



Giant Camels from the Cenozoic  
of North America

JESSICA A. HARRISON

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# Giant Camels from the Cenozoic of North America

*Jessica A. Harrison*



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## ABSTRACT

Harrison, Jessica A. Giant Camels from the Cenozoic of North America. *Smithsonian Contributions to Paleobiology*, number 57, 29 pages, 17 figures, 1985.—Seven genera of giant camels occurred in North America during the interval from the late Clarendonian to the early Holocene. *Aepycamelus* was the first camel to achieve giant size and is the only one not in the subfamily Camelinae. *Blancocamelus* and *Camelops* are in the tribe Lamini, and the remaining giant camels *Megatylopus*, *Titanotylopus*, *Megacamelus*, *Gigantocamelus*, and *Camelus* are in the tribe Camelini. *Megacamelus* is a late Hemphillian giant camel most closely related to *Gigantocamelus*. *Titanotylopus* is reserved for the brachyodont form from the Irvingtonian of Nebraska, and *Gigantocamelus* is reinstated for the broad-chinned, Blancan form.

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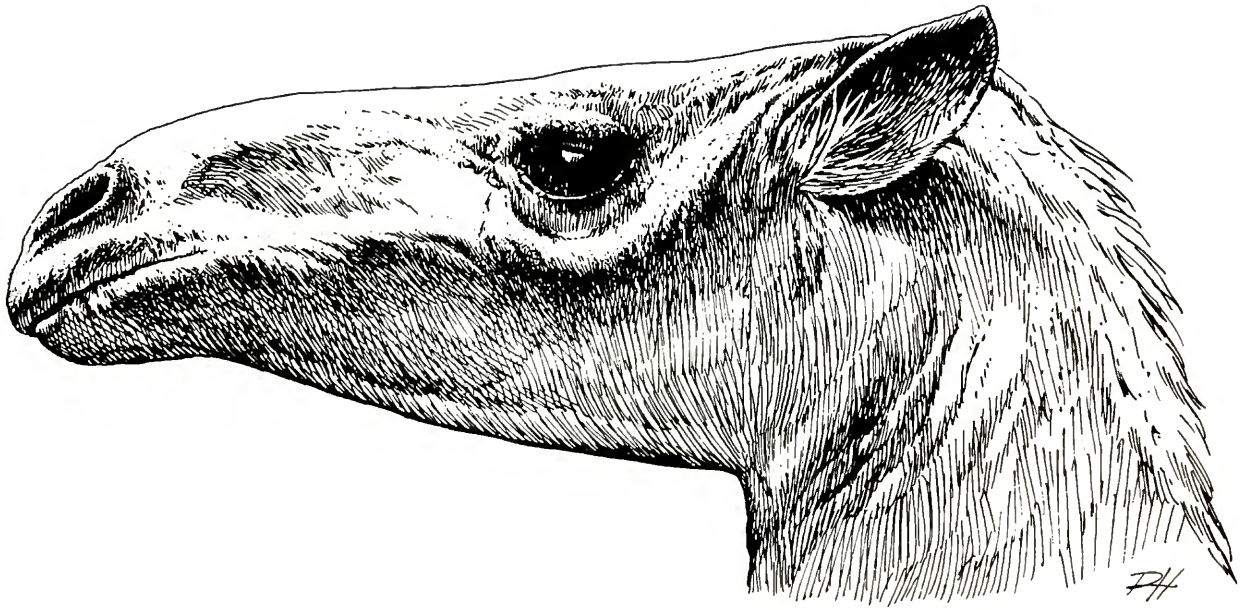
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FRONTISPIECE.—Reconstruction of the head of *Megacamelus merriami*.

# Giant Camels from the Cenozoic of North America

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## Introduction

Throughout the later Cenozoic, camels often figure as an abundant and diverse element of any fauna in which they occur. Until the late Pleistocene, when the group fell on hard times, the Camelidae must be accounted one of the more successful ungulate families. As in many other herbivore families, the earliest members of the Camelidae were of small body size. However, a trend toward gigantism can be observed throughout the later Cenozoic, from the Clarendonian into the Holocene.

Descriptions of very large camels are almost as abundant in the literature as their remains in late Cenozoic faunas. The confusing taxonomic history of the giant camels is such that, for every specific identification, there are many more referrals to "camelid, large, gen. et sp. indet." The purpose of this paper is to provide a temporal, geographic, and systematic framework for the large, late Cenozoic camels.

