychomys torridus pulcher</i> Elliot, 1904				
<i>Onychomys torridus ramona</i> Rhoads, 1893				
<i>Onychomys torridus torridus</i> (Coues, 1874)				

<i>Onychomys torridus yakiensis</i> Merriam, 1904				
<i>Osgoodomys</i> Hooper and Musser, 1964		En		
<i>Osgoodomys banderanus</i> (J. A. Allen, 1897)		En	Po	Co
<i>Osgoodomys banderanus banderanus</i> (J. A. Allen, 1897)				
<i>Osgoodomys banderanus vicinior</i> (Osgood, 1904)				
<i>Peromyscus</i> Gloger, 1841				
<i>Peromyscus aztecus</i> (de Saussure, 1860)			Po	Co
<i>Peromyscus aztecus aztecus</i> (de Saussure, 1860)				
<i>Peromyscus aztecus evides</i> Osgood, 1904				
<i>Peromyscus aztecus oaxacensis</i> Merriam, 1898				
<i>Peromyscus beatae</i> Thomas, 1903		En	Po	Co
<i>Peromyscus beatae beatae</i> Thomas, 1903				
<i>Peromyscus beatae sacarensis</i> Dickey, 1928				
<i>Peromyscus boylii</i> (Baird, 1855)			Po	Co–In
<i>Peromyscus boylii glasselli</i> Burt, 1932	A			
<i>Peromyscus boylii rowleyi</i> (J. A. Allen, 1893)				
<i>Peromyscus bullatus</i> Osgood, 1904	Pr	En	Mo	Co
<i>Peromyscus californicus</i> (Gambel, 1848)			Po	Co
<i>Peromyscus californicus insignis</i> Rhoads, 1895				
<i>Peromyscus caniceps</i> Burt, 1932	Pr	En	Mo	In
<i>Peromyscus carletoni</i> R. D. Bradley, Ordóñez-Garza, Sotero-Caio, Huynh et al., 2014		En	Mo	Co
<i>Peromyscus crinitus</i> (Merriam, 1891)			Po	Co–In
<i>Peromyscus crinitus delgadilli</i> Benson, 1940				
<i>Peromyscus crinitus disparilis</i> Goldman, 1932				
<i>Peromyscus crinitus pallidissimus</i> Huey, 1931	A			
<i>Peromyscus crinitus stephensi</i> Mearns, 1897				
<i>Peromyscus dickeyi</i> Burt, 1932	Pr	En	Mo	In
<i>Peromyscus difficilis</i> (J. A. Allen, 1891)		En	Po	Co
<i>Peromyscus difficilis amplus</i> Osgood, 1904				
<i>Peromyscus difficilis difficilis</i> (J. A. Allen, 1891)				
<i>Peromyscus difficilis felipensis</i> Merriam, 1898				
<i>Peromyscus difficilis petricola</i> Hoffmeister and de la Torre, 1959				
<i>Peromyscus difficilis saxicola</i> Hoffmeister and de la Torre, 1959				
<i>Peromyscus eremicus</i> (Baird, 1857)			Po	Co–In
<i>Peromyscus eremicus alcorni</i> Anderson, 1972				
<i>Peromyscus eremicus anthonyi</i> (Merriam, 1887)				
<i>Peromyscus eremicus avius</i> Osgood, 1909	A			
<i>Peromyscus eremicus cedrosensis</i> J. A. Allen, 1898	A			
<i>Peromyscus eremicus cinereus</i> Hall, 1931	A			

<i>Peromyscus eremicus collatus</i> Burt, 1932	A			
<i>Peromyscus eremicus eremicus</i> (Baird, 1857)				
<i>Peromyscus eremicus insulicola</i> Osgood, 1909	A			
<i>Peromyscus eremicus papagensis</i> Goldman, 1917				
<i>Peromyscus eremicus phaeurus</i> Osgood, 1904				
<i>Peromyscus eremicus polypolius</i> Osgood, 1909	A			
<i>Peromyscus eremicus sinaloensis</i> Anderson, 1972				
<i>Peromyscus eremicus tiburonensis</i> Mearns, 1897	A			
<i>Peromyscus eva</i> Thomas, 1898		En	Po	Co
<i>Peromyscus eva carmeni</i> Townsend, 1912	A			
<i>Peromyscus eva eva</i> Thomas, 1898				
<i>Peromyscus fraterculus</i> (Miller, 1892)			Mo	Co-In
<i>Peromyscus furvus</i> J. A. Allen and Chapman, 1897		En	Mo	Co
<i>Peromyscus gratus</i> Merriam, 1898		En	Po	Co
<i>Peromyscus gratus erasmus</i> Finley, 1952				
<i>Peromyscus gratus gentilis</i> Osgood, 1904				
<i>Peromyscus gratus gratus</i> Merriam, 1898				
<i>Peromyscus gratus zapotecae</i> Hooper, 1957				
<i>Peromyscus guardia</i> Townsend, 1912	P	En	Po	In
<i>Peromyscus guardia guardia</i> Townsend, 1912				
<i>Peromyscus guardia harbisoni</i> Banks, 1967				
<i>Peromyscus guardia mejiae</i> Burt, 1932				
<i>Peromyscus guatemalensis</i> Merriam, 1898			Mo	Co
<i>Peromyscus gymnotis</i> Thomas, 1894			Mo	Co
<i>Peromyscus hooperi</i> Lee and Schmidly, 1977		En	Mo	Co
<i>Peromyscus hylocetes</i> Merriam, 1898		En	Mo	Co
<i>Peromyscus interparietalis</i> Burt, 1932	A	En	Po	In
<i>Peromyscus interparietalis interparietalis</i> Burt, 1932				
<i>Peromyscus interparietalis lorenzi</i> Banks, 1967				
<i>Peromyscus interparietalis ryckmani</i> Banks, 1967				
<i>Peromyscus latirostris</i> Dalquest, 1950		En	Mo	Co
<i>Peromyscus leucopus</i> (Rafinesque, 1818)			Po	Co-In
<i>Peromyscus leucopus affinis</i> (J. A. Allen, 1891)				
<i>Peromyscus leucopus arizonae</i> (J. A. Allen, 1894)				
<i>Peromyscus leucopus castaneus</i> Osgood, 1904				
<i>Peromyscus leucopus cozumelae</i> Merriam, 1901	A			
<i>Peromyscus leucopus incensus</i> Goldman, 1942				
<i>Peromyscus leucopus lachiguiriensis</i> Goodwin, 1956				
<i>Peromyscus leucopus mesomelas</i> Osgood, 1904				
<i>Peromyscus leucopus texanus</i> (Woodhouse, 1853)				
<i>Peromyscus leucopus tornillo</i> Mearns, 1896				
<i>Peromyscus levipes</i> Merriam, 1898		En	Po	Co

<i>Peromyscus levipes ambiguus</i> Álvarez, 1961				
<i>Peromyscus levipes levipes</i> Merriam, 1898				
<i>Peromyscus madrensis</i> Merriam, 1898	A	En	Mo	In
<i>Peromyscus maniculatus</i> (Wagner, 1845)			Po	Co–In
<i>Peromyscus maniculatus assimilis</i> Nelson and Goldman, 1931				
<i>Peromyscus maniculatus blandus</i> Osgood, 1904				
<i>Peromyscus maniculatus cineritius</i> J. A. Allen, 1898	E			
<i>Peromyscus maniculatus coolidgei</i> Thomas, 1898				
<i>Peromyscus maniculatus dorsalis</i> Nelson and Goldman, 1931	A			
<i>Peromyscus maniculatus dubius</i> J. A. Allen, 1898	A			
<i>Peromyscus maniculatus exiguus</i> J. A. Allen, 1898	A			
<i>Peromyscus maniculatus fulvus</i> Osgood, 1904				
<i>Peromyscus maniculatus gambelii</i> (Baird, 1857)				
<i>Peromyscus maniculatus geronimensis</i> J. A. Allen, 1898	A			
<i>Peromyscus maniculatus hueyi</i> Nelson and Goldman, 1932				
<i>Peromyscus maniculatus labecula</i> Elliot, 1903				
<i>Peromyscus maniculatus magdalenae</i> Osgood, 1909	A			
<i>Peromyscus maniculatus margaritae</i> Osgood, 1909	A			
<i>Peromyscus maniculatus rufinus</i> (Merriam, 1890)				
<i>Peromyscus maniculatus sonoriensis</i> (J. L. Le Conte, 1853)				
<i>Peromyscus megalops</i> Merriam, 1898		En	Po	Co
<i>Peromyscus megalops auritus</i> Merriam, 1898				
<i>Peromyscus megalops megalops</i> Merriam, 1898				
<i>Peromyscus mekisturus</i> Merriam, 1898	A	En	Mo	Co
<i>Peromyscus melanocarpus</i> Osgood, 1904		En	Mo	Co
<i>Peromyscus melanophrys</i> (Coues, 1874)		En	Po	Co
<i>Peromyscus melanophrys coahuilensis</i> R. H. Baker, 1952				
<i>Peromyscus melanophrys consobrinus</i> Osgood, 1904				
<i>Peromyscus melanophrys melanophrys</i> (Coues, 1874)				
<i>Peromyscus melanophrys micropus</i> R. H. Baker, 1952				
<i>Peromyscus melanophrys xenurus</i> Osgood, 1904				
<i>Peromyscus melanophrys zamorae</i> Osgood, 1904				
<i>Peromyscus melanotis</i> J. A. Allen and Chapman, 1897		En	Mo	Co
<i>Peromyscus melanurus</i> Osgood, 1909		En	Mo	Co
<i>Peromyscus merriami</i> Mearns, 1896			Po	Co
<i>Peromyscus merriami goldmani</i> Osgood, 1904				
<i>Peromyscus merriami merriami</i> Mearns, 1896				
<i>Peromyscus mexicanus</i> (de Saussure, 1860)			Po	Co
<i>Peromyscus mexicanus angelensis</i> Osgood, 1904				
<i>Peromyscus mexicanus azulensis</i> Goodwin, 1956				
<i>Peromyscus mexicanus mexicanus</i> (de Saussure, 1860)				
<i>Peromyscus mexicanus putlaensis</i> Goodwin, 1964				

<i>Peromyscus mexicanus saxatilis</i> Merriam, 1898				
<i>Peromyscus mexicanus teapensis</i> Osgood, 1904				
<i>Peromyscus mexicanus tehuantepecus</i> Merriam, 1898				
<i>Peromyscus mexicanus totontepecus</i> Merriam, 1898				
<i>Peromyscus nasutus</i> (J. A. Allen, 1891)			Po	Co
<i>Peromyscus nasutus penicillatus</i> Mearns, 1896				
<i>Peromyscus ochraventer</i> R. H. Baker, 1951		En	Mo	Co
<i>Peromyscus pectoralis</i> Osgood, 1904			Po	Co
<i>Peromyscus pectoralis collinus</i> Hooper, 1952				
<i>Peromyscus pectoralis laceianus</i> Bailey, 1906				
<i>Peromyscus pectoralis pectoralis</i> Osgood, 1904				
<i>Peromyscus pambertonii</i> Burt, 1932	E	En	Mo	In
<i>Peromyscus perfulvus</i> Osgood, 1945		En	Po	Co
<i>Peromyscus perfulvus chrysopus</i> Hooper, 1955				
<i>Peromyscus perfulvus perfulvus</i> Osgood, 1945				
<i>Peromyscus polius</i> Osgood, 1904		En	Mo	Co
<i>Peromyscus pseudocrinitus</i> Burt, 1932	A	En	Mo	In
<i>Peromyscus sagax</i> Elliot, 1903		En	Mo	Co
<i>Peromyscus schmidlyi</i> R. D. Bradley, Carroll, Haynie, Muñíz-Martínez, et al. 2004		En	Mo	Co
<i>Peromyscus sejugis</i> Burt, 1932	A	En	Mo	In
<i>Peromyscus simulus</i> Osgood, 1904		En	Mo	Co
<i>Peromyscus slevini</i> Mailliard, 1924	A	En	Mo	In
<i>Peromyscus spicilegus</i> J. A. Allen, 1897		En	Mo	Co
<i>Peromyscus stephani</i> Townsend, 1912	A	En	Mo	In
<i>Peromyscus truei</i> (Shufeldt, 1885)			Po	Co
<i>Peromyscus truei lagunae</i> Osgood, 1909				
<i>Peromyscus truei martirensis</i> (J. A. Allen, 1893)				
<i>Peromyscus winkelmanni</i> Carleton, 1977	Pr	En	Mo	Co
<i>Peromyscus yucatanicus</i> J. A. Allen and Chapman, 1897		En	Po	Co
<i>Peromyscus yucatanicus badius</i> Osgood, 1904				
<i>Peromyscus yucatanicus yucatanicus</i> J. A. Allen and Chapman, 1897				
<i>Peromyscus zarhynchus</i> Merriam, 1898	Pr	En	Mo	Co
<i>Reithrodontomys</i> Giglioli, 1874				
<i>Reithrodontomys bakeri</i> R. D. Bradley, Mendez-Harclerode, Hamilton, and Ceballos, 2004		En	Mo	Co
<i>Reithrodontomys burti</i> Benson, 1939		En	Mo	Co
<i>Reithrodontomys chrysopsis</i> Merriam, 1900		En	Po	Co
<i>Reithrodontomys chrysopsis chrysopsis</i> Merriam, 1900				
<i>Reithrodontomys chrysopsis perotensis</i> Merriam, 1901				
<i>Reithrodontomys fulvescens</i> J. A. Allen, 1894			Po	Co

<i>Reithrodontomys fulvescens amoenus</i> (Elliot, 1905)				
<i>Reithrodontomys fulvescens canus</i> Benson, 1939				
<i>Reithrodontomys fulvescens chiapensis</i> A. H. Howell, 1914				
<i>Reithrodontomys fulvescens difficilis</i> Merriam, 1901				
<i>Reithrodontomys fulvescens fulvescens</i> J. A. Allen, 1894				
<i>Reithrodontomys fulvescens griseoflavus</i> Merriam, 1901				
<i>Reithrodontomys fulvescens helvolus</i> Merriam, 1901				
<i>Reithrodontomys fulvescens infernatis</i> Hooper, 1950				
<i>Reithrodontomys fulvescens intermedius</i> J. A. Allen, 1895				
<i>Reithrodontomys fulvescens mustelinus</i> A. H. Howell, 1914				
<i>Reithrodontomys fulvescens nelsoni</i> A. H. Howell, 1914				
<i>Reithrodontomys fulvescens tenuis</i> J. A. Allen, 1899				
<i>Reithrodontomys fulvescens toltecus</i> Merriam, 1901				
<i>Reithrodontomys fulvescens tropicalis</i> W. B. Davis, 1944				
<i>Reithrodontomys gracilis</i> J. A. Allen and Chapman, 1897		Po		Co-In
<i>Reithrodontomys gracilis gracilis</i> J. A. Allen and Chapman, 1897				
<i>Reithrodontomys gracilis insularis</i> J. K. Jones, 1964	A			
<i>Reithrodontomys gracilis pacificus</i> Goodwin, 1932				
<i>Reithrodontomys hirsutus</i> Merriam, 1901		En	Mo	Co
<i>Reithrodontomys megalotis</i> (Baird, 1857)			Po	Co-In
<i>Reithrodontomys megalotis alticola</i> Merriam, 1901				
<i>Reithrodontomys megalotis amoles</i> A. H. Howell, 1914				
<i>Reithrodontomys megalotis hooperi</i> Goodwin, 1954				
<i>Reithrodontomys megalotis longicaudus</i> (Baird, 1857)				
<i>Reithrodontomys megalotis megalotis</i> (Baird, 1857)				
<i>Reithrodontomys megalotis peninsulae</i> Elliot, 1903				
<i>Reithrodontomys megalotis saturatus</i> J. A. Allen and Chapman, 1897				
<i>Reithrodontomys mexicanus</i> (de Saussure, 1860)			Po	Co
<i>Reithrodontomys mexicanus howelli</i> Goodwin, 1932				
<i>Reithrodontomys mexicanus mexicanus</i> (de Saussure, 1860)				
<i>Reithrodontomys mexicanus riparius</i> Hooper, 1955				
<i>Reithrodontomys mexicanus scansor</i> Hooper, 1950				
<i>Reithrodontomys microdon</i> Merriam, 1901	A		Po	Co
<i>Reithrodontomys microdon albilabris</i> Merriam, 1901				
<i>Reithrodontomys microdon microdon</i> Merriam, 1901				
<i>Reithrodontomys microdon wagneri</i> Hooper, 1950				
<i>Reithrodontomys montanus</i> (Baird, 1855)			Po	Co
<i>Reithrodontomys montanus montanus</i> (Baird, 1855)				
<i>Reithrodontomys spectabilis</i> J. K. Jones and Lawlor, 1965	A	En	Mo	In
<i>Reithrodontomys sumichrasti</i> (de Saussure, 1860)			Po	Co

<i>Reithrodontomys sumichrasti dorsalis</i> Merriam, 1901				
<i>Reithrodontomys sumichrasti luteolus</i> A. H. Howell, 1914				
<i>Reithrodontomys sumichrasti nerterus</i> Merriam, 1901				
<i>Reithrodontomys sumichrasti sumichrasti</i> (de Saussure, 1860)				
<i>Reithrodontomys tenuirostris</i> Merriam, 1901		Mo	Co	
<i>Reithrodontomys zacatecae</i> Merriam, 1901	En	Mo	Co	
SUBFAMILY SIGMODONTINAE Wagner, 1843				
TRIBE ORYZOMYINI Vorontsov, 1959				
<i>Oligoryzomys</i> Bangs, 1900				
<i>Oligoryzomys fulvescens</i> (de Saussure, 1860)		Po	Co	
<i>Oligoryzomys fulvescens engraciae</i> (Osgood, 1945)				
<i>Oligoryzomys fulvescens fulvescens</i> (de Saussure, 1860)				
<i>Oligoryzomys fulvescens lenis</i> (Goldman, 1915)				
<i>Oligoryzomys fulvescens mayensis</i> (Goldman, 1918)				
<i>Oligoryzomys fulvescens pacificus</i> (Hooper, 1952)				
<i>Oryzomys</i> Baird, 1857				
<i>Oryzomys albiventer</i> Merriam, 1901	En	Mo	Co	
<i>Oryzomys alfaroi</i> (J. A. Allen, 1891)		Po	Co	
<i>Oryzomys alfaroi agrestis</i> Goodwin, 1959				
<i>Oryzomys alfaroi gloriaensis</i> Goodwin, 1956				
<i>Oryzomys alfaroi palatinus</i> Merriam, 1901				
<i>Oryzomys chapmani</i> Thomas, 1898	En	Po	Co	
<i>Oryzomys chapmani caudatus</i> Merriam, 1901	Pr			
<i>Oryzomys chapmani chapmani</i> Thomas, 1898				
<i>Oryzomys chapmani dilutior</i> Merriam, 1901				
<i>Oryzomys chapmani huastecae</i> Dalquest, 1951				
<i>Oryzomys couesi</i> (Alston, 1877)		Po	Co-In	
<i>Oryzomys couesi aquaticus</i> J. A. Allen, 1891				
<i>Oryzomys couesi couesi</i> (Alston, 1877)				
<i>Oryzomys couesi cozumelae</i> Merriam, 1901	A			
<i>Oryzomys couesi peragrus</i> Merriam, 1901				
<i>Oryzomys guerrerensis</i> Goldman, 1915	En	Mo	Co	
<i>Oryzomys melanotis</i> Thomas, 1893	En	Po	Co	
<i>Oryzomys melanotis colimensis</i> Goldman, 1918				
<i>Oryzomys melanotis melanotis</i> Thomas, 1893				
<i>Oryzomys nelsoni</i> Merriam, 1898	E	En	Mo	In
<i>Oryzomys fulgens</i> Thomas, 1893		Po	Co	
<i>Oryzomys fulgens aztecus</i> Merriam, 1901				
<i>Oryzomys fulgens fulgens</i> Thomas, 1893	A			
<i>Oryzomys fulgens lambi</i> Burt, 1934				
<i>Oryzomys fulgens mexicanus</i> J. A. Allen, 1897				

