NICORAEPOA (POACEAE, POEAE), A NEW SOUTH AMERICAN GENUS BASED ON POA SUBG. ANDINAE, AND EMENDATION OF POA SECT. PARODIOCHLOA OF THE SUB-ANTARCTIC ISLANDS¹ Robert J. Soreng² and Lynn J. Gillespie³

Abstract

Poa subg. Andinae Nicora (Poaceae, tribe Poae) is raised to the rank of genus, here renamed as Nicoroepoa Soreng & L. J. Gillespie. Six specific and one infraspecific combinations are made: N. andina (Trin.) Soreng & L. J. Gillespie, N. chonotica (Phil.) Soreng & L. J. Gillespie, N. erinacea (Speg.) Soreng & L. J. Gillespie, N. pugionifolia (Speg.) Soreng & L. J. Gillespie, N. robusta (Steud.) Soreng & L. J. Gillespie, N. subenervis (Hack.) Soreng & L. J. Gillespie, and N. subenervis subsp. spegazziniana (Nicora) Soreng & L. J. Gillespie. Lectotypes are designated for N. pugionifolia and N. subenervis. Taxonomy and nomenclature of similar Southern Hemisphere island species of Poa L. and other related genera are discussed. Poa sect. Parodiochloa (C. E. Hubb.) Soreng is emended to include, in addition to P. flabellata (Lam.) Raspail, P. cookii (Hook. f.) Hook. f., P. foliosa (Hook. f.) Hook. f., P. hamiltoni Kirk, P. ramosissima Hook. f., and P. tennantiona Petrie. The one species of Tzvelevia E. B. Alexeev is reunited with Poa, and the genus is transferred to Poa sect. Tzvelevia (E. B. Alexeev) Soreng & L. J. Gillespie. Morphological and anatomical characteristics of Nicoraepoa, Poa sect. Porodiochloa, and similar genera in Poinae are compared. Leaf blade morphology and anatomy were found to be particularly useful in characterizing Nicoraepoa and related taxa. A key is provided to distinguish Nicoraepoa, Poa sect. Parodiochloo, and morphologically similar perennial genera of Poinae.

Key words: Austrofestuco, Bellardiochloa, Festucella, Hookerochloa, Nicoraepoa, Parodiochloa, Poa, Poaceae, Poinae, Tzvelevia.

Poa subgen. Andinae Nicora (Poaceae, tribe Poeae) was described for a group of robust, tough, gynodioecious species from Patagonia having stolons or rhizomes, long staminodes (1–3 mm) in pistillate plants, smooth or scabrous lemmas with glabrous or pilose keel and nerves, and a callus that is glabrous or with rigid hairs to 3 mm long (Nicora, 1977). Some species also have awns at he lemma apex (e.g., P. andina Trin.; Nicora, 1978; Soreng, pers. obs.). Nicora (1978) included six species in Paa subgen. Andinae (P. andina Trin., P. borchersii Phil. (= P. chonotica Phil.), P. erinacea Speg., P. pugionifolia Speg., P. robusta Steud., P. stepparia Nicora), but retained the morphologically similar P. flabellata (Lam.) Raspail and P. subenervis Hack. in subgenus Paa as the latter

two are perfect-flowered. The lead author thought that the subgenus probably needed to he removed from Poa L. since he first saw material of the group in the early 1980s. With new material from collection trips to Chile in 2000–2001 (by R. and N. Soreng) and Argentina in 2001 (by P. M. Peterson, R. J. Soreng, N. Refulio, and M. Belgrano), the opportunity to study types and other specimens, and the continued study of the large genus Poa (ca. 500+ species), the morphological and anatomical distinctness of Poa subgen. Andinae has become clearer, supporting the conclusion that most species of the subgenus and P. subenervis (placed in Poa subgen. Poa by Nicora, 1978) do not belong in Poa. However, there was always uncertainty about whether the group repre-

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sented a new genus or could logically be placed within an existing genus in subtribe Poinae and, if it was distinct, how best to delimit it.

Several genera of Poinae had to be considered in the decision on how to treat Poa subgen. Andinae. Understanding the relationship of P. flabellata (sect. Paradiochloa (C. E. Hubb.) Soreng) to P. subgen. Andinae species was critical in deciding what genus name to use, because that species had been transferred to a new genus, Parodiochloa C. E. Hubh. (Hubbard, 1981). These groups share morphological features such as thick-ribbed leaf blade adaxial surfaces and awned lemma apices. Other genera morphologically similar to Poa subgen. Andinae, or otherwise putatively allied to Poa (Alexeev, 1976, 1985; García-González, 1983; Clayton & Renvoize, 1986), are Austrafestuca (Tzvelev) E. B. Alexeev s. str., Bellardiochloa Chiov., Festucella E. B. Alexeev, Hoakerochloa E. B. Alexeev, and Tzvelevia E. B. Alexeev. As presently circumscribed, these genera include a total of only nine or 10 species that all, except for Bellardiochloa of southeastern Europe and the Middle East, occur in the Southern Hemisphere, in Australia, New Zealand, Tierra del Fuego, and on sub-Antarctic islands. All of the above genera have heen controversial, and, except for Bellardiochloa, all their species were originally described in Festuca L. or other genera other than Poa and have since been placed in the above new genera and, in most cases, also in Paa (Hooker, 1844-1860; Alexeev, 1976, 1985; Clayton & Renvoize, 1986; Tzvelev, 1989; Jacobs, 1990; Watson & Dallwitz, 1992; Sharp & Simon, 2002; Soreng et al., 2003a; Gillespie & Soreng, 2005). Poa flahellata, for example, had also heen placed in Dactylis L., Festuca, Parodiochloa, and Sesleria Scop.

Recent chloroplast DNA (cpDNA) analyses placed species of Poa subgen. Andinae outside of Poa in a clade with (Soreng et al., 2007) or near (Gillespie & Soreng, 2005; Gillespie et al., 2007) the panarctic genus Arctagrostis Griseb., while ITS data support a clade of these two taxa plus two Australian genera, Festucella and Hoakerochloa (Gillespie et al., in prep.). Hunter et al. (2004) provided the first molecular evidence that the latter two genera, placed in Austrafestuca by Jacobs (1990), were closely related to each other and isolated from Poa, and that Austrafestuca s. str. could be included within Poa. The latter result was not unexpected as A. littoralis (Lahill.) E. B. Alexeev is morphologically quite similar to P. macrantha Vasey. Additional molecular studies (Gillespie & Soreng, 2005; Gillespie et al., 2007; Soreng et al., 2007) and morphological comparisons have corroborated this result, and Austrofestuca is now treated as a monotypic section

of Poa (Gillespie & Soreng, 2005). Parodiachloa, which included the single species Paradiochloa flahellata (Lam.) C. E. Hubb., was also shown to helong within Poa and is now placed in its own section within Poa (Soreng et al, 2003a; Gillespie & Soreng, 2005; Gillespie et al., 2007). Resolution of the generic taxonomy problem remained incomplete, however, as there is also a small group of sub-Antarctic island species of Poa that were thought to he closely related to Paradiachloa (Edgar, 1986), and thus potentially related to Poa subgen. Andinae, and no molecular data were available for these or for the monotypic sub-Antarctic island genus Tzvelevia. Another genus of subtribe Poinae, the Eurasian genus Bellardiochlaa, was previously placed in Poa (Clayton & Renvoize, 1986) or was thought to be closely related (García-González, 1983). Molecular analyses have consistently shown that Bellardiochloa does not helong within Poa (Soreng et al., 1990; Davis et al., 1993; Soreng & Davis, 2000), but it has only recently heen demonstrated that this genus is not a sister to or direct ancestor of either Poa or Paa subgen. Andinae (Soreng et al., 2007; Gillespie et al., 2007).

The present paper outlines the nomenclature and taxonomy of a new genus established here to accommodate *Poa* suhgen. *Andinae*. We discuss how it may be distinguished from *Poa* and other genera of Poeae subtribe Poinae, especially those genera thought to be most closely related. A revised taxonomy is also presented for the morphologically similar *Poa* sect. *Parodiochloa* and *Tzvelevia*, which here is reduced to a section in *Poa*. New data on leaf blade structural characters are presented and discussed here. Gillespie et al. (in prep.) will provide details and discussion of our new molecular analyses, including both plastid and nuclear DNA sequences, which support the taxonomic conclusions of this paper.

Materials and Methods

Specimens, including types, were borrowed from the following herbaria: BAA, CONC, K, MO, SGO, and SI. Additional material was reviewed at BAB, HIP, LE, LP, SI, US, and W. Specimens at US were used for anatomical work. Additional representative specimens are cited as vouchers for geographic ranges of the species cited beyond those ranges vouchered in selected published works. Vouchers for taxa depicted in Figures 1–3 are given in the figure legends. Leaf surfaces were examined and photographed using a Philips XL30 ESEM LaB6 microscope (Eindhoven, The Netherlands) at the National Museum of Natural History, Smithsonian Institution. Sections of blades were taken from about 1 cm above the collar of lower culm leaves of herbarium specimens, attached to

stubs, and sputter coated with 12-18 nm of 60:40 gold:palladium alloy in a Cressington Scientific 108 Auto/SE (Watford, U.K.). Both leaf blade abaxial and adaxial surfaces were examined. Leaf trans-sectional anatomy was examined by cutting rough sections with a handheld razor blade and observing these under a dissecting microscope $(50\times)$ or a compound microscope $(100\times)$.

RESULTS AND DISCUSSION

NICORAEPOA-A NEW SOUTHERN SOUTH AMERICAN GENUS

Poa andina, P. chonotica, P. erinacea, P. pugionifolia, P. robusta, and P. subenervis are wetland plants that combine a set of characters that are either absent, rare, or unusual in other species of Poa. These characters include awned lemmas, a crown of callus hairs, basal sheaths becoming coarsely fibrous in age, ciliate ligules, corrugated adaxial leaf blade surfaces, pungent leaf blade tips, and stout rhizomes. None of the species have all these features, but all share several of them (Table 1). The above species are hereafter referred to as Nicoraepoa Soreng & L. J. Gillespie species. All Nicoraepoa species occur in Patagonia and most are restricted to this area. Only *N*. subenervis subsp. spegazziniana (Nicora) Soreng & L. J. Gillespie extends slightly northward in the moderately high parts of the Andes to 32°04'S (San Juan province, Argentina). One species, N. robusta (Steud.) Soreng & L. J. Gillespie, reaches the Falkland/Malvinas Islands. Nicora (1978) provides an excellent key and illustrations to all these species in Flora Patagónica.

CHARACTERS DISTINGUISHING NICORAEPOA FROM POA

SEM anatomical investigations have advanced our understanding of Poa and surrounding genera and bolstered our opinion that Poa subgen. Andinae deserves generic status. Nicoraepoa differ from most Poa and related genera in the leaf blade adaxial surface (Figs. 1-3). Nicoraepoa leaf blade adaxial surfaces are covered by multiple, regularly spaced, tall, costal ridges that are broadly rounded to squareor rectangle-shaped on top and with intercostal valleys between the ridges that are deep, narrow, and smooth or sparsely to moderately densely scabrous (Fig. 1A-F). Poa (Poa type) leaf blades typically have relatively flat adaxial surfaces with only two central grooves, one on either side of the midrib (Fig. 3E, F) (Lewton-Brain, 1904; Vukolov, 1929; Prat, 1932; Metcalfe, 1960; Hernández Cardona, 1978; García-González, 1983). Costal ridges are present in Poa, but, with few exceptions, these are of low relief, narrow, and with intercostal zones

several times wider (Fig. 3E, F). Corrugated adaxial leaf blade surfaces like or approaching those in *Nicoraepoa* are rare in *Poa* but do occur in the following species: *P. cookii* (Hook. f.) Hook. f., *P. flabellata*, *P. foliosa* (Hook. f.) Hook. f., *P. hamiltonii* Kirk, *P. macrantha*, and *P. tennantiana* Petrie (Figs. 2E, F, 3A–C, H). *Nicoraepoa* species lack papillae on both long cells and short cells (Fig. 1A–F), whereas *Poa* species with costal/intercostal topography like or approaching *Nicoraepoa* (Figs. 2A–F, 3A–D) commonly have papillate long cells (Figs. 2E, F, 3A–C).

Nicoraepoa and Poa also differ in leaf blade internal anatomy and blade apex, ligule, and rhizome morphology, in addition to leaf adaxial-surface morphology. In leaf blade transections of *Nicoraepoa*, the adaxial ridges are underlain by sclerenchyma caps, or hy prominent "T"- or "I"-shaped sclerenchyma girders connecting the adaxial surface through the primary vascular bundles to the ahaxial surface (Tables 1, 2). Such broad caps or flared-topped girders have not previously been observed in Poa to our knowledge (Lewton-Brain, 1904; Vukolov, 1929; Prat, 1932; Metcalfe, 1960; Hernández Cardona, 1978; García-González, 1983). The apices of the blades in *Nicoraepoa* are pungent and are not or only obscurely prow-shaped, whereas in Poa they are commonly narrowly to broadly prow-shaped and are rarely pungent. In Poa, hulliform cells (visible in cross sections) occur only in the central two grooves; this is typical of Poinae genera with only two central grooves, including Hookerochloa and Tzvelevia. For those species with multiple broad ridges, we have seen bulliform cells only in the grooves immediately adjacent to the midrib, but more detailed anatomical cross-sectional work is needed to confirm this.

Nicoraepoa ligule apices (Figs. 4C, I, Q, 5B, F, K) are truncate (sometimes slightly raised on the outer margins; Fig. 5B), and ciliolate to ciliate (hairs 0.15–0.6 mm long), whereas in Poa ligule apices vary in shape from truncate to acuminate, while the margins are commonly smooth or scabrid, or infrequently ciliolate. Nicoraepoa species produce thick rhizomes that are mostly unidirectional. Many species of Poa have rhizomes, but apart from the few species of Poa subgen. Arctopoa (Griseb.) Prob., these rhizomes generally are more slender and sometimes have multidirectional shoots (Serebryakova, 1965).