ACKNOWLEDGMENTS.—I am grateful for the use of specimens from the Frick Collection, Department of Vertebrate Paleontology, American Museum of Natural History (F:AM), the University of California Museum of Paleontology

(UCMP), the University of Nebraska State Museum (UNSM), and the University of Kansas Museum of Natural History (KUVF). I very much appreciate careful and constructive reviews by George Corner, Michael Voorhies, John Breyer, and Robert Emry. Drs. Corner and Voorhies were particularly generous in sharing with me their new information on *Titanotylopus*. The frontispiece was done by Robert Hynes.

## Phylogenetic Relationships

The cladogram in Figure 1 summarizes relationships within the Camelinae. It is interesting to note that the trend toward gigantism is far more apparent in the Camelini than in the Lamini. All of the genera comprising the Camelini can be called giants, but only two of the Lamini, *Camelops* and *Blancocamelus*, achieve a formidable body size. *Aepyamelus*, the only noncameline genus, represents the camels' earliest experimentation with gigantism.

The characters appearing at nodes 1 through 35 in the cladogram are listed below. The composition and apomorphies of the Protolabidini are from Honey and Taylor (1978:419–420), whereas those of the Lamini and Camelini appeared in part in Harrison (1979:3–8). More detailed discussion of the characters may also be found in those papers.

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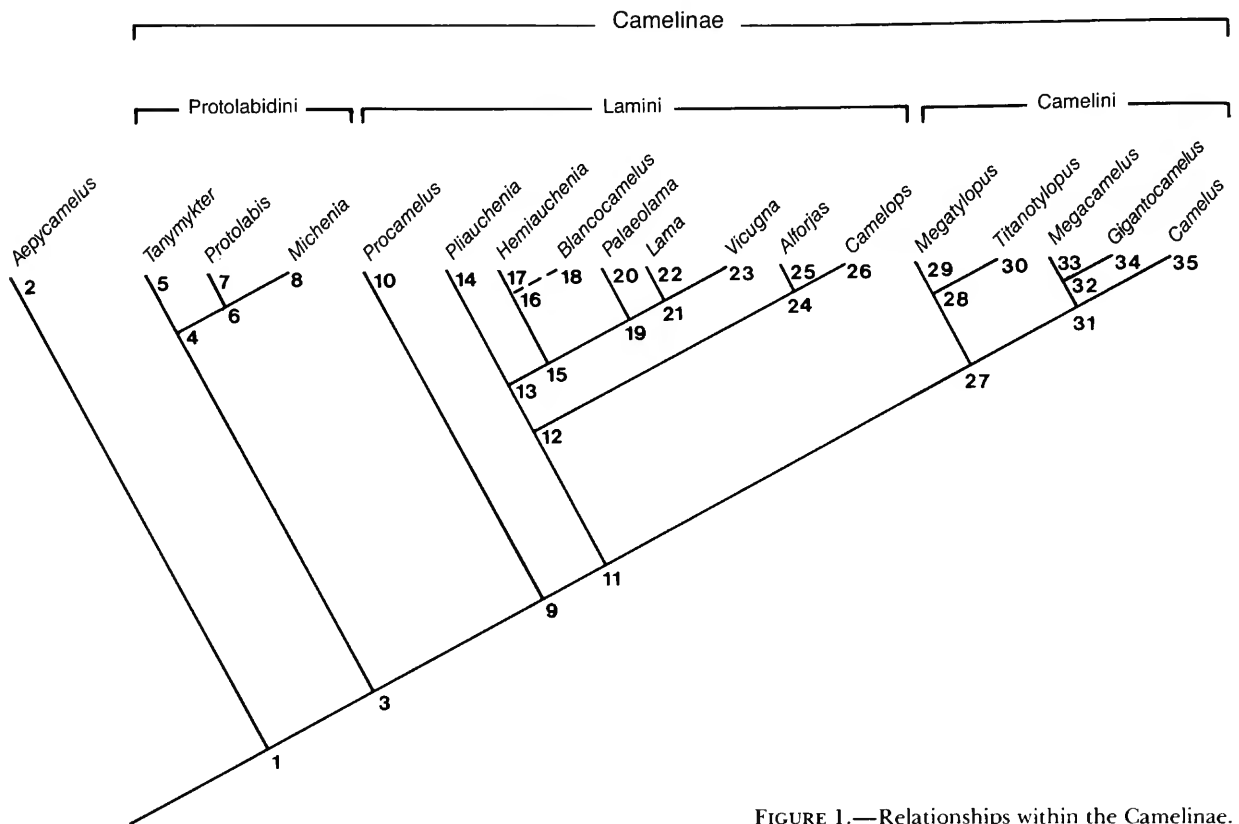


FIGURE 1.—Relationships within the Camelinae.