<i>Oryzomys fulgens regillus</i> Goldman, 1915				
<i>Oryzomys fulgens zygomaticus</i> Merriam, 1901				
<i>Oryzomys peninsulae</i> Thomas, 1897	E	En	Mo	Co
<i>Oryzomys rhabdops</i> Merriam, 1901			Po	Co
<i>Oryzomys rhabdops angusticeps</i> Merriam, 1901				
<i>Oryzomys rostratus</i> Merriam, 1901		En	Po	Co
<i>Oryzomys rostratus carrorum</i> Lawrence, 1947				
<i>Oryzomys rostratus megadon</i> Merriam, 1901				
<i>Oryzomys rostratus rostratus</i> Merriam, 1901				
<i>Oryzomys rostratus yucatanensis</i> Merriam, 1901				
<i>Oryzomys saturator</i> Merriam, 1901			Po	Co
<i>Oryzomys saturator hylocetes</i> Merriam, 1901				
<i>Oryzomys saturator saturator</i> Merriam, 1901				
<i>Oryzomys texensis</i> J. A. Allen, 1894			Mo	Co
TRIBE ICHTHYOMYINI Vorontzov, 1959				
<i>Rheomys</i> Thomas, 1906				
<i>Rheomys mexicanus</i> Goodwin, 1959	Pr	En	Mo	Co
<i>Rheomys thomasi</i> Dickey, 1928			Po	Co
<i>Rheomys thomasi chiapensis</i> Hooper, 1947	Pr			
TRIBE SIGMODONTINI Wagner, 1843				
<i>Sigmodon</i> Say and Ord, 1825				
<i>Sigmodon alleni</i> Bailey, 1902		En	Mo	Co
<i>Sigmodon arizonae</i> Mearns, 1890			Mo	Co
<i>Sigmodon fulviventer</i> J. A. Allen, 1889			Mo	Co
<i>Sigmodon hispidus</i> Say and Ord, 1825			Po	Co
<i>Sigmodon hispidus berlandieri</i> Baird, 1855				
<i>Sigmodon hispidus eremicus</i> Mearns, 1897				
<i>Sigmodon hispidus solus</i> Hall, 1951				
<i>Sigmodon leucotis</i> Bailey, 1902		En	Po	Co
<i>Sigmodon leucotis alticola</i> Bailey, 1902				
<i>Sigmodon leucotis leucotis</i> Bailey, 1902				
<i>Sigmodon mascotensis</i> J. A. Allen, 1897		En	Mo	Co
<i>Sigmodon ochrognathus</i> Bailey, 1902			Mo	Co
<i>Sigmodon planifrons</i> Nelson and Goldman, 1933		En	Mo	Co
<i>Sigmodon toltecus</i> (de Saussure, 1860)		En	Mo	Co
<i>Sigmodon zanjonensis</i> Goodwin, 1932			Mo	Co
SUBFAMILY TYLOMYINAE Reig, 1984				
TRIBE NYCTOMYINI Musser and Carleton, 2005				
<i>Nyctomys</i> de Saussure, 1860				

<i>Nyctomys sumichrasti</i> (de Saussure, 1860)			Po	Co
<i>Nyctomys sumichrasti colimensis</i> Laurie, 1953				
<i>Nyctomys sumichrasti pallidulus</i> Goldman, 1937				
<i>Nyctomys sumichrasti salvinii</i> (Tomes, 1862)				
<i>Nyctomys sumichrasti sumichrasti</i> (de Saussure, 1860)				
<i>Otonyctomys</i> Anthony, 1932				
<i>Otonyctomys hatti</i> Anthony, 1932	A		Mo	Co
TRIBE TYLOMYINI Reig, 1984				
<i>Ototylomys</i> Merriam, 1901				
<i>Ototylomys phyllotis</i> Merriam, 1901			Po	Co
<i>Ototylomys phyllotis connectens</i> Sanborn, 1935				
<i>Ototylomys phyllotis phyllotis</i> Merriam, 1901				
<i>Tylomys</i> Peters, 1866				
<i>Tylomys bullaris</i> Merriam, 1901	A	En	Mo	Co
<i>Tylomys nudicaudus</i> (Peters, 1866)			Po	Co
<i>Tylomys nudicaudus gymnurus</i> Villa, 1941				
<i>Tylomys nudicaudus microdon</i> Goodwin, 1955				
<i>Tylomys nudicaudus nudicaudus</i> (Peters, 1866)				
<i>Tylomys nudicaudus villai</i> Schaldach, 1966				
<i>Tylomys tumbalensis</i> Merriam, 1901	Pr	En	Mo	Co
ORDER CARNIVORA Bowdich, 1821				
SUBORDER FELIFORMIA Kretzoi, 1945				
FAMILY FELIDAE G. Fischer, 1817				
SUBFAMILY FELINAE G. Fischer, 1817				
<i>Herpailurus</i> Severtzov, 1858				
<i>Herpailurus yagouaroundi</i> (É. Geoffroy Saint-Hilaire, 1803)	A		Po	Co
<i>Herpailurus yagouaroundi cacomitli</i> (Berlandier, 1859)				
<i>Herpailurus yagouaroundi fossata</i> (Mearns, 1901)				
<i>Herpailurus yagouaroundi tolteca</i> (Thomas, 1898)				
<i>Leopardus</i> Gray, 1842				
<i>Leopardus pardalis</i> (Linnaeus, 1758)	P		Po	Co
<i>Leopardus pardalis albescens</i> (Pucheran, 1855)				
<i>Leopardus pardalis nelsoni</i> (Goldman, 1925)				
<i>Leopardus pardalis pardalis</i> (Linnaeus, 1758)				
<i>Leopardus pardalis sonoriensis</i> (Goldman, 1925)				
<i>Leopardus wiedii</i> (Schinz, 1821)	P		Po	Co
<i>Leopardus wiedii glauculus</i> (Thomas, 1903)				
<i>Leopardus wiedii oaxacensis</i> (Nelson and Goldman, 1931)				
<i>Leopardus wiedii salvinia</i> Pocock, 1941				
<i>Leopardus wiedii yucatanicus</i> (Nelson and Goldman, 1931)				

Lynx Kerr, 1792

***Lynx rufus* (Schreber, 1777)**

Po Co

Lynx rufus baileyi Merriam, 1890

Lynx rufus californicus Mearns, 1897

Lynx rufus escuinapae J. A. Allen, 1903

Lynx rufus oaxacensis Goodwin, 1963

Lynx rufus peninsularis Thomas, 1898

Lynx rufus texensis J. A. Allen, 1895

Puma Jardine, 1834

***Puma concolor* (Linnaeus, 1771)**

Po Co

Puma concolor cougar (Kerr, 1792)

SUBFAMILY PANTHERINAE Pocock, 1917

Panthera Oken, 1816

***Panthera onca* (Linnaeus, 1758)**

P Po Co

Panthera onca arizonensis (Goldman, 1932)

Panthera onca centralis (Mearns, 1901)

Panthera onca goldmani (Mearns, 1901)

Panthera onca hernandesii (Gray, 1857)

Panthera onca veraecrucis (Nelson and Goldman, 1933)

SUBORDER CANIFORMIA Kretzoi, 1938

INFRAORDER CYNIDEA Flower, 1869

FAMILY CANIDAE G. Fischer, 1817

SUBFAMILY CANINAE G. Fischer, 1817

TRIBE CANINI G. Fischer, 1817

Canis Linnaeus, 1758

***Canis latrans* Say, 1822**

Po Co-In

Canis latrans cagottis (Hamilton Smith, 1839)

Canis latrans clepticus Elliot, 1903

Canis latrans goldmani Merriam, 1904

Canis latrans impavidus J. A. Allen, 1903

Canis latrans jamesi Townsend, 1912

Canis latrans mearnsi Merriam, 1897

Canis latrans microdon Merriam, 1897

Canis latrans peninsulae Merriam, 1897

Canis latrans texensis Bailey, 1905

Canis latrans vigilis Merriam, 1897

***Canis lupus* Linnaeus, 1758**

Po Co

Canis lupus baileyi Nelson and Goldman, 1929

E

TRIBE VULPINI Hemprich and Ehrenberg, 1832

Urocyon Baird, 1857

***Urocyon cinereoargenteus* (Schreber, 1775)**

Po Co-In

Urocyon cinereoargenteus californicus Mearns, 1897

Urocyon cinereoargenteus fraterculus Elliot, 1896

Urocyon cinereoargenteus guatemalae Miller, 1899

Urocyon cinereoargenteus madrensis Burt and Hooper, 1941

Urocyon cinereoargenteus nigrirostris (Lichtenstein, 1830)

Urocyon cinereoargenteus orinomus Goldman, 1938

Urocyon cinereoargenteus peninsularis Huey, 1928

Urocyon cinereoargenteus scottii Mearns, 1891

Vulpes Frisch, 1775

***Vulpes macrotis* Merriam, 1888**

A Po Co

PARVORDER URSIDA Tedford, 1976

SUPERFAMILY URSOIDEA G. Fischer, 1817

FAMILY URSIDAE G. Fischer, 1817

SUBFAMILY URSINAE G. Fischer, 1817

Ursus Linnaeus, 1758

***Ursus americanus* Pallas, 1780**

Po Co

Ursus americanus eremicus Merriam, 1904

P

Ursus americanus machetes Elliot, 1903

***Ursus arctos* Linnaeus, 1758**

Po Co

Ursus arctos horribilis Ord, 1815

E

PARVORDER MUSTELIDA Tedford, 1976

FAMILY MEPHITIDAE Dragoo and Honeycutt, 1997

Conepatus Gray, 1837

***Conepatus leuconotus* (Lichtenstein, 1832)**

Po Co

Conepatus leuconotus leuconotus (Lichtenstein, 1832)

***Conepatus semistriatus* (Boddaert, 1785)**

Po Co

Conepatus semistriatus amazonicus (Lichtenstein, 1838)

Pr

Conepatus semistriatus yucatanicus Goldman, 1943

Mephitis È. Geoffroy Saint-Hilaire and G. Cuvier, 1795

***Mephitis macroura* Lichtenstein, 1832**

Po Co

Mephitis macroura eximius Hall and Dalquest, 1950

Mephitis macroura macroura Lichtenstein, 1832

Mephitis macroura milleri Mearns, 1897

***Mephitis mephitis* (Schreber, 1776)**

Po Co

Mephitis mephitis estor Merriam, 1890

Mephitis mephitis holzneri Mearns, 1897

Mephitis mephitis varians Gray, 1837

Spilogale Gray, 1865

<i>Spilogale angustifrons</i> A. H. Howell, 1902			Po	Co
<i>Spilogale angustifrons angustifrons</i> A. H. Howell, 1902				
<i>Spilogale angustifrons elata</i> A. H. Howell, 1906				
<i>Spilogale angustifrons tropicalis</i> A. H. Howell, 1902				
<i>Spilogale angustifrons yucatanensis</i> Burt, 1938				
<i>Spilogale gracilis</i> Merriam, 1890			Po	Co
<i>Spilogale gracilis leucoparia</i> Merriam, 1890				
<i>Spilogale gracilis lucasana</i> Merriam, 1890				
<i>Spilogale gracilis martirensis</i> Elliot, 1903				
<i>Spilogale putorius</i> (Linnaeus, 1758)			Po	Co
<i>Spilogale putorius interrupta</i> (Rafinesque, 1820)				
<i>Spilogale pygmaea</i> Thomas, 1898	A	En	Po	Co
<i>Spilogale pygmaea australis</i> Hall, 1938				
<i>Spilogale pygmaea intermedia</i> López-Forment and Urbano-V., 1981				
<i>Spilogale pygmaea pygmaea</i> Thomas, 1898				
FAMILY MUSTELIDAE G. Fischer, 1817				
SUBFAMILY LUTRINAE Bonaparte, 1838				
TRIBE LUTRINI Bonaparte, 1838				
<i>Lontra</i> Gray, 1843				
<i>Lontra longicaudis</i> (Olfers, 1818)			Po	Co
<i>Lontra longicaudis annectens</i> (Major, 1897)	A			
TRIBE ENHYDRINI Gray, 1825				
<i>Enhydra</i> Flemming, 1822				
<i>Enhydra lutris</i> (Linnaeus, 1758)	P		Po	Co
<i>Enhydra lutris nereis</i> (Merriam, 1904)	P			
SUBFAMILY MUSTELINAE G. Fischer, 1817				
<i>Eira</i> Hamilton Smith, 1842				
<i>Eira barbara</i> (Linnaeus, 1758)	P		Po	Co
<i>Eira barbara senex</i> (Thomas, 1900)				
<i>Galictis</i> Bell, 1826				
<i>Galictis vittata</i> (Schreber, 1776)	A		Po	Co
<i>Galictis vittata canaster</i> Nelson, 1901				
<i>Mustela</i> Linnaeus, 1758				
<i>Mustela frenata</i> Lichtenstein, 1831			Po	Co
<i>Mustela frenata frenata</i> Lichtenstein, 1831				
<i>Mustela frenata goldmani</i> (Merriam, 1896)				
<i>Mustela frenata latirostra</i> Hall, 1936				
<i>Mustela frenata leucoparia</i> (Merriam, 1896)				

- Mustela frenata macrophonus* (Elliot, 1905)
Mustela frenata neomexicana (Barber and Cockerell, 1898)
Mustela frenata perda (Merriam, 1902)
Mustela frenata perotae Hall, 1936
Mustela frenata tropicalis (Merriam, 1896)
- Taxidea* Waterhouse, 1839
- Taxidea taxus* (Schreber, 1778)** A Po Co
Taxidea taxus berlandieri Baird, 1857
- FAMILY PROCYONIDAE Gray, 1825
 SUBFAMILY BASSARISCINAE Gray, 1869
- Bassariscus* Coues, 1887
- Bassariscus astutus* (Lichtenstein, 1830)** Po Co-In
Bassariscus astutus astutus (Lichtenstein, 1830)
Bassariscus astutus bolei Goldman, 1945
Bassariscus astutus consitus Nelson and Goldman, 1932
Bassariscus astutus flavus Rhoads, 1894
Bassariscus astutus insulicola Nelson and Goldman, 1909 A
Bassariscus astutus macdougalli Goodwin, 1956
Bassariscus astutus palmarius Nelson and Goldman, 1909
Bassariscus astutus saxicola Merriam, 1897 A
- Bassariscus sumichrasti* (de Saussure, 1860)** Pr Po Co
Bassariscus sumichrasti latrans (Davis and Lukens, 1958)
Bassariscus sumichrasti oaxacensis (Goodwin, 1956)
Bassariscus sumichrasti sumichrasti (de Saussure, 1860)
Bassariscus sumichrasti variabilis (Peters, 1874)
- Potos* È. Geoffroy Saint-Hilaire and G. Cuvier, 1795
- Potos flavus* (Schreber, 1774)** Pr Po Co
Potos flavus chiriquensis J. A. Allen, 1904
Potos flavus nocturnus (Wied-Neuwied, 1826)
- SUBFAMILY PROCYONINAE Gray, 1825
- Nasua* Storr, 1780
- Nasua narica* (Linnaeus, 1766)** Po Co-In
Nasua narica molaris Merriam, 1902
Nasua narica narica (Linnaeus, 1766)
Nasua narica nelsoni Merriam, 1901 A
Nasua narica yucatanica J. A. Allen, 1904
- Procyon* Storr, 1780
- Procyon lotor* (Linnaeus, 1758)** Po Co-In
Procyon lotor fuscipes Mearns, 1914
Procyon lotor grinnelli Nelson and Goldman, 1930

<i>Procyon lotor hernandezii</i> J. A. Wagler, 1831				
<i>Procyon lotor insularis</i> Merriam, 1898	P			
<i>Procyon lotor pallidus</i> Merriam, 1900				
<i>Procyon lotor psora</i> Gray, 1842				
<i>Procyon pygmaeus</i> Merriam, 1901	P	En	Mo	In
ORDER ARTIODACTYLA Owen, 1848				
SUBORDER SUIFORMES Jaeckel, 1911				
FAMILY TAYASSUIDAE Palmer, 1897a				
SUBFAMILY TAYASSUINAE Palmer, 1897				
<i>Dicotyles</i> G. Cuvier, 1817				
<i>Dicotyles angulatus</i> Cope, 1889			Po	Co–In
<i>Dicotyles angulatus angulatus</i> Cope, 1889				
<i>Dicotyles angulatus humeralis</i> (Merriam, 1901)				
<i>Dicotyles angulatus sonoriensis</i> Mearns, 1897				
<i>Dicotyles crassus</i> (Merriam, 1901)			Po	Co–In
<i>Dicotyles crassus crassus</i> (Merriam, 1901)				
<i>Dicotyles crassus nanus</i> (Merriam, 1901)				
<i>Dicotyles crassus nelsoni</i> (Goldman, 1926)				
<i>Dicotyles crassus yucatanensis</i> (Merriam, 1901)				
<i>Tayassu</i> G. Fischer, 1814				
<i>Tayassu pecari</i> (Link, 1795)			Po	Co
<i>Tayassu pecari ringens</i> Merriam, 1901	P			
SUBORDER RUMIANTIA Scopoli, 1777				
SUPERFAMILY CERVOIDEA Goldfuss, 1820				
FAMILY ANTILOCAPRIDAE Gray, 1866				
SUBFAMILY ANTILOCAPRINAE Gray, 1866				
<i>Antilocapra</i> Ord, 1818				
<i>Antilocapra americana</i> (Ord, 1815)	P		Po	Co
<i>Antilocapra americana mexicana</i> Merriam, 1901				
<i>Antilocapra americana peninsularis</i> Nelson, 1912				
<i>Antilocapra americana sonoriensis</i> Goldman, 1945				
FAMILY CERVIDAE Goldfuss, 1820				
SUBFAMILY CAPREOLINAE Brookes, 1828				
TRIBE ODOCOILEINI Pocock, 1923				
<i>Mazama</i> Rafinesque, 1817				
<i>Mazama pandora</i> Merriam, 1901		En	Mo	Co
<i>Mazama temama</i> (Kerr, 1792)			Po	Co
<i>Mazama temama temama</i> (Kerr, 1792)				
<i>Odocoileus</i> Rafinesque, 1832				