Floret characters distinguishing *Nicoraepoa* and *Poa* include lemma shape, awn presence, and callus hair distribution and form. *Nicoraepoa* have lemma apices that are sharply pointed, and some are terminated by distinct slender scahrous awns up to 5 mm long. *Poa* typically have slightly hlunt lemma apices that lack awns (except in *P. flabellata*). In

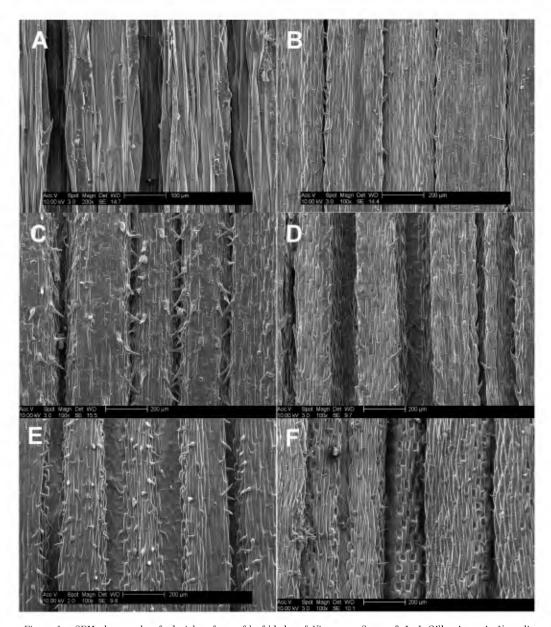


Figure 1. SEM photographs of adaxial surfaces of leaf blades of Nicoraepoa Soreng & L. J. Gillespie. —A. N. andina (Trin.) Soreng & L. J. Gillespie. Chile, Bío-Bío, R. J. & N. L. Soreng 7182 (US). —B. N. chonotica (Phil.) Soreng & L. J. Gillespie. Chile, Los Lagos, R. J. & N. L. Soreng 7236 (US). —C. N. pugionifolia (Speg.) Soreng & L. J. Gillespie. Chile, Magallanes, R. J. & N. L. Soreng 7336 (US). —D. N. robusta (Steud.) Soreng & L. J. Gillespie. Chile, Magallanes, R. J. & N. L. Soreng 7359 (US). —E. N. subenervis (Hack.) Soreng & L. J. Gillespie subsp. subenervis. Chile, Magallanes, R. J. & N. L. Soreng 7334 (US). —F. N. subenervis subsp. spegazziniana (Nicora) Soreng & L. J. Gillespie. Chile, Metropolitana, R. J. & N. L. Soreng 7167 (US).

Nicaraepaa, the calluses of the lemmas are glabrous (P. robusta only) or have a row of slightly curved hairs distributed around the base of the lemma. This row of callus hairs is called a crown, as distinct from a beard (commonly used to describe elongated calluses with short stiff hairs across and down the surface) or a web (an isolated dorsal tuft of hairs known only in the

genus Paa). A few Paa species have callus hairs in a crown (e.g., P. eminens J. Presl [subgenus Arctapoa], P. secunda J. Presl [section Secundae V. L. Marsh ex Soreng], and P. abvallata Steud. [section Diaicapaa E. Desv.]), but Paa species typically have a single dorsal tuft (a "web") of woolly, or plicate (in section Diaicopoa), hairs. Some species of Paa with webs

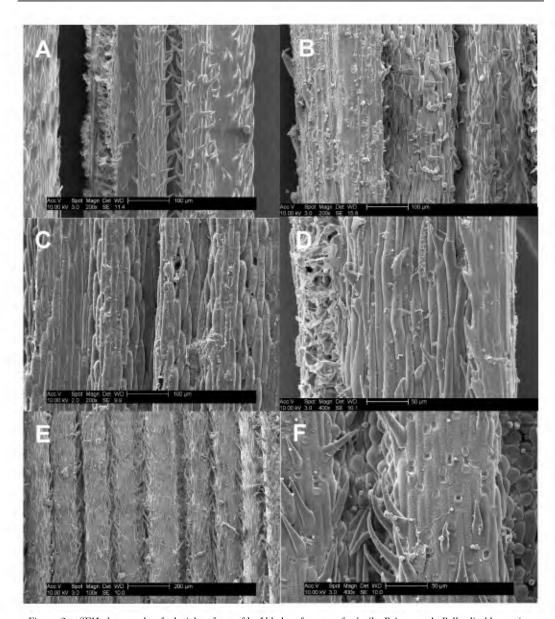


Figure 2. SEM photographs of adaxial surfaces of leaf blades of genera of subtribe Poinae. —A. Bellardiochloa variegata (Lam.) Kerguélen. Left side abaxial epidermis and margin. Greece, Peloponnese, R. J. & N. L. Soreng & L. J. Gillespie 7519 (US). —B. Festucella eriopoda (Vickery) E. B. Alexeev. Left side abaxial epidermis and margin. Australia, New South Wales, E. N. McKie 7449 (US 2010746). —C. Hookerochloa hookeriana (F. Muell. ex Hook. f.) E. B. Alexeev. Australia, Tasmania, R. A. Black (US 1937360). —D. Poa kerguelensis (Hook. f.) Steud. (sect. Tzvelevia (E. B. Alexeev) Soreng & L. J. Gillespie). Right side abaxial epidermis and margin. Indian Ocean, Kerguelen Island, Wilkes Expedition (US 653666). —E. Poa flabellata (Lam.) Raspail (sect. Parodiochloa (C. E. Hubb.) Soreng). South Georgia, Admr. Berggren (US 2042368). —F. Poa flabellata. Close-up of costal intercostal region with papillate cells in intercostal region. South Georgia, Admr. Berggren (US 2042368).

have additional tufts of hairs emerging from the callus at the base of the marginal veins of their lemmas.

Reproductive characteristics also distinguish *Nicaraepaa* from most *Paa* (Nicora, 1977, 1978). All *Nicaraepaa* species, except *N. subenervis*, are gynodioecious with staminodes 1–3 mm long in pistillate

flowers (Fig. 4G, N). In contrast, *Poa* species are mostly perfect-flowered or, when diclinous, they generally have pistillate flowers with rudimentary staminodes 0.1–0.2 mm long (Nicora, 1978; Anton & Connor, 1995; Soreng & Keil, 2003; for exceptions, see Edgar & Connor, 2000; Negritto & Anton, 2000).

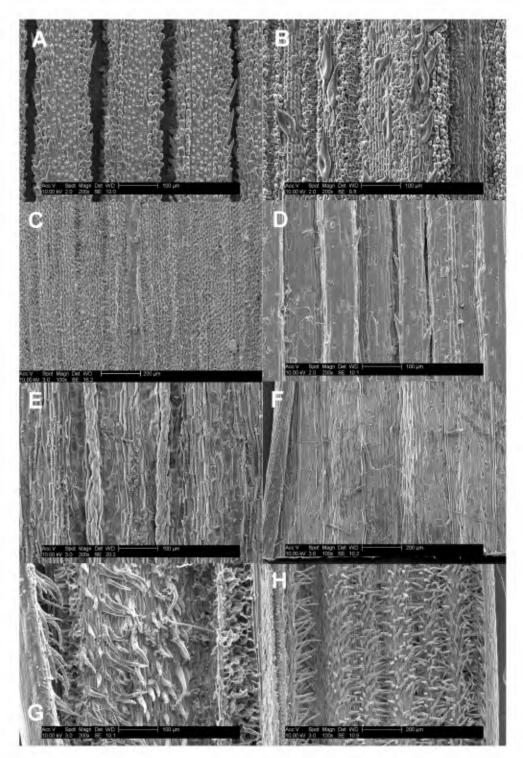


Figure 3. SEM photographs of adaxial surfaces of leaf blades in Poa L. A–D. Poo. sect. Parodiochloa (C. E. Hubb.) Soreng. —A. P. cookii (Hook. f.) Hook f. Indian Ocean, Marion and Prince Edward Islands, B. J. Huntly (US 3000037). —B. P. folioso (Hook. f.) Hook f. New Zealand, The Snares, F. L. S. Demedin s.n. [hb. D. Petrie] (US 947494). —C. P. ramosissima Hook. f. Midrib just left of center. New Zealand, Campbell's Island, J. H. Sorensen (US 2184040). —D. P. tennantiana Petrie. New Zealand, The Snares, B. C. Aston [hb. 31857] (US 2044107 (ex hb. Cheeseman)). —E. P. alpina L. (sect. Alpinae Stapf).

ADDITIONAL TAXONOMIC AND MORPHOLOGICAL CONSIDERATIONS CONCERNING PARTICULAR SPECIES

Nicora (1977, 1978) excluded *Poa subenervis* from her subgenus *Andinae* because it has perfect flowers. However, we consider it to helong in *Nicoraepoa* because it shares the stout rhizomatous habit, the adaxial leaf surface anatomy of hroad ridges and narrow valleys (Fig. 1E, F), sharply pointed leaf apices, ciliate ligules, and calluses with a crown of hairs (Figs. 4E, L, 5D, M, N). DNA analyses from both nuclear and plastid genomes support this relationship (Gillespie et al., 2007; Gillespie et al., in prep.).

Paa stepparia, although placed in Paa subgen. Andinae by Nicora (1977, 1978), needs further study before it can be placed in the new genus. Leaf blade adaxial surfaces of this species are of the Poa type with ridges that are only indistinctly raised. The leaf blade apices are pungent, the lemmas lack awns, calluses are glabrous or the marginal vein and keel hairs run into the callus, and the ligules are slightly raised on the sides with somewhat asperous margins. This species was thought to be related to P. acinaciphylla E. Desv. (Nicora, 1978), and Soreng et al. (2003a) placed both these species and P. planifolia Kuntze in Poa sect. Acutifoliae Pilg. ex Potztal. We concur with M. Negritto (pers. comm.) that P. plicata Hack. (type and sole species of Poa sect. Plicatae Pilg. ex Potztal) also belongs to this section and place Poa sect. Plicatae in synonymy of Poa sect. Acutifoliae (see Taxonomic Treatment section). Analyses of chloroplast and nuclear ITS data confirm P. acinaciphylla as a member of Poa s. str. (Gillespie & Soreng, 2005; Gillespie et al., 2007; Gillespie et al., unpublished data). Poa stepparia, however, is only known from the type and two other collections (Nicora, 1978), and no material has been available for molecular or SEM analyses; its relationships and breeding system need confirmation.

RELATIONSHIPS OF NICORAEPOA

Initial cpDNA results from trnT-trnF (Gillespie & Soreng, 2005; Gillespie et al., 2007), plus new nuclear ribosomal ITS data (Gillespie et al., in prep.), show that the Nicoraepoa taxa align in a clade outside of

Poa and can logically be placed in a separate genus as they do not fall within any other genus. Species sampled for DNA to date are N. andina, N. chanotica, N. pugionifolia, N. robusta, and N. subenervis. Preliminary results resolve Arctagrostis, Festucella, and Hookerochloa as the closest genera to Nicoraepoa. The relationship of this set of genera to other Poinae genera, Arctophila (Rupr.) Rupr. ex Andersson, Bellardiochloa, Dupontia R. Br., and Poa (including Anthochloa Nees & Meyen, Austrofestuca, Dissanthelium Trin., Eremopoa Roshev., Neuropoa Clayton), is unsettled. Molecular relationships among genera of subtribe Poinae and intermixed genera of Poeae subtribes Alopecurinae and Miliinae are complex and are not yet well resolved or understood. Further sampling is needed before firm conclusions about exact relationships can be drawn (Soreng & Davis, 2000; Gillespie & Soreng, 2005; Soreng et al., 2007; Gillespie et al., 2007; Gillespie et al., unpublished data).

Genera of Poinae thought to be related to Nicoraepoa and Poa are discussed below, and their morphological characteristics, including leaf hlade adaxial surface (Figs. 2A-F, 3G), are compared. Festucella and Hookerochloa are two closely related genera of Australia and New Zealand. Although sometimes treated within Austrafestuca (Clayton & Renvoize, 1986; Jacobs, 1990; Sharp & Simon, 2002), recent molecular data support their separate recognition (Hunter et al., 2004; Gillespie et al., 2007). With Nicoraepoa, they share wetland habitats, fibrous basal sheaths, awned lemmas, and a short crown of hairs around the callus, but their leaf blades are Poa-like in adaxial anatomy (Fig. 2B, C). The relationships among the Arctic genus Arctagrastis and the Southern Hemisphere genera Nicoraepaa, Festucella, and Hookerochloa detected in our molecular analyses need further investigation. ITS data place Arctagrostis plus Nicoraepoa as sister to Festuca plus Hookerochloa, but relationships detected with our plastid data are less well resolved at this point. The one-flowered spikelets of Arctagrastis are impossible to confuse with the several-flowered spikelets of the other three genera. Its broad, flat leaf blades with narrow, low costal ridges and broad intercostal zones (not shown) do not resemble the blades of these genera.

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Close-up of central region of blade with rows of hulliform cells (protruding here in the intercostal regions) flanking either side of the midvein. Canada, British Columbia, R. J. & J. E. F. Soreng 5991 (US). —F. P. pratensis L. (sect. Poa). Narrow costal regions with elongated hairs characteristic of the species. Canada, British Columbia, R. J. & J. E. F. Soreng 5987 (US). —G. P. billardierei St.-Yves (sect. Austrofestuca (Tzvelev) Soreng & L. J. Gillespie). New Zealand, South Island, V. D. Zotov (US 2010241). —H. P. macrantha Vasey (sect. Madropoa Soreng). Showing the presence of a more corrugated costal-intercostal surface, a rare exception in Poa (and P. macrantha). U.S.A., Oregon, R. J. Soreng 2959 (US).

Table 1. Morphological comparison of species of Nicoruepou and morphologically similar Poinae genera (except Pou, which is given in Table 2).

