Supplementary Material for

Batrachochytrium salamandrivorans not detected in U.S. survey of pet salamanders

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						Number Imported
Family	Genus	Species	N	Bsal	Bd	2010-2016
.mbystomatidae		gracile	2	0	0	-
		jeffersonianum	2	0	0	=
	Ambystoma	laterale	1	0	0	-
		maculatum	7	0	1	317
		mavortium	8	0	0	-
		mexicanum	18	0	0	1,392
		opacum	5	0	0	3
		talpoideum	1	0	0	-
		tigrinum	65	0	0	50
Plethodontidae	Aneides	aeneus	1	0	0	-
		flavipunctatus	1	0	0	-
		lugubris	2	0	0	-
	Desmognathus	aeneus	2	0	0	-
		folkertsi	1	0	0	-
		ocoee	2	0	0	-
		wrighti	1	0	0	- 2
	Ensatina	eschscholtzii	7	0	0	3
	Eurycea	lucifuga	1	0	0	-
		wilderae 	1	0	0	-
	C	cirrigera	2	0	0	=
	Gyrinophilus	porphyriticus	1	0	0	-
	Hemidactylium	scutatum	2	0	0	-
	Plethodon	cinereus	3	0	0	-
		dunni	5	0	0	=
		glutinosus				-
		shermani	1	0	0	-
		wehrlei	3	0	0	=
		yonahlossee	1	0	0	-
	D 1	serratus	2	0	0	-
	Pseudotriton	ruber	2	0	0	-
	Urspelerpes	brucei	1	0	0	-
Salamandridae	Cynops	chenggongensis	3 42	0	0	2 402
		cyanurus		0	0	3,493
		ensicauda orientalis	16 49	0	1	
			30	0	0	306,176 69,012
	Ichthyosaura	pyrrhogaster	36	0	0	09,012
	Laotriton	alpestris laoensis	1	0	0	30
	Lissotriton	vulgaris	7	0	0	-
	Lissonion	crocatus	15	0	0	325
	Neurergus	kaiseri	28	0	1	90
		strauchii	8	0	0	-
	Notophthalmus	viridescens	17	0	1	469
	11010pmmumus	labiatus	4	0	0	12,162
	Paramesotriton	caudopunctatus	1	0	0	12,102
		chinensis	6	0	0	1,301
		deloustali	1	0	0	8
		hongkongensis	14	0	0	190,011
		nebulosus	2	0	0	170,011
	Pleurodeles	waltl	25	0	2	207
	Salamandra	salamandra	40	0	0	8,509
	Samunana	granulosa	10	0	0	- 0,509
	Taricha	rivularis	7	0	0	-
		sierrae	1	0	0	
		torosa	15	0	0	
	Triturus	carnifex	1	0	0	123
		dobrogicus	29	0	1	-
		ivanbrueschi	4	0	0	
		karelinii	15	0	0	-
		macadonicus	3	0	0	
		marmoratus	24	0	0	16
		pygmaeus	1	0	0	-
	Tylototriton	kweichowensis	11	0	0	1,704
		verrucosus	8	0	0	406
						+00
		yangi	13	0	0	

Supplementary Table S1: All salamander species tested for *Bsal* and *Bd* in U.S. Captive collections. Bolded genera contain species susceptible to *Bsal* and are listed as "Injurious" under the Lacey Act (18 U.S.C. 42).

Supplementary Information: Instructions for swabbing pet salamanders to test for *Batrachochytrium dendrobatidis* and *B. salamandrivorans*

How to swab a salamander for Batrachochytrium salamandrivorans

View an instructional video on how to swab a salamander here: https://www.youtube.com/watch?v=U5h5srxXAaY&feature=youtu.be

Swabs that have been used on listed species under the new ruling must be sent earlier enough to arrive before January 28.

Please check the email that you registered with to obtain the data sheet or email bklocke@masonlive.gmu.edu

- 1) Make sure you have one MW113 swab and empty 1.5ml tube for each animal you plan to swab.
- 2) Pick up the individual salamander from its tank using clean powder-free nitrile disposable gloves, or an inverted new ziplock bag.
- 3) Place the salamander in a clean ziplock bag and gently hold it by its torso, not by its tail. We recommend swabbing the salamander immediately and leaving it in the ziplock bag for no longer than 30 minutes.
- 4) Open the sterile swab packet and remove the swab taking care that the tip does not touch anything other than the salamander or the inside of the ziplock bag. Gently run the swab 10 times back and forth along the belly, five times on each foot, and five unidirectional strokes on the tail starting from the cloaca and gently sliding it towards the tail tip.
- 5) Snap the tip off the swab directly into the 1.5ml tube, allow swab to air dry for 15 minutes once in tube, and then seal it.
- 6) Using a permanent marker label the tube with your name, the date, salamander species and animal number (if you are providing more than one swab).
- 7) Mail the tube and data sheet back to:

Brian Gratwicke

Center for Species Survival

Smithsonian Conservation Biology Institute

MRC 5503 PO Box 37012

Washington DC 20013-7012

Please write down all the information on each tube on a separate sheet of paper and if possible, provide any supplementary information such as where you got the salamander, how long you have had it as a pet, or any other observations about its health.

This project is being conducted by Blake Klocke a student at George Mason University and swabs will be analyzed at the Smithsonian Conservation Biology Institute which is an Amphibian Survival Alliance Partner. bklocke@masonlive.gmu.edu

Supplementary Information: Internet link with basic biosecurity information and registration to receive testing kits for Bd and Bsal

Amphibian Survival Alliance: Be a Salamander Hero

http://www.amphibians.org/salamanderheros/