Odocoileus hemionus* (Rafinesque, 1817)Odocoileus hemionus cerrosensis* Merriam, 1898

P

Po

Co-In

Odocoileus hemionus eremicus (Mearns, 1897)*Odocoileus hemionus fuliginatus* Cowan, 1933*Odocoileus hemionus peninsulae* (Lydekker, 1898)*Odocoileus hemionus sheldoni* Goldman, 1939

A

Odocoileus virginianus* (Zimmermann, 1780)Odocoileus virginianus acapulcensis* (Caton, 1877)*Odocoileus virginianus carminis* Goldman and Kellogg, 1940*Odocoileus virginianus couesi* (Coues and Yarrow, 1875)*Odocoileus virginianus mexicanus* (Gmelin, 1788)*Odocoileus virginianus miquihuanensis* Goldman and Kellogg,
1940*Odocoileus virginianus nelsoni* Merriam, 1898*Odocoileus virginianus oaxacensis* Goldman and Kellogg,
1940*Odocoileus virginianus sinaloae* J. A. Allen, 1903*Odocoileus virginianus texanus* (Mearns, 1898)*Odocoileus virginianus thomasi* Merriam, 1898*Odocoileus virginianus toltecus* (de Saussure, 1860)*Odocoileus virginianus veraecrucis* Goldman and Kellogg,
1940*Odocoileus virginianus yucatanensis* (Hays, 1874)

Po

Co-In

SUPERFAMILY BOVOIDEA Gray, 1821

FAMILY BOVIDAE Gray, 1821

SUBFAMILY CAPRINAE Gray, 1821

TRIBE OVINI Grubb, 2001

SUBTRIBE OVINA Grubb, 2001

Ovis Linnaeus, 1758***Ovis canadensis* Shaw, 1804**

Pr

Po

Co

Ovis canadensis cremnobates Elliot, 1904*Ovis canadensis mexicana* Merriam, 1901*Ovis canadensis weemsi* Goldman, 1937

FAMILY BOVIDAE Gray, 1821

SUBFAMILY BOVINAE Gray, 1821

TRIBE BOVINI Simpson, 1945

SUBTRIBE BOVINA Sokolov, 1953

Bison Hamilton Smith, 1827***Bison bison* (Linnaeus, 1758)**

P

Mo

Co

ORDER PERISSODACTYLA Owen, 1848
 SUBORDER CERATOMORPHA Wood, 1937
 INFRAORDER TAPIROMORPHA Haeckel, 1866
 SUPERFAMILY TAPIROIDEA Gray, 1825
 FAMILY TAPIRIDAE Gray, 1821
Tapirella Palmer, 1903
Tapirella bairdii (Gill, 1865)

P Mo Co

TAXONOMIC CHANGES

ORDER DIDELPHIMORPHIA

Gardner (2005a) included *Philander opossum pallidus* as a synonym of *P. o. fuscogriseus*, which is now the only subspecies of *P. opossum* known to occur in Mexico.

Rossi et al. (2010) compared the holotypes of *Marmosa mayensis* and *M. mexicana*, and concluded that they were conspecific. On the other hand, Gutiérrez et al. (2010), based on a molecular analysis, showed that specimens identified as *M. mexicana* fell into two clearly defined haplogroups, which were not clearly assignable as sister taxa. They identified one as *mexicana* clade A (*mayensis*) and the other as *mexicana* clade B (*mexicana*). Although Gutiérrez et al. (2010) mentioned the need for further study before deciding on which names should be applied, we consider the differences they found to warrant identifying the clades as representing the species *M. mayensis* and *M. mexicana*. *Marmosa mayensis* is monotypic, while *M. mexicana* is represented by at least two subspecies, the nominate subspecies and *M. m. savannarum* known from Panamá (Rossi et al. 2010). Gutiérrez recently affirmed his opinion (pers. comm. to ALG) that *mayensis* and *mexicana* represented separate species.

Gardner (2005a) recognized two subspecies of *Tlacuatzin canescens*: *T. c. canescens* and *T. c. gaumeri*. Based on the morphological differences and geographic isolation, we also recognize the Islas Marias population, *T. c. insularis*, as a valid subspecies thus raising the number of recognized subspecies to three.

Gardner and Dagosto (2008) recognized five subspecies in *Metachirus nudicaudatus*. The name

previously applied by Ramírez–Pulido et al. (2005) to the subspecies occurring in Mexico was *M. n. dentaneus*; however, Gardner and Dagosto (2008) treated *M. n. dentaneus* as a synonym of *M. n. colombianus*, which is the name we use here for the population found in Mexico.

ORDERS CINGULATA AND PILOSA

Gardner (2005b) synonymized *Dasyopus novemcinctus davisii* within *D. n. mexicanus*. In the same report, Gardner listed *Tamandua mexicana hesperia* as a synonym of *T. m. mexicana*. Therefore, we do not recognize *D. n. davisii* and *T. m. hesperia* as valid subspecies.

ORDER SORICOMORPHA

Ohdachi's et al. (2006) phylogenetic assessment of the Soricidae was based on cytochrome-*b* sequences. Their results concern several Mexican soricids: 1) *Notiosorex* and *Megasorex* are recognized as separate genera, as suggested in previous studies, 2) populations from the Baja California Peninsula previously identified as *Notiosorex crawfordi* differ sufficiently from *N. cockrumi* and true *N. crawfordi* to be recognized as a separate species (as yet undescribed); and 3) the Soricinae is subdivided into four tribes, three of which occur in Mexico: Soricini (*Sorex*), Blarinini (*Cryptotis*), and Notiosoricini (*Notiosorex* and *Megasorex*). Carraway's (2007) assessment of Mexican soricids supports the findings reported by Baker et al. (2003), Hutterer (2005a), and Ohdachi et al. (2006).

Carraway's (2007) monograph on Mexican shrews included a number of changes in taxonomic

arrangement, plus descriptions of new species and subspecies, as follows: 1) on the basis of differences in ventral coloration, she recognized *Cryptotis goldmani goldmani* and *C. g. machetes* (the latter was formerly treated as a synonym of *C. goldmani*); 2) in accord with Hutterer (2005a), *Cryptotis goodwini* was treated as monotypic; 3) *Cryptotis parvus tropicalis* was raised to specific level as *C. tropicalis*; and 4) specimens from Guerrero and Oaxaca, previously identified as either *S. saussurei* or *S. veraepacis*, were described as a new species, *Sorex ixtlanensis*. Carraway (2007) also revised the species complex known as *Sorex saussurei* (*sensu lato*), which she divided among three species: 1) *S. saussurei* treated as monotypic and restricted to the population across central Mexico; 2) populations previously assigned to *S. saussurei* from Jalisco, Guerrero, Michoacán, and the state of Mexico, described as a new species, *Sorex mediopua*; 3) the taxon *veraecrucis* was elevated to specific level, with *S. v. cristobalensis*, *S. v. oaxacae*, and her new taxon *S. v. altoensis* recognized as subspecies. Carraway (2007) restricted her study to populations of species occurring in Mexico, and did not treat related taxa from Guatemala. As discussed by Woodman et al. (2012:214), the Guatemalan representative of the complex of shrews previously known as *S. saussurei* in Chiapas is *Sorex salvini* Merriam, 1897, which antedates *S. veraecrucis* and probably represents the taxon Carraway (2007) recognized as *S. veraecrucis cristobalensis*. For purposes of this list, the Mexican representatives should be known as *S. salvini altoensis*, *S. s. cristobalensis*, *S. s. oaxacae*, and *S. s. veraecrucis*. Carraway (2007) did not assign subspecies to Mexican *Sorex monticola*, a species that is believed to occur from Alaska into Mexico; therefore, we treat the Mexican population as the nominate subspecies, *S. m. monticola*. The spelling *Cryptotis parvus*, instead of the more familiar *C. parva*, follows the ruling in Opinion 2164 (ICZN 2006; also see Gardner 2005c). The spelling *monticola* (e.g., *Sorex monticola*), instead of the recent but incorrect spelling *monticolus*, is explained in Woodman (2012:827).

ORDER ERINACEOMORPHA

Suprageneric taxonomy follows Hutterer (2005b). Yates and Salazar-Bravo (2005) analyzed geographic variation in morphological and morphometric characters for moles of the genus *Scapanus* and recommended

specific status for *Scapanus latimanus anthonyi*, which then is the only species of mole endemic to Mexico.

ORDER CHIROPTERA

Family Emballonuridae.—In the former list (Ramírez-Pulido et al. 2005) *Diclidurus albus* was treated as monotypic. Simmons (2005) listed *D. virgo* as a subspecies of *D. albus*, as did Hood and Gardner (2008) who noted that *virgo* is slightly larger than *albus*, but not so large as to warrant recognition at the specific level. Therefore, we consider *Diclidurus albus virgo* to be the subspecies distributed in Mexico. In accord with Simmons (2005), we list *Saccopteryx bilineata* and *Peropteryx macrotis* as monotypic species.

Family Molossidae.—McDonough et al. (2008) recognized four well-developed clades in the *Eumops glaucinus* complex of which *E. ferox* is the only species occurring in Mexico, Central America, and the Caribbean. *E. glaucinus*, formerly listed for Mexico, is now restricted to South America along the eastern slopes of the Andes. *Eumops nanus*, previously treated as a subspecies of *Eumops bonariensis*, is a monotypic species according to Eger (2008) that occurs in Mexico, Central America, and northern South America.

González-Ruiz et al. (2011) found that specimens of *Molossus* from the Yucatan Peninsula, originally allocated to *Molossus sinaloae*, represented a new species, which they named and described as *Molossus alvarezii*. The polytypic *M. sinaloae* occurs along the Pacific versant of Mexico and southward. *Molossus rufus* is polytypic (Simmons 2005), and *M. r. nigricans* is the subspecies occurring in Mexico.

Family Natalidae.—Tejedor (2005) recognized two species of *Natalus* in Mexico, *N. stramineus mexicanus* and a new species, which he described as *Natalus lanatus*. Later, Tejedor (2006, 2011), in his reviews of the status of *N. stramineus* and of the genus *Natalus* in general, restricted *N. stramineus* to the Lesser Antilles north of the St. Lucia Channel. As a result, the two Mexican species became *N. lanatus* and *N. mexicanus*, with *saturatus* a synonym of *mexicanus*. Recently, López-Wilchis et al. (2012) did a genetic and morphological analysis of a sample of Mexican *Natalus*. Despite considerable ambiguity in

their results, they concluded that *N. mexicanus* was the only species of *Natalus* in Mexico. Until the results of López-Wilchis et al. (2012) are confirmed, we prefer to list *N. mexicanus* and *N. lanatus* as separate species.

Family Phyllostomidae.—Although Simmons (2005), McLellan (1984), and Koopman (1994) treated *Carollia perspicillata* as monotypic, we continue to recognize *C. p. azteca* as the subspecies occurring in Mexico (see McLellan and Koopman 2008). Clearly, *C. perspicillata* warrants revision. Treating this species, which occurs from Mexico into Argentina, as monotypic more likely suggests uncertainty by authors about the validity of the many names that have been applied to regional populations of this taxon over the past 256 years than actual monotypy.

In the 2005 list, *Leuconycteris yerbabuena* was treated as a subspecies of *L. curasoae*, as first proposed by Arita and Humphrey (1988). Here we use *L. yerbabuena* as the valid name for the species found in the southwestern USA, Mexico, and northern Central America (see Simmons and Wetterer 2002; Simmons 2005; Cole and Wilson 2006a, 2006b). *L. curasoae* is restricted to the Netherland Antilles and northern Venezuela, including Isla Margarita.

Escobedo-Morales et al. (2006) recorded *Trinycteris nicefori* from Chiapas, a species previously unknown from Mexico. Several authors (Baker 1979; Baker et al. 1981; Simmons and Voss 1998; Burton and Engstrom 2001; Simmons 2005; Williams and Genoways 2008) have treated *Mimon cozumelae* as a species different from *M. bennettii*. Gregorin et al. (2008), however, based on morphometric and morphological evidence, treated *M. cozumelae* as a subspecies of *M. bennettii* as did Goodwin (1942) and Schaldach (1964). We recognize *M. cozumelae* as a valid species separate from *M. bennettii* based on three considerations: 1) the clear differences in pelage coloration, dental morphology, and configuration of the mesopterygoid fossae (Simmons and Voss 1998; Williams and Genoways 2008); 2) the broad area of sympatry extending from at least northwestern Costa Rica into Colombia (Díaz et al. 1986; Gregorin et al. 2008; Williams and Genoways 2008); and 3) their different karyotypes (Baker 1979; Baker et al. 1981).

Porter et al. (2007), based on a molecular analysis, provisionally accepted the species names in *Micronycteris* as currently understood, and sorted them into four subgenera: *Leuconycteris*, new subgenus (*M. brosetti*); *Micronycteris* (*M. megalotis*, *M. microtis*, *M. matses*, and *M. giovanniae*); *Schizonycteris*, new subgenus (*M. minuta*, *M. schmidtorum*, and *M. sanborni*); and *Xenoctenes* (*M. hirsuta*). They pointed out that their results “did not support monophyly of *M. microtis* as the name is currently applied,” and indicated the probable existence of cryptic species in *M. megalotis*, *M. minuta*, and *M. hirsuta*. This arrangement is followed here.

Williams and Genoways (2008) treated *Chrotopterus auritus* as monotypic, pending a revision of the species. We continue to use the epithet *C. auritus auritus* for the population in Mexico.

Guerrero et al. (2004) considered *Artibeus trimylyus* to be a species separate from *A. jamaicensis*; however, Larsen et al. (2007) and Larsen et al. (2010), among others, have pointed out that the population should be treated as a subspecies of *A. jamaicensis*, a criterion followed here. Ever since Davis (1984) separated *Artibeus intermedius* from *A. lituratus* there has been controversy concerning the *Artibeus lituratus*–*intermedius* complex. There are two main opposing opinions: first, that *intermedius* is a synonym of *lituratus* (Koopman 1993, 1994; Hooper et al. 2008); and second, that *intermedius* is a species separate from *lituratus* (Davis 1984; Owen 1987; Wilson 1991; Guerrero et al. 2003). A third, less considered option is that both represent different subspecies within *A. lituratus* (Marques-Aguiar 1994; Simmons 2005; Redondo et al. 2008; Larsen et al. 2010). Recently, Marchán-Rivadeneira et al. (2012) in a morphometric analysis, found two recognizable ecomorphotypes, one pertaining to *intermedius* and the other to *palmarum*; the main differences being skull size, ecological preferences, and distribution. Recognizing them as separate subspecies of *A. lituratus* raises additional problems, mainly the extensive cranial variability, and the difficulty in distinguishing each within their large geographic area of apparent sympatry. Although neither of these options is a clear cut nor satisfactory resolution of their relationship, treating *intermedius* as a synonym of *A. l. palmarum*, as done by Simmons (2005), is the preferred option at this time. Therefore, we recognize *A. l.*

palmarum as the continental subspecies of *A. lituratus* and *A. l. koopmani* as the island form.

Hoffmann et al. (2003) recognized three groups within *Uroderma bilobatum* based on differences in karyotype and molecular genetics, and stated that future work might reveal each group to represent a separate species. Simmons (2005) treated these three groups as subspecies, an arrangement we follow here. *Uroderma bilotatum bilobatum* applies to the $2n = 42$ nominate population in South America; *U. b. convexum* (*molaris* is a junior synonym) applies to the $2n = 38$ population on the Yucatan peninsula (including northern Chiapas, Belize, and the Petén of Guatemala), northern Guatemala, and east and southward through Honduras, Nicaragua, Costa Rica, Panama, and the Pacific versant of Colombia and Ecuador; and *U. b. davisi* is the $2n = 44$ form in El Salvador, southern Guatemala, and Mexico (southeastern Oaxaca and southern Chiapas).

Velazco and Simmons (2011) analyzed geographic variation within *Vampyroides caraccioli* and concluded that differences between the two recognized subspecies (Gardner 2008c) supported treating each as a separate species. *V. major* is the species in Mexico, Central America, and northwestern South America.

For many years, authors have applied the name *Sturnira ludovici* to specimens from Mexico and Central America, and lately as a taxon different from *S. oporaphilum* from South America. However, Gardner (2008b) concluded that *ludovici* is a subspecies of *S. oporaphilum* restricted to South America. The name in current usage for specimens from Mexico and Central America is *S. hondurensis* (Goodwin 1940), which we list for Mexico with two subspecies as *S. h. hondurensis* and *S. h. occidentalis*.

As recommended by Iudica (2000) and confirmed by Velazco and Patterson (2013), we use the name *Sturnira parvidens* for the little yellow-shouldered bat of Mexico and Central America previously treated as a subspecies of *Sturnira lilium*.

Family Vespertilionidae.—Within *Myotis*, Simmons (2005) recognized *Myotis melanorhinus* as a species separate from *M. ciliolabrum*; in agreement we list *M. melanorhinus* for Mexico. We follow Simmons

(2005) and list *M. carteri* as a subspecies of *Myotis nigricans* as originally described by LaVal (1973). We also include *Myotis velifer brevis*, which was overlooked by Ramírez–Pulido et al. (2005).

The taxon listed for Mexico as *Eptesicus brasiliensis andinus* was determined by Simmons and Voss (1998) to be a valid species, but restricted to South America (also see Simmons 2005; Davis and Gardner 2008). Therefore, we list the Mexican populations as *E. b. brasiliensis* pending reevaluation of the samples previously identified as *E. b. andinus* from Mexico and Central America.

Baeodon has been considered as either a separate genus or as a subgenus within *Rhogeessa*. The previous list (Ramírez–Pulido et al. 2005), treated *Baeodon* as a separate genus based on Hooper and Van Den Bussche (2003). The *Baeodon*–*Rhogeessa* problem was reanalyzed by Baird et al. (2008), who found that *gracilis* is closely related to *alleni*, the type species of *Baeodon*, and both species are well separated from the other *Rhogeessa* species. Baird et al. (2008) discussed two ways to deal with their results: 1) keep both *alleni* and *gracilis* within the genus *Rhogeessa*, and 2) recognize *Baeodon* as a separate genus containing *alleni* and *gracilis*. A third option not mentioned is to retain *Baeodon* (with *alleni* and *gracilis*) as a subgenus of *Rhogeessa*. For the purposes of this list, we place all of the species in *Rhogeessa*.

Baird et al. (2012) described *Rhogeessa bickhami* with type locality “23.6 mi N Huixtla, Chiapas, Mexico.” This is the same type locality as that for *Rhogeessa genowaysi*, originally cited as “Chiapas; 23.6 mi by road (Mex. Hwy 200) northwest of Huixtla” (Baker 1984:178), where the two species are sympatric.