	Arctophila fulva Nyman	Bellardiochloa variegata	Dupontia fisheri R. Br.	Festucella eriopoda	Hookeriana hookeriana	Nicoraepoa andina	Nicoraepoa chonotica	Nicoraepoa erinacea	Nicoraepoa pugionifolia	Nicoraepoa robusta	Nicoraepoa subenervis
Habitat	arctic tundra and northerm boreal wetlands, shallow ponds, marshy meadows, swamps, and quiet streams	grassy alpine meadows, commonly over setlimentary rocks	arctic tundra wetlands, mainly coastal, marshy meadows, marginally sub- saline tolerant	montane forest openings nea werlands	nontane rr wetlands	alpine and subalpine riparian and seeps	montane to sea level wetlands, marshes, and lake shores	inland sub-saline flats	inland seasonal of wetlands, and sub-saline flats and seeps	coastal saline I to sub-saline marshes and seeps	montane sub- saline wetlands, about seeps and springs
Habit, culm height	rhiz	densely tufted, 0.09-0.6 m	rhizomatous, 0.05–0.8 m	low tuffs, culms large tuffs, 0.50–1.6 m culms 0.55–1.	85 m	tuffs from a stout little branched rhizome, to 1 m	tufts from a stout little branched rhizome, to 2 m	low tuffs from 1 stout little branched rhizomes, 0.07-0.15 cm	ow tufts from stout little branched rhizomes, 0.2-0.3 m	tuffs from a stout little branched rhizome, to 1 m	low tufts from stout little branched rhizomes, 0.15-0.6 m
Basal sheaths	chartaceous, smooth, retrorsely strigose, to finely fibrous with cross- vein	chartaceous, weakly fibrous	chartaceous, or a bit fibrous, smooth, glabrous to sparingly retrousely strigose	fibrous	fibrous	fibrous, smooth fibrous, smooth		somewhat fibrous, smooth	inmewhat fibrous, smooth	fibrous, smooth somewhat fibrous smooth	somewhat fibrous, smooth
Blade adaxial lateral surface topography	rounded tall to medium height ridges 0.25-1× wider than valleys (similar to abaxial surface)	rounded tall to medium height, medium rounded ridges height ridges 1X wider than 0.25–1X valleys, valleys wider than open valleys (similar to abaxial surface)	flat or faintly undulating	flat or faintly undulating, obscure ridges	flat	ridges 2–4× vider than valleys, valleys closed	rall flat-topped tall flat-topped flat ridges 2-4× ridges 3-5× wider than wider than valleys, valleys, valleys closed valleys closed	flat -	tall flat-topped tall flat-topped middle height ridges 2–3× ridges 2–3× rounded wider than ridges 1–2 valleys, valleys, valleys valleys closed valleys closed valleys or closed or closed or closed valleys or closed valleys.	ridges 2-3× wider than valleys, valleys closed	middle height rounded ridges 1–2× wider than valleys, valleys open or closed

Table 1. Continued.

	Arctophila fulva Nyman	Bellardiochloa variegata	Dupontia fisheri R. Br.	$Festucella \ eriopoda$	Hookerochloa hookeriana	Nicoraepoa andina	Nicoraepoa chonotica	Nicoraepoa erinacea	Nicoraepoa pugionifolia	Nicoraepoa robusta	Nicoraepoa subenervis
Blade lateral adaxial surface vestiture	smooth, glabrous	ridge tops and sides coarsely hispidulous	smooth or moderately scabrous, glabrous	short stout hooks common to very common, glabrous	short stout hooks common over VBs, glabrous	ridge sides with slender hooks occasional, glabrous	ridge sides and sometimes tops with minute hooks common, glabrous	fairly densely hispidulous- strigulose	ridge sides with hooks common, ridge tops with short hooks occasional,	ridge sides and sometimes tops with short hooks common, glabrous	ridge sides with short hooks occasional to common, glabrous
Girdering	I-shaped, thick, present in primaries, keel, and most secondaries, keel girdered abaxially thick, round in cross section	I-shaped, thick, I-shaped, thin, or present in none often primaries, discontinuous, with keel, and narrow caps over most VBs and low secondaries, mounds below, keel girdered, keel VB continuous abaxially to lower side and thick, round with an isolated in cross cap above section	I-shaped, thin, usually continuous, but quite narrow and sometimes broken just s above the VBs (Jacunae below adaxial cpidermis)	discontinuous, Iow basal mounds only	I-shaped primaries, secondaries discontinuous to adaxial side	T-shaped primaries and keel	T-shaped primaries and keel	N/K	caps below ridges, no girders, no marrow isolated U below keel	T-shaped I- primaries and keel	L-shaped, narrow, d present in primaries, keel, and some secondaries
Abaxial sclerenchyma along epidermis	discontinuous 1a	discontinuous	discontinuous	discontinuous	discontinuous	discontinuous	discontinuous	discontinuous? discontinuous	discontinuous	discontinuous	discontinuous
Vascular bundles appearing at adaxial surface	10 to 17/side, alternating primaries and slightly smaller secondaries	3 to 4/side id	5 to 8/side	2/side	6/side (12/side 10 to 11/side abaxially)		24/side	2/side	5/side	7/side	5 to 12/side
Ligule texture, color	membranous, white or bronzy	membranous, white	membranous, bronzy	chartaceous, white	chartaceous, white	membranous- chartaceous, white	membranons- chartaceous, white	membranous- chartaceous, white	membranous- chartaceous, white	membranous- chartaceous, white	membranous, white
Ligule length	2-6(8) mm	2–7 mm	1-2.5 mm	1–3 mm, 2–7 mm on	2-6 mm	1–3 mm	1–3 mm	0.5 mm	0,5-2.5 mm	0.8-1.5 mm	1-1.5 mm

Table 1. Continued.

Arctophila fulva Nyman	Bellardiochloa variegata	Dupontia fisheri R. Br.	Festucella eriopoda	Hookerochloa hookeriana	Nicoraepoa andina	Nicoraepoa chonotica	Nicoraepoa erinacea	Nicoraepoa pugionifolia	Nicoraepoa robusta	Nicoraepoa subenervis
30	smooth or somewhat scabrous, glabrous	smooth, glabrous	with short hooks common, glabrous	with short hooks frequent, glabrous	hispidulous	densely covered smooth or with with short sparse short thick stiff hairs appressed hairs to lisnitulous	smooth or with sparse short stiff hairs	densely covered densely with hooks to with short hisp short thick thick hairs appressed hairs hairs to hispitulious	Jensely covered with short thick appressed hairs to hisroidulous	densely hispidulous
35	smooth or scabrous	smooth, glabrous scabrous	scabrous	smooth or sparsely seabrid	hispidulous	hispidulous	hispidulous	hispidulous	hispidulous	ciliate
open lax, 5— cc 30 cm	contracted or loosely contracted, 5–25 cm	contracted or n open panicle, 3–18 cm	ореп, 4.5–40 ст	open, 18–24 cm	contracted, interrupted, 5–20 cm	contracted and interrupted to open lax, 5-30 cm	contracted, narrow, 2.5- 4.5 cm	contracted, narrow, 3-	contracted, or in viviparous forms loosely contracted	contracted narrow panicle or sometimes lax loose
q	branches densely scabrous	smooth	densely scabrous angled	densely scabrous angled		smooth	densely ciliate scabrid angled	moderately scabrid angled	smooth	smooth
ent or sl occasionally with a cusp to 0.8 mm long	slender cusp or awn 0.5-1.5 mm long, from the apex or a short notch	absent or occasionally with cusp or thin awn up to 0.6 mm long	apiculate to slender awn (0–)0.5–1.5 (–2) mm long, from a minute notch	slender awn s 0.5-2(-3) mm long, from a minute	short slender awn 0.5– 3(–5) mm long	apiculate	apiculate	absent to appropriate	absent to apiculate	apiculate
n crown of w straight hairs around the base, hairs 0.3–0.7 mm	with crown of with crown of hairs straight hairs 0.3–0.6 mm long around the base, hairs 0.3–0.7 mm long	with crown of coarse hairs 0.5–1.2 mm long	with crown of hairs 0.4— 0.6 mm long	with crown of hairs 0.3– 0.6 mm long	with crown of slender hairs 2-3 mm long	with crown of slender hairs 1-2.5 mm long	with crown of hairs 0.2– 0.3 mm long	glabrous	glabrous	with crown of hairs 0.4– 1.5 mm long
Lodicule length, with a lateral ir shape, and lobe margin	irregularly dentate or usually with with 1 to 2 slender a lateral lobes in addition to main slender lobe, glabrous or ciliate	usually with a lateral lobe	0.5 mm, lobed as in a mitten, or shouldered at midpoint, glabrous	0.8 mm, lobed 0.9–1 mm, as in a mitten lanceold or shouldered irregula at midpoint, slender glabrous or lobed, with 1 or 2 glabrous cilia with a c	ate to rly s or	1.5 mm, broadly N/K lanceolate glabrous	N/K	irregularly irregularly lobed or shouldered, glabrous	1.7 mm, broadly lanceolate glabrous	0.5–0.8 mm, lanceolate, irregularly toothed, glabrous

Table 1. Continued.

Bellard varie	Bellardiochloa variegata	Dupontia fisheri R. Br.	Festucella eriopoda		Hookerochloa Nicoraepoa hookeriana andina	Nicoraepoa chonotica	Nicoraepoa erinacea	Nicoraepoa pugionifolia	Nicoraepoa robusta	Nicoraepoa subenervis
cyl	fusoid, sub-cylindrical fusoid,	1soid,	fusoid,	fusoid,	fusoid,	fusoid,	N/K	fusoid,	fusoid,	fusoid,
agol	to subtriagonous,	subtriangular,	subtriangular,	, subtriangular	, subtriangular	subtriangular, subtriangular, subtriangular, subtriangular,		subtriangula	subtriangular, subtriangular, subtriangular,	subtriangular,
idis	sulcus indistinct to	with an	sulcus	sulcus	sulcus	sulcus		sulcus	sulcus	sulcus
ely	moderately distinct	indistinct	distinct	distinct	distinct	distinct		distinct	distinct	distinet
		sulcus								
, 1	8-1/6	.5-2.5 mm long,	1.8-2.5 mm, 1/8-1/6 1.5-2.5 mm long, 3 mm, 0.6 mm, 3.8 mm,	3.8 mm,	2 mm, 0.4 mm,	2 mm, 0.4 mm, 3 mm, 0.8 mm, N/K	N/K	3 mm, 0.4 mm, 3.4 mm,	3.4 mm,	2.2 mm, 0.4 mm,
length of grain.	in.	1/6-1/5 length	narrowly	0.8 mm,	elliptical	narrowly		elliptical	0.6 mm,	elliptical to
Ind	ovoid to punctiform	of grain,	elliptical	elliptical		elliptical			narrowly	broadly
		broadly ovate							elliptical	elliptical
present, firm to		N/K	present, firm	present, firm	present, firm	present, firm	N/K	present, firm to present, firm		present, firm
semi-soft								semi-soft		
Styles, stigma bases with a gap,		bases with a	bases with a	bases with a	bases with a	bases with a	bases with a	bases with a	bases with a	bases with a
gap, plumose plumose $= 42, 63 2n = 14$	2	gap, plumose $2n = 42, 44, 88$.	\geq	gap, plumose N/K	gap, plumose N/K	gap, plumose N/K	gap, plumos N/K	e gap, plumose N/K	gap, plumose K N/K N/K N/K N/K N/K N/K N/K N/K N/K N	gap, plumose N/K
		132								

VB = vascular bundles; N/K = not known.

Table 2. Morphological comparison of species of Poa sect. Parodiochloa and morphologically similar Poa sections.

	Poa (subg. Arctopoa) sect. Aphydris (Griseb.) Tzvelev	P. (subg. & sect. Arctopoa) eminens	P. (sect. Austrofestuca) billardierei	P. (sect. Parodiochloa) cookii	P. (sect. $Parodiochloa$) $flabellata$	P. (sect. $Parodiochloa$) $foliosa$	P. (sect. $Parodiochloa$) $hamiltonii$	P. (sect. $Parodiochloa$) $ramosissima$	P. (sect. Parodiochloa) tennantiana	P. (sect. $Tavelevia$) $kerguelensis$	Poa (other than those separated)
Habitat	alpine, riparian and sub-saline seasonally wet flats, sub- saline springs	coastal, wet meadows, rwcky beaches, sub-saline	coastal sand dunes and sandy to shingly flats	coastal, among rocks and peat mountls, and on slopes, often associated with penguin	coastal, bluffs, omeadows and rocky ledges	coastal slopes, sometimes coastal strand turfy meadows	coastal, among rocks and peat mounds, and on slopes, often associated with penguin colonies	coastal, wet rocks and cliffs, commonly associated with bird colonies	nargins, nargins, clearings in scrub, and on banks	rocky oceanic island slopes 100–400 m	all cool temperate to arctic habitats, generally intolerant of saline conditions
Habit, culm height	rhizomatous or rhizomatous, infrequently to 1 m stooling, to 1.15 m	rhizomatous, to 1 m	dense tufts from of deeply buried culms which may appear like rhizomes, to 0.6 m	obust dense tussoeks, to 0.5 m	massive tussocks, to 1 m	massive tussocks dense tussocks, weak culmed, with short thick to 0.5 m stolonieron stolons, to culms 1.5 m prostrate,	dense tussocks, to 0.5 m	si 5	tufts from stout rhizomes, to 1 m	tuffs from stout dense, low tuffs, pererurial or rhizomes, to to 0.15 m infrequent 1 m annual, the rhizomate stooling	perennial or infrequently annual, tufted, rhizomatous, or stooling
Basal sheaths	fibrous and chartaceous, smooth, retrorsely strigose	chartaceous, smooth, retrorsely strigose	glabrous smooth,	fibrous, pectinate ciliate between ridges	fibrous, f pectinate ciliate between ridges	fibrous, smooth,	fibrous, smooth, papillate, glabrous	s	fibrous, pectinate ciliate between ridges	finely fibrous,	fibrous or rarely becoming fibrous or rarely becoming fibrous, smooth or finely scabrous particularly along ridges, glabrous or infrequently bisesidulous is the particular or infrequently bisesidulous or infrequently bi
Blade adaxial lateral surface topography	flat	flat or slightly undulate	flat or faintly undulating	tall flat-topped ridges 3-4× wider than valleys, valleys closed	rail flat-topped low to medium low flat-topped ridges 3-4× height ridges 1-2× wider than valleys, ridges, 1-2× valleys, valleys, and shallow valleys open (abaxial similarly)	indees 1–2× ridges 1–2× wider than valleys, valleys closed or open and shallow	low flat-topped low flat-topped flat ridges 1–2× ridges 1–2× wider than wider than valleys, valleys, valleys valleys valleys a slightly open slightly open	ow flat-topped ridges 2-3× wider than valleys, valleys open	low flat-topped ridges 1–2× wider than valleys, valleys slightly open	Tja Lja	flat or infrequently ridges slightly raised, ridges 0.2–1× wider than valleys, valleys open

Table 2. Continued.

<i>Poa</i> (other than those separated)	scabrid, or infrequently generally scabrid or hispid puberulent or pilulose, rarely papillate	discontinuous, sometimes discontinuous above or below, primaries and secondaries common	discontinuous or rarely continuous
Poa (a	scabrid scabrid infrequ general scabrid hispid puberu pilulos papillal	I-sk	discontinu rarely contin
P. (sect. Traeleria) kerguelensis	short stout hooks frequent, glabrous	discontinuous, Iow basal mounds only, isolated keel U capped below	discontinuous
$P.~({ m sect}.$ $Parodiochloa)$ $tennantiana$	sparsely papillate, rarely minutely scabrid on ridges, glabrous	I-shaped primaries, thick, secondaries mostly continuous, keel VBs girdered above with mound connected below	discontinuous
P. (sect. Parodiochloa) ramosissima	valleys to ridge margins densely papillate, glabrous	I-shaped primaries, continuous between VBs except in keel, keel, with isolated mound below	discontinuous
P. (sect. $Parodiochloa$) $hamiltonii$	valleys to ridge margins densely papillate, glabrous	T-shaped primaries slender, to I- shaped secondaries	discontinuous
P. (sect. Parodiochloa) foliosa	valleys to ridge margins densely papillate, ridge tops with thick bodied hooks common, glabeous	I-shaped primaries, thick, secondaries free, continuous between VBs in primaries, secondaries free above and below VBs, keel girdered above with isolated mound below	discontinuous
$P.\ ({ m sect}.$ $Parodiochloa)$ flabellata	valleys papillate, ridge sides with slender hooks frequent, glabrous	I-shaped primaries, thick hourglass shape, secondaries slightly narrower, continuous between VBs in primaries and secondaries, keel isolated U shape below	discontinuous
P. (sect. $Parodiochloa$) $cookii$	walleys to ridge margins densely papillate to minutely scabrid, sometimes with additional larger slender hooks occasional, glabrous		discontinuous
P. (sect. Austrofestuca) billardierei	densely coarsely scabrous to stiffly puberulent	I. or A-shaped VB. primaries, and secondaries progressively reduced to margin, narrowest at contact of adaxial epidermis, keel thickest	continuous, thickest below VBs
P. (subg. & sect. Arctopoa)	glabrous	I-shaped primaries, narrow I-shaped secondaries, tertiaries without girders	discontinuous
Poa (subg. Arctopoa) sect. Aphydris (Griseb.) Tzvelev	smooth, glabrous	L-shaped primaries primaries	discontinuous
	Blade lateral adaxial surface vestiture	Girdering	Abaxial sclerenchyma along epidermis

Table 2. Continued.

	Poa (subg. Arctopoa) sect. Aphydris (Griseb.) Tzvelev	P. (subg. & sect. Arctopoa)	P. (sect. Austrofestuca) billardierei	P. (sect. Parodiochloa) cookii	$P.\ ({ m sect}.$ $Parodiochloa)$ $flabellata$	P. (sect. Parodiochloa) foliosa	P. (sect. Parodiochloa) hamiltonii	P. (sect. Parodiochloa) ramosissima	P. (sect. $Parodiochloa$) $tennantiana$	P. (sect. $Txrelevia$) $kerguelensis$	Poa (other than those separated)
Vascular bundles appearing at adaxial surface	5 to 7/side	8 to 9/side apparent, but 18 to 19/side including tertiaries	5/side	16 to 21/side, primaries mainly	10 to 17/side, primaries mainly	12 to 17/side, primaries mainly	6 to 15(21?)/ side, primaries and secondaries	5 to 6/side	15 to 17/side, secondaries frequent	3/side	2 to 11/side
Ligule texture, color	membranous to membranous- membranous- chartaceou chartaceous, yellow-cre off-white to to brown yellowish or	am	chartaceous laterally, membranous- chartaceous centrally, white	chartaceous- membranous, white	chartaceous basally to membranous apically,	chartaceous, white	membranous- chartaceous, white	membranous, white	chartaceous basally to membranous apically,	membranous, white	membranous, white or clear
Ligule length Ligule abaxial surface	1–4(5.5) mm scabrous, glabrous	1-3.5 mm smooth, glabrous	0.7-1 mm moderately scabrous, glabrous	4-7.5 mm smooth, glabrous	6–23 nm with occasional short hooks, glabrous	1–3 mm densely covered with short thick appressed hairs (finely scabrous, f. E.Jone 1095).	4-7.5 mm smooth, glabrous	(1.5)2.5-4 mm densely papillate, glabrous	6–16 mm scabrid to puberulent	ca. 3 mm with occasional short hooks, glabrous	0.2-18 mm smooth, scabrous, or short puberulent
Collar margins	smooth, glabrous or ciliate	smooth, glabrous	smooth, glabrous	smooth, glabrous	smooth, glabrous	smooth, glabrous smooth,	smooth, glabrous	smooth to papillate	smooth to ciliate	smooth or sparsely scabrid,	glabrous or infrequently puberulent
Inflorescence shape, length	open or contracted, 5–35 cm	contracted, 8-30 cm	narrowly contracted, (3.5–)6–25 cm	contracted, 5–25 cm	ightly contracted, dense, branches and axis somewhat contorted, 5-20 cm	contracted, fairly contracted, dense, 10- 5-25 cm 20 cm	contracted, 5-25 cm	narrow, loosely loosely contracted, cont (2)4-5(10) cm 9-16	loosely contracted, n 9-16 cm	giabrous narrowly contracted, 2-4 cm	tightly contracted to wide open and lax

Table 2. Continued.

	Poa (subg. Arctopoa) sect. Aphydris (Griseb.) Tzvelev	P. (subg. & sect. Arctopoa)	P. (sect. Austrofestuca) billardierei	P. (sect. Parodiochloa) cookii	P. (sect. Parodiochloa) flabellata	P. (sect. $Parodiochloa$) $foliosa$	P. (sect. $Parodiochloa$) $hamiltonii$	P. (sect. $Parodiochloa$) ramosissima	P. (sect. $Parodiochloa$) tennantiana	P. (sect. $Tznelevia$) $kerguelensis$	Poa (other than those separated)
Panicle branches smooth to densel, scabro angled	densely scabrous angled	sparsely seabrous	smooth below, coarsely spreading scabrous all around distally and on pedicels (rarely sparsely so)	smooth, densely smooth papillate, sometimes coarsely scabrous, the hooks spreading (and then papillae obscure)	smooth	a few hooks	smooth, densely smooth, densely occasionally papillate papillate commonl sparsely scabrous above an	smooth, densely papillate	oceasionally smooth, commonly sparsely scabrous above and on pedicels	densely scabrid s on and less so between angles	densely seabrid smooth or seabrous on and less so (rarely between papillate), angles hooks diffuse or more often in lines above vascular traces, glabrous or rarely hispidulous
Lemma awn	absent	absent	absent	cuspidate	cuspidate to short stout	cuspidate	cuspidate	cuspidate	cuspidate	absent or cuspidate	absent or rarely cuspidate
Callus	glabrous, or with 1 to several straight hairs (to 1.5 mm long) dorsally and below the marginal	usually with a line of 1–2 mm long slightly simous hairs around base of proximal lemmas, sometimes all glabrous	short crown of phairs around the callus, hairs stiff, 0.5—0.8 mm long	glabrous or with a louse crown of coarse slightly simous hairs	glabrous	dense tuft of crinkled hairs dorsally, sometimes with a few hairs laterally	glabrous	with dorsal tuft of hairs, sometimes with a few hairs laterally, hairs closely simuous to crinkled	glabrous	glabrous	mostly with a dorsal web, sometimes glabrous, infrequently with a crown of hairs or a diffuse web
Lodicule length, shape, and margin	0.9 mm, deeply 1.5 mm, to shallowly obova lobed, with a glabrous latera glabr	1.5 mm, obovate, with a small lateral lobe, glabrous	vith 2 with 2 broadly lanceolate lobes, lobes with a few apical short cilia	0.8–0.9 mm, obovate, margin with or without a few cilia along the upper margins	0.6 mm, with 2 0.4-0.7 mm, lobes, with weakly lo or without 1 glabrous, or 2 cilia infrequent rarely with 2 cilia	weakly lobed, glabruus, infrequently or rarely with 1 or 2 cilia	0.9 mm, infrequently or rarely with 1 or 2 cilia	0.3–1 mm, with 0.5–0.8 mm, a lateral N/K, N/K tooth, glabrous or infrequently with 1 or 2 cilia	0.5-0.3 mm, N/K, N/K	0.8 mm, lobed generally lobed or shouldered as in a mitter at midpoint, infrequently glabrous or rarely with 1 2 citia in so New Zealann species (f. Edgar, 1885	as in a mitten, infrequently glabrous or rarely with I or 2 cilia in some New Zealand species (f. Edgar, 1885)

Table 2. Continued.

	$Poa~(\mathrm{subg.}\\ Arctopoa) \qquad P.~(\mathrm{subg.}~\&\\ \mathrm{sect.}~Aphydris \mathrm{sect.}~Arctopoa)\\ (\mathrm{Griseb.})~\mathrm{Txvelev} \qquad eminens$	P. (subg. & sect. Arctopoa) eminens	P. (sect. Austrofestuca) billardierei	P. (sect. $Parodiochloa$) $cookii$	$P.~({ m sect}.$ $Parodiochloa)$ $flabellata$	P. (sect. $Parodiochloa$) $foliosa$	P. (sect. Parodiochloa) hamiltonii	P. (sect. Parodiochloa) ramosissima	P. (sect. Parodiochloa) tennantiana	P. (sect. $Tzrelevia$) $kergwelensis$	Poa (other than those separated)
Caryopsis shape fusoid, subtriangula sulcus distinct			apically blunt, fu subpentangular, sulcus pronounced	soid, nearly cylindrical, sulcus indistinct	fusoid, sub- triangular, sulcus distinct	fusoid, nearly round to subtriangular, sulcus indistinct	fusoid, nearly cylindrical, sulcus indistinct	fusoid	fusoid, nearly cylindrical, sulcus indistinct	narrowly fusoid, fusoid, subtri- subtriangular, angular, or sulcus nearly distinct cylindrical, sulcus disti	usoid, subtri- augular, or nearly cylindrical, sulcus distinct, or faint
Fruit length, hilum length, hilum shape	27	2.5-4.5 mm, 0.4-0.6 mm, elliptical	P mm, 0.4–? 2.5–4.5 mm, 2.5–4 mm, ca. c mm, elliptical 0.4–0.6 mm, 1 mm, elliptical elliptical	a. 2 mm, ca. 0.3 mm, elliptical	2–2.5 mm, ca. 0.4 mm, elliptical	2–2.7 mm, ca. 0.3 mm, broadly elliptical	ca. 1.5 mm, ca. ca. 1 mm, 0.3 mm, elliptical short < li>length grain	ca. 1 mm, presumably short < 1/2 length of grain	ca. 1.5 mm, ca. 0.2 mm, elliptical to punctiform	ca. 1.8 mm, ca. 1–4 mm, 0.2– 0.25 mm, 0.4 mm, narrowly punctiform elliptical elliptical	1-4 mm, 0.2- 0.4 mm, punctiforn to elliptical
Lipid presence, caryopsis firmness	present, firm present, firm		present, firm	present, firm	present, firm	present, firm	N/K	N/K	present, firm	present, firm	present, firm
Styles, stigma	bases with a bases with a bases with a gap, plumose gap, plumose gap, plumose	bases with a gap, plumose	lose	bases with a ba gap, plumose	bases without a gap between them, slender with only short primary branches	bases with a gap, plumose	bases with a gap, plumose	bases with a gap, plumose	bases with a gap, plumose	es with a bases with a bases with a base gap, plumose gap, plumose gap, plumose	bases with a gap, plumose
Chromosomes	2n = 42, 91, 2n = 28, 42, 97, 62		2n = 28	N/K	N/K	2n = 28	N/K	2n = 28	2n = 56	N/K a	L = x

VB = vascular bundles, N/K = not known.

The single species of Austrofestuca s. str. is now treated as Poa billardierei St.-Yves (≡ A. littoralis (Labill.) E. B. Alexeev) (subg. Poa sect. Austrofestuca (Tzvelev) Soreng & L. J. Gillespie) based on DNA sequence evidence (Hunter et al., 2004; Gillespie & Soreng, 2005; Gillespie et al., 2007). Austrofestuca leaf blades are similar to P. macrantha (subg. Poa sect. Madropoa Soreng) in involution and vestiture (Fig. 3G, H), and both are coastal sand-dune species. Blades of both species have involute margins and are densely covered by prickle hairs ca. 100 µm long, as is typical of Poa sect. Madropoa and many Australian species of Poa. In P. billardierei, the leaf blades have only two distinct grooves on the adaxial surface, as is typical of Poa, whereas P. macrantha can have up to three lateral grooves on each side of the midrib, with low costal ribs up to 2.5× the width of the valleys (a condition not previously reported in any other species of Poa). Poa sect. Austrofestuca is perfect-flowered, however, whereas Poa sect. Madropaa species are diclinous, often dioecious as in P. macrontha.