Piaggio and Perkins (2005) analyzed molecular characteristics in the genus *Corynorhinus*. Among their several conclusions, they retained the five subspecies for *C. townsendii* as proposed by Handley (1959), but revised the known distributions for each. As a result, *C. t. pallascens* (included in Ramírez–Pulido et al. 2005) is now restricted to the USA, and the distribution of *C. t. townsendii* has been modified to now include Mexico where it, along with *C. t. australis*, are the two subspecies of *C. townsendii* we list.

ORDER PRIMATES

Groves (2001) reviewed the taxonomic hierarchy in Primates and assigned *Alouatta* to the subfamily Mycetinae. Brandon–Jones and Groves (2002) and Rylands and Mittermeier (2009) substituted the name Alouattinae for Mycetinae, an arrangement we follow here.

Groves (2001, 2005) stated that *Alouatta palliata* was monotypic, but Rylands et al. (2006) treated it as polytypic, with *A. p. mexicana* as the subspecies occurring in Mexico. Here, we follow Rylands and Mittermeier (2009) and include *A. p. mexicana* on our list.

Some authorities use the name *Alouatta villosa* for the black howler monkey, while others use *A. pigra*. Groves (2005), based on the reviews by Smith (1970) and Hall (1981), among others, concluded that the name *Alouatta villosa* was a *nomen dubium* because the holotype is a skull (no skin exists) of a juvenile and, therefore, identification would be difficult if not impossible. Brandon–Jones (2006) later pointed out that the type specimen is easily recognizable; therefore, *A. villosa* is the valid name for the black howler. *A. pigra* is a junior synonym of *A. villosa*, the name we use here.

ORDER LAGOMORPHA

Hoffmann and Smith (2005) made several changes in the taxonomy of the family Leporidae that have affected the names of the species and subspecies known to be distributed in Mexico as follows:

1) Previously, three subspecies were recognized in *Lepus alleni* (e.g., Best and Henry 1993), but Hoffmann and Smith (2005) reduced the number to two by treating *L. a. palitans* as a junior synonym of the nominate form, *L. a. alleni*. We follow their usage; nevertheless, Hoffmann and Smith (2005) did not provide reasons or references to support their decision and we suggest that a taxonomic revision of *L. alleni* is warranted.

2) Best (1996), in his review of *Lepus californicus*, recognized 16 subspecies, which Hoffmann and Smith (2005) reduced to six (*californicus*, *deserticola*, *insularis*, *magdalenae*, *melanotis*, *texianus*). We tenta-

tively accept the arrangement by Hoffmann and Smith (2005), except for the following: 1) they placed *richardsonii* Bachman, 1839, in the synonymy of *californicus* Gray, 1837, but the name more likely is a synonym of *texianus* Waterhouse, 1848, because the type locality is on the Mexican Plateau (see Gardner 1973:30–33); 2) *xanti* Thomas, 1898 should be recognized as a subspecies of *L. californicus* and not as a synonym of *magdalenae* Nelson, 1907; 3) Hoffmann and Smith (2005) listed *L. insularis* Bryant, 1891 as both a subspecies of *L. californicus* (page 196) and as a separate species (page 200); we suggest that treatment of *insularis* as a subspecies of *L. californicus* is probably correct because there is little morphological (Best 1996; Dixon et al. 1983; Hall 1981), karyotypic (Cervantes et al. 2001), and genetic evidence (Ramírez–Silva et al. 2010) that supports differentiating *insularis* at the species level. Our concept of the taxonomy of subspecies of *L. californicus* (synonyms in parentheses) is as follows: *L. c. californicus* (*bennettii* Gray, 1843; *martirensis* Stowell, 1895; *tularensis* Merriam, 1904; *vigilax* Dice, 1926), *L. c. deserticola* (*wallawalla* Merriam, 1904; *depressus* Hall and Whitlow, 1932), *L. c. insularis*, *L. c. magdalenae*, *L. c. melanotis* (*merriami* Mearns, 1896; *altamirae* Nelson, 1904; *curti* Hall, 1951), *L. c. texianus* (*eremicus* J. A. Allen, 1894; *griseus* Mearns, 1896; *asellus* Miller, 1899; *micropus* J. A. Allen, 1903; *festinus* Nelson, 1904; *richardsonii* Bachman, 1839), and *L. c. xanti* (*sheldoni* Burt, 1933).

3) Hoffmann and Smith (2005) reduced the number of subspecies recognized in *Sylvilagus audubonii* to seven (*audubonii*, *arizonae*, *baileyi*, *confinis*, *goldmani*, *minor*, *warreni*) by treating *sanctidiegi* as a junior synonym of *S. a. arizonae*, and *parvulus* as a synonym of *S. a. minor*. We recognize *arizonae*, *confinis*, *goldmani*, and *minor* as the subspecies in Mexico.

4) Hoffmann and Smith (2005) treated *rosaphagus* as a junior synonym of *Sylvilagus bachmani howelli*.

5) Hoffmann and Smith (2005) treated *pacificus* as a junior synonym of *Sylvilagus cunicularius cunicularius* as we have in this list.

6) Hoffmann and Smith (2005) treated *chiapensis* as a junior synonym of *Sylvilagus floridanus aztecus*.

They also placed *russatus* in the synonymy of *S. f. connectens*; however, *S. f. russatus* has priority and is the valid name because it was described by J. A. Allen, 1904 on February 29, whereas the description of *connectens* Nelson, 1904 was not published until May 18.

7) *Sylvilagus graysoni* was treated as a monotypic species.

Ruedas and Bravo-Salazar (2007) split *Sylvilagus brasiliensis* into two species, *S. gabbi* in Mexico and Central America, and *S. brasiliensis* from eastern Panama southward through South America. Although their hypothesis has merit, we continue to use *S. brasiliensis* for the tapeti rabbits of Mexico pending a broader taxonomic analysis of the *brasiliensis* complex.

ORDER RODENTIA

Family Sciuridae.—Based on molecular and morphological analyses of insular populations related to *Ammospermophilus leucurus*, Álvarez-Castañeda (2007) found that differences were insufficient to maintain *A. insularis* as a separate species. Consequently, we list the taxon as *A. l. insularis* as originally described by Nelson and Goldman (1909).

The New World members of the genus *Spermophilus* F. Cuvier, 1825 were reviewed by Helgen et al. (2009), who elevated the seven North American subgenera to full generic status based on the sum of morphological, cytogenetic, ecological, and behavioral characters. The generic names for these ground squirrels are: *Callospermophilus* Merriam, 1897; *Ictidomys* J. A. Allen, 1877; *Notocitellus* A. H. Howell, 1938; *Otospermophilus* Brandt, 1844; *Poliocitellus* A. H. Howell, 1938; *Urocitellus* Obolenskij, 1927; and *Xerospermophilus* Merriam, 1892; with *Spermophilus*, *sensu stricto*, restricted to northern portions of the Palearctic Region. Five of these genera are found in Mexico: *Notocitellus annulatus* and *N. adocetus* in the western portions of the states of Guerrero, Michoacán, Jalisco, Colima, and Nayarit; *Otospermophilus variegatus*, *O. beecheyi*, and *O. atricapillus* have wide distributions north of the Transvolcanic Belt; *Callospermophilus madrensis* is in southern Chihuahua and northern Durango; *Ictidomys mexicanus* and *I. parvidens* (recognized as a species separate from *I. mexicanus*) occurs

in central Mexico; and *Xerospermophilus perotensis*, *X. spilosoma*, and *X. tereticaudus* are on the Mexican Plateau. This arrangement is followed here.

Fernández (2012) based on an analysis of nuclear and mitochondrial DNA, treated *Xerospermophilus perotensis* as an isolated subspecies of *X. spilosoma* found in the Oriental Basin of Puebla and Veracruz. We agree; recognizing *X. s. perotensis* as a subspecies of *X. spilosoma* was suggested earlier by Harrison et al. (2003), Herron et al. (2004), and Helgen et al. (2009).

Thorington and Hoffmann (2005) treated *Sciurus aberti phaeurus* and *Otospermophilus variegatus tiburonensis* as junior synonyms of *S. a. durangi* and *O. v. grammurus*, respectively. We concur with their arrangement.

The results in Jameson (1999) and Piaggio and Spicer (2001) suggest important changes in the taxonomic position of chipmunks assigned to the genus *Tamias*. Those authors proposed treating *Tamias*, *Neotamias*, and *Eutamias* as genera with *Eutamias* restricted to the Old World. Thorington and Hoffmann (2005), however, retained *Tamias* and used *Neotamias* and *Eutamias* as subgenera. Herein, we use *Neotamias* as the generic name for western chipmunks because Jameson's (1999) evidence from ectoparasites and Piaggio and Spicer's (2001) molecular analysis support this usage.

Thorington and Hoffmann (2005) treated *Neotamias durangae solivagus* as a synonym of *N. durangae durangae*; nevertheless, and contrary to their usage, we treat *solivagus* as a species for the following reasons: 1) the morphological analysis by Levenson et al. (1985) showed that *solivagus* is different from, and on a lineage apart from, *durangae*; 2) Piaggio and Spicer (2001), analyzed cytochrome oxidase subunit II and cytochrome *b* for specimens they identified as *bulleri* (= *solivagus*, because the sample came from Coahuila), and showed in all of their cladograms that *durangae* was basal to and clearly separate from their "*N. bulleri solivagus*"; 3) *N. solivagus* differs from *N. durangae* and *N. bulleri*, especially in color pattern and bacular morphology (Callahan 1980; Bartig et al. 1993; Best et al. 1993); and 4) *solivagus* is geographically isolated from both *N. bulleri* and *N. durangae*, and, in

fact, is found in a different biogeographic region. *N. solivagus* is in the Sierra Madre Oriental, and *N. bulleri* and *N. durangae* occur in the Sierra Madre Occidental with the Mexican Plateau, a broad geographic barrier, separating them.

Family Geomyidae.—Hafner et al. (2008) reexamined the *Cratogeomys castanops–goldmani* complex problem by analyzing mitochondrial and nuclear DNA, distribution patterns, and skull form and size. Those authors found that there are two species in the complex, each with two subspecies, as follows (synonyms in parentheses): *C. c. castanops* (*castanops* Baird, 1852; *clarkii* Baird, 1855; *perplanus* Nelson and Goldman, 1934; *lacrimalis* Nelson and Goldman, 1934; *hirtus* Nelson and Goldman, 1934; *angusticeps* Nelson and Goldman, 1934; *tamaulipensis* Nelson and Goldman, 1934; *convexus* Nelson and Goldman, 1934; *bullatus* Russell and Baker, 1955; *ustulatus* Russell and Baker, 1955; *pratensis* Russell, 1968; *simulans* Russell, 1968; *torridus* Russell, 1968; *parviceps* Russell, 1968; *dalquesti* Hollander, 1990); and *C. c. consitus* (*consitus* Nelson and Goldman, 1934; *excelsus* Nelson and Goldman, 1934; *subsimus* Nelson and Goldman, 1934; *jucundus* Russell and Baker, 1955; *sordidulus* Russell and Baker, 1955; *perexiguus* Russell, 1968; *surculus* Russell, 1968). *Cratogeomys goldmani*: *C. g. goldmani* (*goldmani* Merriam, 1895; *rubellus* Nelson and Goldman, 1934); and *C. g. subnubilus* (*subnubilus* Nelson and Goldman, 1934; *planifrons* Nelson and Goldman, 1934; *peridoneus* Nelson and Goldman, 1934; *elibatus* Russell, 1968; *maculatus* Álvarez and Álvarez–Castañeda, 1996). We follow this arrangement.

Hafner et al. (2004), while reviewing gophers of the *Cratogeomys gymnurus* species group from the Trans-Mexican Volcanic Belt, found five allopatric, well-defined clades that do not coincide with the five currently recognized species (*C. fumosus*, *C. gymnurus*, *C. neglectus*, *C. tylorhinus* and *C. zinseri*). They grouped four of the five clades at the subspecific level within *C. fumosus* as follows (synonyms in parentheses): *C. f. angustirostris* (= *C. t. angustirostris*, *C. f. brevirostris* and *C. zinseri*), *C. f. fumosus* (= *C. g. gymnurus*, *C. g. russelli*, *C. g. tellus*, *C. t. atratus* and *C. t. zodioides*), *C. f. imparilis* (*C. g. imparilis*), and *C. f. tylorhinus* (*C. neglectus* and *C. t. tylorhinus*). The

fifth clade was diagnosed based on multiple genetic, morphologic, chromosomal, and characteristic parasite faunas, and pertains to populations previously recognized as *C. tylorhinus planiceps*, which they raised to specific level, *C. planiceps*. Nevertheless, we note that Hafner et al. (2004) overlooked *Cratogeomys gymnurus russelli*, known only from the type locality, which is geographically close to the type localities of *C. fumosus* and *C. gymnurus*. Therefore, we place *C. gymnurus russelli* in the synonymy of *C. f. fumosus*; otherwise we are in agreement with their conclusions.

Hafner et al. (2005) found three well-defined clades, each representing a separate species, in their study of *Cratogeomys merriami*. Based on mitochondria DNA, diploid number, and analyses of quantitative and qualitative aspects of morphology, they treated each of the three species as monotypic as follows (synonyms and previously used names in parentheses): *C. perotensis* (*C. m. estor*, *C. m. irolonis*, *C. m. peraltus* and *C. m. perotensis*), *C. fulvescens* (*C. m. fulvescens*), and *C. merriami* (*C. m. merriami* and *C. m. saccharalis*).

Desmastes et al. (2003), based on genetic and morphometric analyses of *Pappogeomys alcorni*, concluded that *P. alcorni* was a disjunct subspecies of *P. bulleri* rather than a monotypic species (also see Patton 2005). Hafner et al. (2009) examination of the taxonomy of *Pappogeomys bulleri*, based on cytogenetic and chromosomal evidence, concluded that *Pappogeomys bulleri* consisted of five clades, each one a subspecies, thus reducing the number of subspecies from nine to five as follows: *Pappogeomys bulleri albinasus* (*albinasus* Merriam, 1895; *infuscus* Russell, 1968), *P. b. alcorni* (*alcorni* Russell, 1957); *P. b. bulleri* (*bulleri* Thomas, 1892; *nelsoni* Merriam, 1892; *flammeus* Goldman, 1939; *lagunensis* Goldman, 1939; *amecensis* Goldman, 1939; *lutulentus* Russell, 1968), *P. b. burti* (*burti* Goldman, 1939; *melanurus* Genoways and J. K. Jones, 1969); and *P. b. nayaritensis* (*nayaritensis* Goldman, 1939). We follow this arrangement in the list.

Patton (2005) made several changes in his list of gophers of the genus *Thomomys*. Two taxa previously considered as subspecies of either *T. bottae* or *T. umbrinus* were recognized as belonging to the other species as follows: *T. u. perditus* became *T. b. perditus*,

and *T. b. camargensis* listed as *T. u. camargensis*. Other changes were: *T. b. phasma* treated as a synonym of *T. b. pusillus*; *T. b. santidiegi*, a synonym of *T. b. bottae*; *T. u. sheldoni*, a synonym of *T. u. madrensis*; and *T. u. evexus*, a synonym of *T. u. nelsoni*. Our taxonomic list includes these changes.

Mathis et al. (2013a), in their genetic and morphometric assessment of *Thomomys umbrinus* populations from the Sierra Madre Occidental, resurrected *Thomomys sheldoni* from synonymy under *T. umbrinus madrensis* and recognized *T. sheldoni chihuahuae* from the northern Sierra Madre Occidental of Chihuahua and *T. s. sheldoni* (*T. u. crassidens* is a junior synonym) in the southern Sierra Madre Occidental of Durango, Zacatecas, and Nayarit. Mathis et al. (2013b) described a new species, *Thomomys nayarensis*, from the vicinity of Mesa del Nayar, northwestern Nayarit. This information is incorporated in our list.

Hafner et al. (2011), in their study of the *Thomomys umbrinus* complex in western Mexico, found sufficient genetic, morphological, and biogeographic evidence to elevate *Thomomys umbrinus atrovarius* to species level. *Thomomys atrovarius* J. A. Allen, 1898, as currently recognized, consists of *T. a. atrovarius* (*T. umbrinus extimus* Nelson and Goldman, 1934, and *T. u. musculus* Nelson and Goldman, 1934 are junior synonyms) in the coastal lowlands and adjacent lower slopes of the Sierra Madre Occidental in southern Sinaloa, Durango, Nayarit and northwestern Jalisco; and *T. a. parviceps* Nelson and Goldman, 1934 (formerly treated as a subspecies of *T. umbrinus*) in northeastern Sinaloa and western Durango north of the Río Piaxtla.

Álvarez-Castañeda's (2010) taxonomic assessment of the *Thomomys bottae-umbrinus* complex from Mexico and the western USA suggested the existence of eight monophyletic groups, each of which probably should be treated as a separate species. His interpretation of the species in the *T. bottae-umbrinus* complex is as follows: *Thomomys townsendii* east of the Snake River, *T. laticeps* in northern California, *T. bottae* elsewhere in California, north and west of the Colorado River; *T. fulvus* east of the Colorado River and southward into the Sierra Madre Occidental in Sonora, *T. anitae* south from the Salton Sea into the Baja

California Peninsula, *T. atrovarius* along the coastal region of Sinaloa and Nayarit; *T. chihuahuae* in western Chihuahua and Durango, and *T. umbrinus* from the remainder of the distribution in Mexico. At least partly confirming Álvarez-Castañeda (2010) hypothesis, Hafner et al. (2011) recognized *Thomomys atrovarius* as a species (with two subspecies), and Mathis et al. (2013a) recognized *T. chihuahuae* as a subspecies of *T. sheldoni*. The recognition of additional species within the *bottae-umbrinus* complex, as recommended by Álvarez-Castañeda (2010), must be confirmed.

Trujano-Álvarez and Álvarez-Castañeda (2007), and Ríos and Álvarez-Castañeda (2007) reviewed the taxonomy of populations of *Thomomys bottae* from Baja California Sur south of the Vizcaino Desert previously recognized as *T. b. anitae*, *T. b. alticola*, *T. b. imitabilis*, *T. b. incomptus*, *T. b. litoris*, and *T. b. magdalenae*. We find their results compelling and agree that these six named populations are best treated as the single subspecies, *T. b. anitae*.

Family Heteromyidae.—The taxonomic status of *Dipodomys margaritae* as either a species, or as a subspecies of *D. merriami*, has been unsettled for a long time (e.g., Hall 1981; Ramírez-Pulido et al. 2005). Studies by Williams et al. (1993) and Lidicker (1960), and the review by Patton (2005), have treated the taxon as a subspecies of *D. merriami*, an assignment we follow in this list, despite evidence for recognition as a species. Best and Janecek (1992) suggested that *Dipodomys insularis* was best treated as a subspecies of *D. merriami*; a conclusion subsequently followed by both Patton (2005) and Álvarez-Castañeda et al. (2009), and used here.