Little has been published about the single species of Tzvelevia (Alexeev, 1985), an endemic of the Kerguelen Islands and Heard Island. The genus was not mentioned by Clayton and Renvoize (1986) or Watson and Dallwitz (1992). The combination of Poatype leaf blade adaxial anatomy (Fig. 2D), keeled, unawned, softly pubescent lemmas, glabrous calluses, and acute, entire ligule margins supports its relationship with Poa rather than Nicoraepoa. Tzvelevia does not have papillae in the adaxial or abaxial leaf blade epidermis but does have sparse short hooks adaxially. Scant sclerenchyma is developed between the vascular hundles and the epidermis, and it appears to he discontinuous, not forming girders. For these reasons, we submerge Tzvelevia in Poa as a section (see Taxonomic Treatment section).

The four species of alpine southeastern Europe and the Middle East in the genus Bellardiachloa (type, B. violaceae (Bellardi) Chiov. (= B. variegata (Lam.) Kerguélen)) have been placed in (Clayton & Renvoize, 1986) and out of Paa, sometimes near Paa (Edmondson, 1980), or in a subtribe with Festuca rather than with Poa (Tzvelev, 1983; Mill, 1985). Molecular analyses have consistently shown that B. variegata does not belong within Poa (Soreng et al., 1990; Davis et al., 1993; Soreng et al., 2007; Gillespie et al., 2007). It differs from *Poa* in leaf hlade anatomy, with multiple tall costal ridges and deep valleys (Fig. 2A; Hernández Cardona, 1978; García-González, 1983), slender leaf blade apices, weakly keeled lemmas with awns, calluses with a crown of hairs, ciliate lodicule margins, sometimes semi-soft endosperm, and a crown of hairs at rachilla joint apices. With Nicoraepoa, the genus Bellardiochloa shares the deeply corrugated

leaf hlade adaxial epidermis and short-awned lemmas, but differs in the narrow costal ridges, weakly keeled lemmas, ciliate lodicule margins, and a crown of hairs at rachilla joint apices.

AN EXPANDED SUB-ANTARCTIC POA SECT, PARODIOCHLOA

Analyses of cpDNA and ITS data place Parodiochloa flabellata within Poa, above the hasal split of the remainder of Poa from section Sylvestres V. L. Marsh ex Soreng (and, in cpDNA analyses, subgenus Arctopaa (Griseb.) Prob.). The species is placed within the subgenus Ochlopoa (Asch. & Graebn.) Hyl. clade in cpDNA analyses, and between the subgenus Ochlapoa clade and the large higher Poa clade including suhgenera Poa, Pseudopoa (K. Koch) Stapf, and Stenapoa (Dumort.) Soreng & L. J. Gillespie in ITS analyses (Gillespie & Soreng, 2005; Gillespie et al., 2007; Gillespie et al., in prep.). The single species of Paradiochloa was elevated within Paa to Poa sect. Parodiochloa (Gillespie & Soreng, 2005). This species is morphologically unusual in *Poa* and differs from all other Poa species (Hubbard, 1981) in having the styles terminal with no gap between them at the base, and stigmas that are elongated and simple with only short primary branches. In all other Poa species (Tzvelev, 1983; Watson & Dallwitz, 1992; Soreng, pers. ohs.), the styles have a distinct gap between them at the hase, and the stigmas are plumose with secondary branching. The species also differs from all Poa species in having apical cusps grading to short, stout apical awns. Parodiochlaa differs from Nicoraepoa in its caespitose habit, elongated, deeply lacerate, glahrous ligules, and asymmetrically curved florets (Fig. 5S). Its adaxial leaf blade anatomy (Fig. 2E, F) is intermediate between that of Poa and Nicoraepoa in the ridge and valley adaxial epidermal topography, but the intercostal zones are covered by globose papillae, a condition unknown in Poa (except in five other suh-Antarctic island species; see next paragraph) and absent in Nicoraepoa. A few other species of Poa (e.g., P. arida Vasey and P. keckii Soreng) sometimes have single acentrically positioned papillae on some long cells of the abaxial leaf hlade surfaces (Soreng, pers. obs.), a feature characteristic of Puccinellia Parl. and other genera of subtribe Puccinelliinae Soreng & J. I. Davis (Soreng et al., 2003h).

Five sub-Antarctic island species of *Poa* approach *Nicoraepoa* and *P. flabellata* in leaf hlade anatomy and other characters. Edgar (1986), in a revision of New Zealand *Poa*, considered *P. cookii* and *P. ramosissima* Hook. f. to be a natural group that is likely allied to *P. flabellata*. *Poa flabellata* grows on oceanic islands adjacent to South America, *P. cookii*

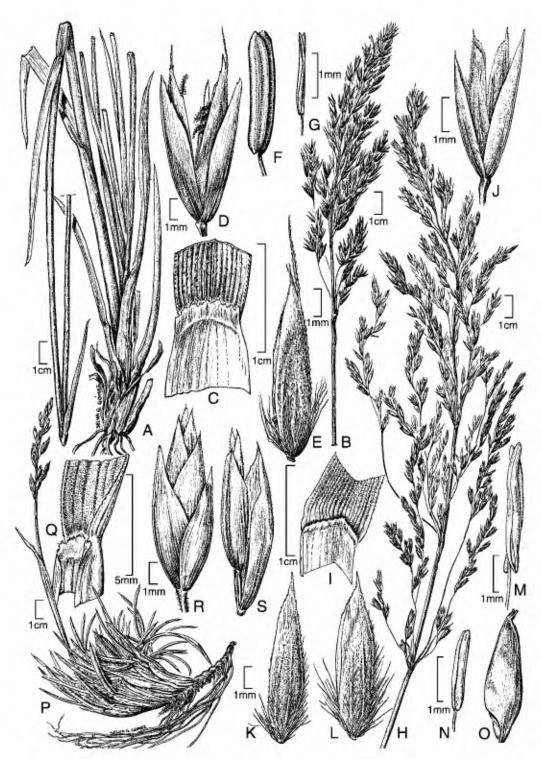


Figure 4. A–G. Nicoraepoa andina (Trin.) Soreng & L. J. Gillespie. —A. Habit. —B. Inflorescence. —C. Ligule. —D. Spikelet. —E. Floret. —F. Anther (fertile). —G. Anther (sterile). (Drawn by S. G. Gómez. Reprinted from Flora Patagónica vol. 8(3): fig. 109, with permission from INTA.) H–O. Nicoraepoa chonotica (Phil.) Soreng & L. J. Gillespie. —H. Inflorescence. —I. Ligule. —J. Spikelet (perfect). —K. Floret (perfect). —L. Floret (pistillate). —M. Anther (fertile). —N. Anther (sterile) —O. Caryopsis. (Drawn by S. G. Gómez. Reprinted from Flora Patagónica vol. 8(3): fig. 113, with permission

on islands in the southern Indian Ocean, and P. ramosissima on islands south of New Zealand's South Island. Poa cookii shares many characteristics of P. flahellata. A close relationship between P. flahellata and P. ramosissima has now heen demonstrated based on ITS sequence analyses (Gillespie et al., in prep.). Three additional New Zealand sub-Antarctic island species are considered closely related to P. ramosissima and P. cookii: P. hamiltonii, P. foliosa, and P. tennantiana. Edgar (1986) grouped the latter two species and treated P. hamiltonii as a synonym of P. cookii. We tentatively consider P. hamiltonii distinct from P. cookii, but agree there are few differences (see Notes in Taxonomic Treatment section under P. hamiltonii). This set of morphologically similar, southern-ocean island species has neither been united in an infrageneric classification nor formally grouped with P. flabellata. These five species are here placed in Poa sect. Parodiochloa with P. flabellata. Molecular relationships of the latter four species remain to be studied.

Several morphological features distinguish this newly expanded Poa sect. Parodiochloa (Table 2). The species are mostly large tufted grasses (except P. ramosissima), sometimes rhizomatous (P. foliosa, P. tennantiana), sometimes with pungent (P. flahellata) or semi-pungent leaf hlades (P. cookii, P. foliosa, P. hamiltonii). Adaxial leaf blade surfaces are densely papillate on the costal or intercostal regions or both (all species, Figs. 2E, F, 3A-C; also noted by Edgar & Connor, 2000) (sometimes absent in P. tennantiana, Fig. 3D), and are otherwise mostly smooth (P. ramosissima, P. tennantiana) or quite scahrous (P. cookii, P. flabellata, P. foliosa, P. hamiltonii). The five to 42 lateral costal ridges (occasionally obscured by intercostal papillae) are tall (P. cookii) or have moderate or low adaxial epidermal relief in other species, and are one to three times wider than the intercostal valleys. The ligules are deeply and regularly lacerate (P. cookii, P. hamiltonii, P. ramosissima), elongate and entire to irregularly lacerate at the apex (P. flabellata), or short, truncate, and densely covered with short thick appressed hairs (P. foliosa, P. tennantiana). Panicle branches are mostly smooth but can be scabrid to hispid (some P. cookii). Lemma apices are generally slightly twisted and sharply pointed, with a short, somewhat thick attenuated cusp (P. cookii, P. foliosa, P. hamiltonii, P. ramosissima, P. tennantiana) grading to an awn up to 3 mm long in P. flahellata. The species are gynomonoecious (P. cookii, P.

hamiltonii, P. ramosissima), dioecious (P. foliosa), or perfect-flowered (P. tennantiana) (Edgar, 1986; Edgar & Connor, 2000). Although P. flabellata was reported to be perfect-flowered (Anton & Connor, 1995), this needs further study. The stigmas are plumose and the styles are distinctly separate at the base, as is typical of Poeae, or the stigmas are slender and short-branched and the styles have no gap between them at the base (P. flabellata). The floret calluses may be glabrous (four species) or distinctly webhed (P. foliosa and most P. ramosissima). The glabrous state for calluses appears to be derived in this group, and thus the presence of a dorsal web is still an autapomorphy of Poa.

KEY TO THE GENUS NICORAEPOA AND SELECTED GENERA AND SPECIES OF SUBTRIBE POINAE

Genera of Poeae subtribe Poinae included in the key are: Arctophila, Bellardiochloa, Dupontia, Festucella, Hookerochloa, Hyalopoa (Tzvelev) Tzvelev, Nicoraepoa, and Poa (including sections Austrofestuca, Madropoa, p.p., Paradiochloa, Siphonocoleus Hitchc., and Tzvelevia). These genera share a perennial habit, multiple-flowered spikelets that are lanceolate in shape, and acute to acuminate glumes reaching over three fourths the length of the adjacent florets. Their caryopses have a non-linear, short hilum up to one third the length of the grain and are apically glabrous. The Poinae genus Arctagrostis is not included because it can be easily distinguished by its single-flowered spikelets. Likewise, genera of Alopecurinae and Miliinae may be distinguished by single-flowered spikelets. Genera of subtribe Puccinelliinae (Soreng et al., 2003b) similar to Poa in gross or overall morphology are excluded by their generally shorter glumes in combination with papillate leaf surfaces or single-flowered spikelets (Catahrasa P. Beauv., Catahrosella (Tzvelev) Tzvelev, Oreopoa H. Scholz & Parolly, Paracolpodium (Tzvelev) Tzvelev, Phippsia (Trin.) R. Br., Pseudosclerochloa Tzvelev, Puccinellia, Sclerochloa P. Beauv.).

Various combinations of characters can be used to discriminate among the above set of Poinae genera, but none holds up for all species. Thus, producing a key to the genera could be approached in a variety of ways depending on which characters are employed in which order. Leaf characteristics, including adaxial leaf blade surface morphology and ligule shape, seem to be the most consistently diagnostic for *Nicoraepoa*.

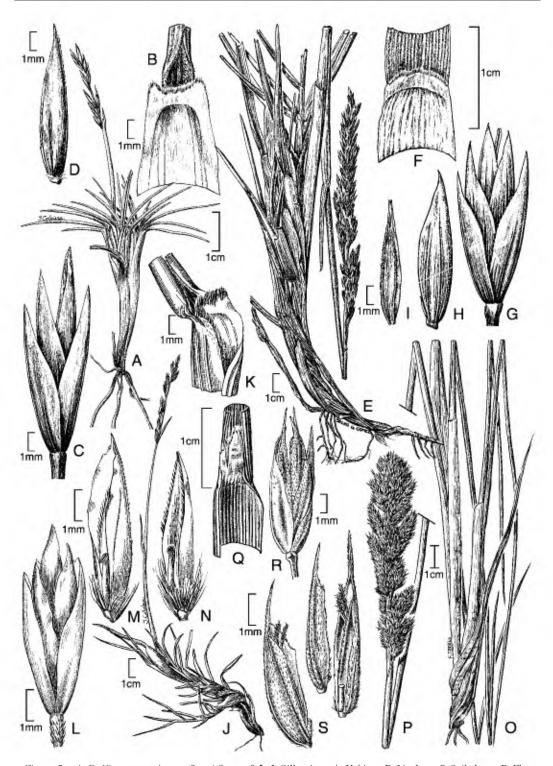


Figure 5. A–D. Nicoraepoa erinacea (Speg.) Soreng & L. J. Gillespie. —A. Habit. —B. Ligule. —C. Spikelet. —D. Floret. (Drawn by I. Coloiera. Reprinted from Flora Patagónica vol. 8(3): fig. 110, with permission from INTA.) E–I. Nicoraepoa robusta (Steud.) Soreng & L. J. Gillespie. E. Habit. —F. Ligule. G. Spikelet. —H. Floret. —I. Palea. (Drawn by I. Coloiera. Reprinted from Flora Patagónica vol. 8(3): fig. 108, with permission from INTA.) J–N. Nicoraepoa subenervis (Hack.) Soreng & L. J. Gillespie subsp. subenervis. —J. Habit. —K. Ligule. —L. Spikelet. —M. Floret. —N. Palea. (Drawn by