The assessment of the systematics and biogeography of *Dipodomys phillipsii* by Fernández et al. (2012) showed that the northern population on the Mexican Plateau is markedly different from those from the subtropical deserts of the Mexican Transvolcanic Belt. The differences support recognizing the northern Mexican Plateau population as *D. ornatus*, which is monotypic. The southern populations are represented by *D. phillipsii* and consist of *D. p. phillipsii*, *D. p. perotensis*, and *D. p. oaxacae*.

There has been recent controversy over the taxonomic status of *Liomys* as either a separate genus or as a synonym of *Heteromys*. Although most authors follow Goldman (1911) in treating *Liomys* as distinct from *Heteromys*, Hafner et al. (2007), following up on research by Anderson et al. (2006), confirmed that *Liomys* is paraphyletic relative to *Heteromys* as presently understood. Hafner et al. (2007) evaluated the taxonomic hierarchy within the Heteromyidae and concluded that *Liomys* is a synonym of *Heteromys*. Rogers and González (2010:915), however, retained *Liomys* as a separate genus pending a more inclusive evaluation of heteromyine taxa. A third option not mentioned previously is to recognize the *Liomys salvini* and *L. adspersus* clade as a separate genus for which *Schaeferia*, described by Lehmann and Schaefer (1979), is available. Nevertheless, the evidence presented by Hafner et al. (2007) is sufficiently robust that we follow their lead in treating *Liomys* as a synonym of *Heteromys*.

In the past we listed *Heteromys desmarestianus goldmani* as a subspecies, but recent molecular (Hafner et al. 2007; Rogers and González 2010) and morphological (Espinoza et al. 2011) studies have shown that *goldmani* warrants recognition as the species *H. goldmani*. We recognize *H. temporalis* (usually cited as *Heteromys desmarestianus temporalis*) as a separate species based on differences between the population in Oaxaca and Veracruz and the subspecies recognized in *H. desmarestianus*. We also suggest that *H. temporalis* represents the species cited as “*Heteromys* candidate species B” by Rogers and González (2010:930).

Alexander and Riddle (2005) evaluated phylogenetic relationships within the Heteromyidae, concluding that five of the six genera composing the family were distributed into three clearly monophyletic units, the subfamilies Dipodomysinae and Heteromyinae, and *Chaetodipus*. They found no support for a monotypic Perognathinae (*Chaetodipus* and *Perognathus*), which they explained might be due to the use of mitochondrial genes in the analysis. Hafner et al. (2007), however, who also used mitochondrial DNA in their assessment, concluded that *Chaetodipus* and *Perognathus* were sister taxa comprised within the Perognathinae, which is the classification used by Simpson (1945) and Patton (2005), and used here.

Álvarez–Castañeda and Rios (2011) used pelage characteristics, morphometrics, and genomic DNA in their analysis of the *Chaetodipus arenarius* species complex. Previously, the species group was believed to consist of only two species: *C. dalquesti* (monotypic; see Riddle et al. 2000; Patton 2005; Williams et al. 1993; Patton and Álvarez–Castañeda 1999), and *C. arenarius* with 12 subspecies (*arenarius*, *albescens*, *albulus*, *ambiguus*, *ammophilus*, *helleri*, *mexicalis*, *paralios*, *ramirezpulidoi*, *sabulosus*, *siccus*, *sublucidus*; see Álvarez–Castañeda and Cortés–Calva 2004; Hall 1981; Patton 2005). Three of the taxonomic conclusions reached by Álvarez–Castañeda and Rios (2011) are important for our list: 1) recognizing *C. arenarius*, *C. dalquesti*, and *C. siccus* as phylogenetically distinct species; 2) assigning *C. arenarius ammophilus* and *C. a. sublucidus* to *C. dalquesti*; 3) and advising no changes to the taxonomic position of the other subspecies in *C. arenarius*. We are following the taxonomic conclusions reached by Álvarez–Castañeda and Rios (2011) with two exceptions. First, we recognize *C. arenarius sabulosus* as a valid subspecies of *C. arenarius*; this taxon was not mentioned by Álvarez–Castañeda and Rios (2011) although they must have examined specimens of *sabulosus* based on geography. Second, when the species they identified as *C. dalquesti* (Roth 1976) was reconstituted to also include *ammophilus* Osgood, 1907, and *sublucidus* Nelson and Goldman, 1929, the name for that species must be changed to *Chaetodipus ammophilus* (Osgood 1907), which has priority as the oldest name.

Hafner and Hafner (1983), and Williams et al. (1993) suggested that *Chaetodipus anthonyi*, found only on Isla Cedros, was an isolated subspecies of *C. fallax*. Based on additional evidence presented by Ríos and Álvarez–Castañeda (2010), we list the taxon as a subspecies of *C. fallax*. We continue to treat *Chaetodipus lineatus* as a monotypic species as treated by Hall (1981), Jones et al. (1986), Williams et al. (1993), and Patton (2005).

Andersen and Light (2012) reexamined the taxonomic status of subspecies of *Chaetodipus hispidus* based on genetic and morphological analyses. Their results, which greatly altered the previously derived distributional pattern of the subspecies, showed that there are three, instead of four subspecies (*C. h. zacate-*

cae treated as a synonym of *C. h. hispidus*). We list two subspecies (*C. h. conditi* and *C. h. hispidus*) for Mexico. The distribution map in Andersen and Light (2012:1204) includes northern-most Coahuila within the distribution of *C. h. paradoxus*, which otherwise is entirely within the USA, but we are not aware of any locality records of specimens from that area assignable to *paradoxus*.

Lee et al. (1996), based on an analysis of mitochondrial DNA, treated *Chaetodipus penicillatus eremicus* as a species, *C. eremicus*, with two subspecies: the nominate form and *C. e. atrodorsalis*, which also was formerly considered to be a subspecies of *C. penicillatus* (see Patton 2005). We follow this arrangement.

McKnight (2005) analyzed mitochondrial cytochrome-*b* in his review of the taxonomy of the *Perognathus longimembris* species group, which consists of *P. longimembris*, *P. amplus*, and *P. inornatus*. The first two species occur in Mexico. McKnight (2005) suggested that *P. longimembris* may ultimately prove to be two species, but did not propose any change in its nomenclature. Therefore, we follow Williams et al. (1993) and Patton (2005) by continuing to recognize *P. l. aestivus*, *P. l. bombycinus*, *P. l. internationalis*, *P. l. kinoensis* and *P. l. venustus* as subspecies occurring in Mexico. McKnight (2005) treated *P. amplus* as consisting of three subspecies of which *P. a. amplus* and *P. a. taylori* occur in Mexico.

Family Cricetidae.—Hall (1981) recognized *Ondatra zibethicus bernardi* as the muskrat found in the lower Colorado River drainage; however, we use *Ondatra zibethicus pallidus* for that subspecies, based on Hoffmeister (1986).

Patton et al. (2008) did an extensive and detailed analysis of the *Neotoma lepida* complex resulting in several major revisory changes as follow: 1) *Neotoma devia* treated as monotypic with the former subspecies *N. d. aureotunicata* and *N. d. bensoni* now synonyms of *N. devia*; 2) *N. anthonyi* and *N. martinensis* treated as Mexican subspecies of *N. bryanti*; 3) *N. bunkerii* now a synonym of *N. b. bryanti*; 4) *N. lepida lepida* is the subspecies of *N. lepida* in Mexico, while most of the subspecies previously assigned to *N. lepida* are now either subspecies or synonyms of *N. bryanti*; 5) there are five subspecies of *N. bryanti* in Mexico (*N. b.*

anthonyi, *N. b. bryanti*, *N. b. intermedia*, *N. b. marcosensis*, and *N. b. martinensis*); 6) 14 names previously listed as subspecies of *N. lepida* (*abbreviata*, *arenacea*, *aridicola*, *felipensis*, *latirostra*, *molagrandis*, *notia*, *nudicauda*, *perpallida*, *pretiosa*, *ravida*, and *vicina*) are now synonyms of *N. bryanti bryanti*, and two additional names previously listed as subspecies of *N. lepida* (*egressa* and *gilva*) are now considered to be synonyms of *N. bryanti intermedia*; and 6) *N. lepida insularis* is now the monotypic species *N. insularis*.

In accord with Bogan (1997), Musser and Carleton (2005), and Álvarez-Castañeda and Rios (2010), we list *Neotoma varia* as a subspecies of *N. albigula*. *Neotoma albigula sheldoni*, included in Ramírez-Pulido et al. (2005), is a junior synonym of *N. a. mearnsi* and not listed here.

Romo-Vázquez et al. (2005) described a new species of *Habromys* from Guerrero and the southern state of Mexico, naming it *Habromys schmidlyi*. Bradley et al. (2007) brought needed attention to the many anomalies and taxonomic problems in the currently accepted taxonomic arrangement of species and species groups in *Peromyscus*. They discussed two possible arrangements: 1) integrate *Habromys*, *Megadontomys*, *Neotomodon*, *Osgoodomys*, and *Podomys* as a clade within *Peromyscus*, thus returning the genus to near its earlier structure *sensu lato*; 2) or maintain *Habromys*, *Megadontomys*, *Neotomodon*, *Osgoodomys* and *Podomys* as separate genera and recognize several species and species groups now within *Peromyscus*, as additional genera. Miller and Engstrom (2008) introduced another phylogenetic hypothesis for peromyscine rodents based on a broader array of genetic characters and reached conclusions similar to those of Bradley et al. (2007). As currently constituted, *Peromyscus* is polyphyletic as demonstrated by several studies (e.g., Bradley et al. 2007; Miller and Engstrom 2008; Rogers et al. 2005). Whatever the definitive treatment of *Peromyscus*, the *leucopus*–*maniculatus* species group must remain in *Peromyscus* because *leucopus* is the type species. Because of the current lack of consensus on peromyscine taxonomy, our list of taxa differs little, except in sequence, from that in Ramírez-Pulido et al. (2005).

Following Bradley et al. (2000), we list *P. beatae* with two subspecies (*P. b. beatae* and *P. b. sacarensis*). The first is distributed in the Sierra Madre Oriental

within the states of Hidalgo, Puebla and Veracruz, as well as in the Sierra Madre del Sur in Guerrero and Oaxaca. The second occurs in the Sierra Madre del Sur from Chiapas into El Salvador and Honduras. Musser and Carleton (2005) listed *P. beatae* as monotypic.

Bradley et al. (2014) found that specimens of the *Peromyscus boylii* species group from Nayarit, originally allocated to *Peromyscus levipes*, represented a new species, which they described and named as *Peromyscus carletoni*. According to Bradley et al. (2004, 2014) and Tiemann-Boege et al. (2000), several taxa in the *Peromyscus boylii* species group in western Mexico have yet to be described.

Musser and Carleton (2005) treated *P. guatemalensis*, *P. sagax* and *P. hyllocetes* as monotypic based on, respectively, Huckaby (1960), Bradley et al. (1996), and Sullivan et al. (1997). We treat *P. aztecus* and *P. levipes* as polytypic, but the taxonomy of higher elevation populations of *P. aztecus* referred to as *P. a. evides* and *P. a. oaxacensis* is not clear.

Bradley et al. (2004) described *Peromyscus schmidlyi* as a new species found in Durango and eastern Sinaloa. Cabrera et al. (2007) and López-González et al. (2013) recently have recorded *P. schmidlyi* in Chihuahua, Jalisco, Sonora, and Zacatecas.

Ávila-Valle et al. (2012) studied genetic variation in *Peromyscus furvus* and found a clear division between populations in San Luís Potosí and Querétaro, and those populations from farther south in Hidalgo, Puebla and Veracruz. The San Luís Potosí and Querétaro populations now bear the name *Peromyscus latirostris*, and *P. furvus* is restricted to the southern populations. Recognizing that *P. furvus* was a composite of two species was mentioned earlier by Harris and Rogers (1999), Harris et al. (2000), and Rogers and Skoy (2011).

Musser and Carleton (2005) did not indicate the status of *Peromyscus mexicanus saxatilis*, but Musser later confirmed its validity as a subspecies (personal communication to JRP).

Arellano et al. (2005), in agreement with Hooper's (1952) monograph on *Reithrodontomys*,

found clear separation between the two subgenera (*Reithrodontomys* and *Aporodon*). An analysis of cytochrome-*b*, however, indicated problems within and between named species. They advocated additional study covering more taxa and larger sample sizes. We follow the taxonomy of Hooper (1952), and have added the recently described species.

Voss et al. (2002), upon finding that two allopatric species from Colombia (*Aepeomys fuscatus* and *Oryzomys intectus*) shared a series of unique external and cranial characters, described *Handleyomys* as a new genus for them with *Aepeomys fuscatus* as type species. Later, Weksler et al. (2006) separated certain species, previously included in *Oryzomys*, into ten new genera (*Aegialomys*, *Cerradomys*, *Eremoryzomys*, *Euryoryzomys*, *Hylaeamys*, *Mindomys*, *Nephelomys*, *Oreoryzomys*, *Sooretamys* and *Transandinomys*), all of them morphologically different and ecogeographically distinct, none of which is found in Mexico. Weksler et al. (2006:2) provisionally transferred species included in the *Oryzomys alfaroi* species group (*O. alfaroi*, *O. chapmani*, *O. melanotis*, *O. rhabdops*, *O. rostratus*, and *O. saturator*) to *Handleyomys*. There are ongoing studies to resolve the taxonomy of the *alfaroi* group and, although we acknowledge that the *O. alfaroi* species group is composite, we continue to use *Oryzomys* for these species because a definitive analysis of their generic affinity has yet to be published.

Carleton and Arroyo-Cabrales (2009), in their review of the *Oryzomys couesi* complex from western Mexico, elevated *O. c. albiventer* and *O. c. peninsularae* to species level and reduced *O. c. lambi* and *O. c. bulleri* to the synonymy of *O. c. mexicanus*. Carleton and Arroyo-Cabrales (2009) expressed doubt about the validity of the subspecies *O. c. crinitus*, *O. c. aztecus*, and *O. c. regillus*; nevertheless, we retain the last two names in the list and consider *crinitus* to be a synonym of *O. fulgens* (see below).

In their study of the *Oryzomys palustris* species complex, Hanson et al. (2010) relied on nucleotide sequence data from three independent gene regions and recommended that the complex actually consisted of six species instead of the two species (*O. palustris* and *O. couesi*) as the group had been treated previously, insular forms excepted. Hanson et al. (2010) reached

four general conclusions, paraphrased as follows: 1) they confirmed reciprocal monotypy of *O. palustris* and *O. couesi*; 2) populations previously identified as *O. palustris* (*sensu lato*) consist of two clades (*O. palustris* and *O. texensis*); 3) *O. couesi* (*sensu lato*) also consists of two clades (*O. mexicanus* along the western [Pacific] side of Mexico, Guatemala, and El Salvador; and *O. couesi* along the eastern [Caribbean] side of Mexico, northern Guatemala, Belize, Honduras, and Nicaragua) along with an unassigned, but distinct population in Costa Rica and another in southern Panama. All of the specimens Hanson et al. (2010) identified as *O. texensis* are from north of the USA-Mexican border (also see Davis and Schmidly 1994; Hall 1960), but later Indorf and Gaines (2013) assigned specimens from Matamoros, Tamaulipas to *O. texensis*, thus confirming the species for Mexico. Although the divergence level found by Hanson et al. (2010) was lower between *O. mexicanus* and *O. couesi* than that between *O. palustris* (*sensu lato*) and *O. couesi* (*sensu lato*), it was high enough (4.41 ± 0.49) to support recognizing the western Mexican taxon (see Carleton and Arroyo-Cabrales 2009) as *O. mexicanus*, as we have done in this list (but see the next discussion).

Oryzomys fulgens has been a nomenclatorial problem because the type locality has been in doubt (see discussion in Carleton and Arroyo-Cabrales 2009). Merriam (1901; not Thomas 1893a) restricted the type locality to “in southern Mexico, probably in or near the Valley of Mexico”, which has been repeated in subsequent literature (e.g., Hall 1981). We have two additional pieces of evidence supporting the Valley of Mexico as the restricted type locality: 1) We (NGR and JAC) examined the holotype of *O. fulgens* (BMNH 70.6.20.3) and found it to closely resemble the Pacific versant population morphologically, and 2) according to Thomas (1893b), the holotype of *O. fulgens* was probably collected at the same locality as was the holotype of *Cratogeomys merriami*, whose type locality Merriam (1895) also restricted to Valley of Mexico. In fact, the geographical distribution of *C. merriami* basically is restricted to the Valley of Mexico. Based on this evidence, we have reached the following conclusions. First, we now treat *crinitus* (type locality Tlalpam, Federal District, Mexico) as a junior synonym of *fulgens*. Second, based on the recent taxonomic changes by Hanson et al. (2010) in which

O. mexicanus was restricted to the Pacific versant of Mexico, and *O. couesi* restricted to the eastern zone of Mexico (see above), we conclude that the name for the representative of the *Oryzomys couesi* species group distributed along the Pacific versant of Mexico (except for *O. albiventer*) must bear the name *O. fulgens* Thomas, 1893, which has priority over *O. mexicanus* J. A. Allen, 1897.

Almendra et al. (2014) split *Oryzomys chapmani* into two allopatric species, *O. chapmani* (*sensu stricto*) from the Sierra Madre Oriental and northern Oaxaca highlands, and *O. guerrerensis* on the Sierra Madre del Sur highlands from central Guerrero to Oaxaca. We recognize *O. guerrerensis* as a monotypic species and *O. chapmani* as polytypic and containing the subspecies *caudatus*, *dilutior*, and *huastecae*.

Carroll et al. (2005), building on previous work (Carroll and Bradley 2005; Peppers et al. 2002), mapped *Sigmodon toltecus* as occurring along the Gulf Coast versant of Mexico and the Yucatan Peninsula including Belize and northern Guatemala, a restriction that would include the following subspecies previously placed under *S. hispidus*: *S. hispidus furvus*, *S. h. microdon*, *S. h. saturatus*, *S. h. solus* and *S. h. toltecus*. Here, we are treating *S. toltecus* as monotypic because we have no evidence at hand that indicates which of the named subspecies taxa are actually valid and which should be reduced to synonymy.

Carroll et al. (2005), Bradley et al. (2008), and Henson and Bradley (2009), identified the cotton rats occurring in Chiapas and southeastern Oaxaca as *Sigmodon hirsutus*. However, we think that *S. hirsutus* (*sensu* Carroll et al. 2005) consists of two species, the South American *S. hirsutus* (originally *Lasiomys hirsutus* described by Burmeister [1854]; see synonymy in Voss 1992:26) and the Mexican and Central American *S. zanjonensis* Goodwin 1932 (*S. hispidus villae* Goodwin 1958 is a synonym). Musser and Carleton (2005) recognized both *S. planifrons* and *S. zanjonensis* as species. We follow this arrangement.

In this list, we recognize three subspecies of *Sigmodon hispidus* in Mexico: *S. h. berlandieri*, *S. h. eremicus* and *S. h. solus*. The status of *S. h. berlandieri* and *S. h. solus* is unresolved and requires additional

study. Carroll et al. (2005) did not clarify the status of named population and relationships much beyond the information in Peppers and Bradley (2000) and Peppers et al. (2002). The map in Carroll et al. (2005: fig. 1) implies that populations previously known as *S. h. berlandieri* and *S. h. solus* are subsumed under *S. toltecus*, and they did not identify the population of *S. hispidus* from southwestern Arizona and northwestern Sonora.

Family Erethizontidae.—Bonvicino et al. (2002) and Woods and Kilpatrick (2005) recognized *Sphiggurus* and *Coendou* as valid genera, and Woods and Kilpatrick (2005) used the name combination *Sphiggurus mexicana* for the Mexican prehensile-tailed porcupine. However, we follow Voss (2011) and Voss et al. (2013) by treating *Sphiggurus* as a synonym of *Coendou* and use *Coendou mexicanus* for the species in Mexico.

ORDER CARNIVORA

Wozencraft (2005) recognized only six subspecies of *Puma concolor* from throughout its distribution. He listed the subspecies in Mexico as *P. c. cougar* under which the previously recognized names for Mexican *P. concolor* (*P. c. azteca*, *P. c. browni*, *P. c. californica*, *P. c. improcera*, *P. c. mayensis*, and *P. c. stanleyana*) are treated as synonyms.

Wozencraft (2005) separated *Spilogale angustifrons* from *S. putorius* and recognized four subspecies of *S. angustifrons* from Mexico: *S. a. angustifrons*, *S. a. elata*, *S. a. tropicalis*, and *S. a. yucatanensis*. Wozencraft (2005) also treated *Bassariscus sumichrasti campechensis* as a junior synonym of *B. s. sumichrasti*. We follow these taxonomic changes.

Helgen and Wilson (2005) assessed the taxonomic status of named forms of *Procyon lotor* including those from Mexico and Central America. They designated a neotype for *P. l. hernandezii*, treated both *P. l. mexicanus* and *P. l. shufeldti* as synonyms of *P. l. hernandezii*, and treated *P. insularis* as a subspecies of *P. lotor*.

Álvarez-Castañeda (2000) listed *Lontra canadensis* from Baja California based on records from Grinnell (1914a). During a careful reading of Grinnell's publication we did not find any records listed for the Colorado River or for Mexico. Also, there is

no mention of any record of *L. canadensis* from the lower Colorado River or from Mexico in Grinnell (1914b), although Grinnell acknowledged that river otters probably occurred along the Colorado. Ceballos and Arroyo-Cabrales (2013) recently listed the species for Mexico, but we have yet to find objective evidence verifying its presence.

Pacheco et al. (2000, 2002) published the initial results for the reintroduction of the black-footed ferret *Mustela nigripes* into northwestern Chihuahua, particularly in the Janos Biosphere Reserve–Casas Grandes region. López-González and García-Mendoza (2012) listed the species for Chihuahua as an introduction, not as a reintroduction. We do not include *M. nigripes* in this list and little has been reported regarding the status of the introduction (Belant et al. 2008; List et al. 2010). We follow Wozencraft (2005) in treating *Vulpes macrotis* as a species separate from *V. velox*.

ORDER ARTIODACTYLA

Góngora and Morán (2005), based on a study of mitochondrial and nuclear DNA in the three recognized Recent genera in the family Tayassuidae (*Pecari*, *Tayassu* and *Catagonus*), suggested that the complex consisted of only two genera, *Pecari* for the collared peccaries, and *Tayassu* for the white-lipped and Chacoan peccaries. Grubb (1993, 2005), continued recognizing three genera, two of which occur in Mexico (*Pecari* [now *Dicotyles*—see below] and *Tayassu*).

Groves and Grubb (2011), based on finding consistent differences in dental morphology, shape of the nasals and malar crest, form of the fossa above the diastema, and pelage coloration between the peccaries from North and South America, restricted *Pecari tajacu* to South America and recognized *P. angulatus* and *P. crassus* in the USA and Mexico. Groves and Grubb (2011) gave the distribution of *P. angulatus* as western Mexico and southwestern USA; with *P. crassus* distributed from Texas, USA, southward through eastern Mexico, Central America, and into South America along the Pacific lowlands of Colombia and Ecuador. In this list we use the species-group taxa as detailed by Groves and Grubb (2011); however, as in many other groups, further study is needed to identify diagnostic characters separating both species.

Grubb (2005) and Groves and Grubb (2011) used the name *Pecari* Reichenbach, 1835, (type species *Sus torquatus* G. Cuvier, 1816, by monotypy), for the collared peccary based on the understanding that the generic name *Dicotyles* was unavailable for collared peccaries because the type species of *Dicotyles* was *Dicotyles labiatus* G. Cuvier 1816, by subsequent selection by Miller (1912:384). Miller (1912), however, as he later acknowledged (Miller 1914), had overlooked the designation of *Dicotyles torquatus* as the type species of *Dicotyles* by Palmer (1904:930). Both generic names have the same type species; therefore, *Dicotyles* G. Cuvier 1816 has priority over *Pecari* Reichenbach, 1835, and the collared peccaries (*sensu lato*) take the names *Dicotyles angulatus*, *D. crassus*, and *Dicotyles tajacu* (the latter based on *Sus tajacu* Linnaeus 1758). *Tayassu* G. Fischer 1814 is the correct generic name for the white-lipped peccary, *T. pecari*.

Grubb (2005) and Groves and Grubb (2011) restricted *Mazama americana* to South America, and recognized *M. temama* as distributed in Mexico, Central America, and northwestern Colombia. We recognize the two brocket deer from Mexico as *Mazama pandora* and *M. temama*.

ORDER PERISSODACTYLA

Groves and Grubb (2011), based on the genetic work by Ashley et al. (1996), discuss the differences among the different species of tapir, concluding that the South American tapirs (*Tapirus pinchaque* and *T. terrestris*) and the Mesoamerican tapir (*T. bairdii*) are sufficiently distinct to treat them as different genera. Consequently, they used *Tapirus* for the South American tapirs and *Tapirella* for the Mesoamerican species, as we do here.

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LITERATURE CITED

- Alexander, L. F., and B. R. Riddle. 2005. Phylogenetics of the New World rodent family Heteromyidae. *Journal of Mammalogy* 86:366–379.
- Allen, J. A. 1904. Mammals from southern Mexico and Central and South America. *Bulletin of the American Museum of Natural History* 20:29–80.
- Almendra, A. L., D. S. Rogers, and F. X. González-Cózatl. 2014. Molecular phylogenetics of the *Handleyomys chapmani* complex in Mesoamerica. *Journal of Mammalogy* 95:26–40.
- Álvarez-Castañeda, S. T. 2000. Familia Mustelidae. Pp. 731–757 in *Mamíferos del Noroeste de México II* (S. T. Álvarez-

- Castañeda, and J. L. Patton, eds.). Centro de Investigaciones Biológicas del Noroeste, S. C. xii+587–873 pp.
- Álvarez–Castañeda, S. T. 2007. Systematics of the antelope ground squirrel (*Ammospermophilus*) from islands adjacent to the Baja California Peninsula. *Journal of Mammalogy* 88:1160–1169.
- Álvarez–Castañeda, S. T. 2010. Phylogenetic structure of the *Thomomys bottae–umbrinus* complex in North America. *Molecular Phylogenetics and Evolution* 54:671–679.
- Álvarez–Castañeda, S. T., and P. Cortés–Calva. 2004. A new subspecies of sand pocket mouse *Chaetodipus arenarius* (Rodentia: Heteromyidae) from Baja California Sur. Pp. 33–40 in *Homenaje a la trayectoria mastozoológica de José Ramírez Pulido* (A. Castro Campillo and J. Ortega, eds.). Universidad Autónoma Metropolitana, Unidad Iztapalapa, xiv+234 pp.
- Álvarez–Castañeda, S. T., and E. Ríos. 2010. A phylogenetic analysis of *Neotoma varia* (Rodentia: Cricetidae), a rediscovered, endemic, and threatened rodent from Datil Island, Sonora, Mexico. *Zootaxa* 2647:51–60.
- Álvarez–Castañeda, S. T., and E. Ríos. 2011. Revision of *Chaetodipus arenarius* (Rodentia: Heteromyidae). *The Zoological Journal of the Linnean Society* 160: 213–228.
- Álvarez–Castañeda, S. T., W. Z. Lidicker, and E. Ríos. 2009. Revision of the *Dipodomys merriami* complex in the Baja California, Peninsula, Mexico. *Journal of Mammalogy* 90:992–1008.
- Andersen, J. J., and J. E. Light. 2012. Phylogeography and subspecies revision of the hispid pocket mouse, *Chaetodipus hispidus* (Rodentia: Heteromyidae). *Journal of Mammalogy* 93:1195–1215.
- Anderson, R. P., M. Weksler, and D. S. Rogers. 2006. Phylogenetic analyses of pocket mice (Heteromyidae: Heteromyinae) based on allozymic and morphological data. *Journal of Mammalogy* 87:1218–1233.
- Arellano, E., F. X. González–Cozátl, and D. S. Rogers. 2005. Molecular systematics of Middle American harvest mice *Reithrodontomys* (Muridae), estimated from mitochondrial cytochrome *b* gene sequences. *Molecular Phylogenetics and Evolution* 37:529–540.
- Arita T., H., and S. R. Humphrey. 1988. Revisión taxonómica de los murciélagos magueyeros del género *Leptonycteris* (Chiroptera: Phyllostomidae). *Acta Zoológica Mexicana* (nueva serie) 29:1–60.
- Ashley, M., J. Norman, and L. Stross. 1996. Phylogenetic analysis of the Perissodactyla family Tapiridae using mitochondrial cytochrome c oxidase (COII) sequences. *Journal of Mammalian Evolution* 3:315–326.
- Ávila–Valle, Z. A., A. Castro–Campillo, L. León–Paniagua, I. H. Salgado–Ugalde, A. G. Navarro–Sigüenza, B. E. Hernández–Baños, and J. Ramírez–Pulido. 2012. Geographic variation and molecular evidence of the blackish deer mouse complex (*Peromyscus furvus*, Rodentia: Muridae). *Mammalian Biology* 77:166–177.
- Baird, A. B., D. M. Hills, J. C. Patton, and J. W. Bickham. 2008. Evolutionary history of the genus *Rhogeessa* (Chiroptera: Vespertilionidae) as reevaluated by mitochondrial DNA sequences. *Journal of Mammalogy* 89:744–754.
- Baird, A. B., M. R. Marchán–Rivadeneira, S. G. Pérez, and R. J. Baker. 2012. Morphological analysis and description of two new species of *Rhogeessa* (Chiroptera: Vespertilionidae) from the neotropics. *Occasional Papers, Museum of Texas Tech University* 307:1–25.
- Baker, R. J. 1979. Karyology. Pp. 107–176 in *Biology of bats of the New World family Phyllostomatidae*. Part. III (R. J. Baker, J. K. Jones, Jr., and D. C. Carter, eds.). Special Publications The Museum, Texas Tech University 16:1–441.
- Baker, R. J. 1984. A sympatric cryptic species of mammal: a new species of *Rhogeessa* (Chiroptera: Vespertilionidae). *Systematic Zoology* 33:178–183.
- Baker, R. J., H. H. Genoways, and P. A. Seyfarth. 1981. Results of the Alcoa Foundation–Suriname Expeditions. VI. Additional chromosomal data for bats (Mammalia: Chiroptera) from Suriname. *Annals of Carnegie Museum* 50:333–344.
- Baker, R. J., M. B. O’Neill, and L. R. McAliley. 2003. A new species of desert shrew, *Notiosorex*, based on nuclear and mitochondrial sequence data. *Occasional Papers, Museum of Texas Tech University* 222:1–12.
- Bartig, J. L., T. L. Best, and S. L. Burt. 1993. *Tamias bulleri*. *Mammalian Species* 438:1–4.
- Belant, J., P. Gober, and D. Biggins. 2008. *Mustela nigripes* in IUCN 2013. IUCN Red List of Threatened Species. Version 2013.1. <www.iucnredlist.org>. accessed date 12 July 2013.
- Best, T. L. 1996. *Lepus californicus*. *Mammalian Species* 530:1–10.
- Best, T. L., and T. H. Henry. 1993. *Lepus alleni*. *Mammalian Species* 424:1–8.
- Best, T. L., and L. L. Janecek. 1992. Allozymic and morphologic variation among *Dipodomys insularis*, *Dipodomys nitratoides*, and two populations of *Dipodomys merriami* (Rodentia: Heteromyidae). *The Southwestern Naturalist* 37:1–8.
- Best, T. L., S. L. Burt, and J. L. Bartig. 1993. *Tamias durangae*. *Mammalian Species* 437:1–4.
- Bogan, M. A. 1997. On the status of *Neotoma varia* from Isla Datil, Sonora. Pp. 81–88 in *Life among the muses: papers in honor of James S. Findley* (T. L. Yates, W. L. Gannon, and D. Wilson, eds.). Special Publication, The Museum of Southwestern Biology 3:1–290.

- Bonvicino, C. R., V. Penna-Firme, and E. Braggio. 2002. Molecular and karyologic evidence of the taxonomic status of *Coendou* and *Sphiggurus* (Rodentia: Hystricognathi). *Journal of Mammalogy* 83: 1071–1076.
- Bradley, R. D., D. J. Schmidly, and C. W. Kilpatrick. 1996. The relationships of *Peromyscus sagax* to the *P. boylii* and *P. truei* species groups in Mexico based on morphometric, karyotypic, and allozymic data. Pp. 95–106 in *Contributions in Mammalogy: A Memorial Volume Honoring Dr. J. K. Jones, Jr.* (H. H. Genoways, ed.). Museum of Texas Tech University il+315 pp.
- Bradley, R. D., I. Tiemann–Boege, C. W. Kilpatrick, and D. J. Schmidly. 2000. Taxonomic status of *Peromyscus boylii sacarensis*: interferences from DNA sequences of the mitochondrial cytochrome-*b* gene. *Journal of Mammalogy* 81:875–884.
- Bradley, R. D., D. S. Carroll, M. L. Haynie, R. Muñiz Martínez, M. J. Hamilton, and W. L. Kilpatrick. 2004. A new species of *Peromyscus* from western Mexico. *Journal of Mammalogy* 85:1184–1193.
- Bradley, R. D., N. D. Durish, D. S. Rogers, J. R. Miller, M. D. Engstrom, and C. W. Kilpatrick. 2007. Toward a molecular phylogeny for *Peromyscus*: evidence from mitochondrial cytochrome-*b* sequences. *Journal of Mammalogy* 88:1146–1159.
- Bradley, R. D., D. D. Henson, and N. D. Durish. 2008. Re-evaluation of the geographic distribution and phylogeography of the *Sigmodon hispidus* complex based on mitochondrial DNA sequences. *The Southwestern Naturalist* 53:301–310.
- Bradley, R. D., N. Ordóñez-Garza, C. G. Sotero-Caio, H. M. Huynh, C. W. Kilpatrick, L. I. Iñiguez-Dávalos, and D. J. Schmidly. 2014. Morphometric, karyotypic, and molecular evidence for a new species of *Peromyscus* (Cricetidae: Neotominae) from Nayarit, Mexico. *Journal of Mammalogy* 95:176–186.
- Brandon-Jones, D. 2006. Apparent confirmation that *Alouatta villosa* (Gray, 1845) is a senior synonym of *A. pigra* Lawrence, 1933 as the species-group name for the black howler monkey of Belize, Guatemala and Mexico. *Primate Conservation* 21:41–43.
- Brandon-Jones, D., and C. P. Groves. 2002. Neotropical primate family-group names replaced by Groves (2001) in contravention of Article 40 of the International Code of Zoological Nomenclature. *Neotropical Primates* 10:113–115.
- Burmeister, H. 1854. Untitled [Ueber einer neue Ratte, *Lasiomys hirsutus*, von Maracaibo]. *Abhandlungen der Naturforschenden Gesellschaft zu Halle, Sitzungsberichte* 2:15–17.
- Burton, K. L., and M. D. Engstrom. 2001. Species diversity of bats (Mammalia: Chiroptera) in Iwokrama Forest, Guyana, and the Guianan subregion: implications for conservation. *Biodiversity and Conservation* 10:613–657.
- Cabrera, H., S., T. Álvarez-Castañeda, N. González-Ruiz, and J. P. Gallo. 2007. Distribution and natural history of Schmidly's deer mouse (*Peromyscus schmidlyi*). *The Southwestern Naturalist* 52:620–623.
- Callahan, J. R. 1980. Taxonomic status of *Eutamias bulleri*. *The Southwestern Naturalist* 25:1–8.
- Carleton, M. D., and J. Arroyo-Cabrales. 2009. Review of the *Oryzomys couesi* complex (Rodentia: Cricetidae: Sigmodontinae) in western Mexico. Pp. 93–127 in *Systematic Mammalogy: contributions in honor of Guy G. Musser* (R. S. Voss and M. D. Carleton, eds.). *Bulletin of the American Museum of Natural History* 331:1–450.
- Carraway, L. N. 2007. Shrews (Eulypotyphla: Soricidae) of Mexico. *Monographs of the Western North American Naturalist* 3:1–91.
- Carroll, D. S., and R. D. Bradley. 2005. Systematics of the genus *Sigmodon*: DNA sequences from beta-fibrinogen and cytochrome-*b*. *The Southwestern Naturalist* 50:342–349.
- Carroll, D. S., L. L. Peppers, and R. D. Bradley. 2005. Molecular systematics and phylogeography of the *Sigmodon hispidus* species group. Pp. 87–100 in *Contribuciones mastozoológicas en homenaje a Bernardo Villa* (V. Sánchez-Cordero and R. A. Medellín, eds.). Instituto de Biología, UNAM; Instituto de Ecología, UNAM; CONABIO, México, 706 pp.
- Ceballos, G., and J. Arroyo-Cabrales. 2013. Lista actualizada de los mamíferos de México 2012. *Revista Mexicana de Mastozoología, nueva época* 2:27–80.
- Cervantes, F. A., A. Castro-Campillo, and J. Ramírez-Pulido. 1994. Mamíferos terrestres nativos de México. *Anales del Instituto de Biología, Universidad Nacional Autónoma de México, Serie Zoológica* 65:177–190.
- Cervantes, F. A., A. Rojas-Viloria, C. Lorenzo, and S. T. Álvarez-Castañeda. 2001. Chromosomal differentiation between the jackrabbits *Lepus insularis* and *Lepus californicus* from Baja California Sur, México. *Revista Mexicana de Mastozoología* 4:41–53.
- Cole, F. R., and D. E. Wilson. 2006a. *Leptoncyteris curasoae*. *Mammalian Species* 796:1–3.
- Cole, F. R., and D. E. Wilson. 2006b. *Leptoncyteris yerbabuenae*. *Mammalian Species* 797:1–7.
- Davis, W. B. 1984. Review of the large fruit-eating bats of the “*Artibeus lituratus*” complex (Chiroptera: Phyllostomidae) in Middle America. *Occasional Papers, Museum of Texas Tech University* 93:1–16.
- Davis, W. B., and A. L. Gardner. 2008. Genus *Eptesicus* Rafinesque, 1820. Pp. 440–450 in *Mammals of South America. Vol. 1: Marsupials, xenarthrans, shrews, and bats* (A. L. Gardner, ed.). University of Chicago Press 1:xxi+1–669.