1a. Adaxial leaf blade lateral surfaces prominently corrugated with multiple ridges and valleys, the ridges 2-5× wider than the valleys, without papillae; ligules 0.2-3 mm long, apices truncate (to slightly higher at the margins), margins entire or irregularly crose, never lacerate, ciliate or ciliolate (cilia 0.1-0.6 mm long); calluses glabrous or with a crown of straight to slightly curved hairs to 0.2-3 mm long, never webbed; plants usually of wet or seasonally wet, sometimes saline or subsaline habitats in southern Snuth America	7
valleys and the ridges up to 1-5× wider than the valleys, then the adaxial surface papillate in part (except some Poa tennantiana) and ligules commonly longer than 3 mm and sometimes deeply lacerate; ligules 0.2-25 mm long, apices truncate to acuminate, margins entire to deeply and regularly lacerate, smooth or infrequently asperous or cilinlate (cilia to 0.1 mm long); calluses glabrous or with a crown or dorsal tuft of hairs (webbed); plants of various	
habitats, not from South America (except Nicaraepaa erinacea and some Poa) 2a. Ligules deeply and more or less regularly lacerate 3a. Calluses with a crown of hairs 0.3–0.7 mm long; lemmas 3-nerved; adaxial leaf blade surfaces not papillate; plants of Northern Hemisphere high-latitude wetlands Arctaphila	3
3b. Calluses glabrous or webbed, infrequently with a few isolated hairs around the sides of the callus; lemmas 3- to 5-nerved	1
 4a. Culms and nodes strongly compressed; sheaths fused to near their collars; adaxial leaf blade surfaces not papillate; plants of uplands in the Hawaiian Islands)
papillate; plants of sub-Antarctic island coastal reginns (Poa sect. Paradiachloa, p.p.)	
5b. Culms stout, spreading to erect; plants caespitose, forming large dense tufts; leaf blades	ı
moderately firm, 2–10 mm wide, flat or folded, abaxially smooth, adaxially papillate in part;	5
6a. Fibrous ridges of basal sheaths smooth or irregularly sparsely scabrous; leaf blades adaxially nearly smooth or with scattered thick-based low hooks over the veins Poa hamiltoni	÷
6b. Fibrous ridges of basal sheaths regularly and closely pectinately scabrous (as in a sawfish's	v
hill); leaf hlades adaxially papillate and sometimes scabrous on the sides of the ridges with narrow-based spreading tipped hnnks	,
2h. Ligules entire or infrequently with few irregular, mostly shallow lacerations	ι 7
7a. Adaxial leaf hlade surfaces regularly corrugated with ridges 1–3× as wide as adjacent intercostal valleys 8	
8a. Lemmas munded to weakly keeled across the back, apex usually slender short-awned; leaf blades	
slender, 0.2–0.7(–1) nm wide, tightly involute; calluses with a crown of hairs often persisting at the tips of rachilla joints; panicle branches densely scabrous angled; caryopses semi-soft; alpine plants of	
southern Europe and the Middle East	1
8b. Lemmas keeled across the back, apex cuspidate to stiffly short-awned; leaf blades broader, 2–10 mm	
wide, flat or folded, or the outer margins inrolled; calluses glabrous or webbed; panicle branches	
smooth or finely scabrous angled; caryopses firm; plants of suh-Antarctic islands and coasts of Tierra del Fuego, South America, and Shetland Islands (Poa sect. Paradiochloa, p.p.))
9a. Panicles tightly contracted; lemmas and calluses glahrous; ligules 6-23 mm long, abaxially	
smooth or sparsely scabrous	1
wehhed; ligules 1–16 mm long, ahaxially densely scahrid to puberulent)
10a. Ligules 1-3 mm; fibrous ridges of basal sheaths smooth; calluses webbed; panicles	
contracted	ı
loosely contracted	ı
7b. Adaxial leaf blade surfaces generally with only two central grooves, or with low ridges not more than 1× as	
wide as adjacent shallow valleys	
12a. Lemmas with a terminal or slightly subterminal awn; plants with coarsely fibrous old basal	-
sheaths; panicles open, branches scabrous-angled; calluses with a crown or beard of short hairs to 1 mm long	2
13a. Leaf blades flat or folded, 2–7 mm wide, surfaces smooth or nearly so, margins	,
scabrous	7

	13b.	. Leaf blades involute, 0.5–1.5 mm wide, surfaces and margins scabrous Festucelle
12b.		nmas unawned; plants with or without fibrous old basal sheaths; panicles contracted or open,
	brar	nches smooth or scabrous (rarely hispidulous), terete nr angled; calluses with a crown or
	bear	rd of short or longer hairs to 5 mm long
	14a.	. Ligules short-truncate, tn 1 mm long, margins ciliolate tn ciliate; callus with a dense
		crown or beard of short bairs to 1 mm long; leaf blades ca. 1-2 mm wide, tightly folded
		with involute margins, firm to moderately firm, adaxially densely scabrid to coarsely
		hispidulous or soft-pubescent on and between the veins, abaxially smooth, glabrous;
		panicles contracted, dense or with few spikelets; stnloniferous to rhizomatous plants 13
		15a. Panicles simple, sub-spicate, with fewer than 8 spikelets; plants forming low dense
		cushions to 25 cm tall; leaf blades up to 3(4) cm long, abaxially softly puberulent,
		stiff and sharply pungent; plants of interior sub-saline seasonally wet habitat in
		Patagonia, Argentina
		15b. Panicles branched, densely contracted, with more than 10 spikelets; plants dense to
		widely spreading, usually forming larger tufts, 20-100 cm tall; some or many leaf
		blades over 5 cm long, abaxially coarsely scabrous to stiffly puberulent, stiff and
		sharply pungent to moderately firm and weakly pungent; plants of coastal sands 10
		16a. Leaf blades firm, sharply pungent; flowers perfect; plants of Australia and
		New Zealand Poa billardierei (sect. Austrofestuca
		16b. Leaf blades moderately firm, weakly pungent; flowers unisexual; plants of
		the Pacific coast of United States and Canada
		Poa sect. Madropoa, p.p., P. douglasii Nees, and P. macranthe
14	14b.	Ligules obtuse to acuminate, 1-18 mm long, margins glabrous; callus glabrous or with
		a sbort or longer crown of hairs to 5 mm long; leaf blades to 1.5–10 mm wide, flat, folded
		nr involute, snft, adaxially smooth or scabrous mainly over the veins, abaxially smooth or
		scabrnus, glabrnus or infrequently stiffly puberulent; panicles contracted or open; plants of
		various babits
		17a. Palea keels scabrid at least in part; panicle branches smnoth or scabrous Poa, p.p.
		17b. Palea keels smooth nr nearly so; panicle branches smooth
		18a. Lemmas distinctly keeled; callus with a diffuse crown of fine soft weakly
		woolly hairs 1–2.5 mm long
		18b. Lemmas weakly keeled; callus with a tuft of straight or sinuous hairs on the
		surrounding lemma base
		19a. Leaf blade abaxial surface strongly ribbed, flat; glumes usually much
		sborter than adjacent florets; paleae subequal to lemmas; florets 2 to 6;
		lemmas glabrous, apices obtuse
		19b. Leaf blade abaxial surface smooth or weakly ribbed, flat or weakly
		involute; glumes subequal nr longer than adjacent florets; florets 1 to
		2; paleae distinctly shorter than lemmas; lemmas glabrous or softly
		pubescent, apices acute to acuminate

TAXONOMIC TREATMENT

Nicoraepoa Soreng & L. J. Gillespie, gen. et nom. et stat. nov. Replaced name: Poa subgen. Andinae Nicora, Hickenia 1(18): 99. 1977. TYPE: Nicoraepoa andina (Trin.) Soreng & L. J. Gillespie.

Perennials; gynodioecious to weakly dioecious, or synecious; stoutly rhizomatous, rhizomes 3 mm diam., short or long; major roots 1–2 mm diam.; vegetative branching intra- and extra-vaginal; sheaths of basal leaves becoming coarsely fibrous in age, smooth or sparsely scabrous between veins; sheaths of upper culm leaves open to near base or margins joined hy a narrow hyaline membrane up to 1/4 the sheath in length; ligules 0.2-3 mm long, thickly membranous to chartaceous, abaxially smooth or pubescent with appressed to loose hairs, apices truncate or longest near the margins, irregularly erose ciliolate to ciliate (with hairs to 0.5 mm long); blades firm (fairly soft in N. subenervis), flat, folded or involute, 2-15 mm wide,

abaxially smooth or the keel scabrid near apices, glabrous, adaxially with (2)4 to 48 deep to moderately deep intercostal valleys that are smooth or more often scabrous, separated with blocky flat-topped or broadly rounded costal ridges (anatomically with adaxial epidermis underlain by sclerenchyma caps or with "T"- or "I"-shaped girders extending to the abaxial epidermis in cross section), 1-5× broader than the valleys, without papillae, smooth or commonly scabrous especially on the sides of the ridges, glabrous or rarely short stiff puherulent (N. erinacea), apices pungent (weakly so in N. subenervis). Panicle axis and branches smooth or less often scabrous, pedicels smooth or scahrous, glabrous (or rarely coarsely hispidulous). Spikelets 2- to 5-flowered; rachilla internodes terete, usually scahrous; glumes suhequal, smooth or keels scahrous, chartaceous throughout or apically scarious, 1- or 3-veined, lateral veins faint or distinct, subequal to the adjacent lemmas; calluses blunt, glabrous or with a crown of 0.2-3 mm long, straight or slightly curved hairs;

lemmas keeled, apices symmetrical and not twisted, acutely pointed, mucronate, or awned from a sinus up to 0.2 mm deep, glabrous or with stiff hairs on the keel to 1 mm long and sometimes on the marginal veins at the base, smooth throughout or scabrous over much of the surface, weakly veined, awnless or with a slender, erect, briefly divergent or weakly sinuous, slender, scabrid, 0.2-3(5) mm long awn from the keel vein (without lateral merging veins); palea keels scabrous or coarsely ciliate; lodicules 2, 1-2 mm long, membranous, lanceolate, apically entire or irregularly short-toothed; anthers 3, 2-3 mm long, staminodes in pistillate flowers 1–3 mm long. Caryopsis glabrous, firm or less often semi-soft, lipid present; hilum round to narrowly elliptical, 1/5-1/3 the grain in length; style hases terminal, with a gap between them, stigmas plumose, white.

Distribution and habitat. Argentina and Chile, from moderately high elevations in the Andes at about 32°S in the province of San Juan, Argentina, and Region V, Chile, south through Patagonia to coastal and interior regions of Tierra del Fuego, Isla de Estados, and Falkland/Malvinas Islands (Nicora, 1978; Moore, 1983). Perennially or seasonally wet ground, or in shallow water, in fresh water or frequently in subsaline, saline, or mineralized soils of coastal sands and estuaries, springs, hogs, marshes, springy meadows, wet rocks, and riparian meadows, from sea level to over 3200 m.

Notes. The new genus name commemorates the late Elisa G. Nicora, Argentine specialist in Poa, author of Poa subg. Andinae, and influential, longtime student of South American Poaceae who passed away in 2001 (Rúgolo de Agrasar, 2001). The subgenus epithet Andinae is a plural adjective and cannot he satisfactorily employed as a plural Latinized noun. Also, there is a genus Andinia (Luer) Luer in Orchidaceae. Nicoraepoa has six species and two subspecies. A good key to the species is available in Nicora (1978).

- Nicoraepoa andina (Trin.) Soreng & L. J. Gillespie, comb. nov. Basionym: Poa andina Trin., Linnaea 10(3): 306. 1836. TYPE: Chile. [Region Bío-Bío, province Bío-Bío]: "austr. in alp. frigid. mont. igniv. Antuco," [1829], E. Päppig [D.-907] (holotype, LE [TRIN-2578.01a]!; isotypes, LE [TRIN-2578.01a] fragment at BAA-2450b!, LE!). Figure 4A-G.
- Poa acrochaeta Hack., Repert. Spec. Nov. Regni Veg. 10(243–247): 172. 1911. TYPE: Chile. [Region Maule, province Curicó]: Vulkan Peteroa, s.d., comm. K. F. Reiche s.n. [collector questionable: 448 in Philippi script on label (possibly M. Vidal Gormaz in 1891 according to E. Nicora annotation)] (holotype, SGO

- [PHIL-448] W 39455!; isotypes, SGO [PHIL-448] fragment and photo at US 1723707!, W 39455 fragment at US 88711!).
- Poa oristata Phil., Anales Univ. Chile 43: 574. 1873. TYPE: Chile. [Region Los Lagos, province Osorno]: Volcán de Osorno, [1872], don Carlos Juliet s.n. (holotype, SGO [PHIL-439] W 39450!; isotypes, BAA-2457!, SGO [PHIL-439] fragment and photo at US 89700!).
- Deschampsia latifolia Phil., Linnaea 29(1): 91. 1858, non Deschampsia latifolia Hochst. ex A. Rich., 1850. TYPE: Chile. [Region Maule, province Linares]: in Andibus dep. [Cordillera] Linares, Germain s.n. (holotype, SGO [PHIL-197] W not seen; isotypes, SGO 63483!, SGO [PHIL-197] fragment and photo at US 556494!).

Distribution and habitat. Argentina (Neuquén), Chile (Araucanía, Bío-Bío, Los Lagos, Maule), in the central Patagonian Andes (Nicora, 1978). Stream banks and wet meadow at high elevations.

Notes. Many SGO Philippi types of Poaceae were loaned to US ca. 1904 (where fragments were retained and specimens photographed; see U.S. National Herbarium Type Specimen Register, http://ravenel.si.edu/hotany/types/), returned to SGO, and then sent to Edward Hackel in Vienna, where they may be seen today, generally with W accession numbers on the SGO sheets.

Additional specimen examined. CHILE. Araucanía: Laguna Captrén, R. J. & N. L. Soreng 7193 (CONC, US).