- Davis, W. B., and D. J. Schmidly. 1994. The mammals of Texas. Texas Parks and Wildlife Press.
- Desmastes, J. W., A. L. Butt, M. S. Hafner, and J. E. Light. 2003. Systematics of a rare species of pocket gopher *Papogeomys alcorni*. *Journal of Mammalogy* 84:753–761.
- Díaz, A. C., J. Hernández Camacho, and A. Cadena G. 1986. Lista actualizada de los mamíferos de Colombia. Anotaciones sobre su distribución. *Caldasia* 15:471–501.
- Dixon, K. R., J. A. Chapman, G. R. Willner, D. E. Wilson, and W. López-Forment. 1983. The new world jackrabbits and hares (Genus *Lepus*).—2. Numerical taxonomic analysis. *Acta Zoologica Fennica* 174:53–56.
- Eger, J. L. 2008. Family Molossidae P. Gervais, 1856. Pp. 399–439 in *Mammals of South America*. Vol. 1: Marsupials, xenarthrans, shrews, and bats (A. L. Gardner, ed.). University of Chicago Press 1:xxi+1–669.
- Escobedo-Morales, L. A., L. León-Paniagua, J. Arroyo-Cabrales, and F. Greenaway. 2006. Distributional records for mammals from Chiapas, Mexico. *The Southwestern Naturalist* 51:269–272.
- Espinoza, J., C. Lorenzo, and E. Rios. 2011. Variación morfológica y morfométrica de *Heteromys desmarestianus* en Chiapas, México. *Therya* 2:139–154.
- Fernández, J. A. 2012. Phylogenetics and biogeography of the microendemic rodent *Xerospermophilus perotensis* (Perote ground squirrel) in the Oriental Basin of Mexico. *Journal of Mammalogy* 93:1431–1439.
- Fernández, J. A., F. A. Cervantes, and M. S. Hafner. 2012. Molecular systematics and biogeography of the Mexican endemic kangaroo rat, *Dipodomys phillipsii* (Rodentia: Heteromyidae). *Journal of Mammalogy* 93:560–571.
- Gardner, A. L. 1973. The systematics of the genus *Didelphis* (Marsupialia: Didelphidae) in North and Middle America. Special Publications The Museum, Texas Tech University 4:1–81.
- Gardner, A. L. 2005a. Order Didelphimorphia. Pp. 3–20 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxxv+1–743 pp.
- Gardner, A. L. 2005b. Order Cingulata. Pp. 94–99 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxxv+1–743 pp.
- Gardner, A. L. 2005c. Case 3328. *Didelphis* Linnaeus, 1758 (Mammalia, Didelphidae): proposed correction of gender, and *Cryptotis* Pomel, 1848 (Mammalia, Soricidae): proposed fixation of gender. *Bulletin of Zoological Nomenclature* 62:142–145.
- Gardner, A. L., 2008a. *Mammals of South America*. Vol. 1: Marsupials, xenarthrans, shrews, and bats. University of Chicago Press 1:xxi+1–669.
- Gardner, A. L. 2008b. Tribe Sturnirini [Miller, 1907]. Pp. 363–376 in *Mammals of South America*. Vol. 1: Marsupials, xenarthrans, shrews, and bats (A. L. Gardner, ed.). University of Chicago Press 1:xxi+1–669.
- Gardner, A. L. 2008c. Genus *Vampyroides* O. Thomas, 1900. Pp. 355–357 in *Mammals of South America*. Vol. 1: Marsupials, xenarthrans, shrews, and bats (A. L. Gardner, ed.). University of Chicago Press 1:xxi+1–669.
- Gardner, A. L., and M. Dagosto. 2008. Tribe Metachirini Reig, Kirsch, and Marshall, 1985. Pp. 35–39 in *Mammals of South America*. Vol. 1: Marsupials, xenarthrans, shrews, and bats (A. L. Gardner, ed.). University of Chicago Press 1:xxi+1–669.
- Godínez, E. G., N. González-Ruiz, and J. Ramírez-Pulido. 2011. Actualización de la lista de los mamíferos de Jalisco, México: implicaciones de los cambios taxonómicos. *Therya* 2:7–35.
- Goldman, E. A. 1911. Revision of the spiny pocket mice (genera *Heteromys* and *Liomys*). *North American Fauna* 34:1–70, 3 plates.
- Góngora, J., and C. Morán. 2005. Nuclear and mitochondrial evolutionary analyses of collared, white-lipped, and chacoan peccaries (Tayassuidae). *Molecular Phylogenetics and Evolution* 34:181–189.
- González-Ruiz, N., J. Ramírez-Pulido, and J. Arroyo-Cabrales. 2011. A new species of mastiff bat (Molossidae: Molossus) from Mexico. *Mammalian Biology* 76:461–469.
- Goodwin, G. G. 1940. Three new bats from Honduras and the first record of *Enchisthenes harti* (Thomas) for North America. *American Museum Novitates* 1075:1–4.
- Goodwin, G. G. 1942. *Mammals of Honduras*. *Bulletin of the American Museum of Natural History* 79:107–195.
- Gregorin, R., G. L. Capusso, and V. R. Furtado. 2008. Geographic distribution and morphological variation in *Mimon bennettii* (Chiroptera, Phyllostomidae). *Iheringia, Série Zoologia, Porto Alegre* 98:404–411.
- Grinnell, J. 1914a. An account of the mammals and birds of the lower Colorado valley with special reference to the distributional problems presented. University of California, Publications in Zoology 12:51–294.
- Grinnell, J. 1914b. Distribution of river otters in California. University of California, Publications in Zoology 12:305–310, 1 plate.
- Groves, C. P. 2001. *Primate taxonomy*. Smithsonian Institution Press, Washington, D.C. 350 pp.
- Groves, C. P. 2005. Order Primates. Pp. 111–184 in *Mammal species of the world. A taxonomic and geographic refer-*

- ence (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxv+1–743 pp.
- Groves, C., and P. Grubb. 2011. Ungulate taxonomy. Baltimore: The Johns Hopkins University Press ix+317 pp.
- Grubb, P. 1993. Order Artiodactyla. Pp. 377–414 in Mammal species of the world. A taxonomic and geographic reference (D. E. Wilson and A. M. Reeder, eds.), Second edition. Smithsonian Institution Press, Washington, D.C. xvii+1207 pp.
- Grubb, P. 2005. Order Artiodactyla. Pp. 637–722 in Mammal species of the world. A taxonomic and geographic reference (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxv+1–743 pp.
- Guerrero, J. A., E. de Luna, and C. Sánchez–Hernández. 2003. Morphometrics in the quantification of character state identity for the assessment of primary homology: an analysis of character variation of the genus *Artibeus* (Chiroptera: Phyllostomidae). *Biological Journal of the Linnean Society* 80:45–55.
- Guerrero, J. A., E. de Luna, and D. González. 2004. Taxonomic status of *Artibeus jamaicensis triomylus* inferred from molecular and morphometric data. *Journal of Mammalogy* 85:866–874.
- Gutiérrez, E. E., S. A. Jansa, and R. S. Voss. 2010. Molecular systematics of mouse opossums (Didelphidae: *Marmosa*): assessing species limits using mitochondrial DNA sequences, with comments on phylogenetic relationships and biogeography. *American Museum Novitates* 3692:1–22.
- Hafner, D. J., and M. S. Hafner. 1983. Evolutionary relationships of heteromyid rodents. *Great Basin Naturalist Memoirs* 7:3–29.
- Hafner, D. J., M. S. Hafner, G. L. Hasty, T. A. Spradling, and J. W. Demastes. 2008. Evolutionary relationships of pocket gophers (*Cratogeomys castanops* species group) of the Mexican Altiplano. *Journal of Mammalogy* 89:190–208.
- Hafner, M. S., J. E. Leight, D. J. Hafner, S. V. Brant, T. A. Spradling, and J. W. Demastes. 2005. Cryptic species in the Mexican pocket gopher *Cratogeomys merriami*. *Journal of Mammalogy* 86:1095–1108.
- Hafner, M. S., J. C. , J. E. Light, D. J. Hafner, M. S. Hafner, E. Reddington, D. S. Rogers, and B. R. Riddle. 2007. Basal clades and molecular systematic of heteromyid rodents. *Journal of Mammalogy* 88:1129–1145.
- Hafner, M. S., T. A. Spradling, J. E. Light, D. J. Hafner, and J. R. Demboski. 2004. Systematic revision of pocket gophers of the *Cratogeomys gymnurus* species group. *Journal of Mammalogy* 85:1170–1183.
- Hafner, M. S., D. J. Hafner, J. W. Demastes, G. L. Hasty, J. E. Light, and T. A. Spradling. 2009. Evolutionary relationships of pocket gophers of the genus *Pappogeomys* (Rodentia: Geomyidae). *Journal of Mammalogy* 90:47–56.
- Hafner, M. S., A. R. Gates, V. Mathis, J. W. Demastes, and D. J. Hafner. 2011. Redescription of the pocket gopher *Thomomys atrovarius* from the Pacific coast of mainland Mexico. *Journal of Mammalogy* 92:1367–1382.
- Hall, E. R. 1960. *Oryzomys couesi* only subspecifically different from the marsh rice rat, *Oryzomys palustris*. *The Southwestern Naturalist* 5:171–173.
- Hall, E. R. 1981. The mammals of North America. John Wiley and Sons, vol. 1:xv+600+90, vol. 2:vi+601–1181+90.
- Handley, C. O., Jr. 1959. A revision of American bats of the genera *Euderma* and *Plecotus*. *Proceedings of the United States National Museum* 110:95–246.
- Hanson, J. D., J. L. Indorf, V. J. Swier, and R. D. Bradley. 2010. Molecular divergence within the *Oryzomys palustris* complex: evidence for multiple species. *Journal of Mammalogy* 91:336–347.
- Harris, D., and D. S. Rogers. 1999. Species limits and phylogenetic relationships among populations of *Peromyscus furvus*. *Journal of Mammalogy* 80:530–544.
- Harris, D., D. S. Rogers, and J. Sullivan. 2000. Phylogeography of *Peromyscus furvus* (Rodentia; Muridae) based on cytochrome *b* sequence data. *Molecular Ecology* 9:2129–2135.
- Harrison, R. G., S. M. Bogdanowicz, R. S. Hoffmann, E. Yensen, and P. W. Sherman. 2003. Phylogeny and evolutionary history of the ground squirrels (Rodentia: Marmotinae). *Journal of Mammalian Evolution* 10:249–276.
- Helgen, K. M., and D. E. Wilson. 2005. A systematic and zoogeographic overview of the raccoons of Mexico and Central America. Pp. 221–236 in *Contribuciones mastozoológicas en homenaje a Bernardo Villa* (V. Sánchez–Cordero and R. A. Medellín, eds.). Instituto de Biología, UNAM; Instituto de Ecología, UNAM; CONABIO, México 706 pp.
- Helgen, K. M., F. R. Cole, L. E. Helgen, and D. E. Wilson. 2009. Generic revision in the Holarctic ground squirrel genus *Spermophilus*. *Journal of Mammalogy* 90:270–305.
- Henson, D. D., and R. D. Bradley. 2009. Molecular systematics of the genus *Sigmodon*: results from mitochondrial and nuclear gene sequences. *Canadian Journal of Zoology* 87:211–220.
- Herron, M. D., T. A. Castoe, and C. L. Parkinson. 2004. Sciurid phylogeny and the paraphyly of Holarctic ground squirrels (*Spermophilus*). *Molecular Phylogenetics and Evolution* 31:1015–1030.
- Hoffmann, F. G., J. G. Owen, and R. J. Baker. 2003. mtDNA perspective of chromosomal diversification and hybridization in Peters' tent-making bat (*Uroderma bilobatum*: Phyllostomidae). *Molecular Ecology* 12:2981–2993.

- Hoffmann, R. S., and A. T. Smith. 2005. Order Lagomorpha. Pp. 185–211 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxxv+1–743 pp.
- Hoffmeister, D. F. 1986. *Mammals of Arizona*. The University of Arizona Press and The Arizona Game and Fish Department xx+602 pp.
- Hood, C., and A. L. Gardner. 2008. Family Emballonuridae Gervais, 1856. Pp. 188–207 in *Mammals of South America. Vol. 1: Marsupials, xenarthrans, shrews, and bats* (A. L. Gardner, ed.). University of Chicago Press, 1:xxi+1–669.
- Hooper, S. R., and R. A. Van Den Bussche. 2003. Molecular phylogenetics of the chiropteran family Vespertilionidae. *Acta Chiropterologica* 5 (supplement):1–63.
- Hooper, S. R., S. Solari, P. A. Larsen, R. D. Bradley, and R. J. Baker. 2008. Phylogenetics of the fruit-eating bats (Phyllostomidae: *Artibeina*) Inferred from mitochondrial DNA sequences. *Occasional Papers, Museum of Texas Tech University* 277:1–15.
- Hooper, E. T. 1952. A systematic review of the harvest mice (genus *Reithrodontomys*) of Latin America. *Miscellaneous Publications of the Museum of Zoology, University of Michigan* 77:1–225.
- Huckaby, D. G. 1980. Species limits in the *Peromyscus mexicanus* group (Mammalia: Rodentia: Muroidea). *Contributions in Science, Natural History Museum of Los Angeles County* 326:1–24.
- Hutterer, R. 2005a. Order Soricomorpha. Pp. 220–311 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxxv+1–743 pp.
- Hutterer, R. 2005b. Order Erinaceomorpha. Pp. 212–219 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxxv+1–743 pp.
- Indorf, J. L., and M. S. Gaines. 2013. Genetic divergence of insular marsh rice rats in subtropical Florida. *Journal of Mammalogy* 94(4):897–910.
- International Commission on Zoological Nomenclature (ICZN). 2006. Opinion 2164 (case 3328). *Didelphis* Linnaeus, 1758 (Mammalia, Didelphidae): gender corrected to feminine, and *Cryptotis* Pomel, 1848 (Mammalia, Soricidae): gender fixed as masculine. *Bulletin of Zoological Nomenclature* 63:282–283.
- Iudica, C. A. 2000. Systematic revision of the neotropical fruit bats of the genus *Sturnira*: a molecular and morphological approach. Unpublished Ph.D. dissertation, University of Florida, Gainesville xii+284 pp.
- Jameson, E. W., Jr. 1999. Host–parasite relationships among North American chipmunks. *Acta Theriologica* 44:225–231.
- Jones, J. K., Jr., D. C. Carter, H. H. Genoways, R. S. Hoffmann, D. W. Rice, and C. Jones. 1986. Revised checklist of North American mammals north of Mexico, 1986. *Occasional Papers, Museum of Texas Tech University* 107:1–22.
- Koopman, K. F. 1993. Order Chiroptera. Pp. 137–241 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Second edition. Smithsonian Institution Press, Washington, D.C. xvii+1207 pp.
- Koopman, K. F. 1994. Chiroptera: Systematics. *Handbook of Zoology*, vol. VIII, part 60. Walter de Gruyter, Berlin and New York 217 pp.
- Larsen, P. A., S. R. Hooper, M. C. Bozeman, S. C. Pedersen, H. H. Genoways, C. J. Phillips, D. E. Pumo, and R. J. Baker. 2007. Phylogenetics and phylogeography of the *Artibeus jamaicensis* complex based on cytochrome-*b* DNA sequences. *Journal of Mammalogy* 88:712–727.
- Larsen, P. A., M. R. Marchán-Rivadeneira, and R. J. Baker. 2010. Taxonomic status of Andersen's fruit-eating bat (*Artibeus jamaicensis aequatorialis*) and revised classification of *Artibeus* (Chiroptera: Phyllostomidae). *Zootaxa* 2648:45–60.
- LaVal, R. K. 1973. A revision of the neotropical bats of the genus *Myotis*. *Natural History Museum of Los Angeles County, Science Bulletin* 15:1–54.
- Lee, T. E., Jr., B. R. Riddle, and P. L. Lee. 1996. Speciation in the desert pocket mouse (*Chaetodipus penicillatus* Woodhouse). *Journal of Mammalogy* 77:58–68.
- Lehmann, E. von, and H. -E. Schaefer. 1979. Cytologisch-taxonomische Studien an einer Kleinsaugeraufsammlung aus Honduras (Spermienmorphologie und vergleichende Cytochemie). *Zeitschrift für Zoologische Systematik und Evolutionsforschung* 17: 226–236.
- Levenson, H. R., R. S. Hoffman, C. F. Nadler, L. Deutsch, and S. D. Freeman. 1985. Systematics of the holarctic chipmunks (*Tamias*). *Journal of Mammalogy* 66:219–242.
- Lidicker, W. Z., Jr. 1960. An analysis of intraspecific variation in the kangaroo rat *Dipodomys merriami*. *University of California Publications in Zoology* 67:125–218.
- List, R., J. Pacheco, E. Ponce, R. Sierra-Corona, and G. Ceballos. 2010. The Janos Biosphere Reserve, northern Mexico. *International Journal of Wilderness* 16:35–41.
- López-González, C., and D. F. García-Mendoza. 2012. A checklist of the mammals (Mammalia) of Chihuahua, Mexico. *Check List* 8:1122–1133.
- López-González, C., D. F. García-Mendoza, and M. A. Correa-Ramírez. 2013. Morphologic characterization of