- 2. Nicoraepoa chonotica (Phil.) Soreng & L. J. Gillespie, comb. nov. Basionym: Poa chanotica Phil., Linnaea 29(1): 97. 1858. TYPE: Chile. [Region Aysén, province Aysén]: [Chonos-Islands], "in monte Cerro de Chonos ad circa 1290 ped. s.m.," Dr. Fank s.n. (holotype, SGO [PHIL-410] W s.n.!; isotypes, SGO [PHIL-410] fragment at BAA!, SGO [PHIL-410] fragment and photo at US 39683!). Figure 4H–O.
- Pou berningeri Pilg., Notizbl. Bot. Gart. Berlin-Dahlem 10(97): 761. 1929. TYPE: Chile. [Region Los Lagos], province Llanquihue: Volcán Yates, ca. 1200 m, "vereinzelt zwischen Nothofagus an der Waldgrenze," Mar. 1925, Werdermann 1698 (holotype, B not seen; isotypes, B fragment at BAA!, B fragment at US 89693! [pistillate]).
- Poa borchersii Phil., Anales Univ. Chile 94: 172. 1896. TYPE: Chile. [Region Bío-Bío, province Ñuble]: "prope thermas chillanenses," [Baños de Chillan, Jan. 1883], Augustus Borchers s.n. (holotype, SGO [PHIL-431] W 39457!; isotypes, US fragment at BAA!, SGO [PHIL-431] fragment and photo US 89691a!, W 29457 fragment at US 89691h!).
- Poa chubutensis Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 7: 196. 1902. TYPE. Argentina. Province Chuhut: [region Río Corcovado, 43°S, 71°W], "in rupestrihus collinis prope Teka-choique, [1–15 Feb.] aest. 1901," N. Illin [90] (holotype, LP!; isotypes, BAA 2518!, SI!, US 916986!, LP [SPEG] US 1503958!).
- Poa lotifolia Phil., Linnaea 29(1): 97. 1858, non Poa latifolia G. Forst., Fl. Ins. Austr. 4, no. 44. 1786. TYPE: Chile.

[Region Aysén, province Aysén]: [Chonos-Islands], "in monte Cerro de Chonos ad circa 1290 ped. s.m.," *Dr. Fonk s.n.* (holotype, SGO [PHIL-414] W 39453!; isotypes, SGO fragment at BAA-2612!, SGO 63493!, SGO [PHIL-414] fragment and photo at US 88768!).

Poo robusta Phil., Anales Univ. Chile 43: 574. 1873, non Poa robusta Steud., 1854. TYPE: Chile. [Region Los Lagos, province Llanquihue]: Volcán de Calbuco, [1872], Carlos Juliet s.n. (holotype, SGO [PHIL-443] W 39449!; isotype, SGO [PHIL-443] fragment and photo at US 88733!).

Distribution and habitat. Argentina (Chubut, Neuquén, Río Negro), Chile (Araucanía, Aysén, Los Lagos, Magallanes), in central and southern Patagonian Andes and coastal reaches of southwestern Chile. Low to upland marshes, riverbanks, and wet rocks.

Nates. This species was called Paa borchersii (Nicora, 1978), but Soreng et al. (2003a) determined that the older name P. chonatica applies to the species. Viviparous specimens of this species are common but can be separated from Nicoraepoa robusta by their looser or open panicles, shorter glumes, and frequent presence of a few hairs around the callus.

Additional specimens examined. CHILE. Araucanía: Lonquimay, A. Pfister 7949 (CONC). Aysén: Seno Ventisquero, R. J. & N. L. Soreng 7254 (CONC, US). Magallanes: Última Esperanza, Río Pingo, R. J. & N. L. Soreng 7349 (CONC, US).

Nicoraepoa erinacea (Speg.) Soreng & L. J. Gillespie, comh. nov. Basionym: Poa erinacea Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 7: 198. 1902. TYPE: Argentina. [Province Chuhut]: "in aridissimis subsalsis secus Río Chubut," Dec. 1899, N. Illin s.n. (holotype, LP 67!; isotypes, LP BAA 2534!, LP fragment at US 88783!). Figure 5A-D.

Distribution and habitat. Argentina (Chubut), in arid parts of Patagonia (Nicora, 1978). Subsaline springs, at mid-elevations.

Notes. This is one of the odder species of the new genus, as the leaf blades are narrow and softly hairy, with few intercostal valleys on the adaxial surface. However, as in other *Nicaraepoa* species, the callus has a distinct crown of hairs, the short stiff leaf blades are apically pungent, and the ligules are densely ciliate.

4. Nicoraepoa pugionifolia (Speg.) Soreng & L. J. Gillespie, comb. nov. Basionym: Poa pugionifolia Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 7: 199. 1902. TYPE: Argentina. [Province Santa Cruz]: "Párr-aik (Río Sehuen), Feb 1898,"

C. Amegbino s.n. (lectotype, designated here, LP [SPEG-65]!); isotype, [LP-SPEG-65 fragment] BAA-2672!. Figure 4P-S.

Poo acutissima Pilg., Repert. Spec. Nov. Regni Veg. 12: 306. 1913. TYPE: Argentina. Süd-Patagonien, [1907–1908], C. Skottsberg s.n. (holotype, B not seen; isotypes, BAA 2442!, BAA 2443!, B fragment at US 88712!).

Distribution and habitat. Argentina (Santa Cruz, Tierra del Fuego), Chile (Magallanes), in Patagonia. Low- to mid-elevation subsaline meadows and open ground in semi-arid Patagonia.

Nates. LP-SPEG-65 is unambiguous as a lectotype. The two original specimens in Speggazini's herharium are taxonomically indistinguishable, but the place, river, and month recorded on the labels are arranged differently than in the original protologue. The protologue stated, "in rupestribus porphyricis Párr-aik secus Río Chico et Boron-aik secus Río Sehuen, Jan. et Feb. 1898, C. Ameghino s.n." The protologue has the localities and rivers and dates switched, as on a map we located Párr-aik south of the Río Sehuen, which is well south of the Río Chico. This is in agreement with the chosen lectotype and its isotype BAA-2672, "Párr-aik (Rio Sehuen)," which gives the date as February, "2-1898." We have not been able to locate Boron-aik on a map, but the syntype at LP [SPEG-66] says Boron-aik (Rio Chico), Jan. 1898.

Additional specimens examined. ARGENTINA. Santa Cruz: Güer Aike, 9 km S of Río Gallegos, P. M. Peterson et al. 17062 (SI, US); Buenos Aires, P. M. Peterson et al. 17254 (SI, US). CHILE. Magallanes: Última Esperanza, Sierra Baguales, R. J. & N. L. Soreng 7336 (CONC, US).

5. Nicoraepoa robusta (Steud.) Soreng & L. J. Gillespie, comb. nov. Basionym: Poa robusta Steud., Syn. Pl. Glumac. 1: 426. 1854. TYPE: Chile. [Region] Magallanes, [province Magallanes]: "Punta Arenas, paludihus salsis lit. maris prope, Dec.," W. Lechler [Hohenacker exsiccatae] 1191 (holotype, P [STEUD] not seen; isotypes, G fragment at BAA!, P fragment at BAA!, G not seen, LE!, K!, W, K, P fragments at US 81586!, LE fragment at US 946984!, W not seen). Figure 5E–I.

Festuca arenorio Lam., Encycl. 1: 191. 1791, non Festuca arenaria Osbeck, 1788. Poa arenicola St.-Yves, Candollea 3: 282. 1927, non Poo arenaria Lam., Tabl. Encycl. 1: 183. 1771. TYPE: [Chile. Region Magallanes, either province Magallanes or Tierra del Fuego]: E. Region of Magellan, in arenis maritimis, M. Commerson s.n. (holotype, P [LAM] not seen; isotypes, MPU not seen, P [LAM] fragment at US 2875377b!, MPU fragment and photo at US 2875377a! [perfect-flowered]).

Distribution and habitat. Argentina (Santa Cruz, Tierra del Fuego), Chile (Magallanes), in southern Patagonia, Tierra del Fuego, Isla de los Estados, and Falkland/Malvinas Islands. Coastal wet meadows and beaches.

Notes. In this species there are normal-flowered and viviparous forms. The normal-flowered forms always have glabrous lemmas and calluses. In the viviparous forms, these structures may be glabrous, or, more commonly, there are some short sparse hairs on the callus. The latter plants are separable from Nicoraepoa chonotica viviparous plants by their narrowly lanceolate panicles and long glumes. Reports from farther north in Chile, Aysén Region, represent viviparous plants of N. chonotica.

6. Nicoraepoa subenervis (Hack.) Soreng & L. J. Gillespie, comb. nov. Basionym: Poa subenervis Hack., Ark. Bot. 7(2): 7, t. 2, f. 3. t. 7, f. 1908. TYPE: Argentina. [Province Santa Cruz]: Patagonia, "in paludosis inter, Lago Viedma et Laguna Tar, ca. 1000 m, 27 Feb. 1905," P. K. H. Dusén 6021 (lectotype, designated here, W not seen; isotypes, K fragment at BAA 2704-1!, US fragment at BAA 2704-2!, W fragment at US 88723!).

Distribution and habitat. Argentina, Chile, in Patagonia. Upland vegas and subsaline springs in arid mountains.

Notes. The protologue reads: "Argentina. E. & S. Patagonia: 'in montanis inter lacum Lago Viedma et lacum Laguna Tar ad marginem paludis in alt c 1000 m; ad flumen Río Fosiles in montanis uliginosis c 800 m,' Dusén." Three specimens agree with the type protologue: (1) "inter lacum Lago Viedma et lacum Laguna Tar," P. K. H. Dusén 5927 (syntype, comm. J. F. Mullins in 1922 BAA 2703!); (2) "Lago San Martin, Río Fosiles in uliginosis c. 800 m, 1 'A.' 1905," P. K. H. Dusén (syntype, S not seen); and (3) P. K. H. Dusén 6021 (cited above as the lectotype). P. K. H. Dusén 6021 was selected as lectotype because duplicates and fragments are present in several herbaria.

6a. Nicoraepoa subenervis subsp. subenervis. Basionym: Poa subenervis Hack. var. subenervis.

Distribution and habitat. Argentina (Santa Cruz), Chile (Magallanes) (Nicora, 1978). Southern Patagonia. Figure 5J–N.

6b. Nicoraepoa subenervis subsp. spegazziniana (Nicora) Soreng & L. J. Gillespie, comb. nov. Basionym: Poa subenervis var. spegazziniana Nicora, Hickenia (18): 103. 1977. Poa spegazziniana Parodi, Dansk Bot. Ark. 22(1): 67. 1963, nom. nud. TYPE: Argentina. Province Mendoza: Dpto. San Rafael, Atuel Valle, a Arroyo Colorado [lat. 34°, 2200 m, 19 Nov. 1955], T. W. Böcher, Hjerting & K. Rahn 1306 (holotype, BAA 2706-1!).

Distribution and babitat. Argentina (Mendoza, Neuquén [Nicora, 1978], and San Juan), Chile, (Libertador General Bernardo O'Higgins, Metropolitana), in the northern Patagonian Andes.

Additional specimens examined. ARGENTINA. San Juan: Río del Agua Negra, P. M. Peterson et al. 19296 (SI, US). CHILE. Libertador General Bernardo O'Higgins: Sewell, Río Coya, Jiles s.n. (CONC 134734). Metropolitana: above Embalse del Río Yeso, R. J. & N. L. Soreng 7155 (CONC, US).

SIX SUB-ANTARCTIC ISLAND POA SPECIES ARE PLACED IN POA SECT. PARODIOCHLOA

Poa sect. Parodiochloa (C. E. Hubh.) Soreng, Contr. U.S. Natl. Herb. 48: 579. 2003. Basionym: Parodiochloa C. E. Hubh., Bull. Brit. Mus. (Nat. Hist.), Bot. 8: 395. 1981, non Parodiochloa A. M. Molina, 1986. TYPE: Festuca flobellato Lam. (≡ Poa flabellata (Lam.) Hook. f.).

Notes. The section as here emended has six species.

Poa cookii (Hook. f.) Hook. f., Philos. Trans. 168:
 1879. Basionym: Festuca cookii Hook. f., Fl. Antarct.
 182. 382, t. 139. 1846. TYPE: [France. Kerguelen Islands]: Kerguelen's Land, Christmas Harbour, on rocks and in moist places always near the sea, abundant, May-Aug. 1840, J. D. Hooker 762 (lectotype, designated by Edgar, 1986, New Zealand J. Bot. 24: 433, K [H2003/00969-290!]; isotype, LE not seen).

Distribution and babitat. Sub-Antarctic islands of the southern Indian Ocean: Crozet Islands, Heard Island, Kerguelen Islands, Marion Island, and Prince Edward Islands (Edgar, 1986, without voucher citations). Edgar's (1986) records for Poa cookii s.l. on Macquarie Island are referred to P. bamiltonii. Specimens reported by the Australian Antarctic Data Center (2007) from Crozet Islands (e.g., Possession Islands, W. A. Proctor 00257 [BM not seen]) and Heard Island (e.g., J. M. R. Hughes & R. D. Seppelt 24056 [ADT not seen]) are presumed to be P. cookii s. str. based on Edgar's comments about differences in P. cookii s.l. plants from Macquarie Island from the rest of the species. Coastal wet rocks and bluffs, and wet slopes, to 100 m, often associated with penguin colonies. The Australian Antarctic Data Center (2007) reported P. cookii from Gough Island based on E. W. Groves collections, but these (and other specimens

seen by us) were verified by C. E. Huhbard (Groves, 1981) as *P. flabellata*. Two other specimens were cited in Hooker's original protologue: (1) Hah. Kerguelen's Land, *Anderson* (in Cook's Voyage) *s.n.* (syntype, not seen), and (2) Kerguelen's Land, *D. Lyall s.n.* (syntype, K [H2003/00969-289]!).

Additional specimens examined. SOUTH AFRICA. Prince Edward Islands: Marion Island, S. Fugler 125 (US); Prince Edward Island, B. J. Huntly 975 (K).

Poa flabellata (Lam.) Raspail, Ann. Sci. Observ.
 86, t. 4, f. 11. 1829. Basionym: Festuca flabellata Lam., Encycl. 2: 462. 1788. Paradiachloa flabellata (Lam.) C. E. Hubh., Bull. Brit. Mus. (Nat. Hist.), Bot. 8: 396. 1981. TYPE: [Chile. Region Magallanes, province Magallanes or Tierra del Fuego]: Cette plante a été trové au détroit de Magellan, [Dec. 1767–Jan 1768], M. Cammerson s.n. (holotype, P [LAM] not seen; isotype, P [LAM] two fragments at US 2875414!). Figure 50–S.

Dactylis caespitosa G. Forst., Fasc. Pl. Magell. 12. 1788.
Festuca antarctica Spreng., Pl. Min. Cogn. Pug. 2: 23.
1815. Festuca caespitosa (G. Forst.) Roem. & Schult.,
Syst. Veg. 2: 732. 1817, non Festuca caespitosa Desf.,
1798. Poa forsteri Steud., Syn. Pl. Glumac. 1: 260.
1854. Poa caespitosa (G. Forst.) Hook. ex Speg., Anales
Mus. Nac. Hist. Nat. Buenos Aires 5: 91. 1896, non
Poa caespitosa Poir., 1804. TYPE: [Argentina. Province
Tierra del Fuego]: [Isla de los Estados], Novi Anni
insulis, s.d., Forster s.n. (types, BM not seen, K!, KIEL
not seen, LE [TRIN-2791.2]!, UPS [T-2347] not seen,
K [FORST] fragment at US 2851271!, W not seen).

Festuca urvilleana Steud., Syn. Pl. Glumac. 1: 312. 1854. TYPE: [Falkland/Malvinas Islands.] Ins. Maclov., "sec. specimen ex Herbo. Urville suh F. flabellata Lam.," d'Urville s.n. (holotype, P [URVILLE] not seen).

Poa controversa Steud., Syn. Pl. Glumac. 1: 260. 1854. Poa controversa var. minor Steud., Syn. Pl. Glumac. 1: 260. 1854. TYPE: Falkland Islands. Port William, "sub Dactylis caespitosa," [4 Sep. 1850], W. Lechler [(Hohenacker exsiccata) 106] (holotype, P [STEUD-385] not seen; isotype, P [STEUD] fragment at BAA not seen, P [STEUD-385] fragment at US 2851272!).

Sesleria americana Nees ex Steud., Syn. Pl. Glumac. 1: 296. 1854. TYPE: Argentina. [Province Tierra del Fuego]: [Isla de los Estados], Staten Island, Sep. [1829], Chanticleer [the ship], [W. H. B. Webster s.n.] (holotype, CGE [LINDL] not seen).

Distribution and habitat. South Atlantic Ocean and Drake Passage to the Pacific Ocean, mainly or always on islands. Argentina (Isla de los Estados, SE Tierra del Fuego), Chile (Tierra del Fuego, Isla Diego Ramirez), Falkland Islands/Islas Malvinas, South Georgia Island, Tristan da Cunha (Gough Island) (Nicora, 1978; Groves, 1981; Moore, 1983). It was introduced and established in the Shetland Islands (Edmondson, 1980; Groves, 1981) and was probably introduced to Gough Island (Groves, 1981). Coastal

wet rocks, hluffs, and wet rocky slopes to 100 m, often associated with penguin colonies.

Nates. Reported from mainland Chile along the north side of the Straits of Magellan hased on the Cammerson type collection for Festuca flabellata. Moore (1983) indicates Commerson's collections from the Tierra del Fuego area were all from along the north shore of the Straits of Magellan and the adjacent Isla Isabella, in Chile. It is impossible to plot this collection precisely; moreover, the species has not been collected in this area since. Possibly the original location given on the Cammerson label was in error. Only four other collection localities for the species from west of Isla de los Estados are known to us (three cited above, and one cited by Nicora, 1978).

Additional specimens examined. ARGENTINA. Tierra del Fuego: [SE end of Isla Grande], Policarpo, R. N. P. Goodall 2272 (BAB, NA, SI). CHILE. Magallanes: Cape Horn, Hermit Island, J. D. Hooker 68 (K); Isla Diego Ramírez, E. E. Pisano-Valdés 3405 (CONC). UNITED KINGDOM. South Georgia Island: J. Smith M-1148 (K). Tristan da Cunha Islands: Gough Island, N. M. Wasce G207 (K).

3. Poa foliosa (Hook. f.) Hook. f., Handb. N. Zeal. Fl. 338. 1864. Basionym: Festuca faliasa Hook. f., Fl. Antarct. 1: 99, t. 55. 1844 [1845]. TYPE: [New Zealand.] Lord Auckland's Islands: abundant, especially in rocky places near the sea, on the ground forming large green tufts on the cliffs never far from the sea, Dec. 1840, Antart. Exp. 1839–1843, J. D. Hooker s.n. (holotype, K [H203/00969-288]!; isotypes, fragment CHR 278601 not seen, LE not seen).

Distribution and babitat. New Zealand. Sub-Antarctic Islands: islets around Stewart Island, Solander Islands, Antipodes Islands, Auckland Islands, Campbell Island, and Macquarie Island (Edgar, 1986). Coastal wet rocks, bluffs, and adjacent wet slopes.

Nates. Paa faliasa is similar to P. ramasissima in its leaf blade architecture, as the adaxial valleys are very shallow between the costal ridges and are covered by papillate long cells. It is presumed to be closely related to P. caakii, which also has papillate long cells between its veins. Paa faliasa has a web on the callus of the lemma, whereas P. caakii does not. Chromosome number: 2n = 28 (Edgar & Connor, 2000).

4. Poa hamiltonii Kirk, Trans. & Proc. New Zealand Inst. 27: 353. 1895. TYPE: New Zealand. Sub-Antarctic Islands, Macquarie Island, 1894, A. Hamilton s.n. (holotype, K [H2003/00969–295]!; isotype, WELT 66728 not seen). Distribution and habitat. New Zealand sub-Antarctic Islands: Macquarie Island (Edgar, 1986). Coastal wet rocks, bluffs, and adjacent wet slopes.

Notes. There are two collections at Kew with the same data. The holotype, K [H2003/00969-295], is a spindly plant that has the lacerate ligules of Poa cookii, and keys to this species, but in some respects approaches P. ramosissima. This sheet has "Poa Hamiltonii n.s." [in Kirk's handwriting?] and a diagnosis. A second sheet with the same collection information, K [H2003/00969-336]!, has only the determination "Poa foliosa," and is that species.

Edgar (1986) placed *Poa hamiltonii* in *P. cookii* but noted that, in the Macquarie Island plants, the panicles are overtopped by the leaves and are somewhat narrower, and spikelets are on average slightly shorter. From the specimens on the four sheets we have seen from Macquarie Island, in addition to the differences noted by Edgar (1986), it appears that *P. hamiltonii*, unlike *P. cookii*, lacks prickle hairs on the sheaths and adaxial surfaces of the leaf blades. Thus, until we have reviewed other collections from Macquarie Island, we prefer to maintain this as a separate species closely related to *P. cookii*.

5. Poa ramosissima Hook. f., Flora Antaretica 1(1): 101. 1844. [1845.] TYPE: [New Zealand]. Hab. Lord Auckland's Islands, "very common on the rocks overhanging the sea, trailing over banks, &c.," [hangs down from the cliffs and rocks near the sea, common, very stoloniferous, Nov. 1840], J. D. Hooker s.n. (holotype, K [H2003/00969–298]!).

Distribution and habitat. New Zealand. Sub-Antarctic Islands, Auckland Islands, and Campbell Island (Edgar, 1986). Wet rocks and bluffs by the coast.

Notes. The specimens seen entirely lack prickle hairs, and both surfaces of the leaf blades are covered in papillate long cells. Papillate long cells are an anatomical feature characteristic of the adaxial blade surfaces of $Poa\ cookii$, $P.\ foliosa$, and $P.\ hamiltonii$ but are not always expressed in $P.\ tennantiana$, and only weakly so in material examined of $P.\ flabellata$. Hooker identified two pubescence variations in the florets of $P.\ ramosissima$ —one with a web on the callus and one without—hut most material from both islands is webhed (Edgar, 1986). ITS DNA data place this species with $P.\ flabellata$ (Gillespie & Soreng, unpublished data). Chromosome number: 2n=28 (Hair, 1968).

6. Poa tennantiana Petrie, Subantarctic Is. N. Z. 2: 467. 1909. *Poa foliosa* var. *tennantiana* (Petrie)

Cheeseman, Man. N. Z. Fl. (ed. 2) 188. 1925. TYPE: New Zealand. The Snares, 9 Jan. 1890, T. Kirk (lectotype, designated by Edgar, 1986, New Zealand J. Bot. 24: 188, WELT 36063 not seen).

Distribution and habitat. New Zealand South Island and New Zealand sub-Antarctic Islands: islets off Stewart Island, Snares Islands, Antipodes Islands, and Auckland Islands (Edgar, 1986). Coastal wet rocks and adjacent slopes.

Notes. Poa tennantiana, unlike P. cookii, P. hamiltonii, P. foliosa, and P. ramosissima, sometimes lacks papillae on the long cells of the leaf hlade surfaces. (Edgar [1986] noted the adaxial leaf hlade surfaces of this species were minutely papillose, but papillae were not seen in the material we examined.) Like the first two species, but unlike the latter two, it has glabrous calluses. It also has moderately deep narrow valleys and broad blocky ridges on leaf hlade adaxial surfaces and pectinately scabrous sheaths as in P. cookii and P. flahellata. Chromosome number: 2n = 28 (Hair, 1968).

TZVELEVIA REDUCED TO A SECTION IN POA

Poa sect. Tzvelevia (E. B. Alexeev) Soreng & L. J. Gillespie, comb. et stat. nov. Basionym: Tzvelevia E. B. Alexeev, Byull. Moskovsk. Obshch. Isp. Prir., Otd. Biol. 90(5): 103. 1985. TYPE: Poakerguelensis (Hook. f.) Steud.

Notes. Monotypic.

Poa kerguelensis (Hook. f.) Steud., Syn. Pl. Glumac. 1: 257. 1854. Basionym: Triodia kerguelensis Hook. f., Fl. Antarct. 379, t. 138. 1847. Festuca kerguelensis (Hook. f.) Hook. f., Philos. Trans. 168: 22. 1879. Tzvelevia kerguelensis (Hook. f.) E. B. Alexeev, Byull. Moskovsk. Obshch. Isp. Prir., Otd. Biol. 90(5): 103. 1985. TYPE: [France. Kerguelen Islands]: Kerguelen's land, "rocky places, at 300–1200 ft." [on the debris of the rocks common "perennial" and alpine, Christmas Harhour, May 1840], J. D. Hooker 761 (holotype, K [H2003/00969–293]!; isotypes, CN not seen, CN CGE fragment at US 1127149!).

Distribution and habitat. Sub-Antarctic Islands of Indian Ocean: Heard Island and Kerguelen Islands. Wet rocky upland to alpine slopes. Reported also from Heard Island (J. M. R. Hughes & R. D. Seppelt 24053 [ADT not seen]; fide Australian Antarctic Data Center, 2007).

Notes. Paa kerguelensis was placed in its own genus, Tzvelevia, principally based on its having a long narrow hilum (Alexeev, 1985). The hilum seen in US material is actually between one fifth and one fourth the length of the fruit and narrowly ovoid, just at the long end of hilum lengths in species of Paa. We return it to Paa, as there is nothing else to distinguish the genus by, hut note that it needs further study as to its position in the genus.

POA SECT. PLICATAE FROM ARGENTINA AND CHILE IS PLACED IN POA SECT. ACUTIFOLIAE

Poa sect. Acutifoliae Pilg. ex Potztal, Willdenowia 5(3): 473. 1969. TYPE: Poa acutifalia Hauman [= P. planifalia Kuntze].

Poa sect. Plicatae Pilg. ex Potztal, Willdenowia 5(3): 472. 1969. TYPE: Poa plicata Hack.

Distributian ond habitat. Northwestern Patagonia from the province of Neuquén, Argentina, and adjacent valleys of the Andes of Chile (Coquimbo, Metropolitana, and Valparaíso), and Argentina north to the province of Salta, Argentina. Poa acinaciphylla (syn. Paa villaroelii Phil., fide Nicora, 1978; Soreng et al., 2003a): Argentina (Mendoza), Chile (Coquimbo, Metropolitana, and Valparaíso). Poa planifalia: Argentina (Mendoza), Chile (Metropolitana). Paa plicata: Argentina (Catamarca, La Rioja, Salta, and Tucumán; Negritto & Anton, 2000). Poa stepparia: Argentina (Neuquén; Nicora, 1978). Alpine stream margins and wet meadows and steppe, sometimes in subsaline conditions.

Nates. Paa plicata, the sole species of section Plicatae, is here formally placed in section Acutifaliae, which now contains four species: P. acinaciphylla, P. planifalia, P. plicata, and P. stepparia. The species have pungent leaf apices hut otherwise have ligule margins and leaf hlade surface anatomy typical of Poa (except for P. stepparia). Callus hairs, when present, arise from the dorsal side of the lemma as is typical for Paa (callus hairs in P. stepparia are not clearly distinct from the keel and marginal vein hairs), and the lemma surfaces are glabrous (except for hairs at the base of the keel and marginal veins in P. stepparia). The placement of P. stepparia needs further investigation.

Additional specimens examined. Poa acinaciphylla: AR-GENTINA. Mendoza: Laguna de Solsneado, P. M. Peterson & C. R. Annoble 11380 (US). CHILE. Coquimbo: Cordillera de Combarbala, Potrero Grande, Jiles 4829 (CONC). Metropolitana: Río Yerba Loca, R. J. & N. L. Soreng 7169 (CONC, US). Valparaíso: Laguna Castro, Zoellner 932 (CONC).

Poa planifolio: CHILE. **Metropolitanas:** above Embalse del Río Yeso, R. J. & N. L. Soreng 7160 (CONC, US).

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