- Peromyscus schmidlyi* (Rodentia: Cricetidae), an endemic of the Sierra Madre Occidental, Mexico. *Journal of Mammalogy* 94:923–937.
- López-Wilchis, R., L. M. Guevara-Chumacero, N. P. Ángeles, J. Juste, C. Ibáñez, and I. D. L. A. Barriga-Sosa. 2012. Taxonomic status assessment of the Mexican populations of funnel-eared bats, genus *Natalus* (Chiroptera: Natalidae). *Acta Chiropterologica* 14:305–316.
- Marchán-Rivadeneira, M. R., P. A. Larsen, C. J. Phillips, R. E. Strauss, and R. J. Baker. 2012. On the association between environmental gradients and skull size variation in the great fruit-eating bat, *Artibeus lituratus* (Chiroptera: Phyllostomidae). *Biological Journal of the Linnean Society* 105:623–634.
- Marques-Aguiar, S. A. 1994. A systematic review of large species of *Artibeus* Leach, 1821 (Mammalia: Chiroptera), with some phylogenetic inferences. *Boletín do Museu Paraense, Emilio Goeldi, Serie Zoologia* 10:3–83.
- Mathis, V. L., M. S. Hafner, D. J. Hafner, and J. W. Demastes. 2013a. Resurrection and redescription of the pocket gopher *Thomomys sheldoni* from the Sierra Madre Occidental of Mexico. *Journal of Mammalogy* 94:544–560.
- Mathis, V. L., M. S. Hafner, D. J. Hafner, and J. W. Demastes. 2013b. *Thomomys nayarensis*, a new species of pocket gopher from the Sierra del Nayar, Nayarit, Mexico. *Journal of Mammalogy* 94:983–994.
- McDonough, M. M., L. K. Ammerman, R. M. Timm, H. H. Genoways, P. A. Larsen, and R. J. Baker. 2008. Speciation within bonneted bats (genus *Eumops*): the complexity of morphological, mitochondrial, and nuclear data sets in systematics. *Journal of Mammalogy* 89:1306–1315.
- McKenna, M., and S. K. Bell. 1997. *Classification of mammals above species level*. Columbia University Press, New York xiii+631 pp.
- McKnight, M. L. 2005. Phylogeny of the *Perognathus longimembris* species group based on mitochondrial cytochrome-*b*: how many species? *Journal of Mammalogy* 86:826–832.
- McLellan, L. J. 1984. A morphometric analysis of *Carollia* (Chiroptera, Phyllostomidae). *American Museum Novitates* 2791:1–35.
- McLellan, L. J., and K. F. Koopman. 2008. Subfamily Carollinae Miller, 1924, Pp. 208–218 in *Mammals of South America*. Vol. 1: Marsupials, xenarthrans, shrews, and bats (A. L. Gardner, ed.). University of Chicago Press 1:xxi+1–669.
- Merriam, C. H. 1901. Synopsis of the rice rats (genus *Oryzomys*) of the United States and Mexico. *Proceedings of the Washington Academy of Sciences* 3:273–295.
- Merriam, C. 1895. Monographic revision of the pocket gophers family Geomyidae (exclusive of the species of *Thomomys*). *North American Fauna* 8:1–258.
- Miller, G. S., Jr. 1912. List of North American land mammals in the United States National Museum, 1911. *Bulletin of the United States National Museum* 79:xiv+455 pp.
- Miller, G. S., Jr. 1914. The generic name of the collared peccaries. *Proceedings of the Biological Society of Washington* 27:215.
- Miller, J. R., and M. D. Engstrom. 2008. The relationships of major lineages within peromyscine rodents: a molecular phylogenetic hypothesis and systematic reappraisal. *Journal of Mammalogy* 89:1279–1295.
- Musser, G. G., and M. D. Carleton. 2005. Superfamily Muroidea. Pp. 894–1534 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 2:xvii+745–2142 pp.
- Nelson, E. W. 1904. Descriptions of seven new rabbits from Mexico. *Proceedings of the Biological Society of Washington* 17:103–110.
- Nelson, E. W., and E. A. Goldman. 1909. Eleven new mammals from Lower California. *Proceedings of the Biological Society of Washington* 22:23–28.
- Norma Oficial Mexicana. 2010. NOM-059-SEMARNAT-2010, Protección ambiental-Especies nativas de México de flora y fauna silvestres-Categorías de riesgo y especificaciones para la inclusión, exclusión o cambio-Lista de especies en riesgo. *Diario Oficial de la Federación*, 30 de diciembre de 2010. México, D.F. 93 pp.
- Ohdachi, S. D., M. Hasegawa, M. A. Iwasa, P. Vogel, T. Oshida, L.-K. Lin, and H. Abe. 2006. Molecular phylogenetics of soricid shrews (Mammalia) based on mitochondrial cytochrome-*b* gene sequences: with special reference to the Soricinae. *Journal of Zoology* 270:177–191.
- Osgood, W. H. 1907. Four new pocket mice. *Proceedings of the Biological Society of Washington* 20:19–22.
- Owen, R. D. 1987. Phylogenetic analyses of the bat subfamily Stenodermatinae (Mammalia: Chiroptera). *Special Publications The Museum, Texas Tech University* 26:1–65.
- Pacheco, J., G. Ceballos, and R. List. 2000. Los mamíferos de la región de Janos-Casas Grandes, Chihuahua, México. *Revista Mexicana de Mastozoología* 4:71–85.
- Pacheco, J., G. Ceballos, and R. List. 2002. Reintroducción del hurón de patas negras en las praderas de Janos, Chihuahua. *Biodiversitas* 42:1–5.
- Palmer, T. S. 1904. Index generum mammalium: A list of the genera and families of mammals. *North American Fauna* 23:1–984.
- Patton, J. L. 2005. Family Geomyidae. Pp. 859–870 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 2:xvii+745–2142 pp.

- Patton, J. L., and S. T. Álvarez-Castañeda. 1999. Family Heteromyidae. Pp. 351–443 in Mamíferos del Noroeste de México (S. T. Álvarez-Castañeda and J. L. Patton, eds.). Centro de Investigaciones Biológicas del Noroeste, S. C. 583 pp.
- Patton, J. L., D. G. Huckaby, and S. T. Álvarez-Castañeda. 2008. The evolutionary history and a systematic revision of woodrats of the *Neotoma lepida* Group. University of California Publications in Zoology 135:1–411.
- Peppers, L., and R. D. Bradley. 2000. Cryptic species in *Sigmodon hispidus*: evidence from DNA sequence. Journal of Mammalogy 81:332–343.
- Peppers, L., D. S. Carroll, and R. D. Bradley. 2002. Molecular systematics of the genus *Sigmodon* (Rodentia: Muridae): evidence from the mitochondrial cytochrome-*b* gene. Journal of Mammalogy 83:396–407.
- Piaggio, A. J., and S. L. Perkins. 2005. Molecular phylogeny of North American long-eared bats (Vespertilionidae: *Corynorhinus*); inter- and intraspecific relationships inferred from mitochondrial and nuclear DNA sequences. Molecular Phylogenetics and Evolution 37:762–775.
- Piaggio, A. J., and G. S. Spicer. 2001. Molecular phylogeny of the chipmunks inferred from mitochondrial cytochrome *b* and cytochrome oxidase II gene sequences. Molecular Phylogenetics and Evolution 20:335–350.
- Porter, C. A., S. R. Hooper, C. A. Cline, F. G. Hoffmann, and R. J. Baker. 2007. Molecular phylogenetics of the phyllostomid bat genus *Micronycteris* with descriptions of two new subgenera. Journal of Mammalogy 88:1205–1215.
- Ramírez-Pulido, J., R. López Wilchis, C. Müdspacher, and I. Lira. 1982. Catálogo de los mamíferos terrestres nativos de México. Universidad Autónoma Metropolitana-Iztapalapa, México 126 pp.
- Ramírez-Pulido, J., R. López Wilchis, C. Müdspacher, and I. Lira. 1983. Lista y bibliografía reciente de los mamíferos de México. Universidad Autónoma Metropolitana-Iztapalapa, México 5+xii+363 pp.
- Ramírez-Pulido, J., A. Castro-Campillo, J. Arroyo-Cabrales, and F. A. Cervantes. 1996. Lista taxonómica de los mamíferos terrestres de México: A taxonomic list of the terrestrial mammals of Mexico. Occasional Papers, Museum of Texas Tech University 158:1–62.
- Ramírez-Pulido, J., J. Arroyo-Cabrales, and A. Castro-Campillo. 2005. Estado actual y relación nomenclatural de los mamíferos terrestres de México. Acta Zoológica Mexicana (nueva serie) 21:21–82.
- Ramírez-Silva, J. P., F. X. González-Cózatl, E. Vázquez-Domínguez, and F. A. Cervantes. 2010. Phylogenetic position of Mexican jackrabbits within the genus *Lepus* (Mammalia: Lagomorpha): a molecular perspective. Revista Mexicana de Biodiversidad 81:721–731.
- Redondo, R. A. F., L. P. S. Brina, R. F. Silva, A. D. Ditchfield, and F. R. Santos. 2008. Molecular systematics of the genus *Artibeus* (Chiroptera: Phyllostomidae). Molecular Phylogenetics and Evolution 49:44–58.
- Riddle, B. R., D. J. Hafner, and L. F. Alexander. 2000. Comparative phylogeography of Baileys' pocket mouse (*Chaetodipus baileyi*) and the *Peromyscus eremicus* species group: historical vicariance of the Baja California Peninsular desert. Molecular Phylogenetics and Evolution 17:161–172.
- Ríos, E., and S. T. Álvarez-Castañeda. 2007. Environmental responses to altitudinal gradients and subspecific validity in pocket gophers (*Thomomys bottae*) from Baja California Sur, Mexico. Journal of Mammalogy 88:926–934.
- Ríos, E., and S. T. Álvarez-Castañeda. 2010. Phylogeography and systematics of the San Diego pocket mouse (*Chaetodipus fallax*). Journal of Mammalogy 91:293–301.
- Rogers, D. S., and M. W. González. 2010. Phylogenetic relationships among spiny pocket mice (*Heteromys*) inferred from mitochondrial and nuclear sequence data. Journal of Mammalogy 91:914–930.
- Rogers, D. S., and J. A. Skoy. 2011. *Peromyscus fuvvus* (Rodentia: Cricetidae). Mammalian Species 43: 209–215
- Rogers, D. S., M. D. Engstrom, and E. Arellano. 2005. Phylogenetic relationships among peromyscine rodents: allozyme evidence. Pp. 437–450 in Contribuciones mastozoológicas en homenaje a Bernardo Villa (V. Sánchez-Cordero and R. A. Medellín, eds.). Instituto de Biología, UNAM; Instituto de Ecología, UNAM; CONABIO, México 706 pp.
- Romo-Vázquez, E., L. León-Paniagua, and O. Sánchez. 2005. A new species of *Habromys* (Rodentia: Neotominae). Proceedings of the Biological Society of Washington 118:605–618.
- Rossi, R. V., R. S. Voss, and D. P. Lunde. 2010. A revision of the didelphid marsupial genus *Marmosa*. Part 1. The species in Tate's "mexicana" and "Mitis" sections and other closely related forms. Bulletin of the American Museum of Natural History 334:1–83.
- Roth, E. L. 1976. A new species of pocket mouse (*Perognathus*: Heteromyidae) from the Cape Region of Baja California Sur, Mexico. Journal of Mammalogy 57:562–566.
- Ruedas, L. A., and J. Salazar-Bravo. 2007. Morphological and chromosomal taxonomic assessment of *Sylvilagus brasiliensis gabbi* (Leporidae). Mammalia 71:63–67.
- Rylands, A. B., and R. A. Mittermeier. 2009. The diversity of the New World primates (Platyrrhini). An annotated taxonomy. Pp. 23–54 in South American Primates: Comparative perspectives in the study of behavior, ecology, and conservation (P. A. Garber, A. Estrada, J. C. Bicca-Marques, E. W. Heymann, K. B. Strier, eds.). New York: Springer 580 pp.

- Rylands, A. B., C. P. Groves, R. A. Mittermeier, L. Cortés-Ortiz, and J. J. Hines. 2006. Taxonomy and distributions of Mesoamerican primates. Pp. 29–79 in *New perspectives in the study of Mesoamerican primates: Distribution, ecology, behavior and conservation* (A. Estrada, P. Garber, M. Pavelka, and L. Luecke, eds). New York: Springer, xvi+600 pp.
- Schaldach, W. J., Jr. 1964. Notas breves sobre algunos mamíferos del sur de México. *Anales del Instituto de Biología, Universidad Nacional Autónoma de México* 35:129–137.
- Simmons, N. B. 2005. Order Chiroptera. Pp. 312–529 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson, and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxv+743 pp.
- Simmons, N. B., and R. S. Voss. 1998. The mammals of Paracou, French Guiana: a neotropical lowland rainforest fauna part I. Bats. *Bulletin of the American Museum of Natural History* 237: 1–219.
- Simmons, N. B., and A. L. Wetterer. 2002. Phylogeny and convergence in cactophilic bats. Pp. 87–121 in *Columnar Cacti and their Mutualists: Evolution, Ecology, and Conservation* (T. H. Fleming and A. Valiente-Banuet, eds.). The University of Arizona Press xiii+371 pp.
- Simpson, G. G. 1945. The principles of classification and a classification of mammals. *Bulletin of the American Museum of Natural History* 85:xvi+350.
- Smith, J. D. 1970. The systematic status of the black howler monkey, *Alouatta pigra* Lawrence. *Journal of Mammalogy* 51:358–369.
- Sullivan, J., J. A. Markert, and C. W. Kilpatrick. 1997. Phylogeography and molecular systematics of the *Peromyscus aztecus* species group (Rodentia: Muridae) inferred using parsimony and likelihood. *Systematic Biology* 46:426–440.
- Tejedor, A. 2005. A new species of funnel-eared bat (Natalidae: *Natalus*) from Mexico. *Journal of Mammalogy* 86:1109–1120.
- Tejedor, A. 2006. The type locality of *Natalus stramineus* (Chiroptera: Natalidae): implications for the taxonomy and biogeography of the genus *Natalus*. *Acta Chiropterologica* 8:361–380.
- Tejedor, A. 2011. Systematics of funnel-eared bats (Chiroptera: Natalidae). *Bulletin of the American Museum of Natural History* 353:1–140.
- Thomas, O. 1893a. Notes on some Mexican *Oryzomys*. *Annals and Magazine of Natural History Series* 6, 11:402–405.
- Thomas, O. 1893b. On some of the larger species of *Geomys*. *Annals and Magazine of Natural History, Series* 6 12:269–273.
- Thorington, R. W., and R. S. Hoffmann. 2005. Family Sciuridae. Pp. 754–818 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 2: xvii+745–2142 pp.
- Tiemann-Boege, I., C. W. Kilpatrick, D. J. Schmidly, and R. D. Bradley. 2000. Molecular phylogenetics of *Peromyscus boylii* species group (Rodentia: Muridae) based on mitochondrial cytochrome b sequences. *Molecular Phylogenetics and Evolution* 16:366–378.
- Trujano-Álvarez, A. L., and S. T. Álvarez-Castañeda. 2007. Taxonomic revision of *Thomomys bottae* in the Baja California Sur lowlands. *Journal of Mammalogy* 88:343–350.
- Velazco, P. M., and B. D. Patterson. 2013. Diversification of the yellow-shouldered bats, genus *Sturnira* (Chiroptera, Phyllostomidae), in the New World tropics. *Molecular Phylogenetics and Evolution* 68:683–698.
- Velazco, P. M., and N. B. Simmons. 2011. Systematics and taxonomy of great striped-faced bats of the genus *Vampyrodes* Thomas, 1900 (Chiroptera: Phyllostomidae). *American Museum Novitates* 3710:1–35.
- Voss, R. S. 1992. A revision of the South American species *Sigmodon* (Mammalia: Muridae) with notes on their natural history and biogeography. *American Museum Novitates* 3050:1–56.
- Voss, R. S. 2011. Revisionary notes on neotropical porcupines (Rodentia: Erethizontidae) 3. An annotated checklist of the species of *Coendou* Lacépède, 1799. *American Museum Novitates*, 3720:1–36.
- Voss, R. S., M. Gómez-Laverde, and V. R. Pacheco Torres. 2002. A new genus for *Aepeomys fuscatus* Allen, 1912, and *Oryzomys intectus* Thomas, 1921: enigmatic murid rodents from Andean cloud forests. *American Museum Novitates* 3373:1–42.
- Voss, R. S., C. Hubbard, and S. A. Jansa. 2013. Phylogenetic relationships of New World porcupines (Rodentia, Erethizontidae): Implications for taxonomy, morphological evolution, and biogeography. *American Museum Novitates* 3769:1–36.
- Weksler, M., A. R. Percequillo, and R. S. Voss. 2006. Ten new genera of oryzomyine rodents (Cricetidae: Sigmodontinae). *American Museum Novitates* 3537:1–29.
- Williams, D. F., H. H. Genoways, and J. K. Braun. 1993. Taxonomy. Pp. 38–196 in *Biology of the Heteromyidae* (H. H. Genoways and J. H. Brown, eds.). American Society of Mammalogists, Special Publication 10: xii+719.
- Williams, S. L., and H. H. Genoways. 2008. Subfamily Phyllostominae Gray, 1825. Pp. 255–300 in *Mammals of South America. Vol. 1: Marsupials, xenarthrans, shrews, and bats* (A. L. Gardner, ed.). University of Chicago Press 1:xxi: 1–669.

- Wilson, D. E. 1991. Mammals of the Tres Marias Islands. *Bulletin of the American Museum of Natural History* 206:214–250.
- Wilson, D. E., and D. M. Reeder, ed. 2005. *Mammal species of the world. A taxonomic and geographic reference*, Third edition. The Johns Hopkins University Press, Baltimore, vol. 1:xxxv+743, vol. 2:xvii+745–2142 pp.
- Woodman, N. 2012. Taxonomic status and relationships of *Sorex obscurus parvidens* Jackson, 1921, from California. *Journal of Mammalogy* 93:826–838.
- Woodman, N., J. O. Matson, J. J. McCarthy, R. P. Eckerlin, W. Bulmer, and N. Ordóñez-Garza. 2012. Distributional records of shrews (Mammalia, Soricomorpha, Soricidae) from northern Central America with the first records of *Sorex* from Honduras. *Annals of Carnegie Museum* 80:207–237.
- Woods, C.A., and C.W. Kilpatrick. 2005. Infraorder Hystricognathi Brandt, 1855. Pp. 1538–1600 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 2: xvii+745–2142 pp.
- Wozencraft, W. C. 2005. Order Carnivora. Pp. 512–628 in *Mammal species of the world. A taxonomic and geographic reference* (D. E. Wilson and D. A. M. Reeder, eds.), Third edition. The Johns Hopkins University Press, Baltimore 1:xxxv+743 pp.
- Yates, T. L., and J. Salazar-Bravo. 2005. A revision of *Scapanus latimanus*, with the revalidation of a species of Mexican mole. Pp. 489–506 in *Contribuciones mastozoológicas en homenaje a Bernardo Villa* (V. Sánchez-Cordero and R. A. Medellín, eds.). Instituto de Biología, UNAM; Instituto de Ecología, UNAM; CONABIO, México 706 pp.

*Addresses of authors:***JOSÉ RAMÍREZ-PULIDO**

*Universidad Autónoma Metropolitana
Unidad Iztapalapa, División C.B.S.
Departamento de Biología
Apartado Postal 55–535
México 09340, D.F.*

NOÉ GONZÁLEZ-RUIZ

*Universidad Autónoma Metropolitana
Unidad Iztapalapa, División C.B.S.
Departamento de Biología
Apartado Postal 55–535
México 09340, D.F.*

ALFRED L. GARDNER

*USGS–Patuxent Wildlife Research Center
Biological Survey Unit
National Museum of Natural History
Washington, DC 20560–0111*

JOAQUÍN ARROYO-CABRALES

*Laboratorio de Arqueozoología “M. en C. Ticul
Álvarez Solórzano”
Subdirección de Laboratorios y Apoyo Académico
Instituto Nacional de Antropología e Historia
Moneda # 16, Col. Centro
06060 México, D.F.*