- Node 1. *Aepycamelus* shares with the Camelinae
- a. metastylid present on the lower molars
- Node 2. *Aepycamelus* is distinguished by
- a. extremely elongate limbs
  - b. extremely elongate cervical vertebrae
  - c. metapodials longer than the basal length of the skull
- Node 3. The Camelinae are united by
- a. weak buccinator fossa
  - b. elongate rostrum
- Node 4. The Protolabidini are united by
- a. narrow rostrum
  - b. laterally expanded anterior nares
- Node 5. *Tanymyktar* is distinguished by
- a. closely appressed  $P_1^1$  roots
- Node 6. *Protolabis* and *Michenia* are derived relative to *Tanymyktar* by
- a. the absence of elongate basioccipital tuberosities
  - b.  $P^2$  without strong, continuous lingual cingulum
  - c. auditory bulla less inflated with medial plates more compressed
  - d. moderate to strong buccinator fossa
- Node 7. *Protolabis* is distinguished by
- a. hypsodont molars
  - b. anteroposteriorly elongate  $M_3^3$
  - c. very weak to absent metastylid on lower molars
  - d. ventrally produced mandibular angle with weak to strong lateral flare
  - e. fused metapodials
  - f. elongate proximal phalanx with distal articular surface anteriorly extended
- Node 8. *Michenia* is derived relative to *Protolabis* in having
- a. short braincase
  - b. weak  $I^3-C^1$ , small  $C_1$
  - c. shallow symphysis
  - d. inflection of mandibular angle suppressed
- Node 9. The Camelinae exclusive of the Protolabidini are united by
- a. metacarpal length exceeds metatarsal length
  - b. metapodials completely fused
  - c.  $I^1$  absent
- Node 10. *Procamelus* is the sister taxon to the remaining camelines. It retains several primitive characters but has almost completed the loss of  $I^2$



- Node 11. The Lamini share with the Camelini
- $I^2$  absent
  - $P_2^2$  absent
  - raised posterolateral edges on the proximal end of the proximal phalanx
- Node 12. The Lamini are united by
- in cross section the anterior end of the nasals form a high, bilobed arch (Harrison, 1979, fig. 3)
  - anteroexternal style (= llama buttress) present on lower molars
- Node 13. *Pliauchenia*, *Hemiauchenia*, (?)*Blancocamelus*, *Palaeolama*, *Lama*, and *Vicugna* share
- reduced lacrimal vacuity
  - shortened rostrum
- Node 14. *Pliauchenia* is primitive in all known characters to the remaining Lamini.
- Node 15. *Hemiauchenia*, (?)*Blancocamelus*, *Palaeolama*, *Lama*, and *Vicugna* are united by
- small  $P_1^1$
  - small or absent  $P_3$
- Node 16. *Hemiauchenia* and *Blancocamelus* share
- extremely elongated metapodials
- Node 17. *Hemiauchenia* is distinguished by
- extremely elongated cervical vertebrae
- Node 18. *Blancocamelus* is distinguished by
- great size
- Node 19. *Palaeolama*, *Lama*, and *Vicugna* share
- $P_1^1$  absent
  - reduced maxillary fossa
  - moderate to strong anteroexternal style on lower molars
- Node 21. *Lama* and *Vicugna* are derived relative to *Palaeolama* in having
- $P_3$  absent
  - metacarpal length subequal to metatarsal length
  - strong anteroexternal style on lower molars
  - greatly reduced lacrimal vacuity
  - extremely retracted nasals
  - greatly reduced  $P_4$
- Node 22. *Lama* is distinguished by
- callosities on the inner foreleg
- Node 23. *Vicugna* is distinguished by
- hypsodont lower incisors
- Node 24. *Alforjas* and *Camelops* are derived relative to other lamines in having
- moderately hypsodont to very hypsodont molars
  - cheek teeth narrow in relation to length
- Node 25. *Alforjas* is primitive in all known characters relative to *Camelops*
- Node 26. *Camelops* is derived relative to *Alforjas* in having
- cheek teeth much more hypsodont
  - $P_1^1$  absent
  - $P_3$  absent
  - dorsal surface of the mandibular condyle transversely concave
  - suspensory ligament scar extends to center of proximal phalanx and has a raised center
- Node 27. The Camelini are united by
- angular process on mandible enlarged and strongly inflected
  - large postglenoid foramen
  - long postglenoid process on skull with correspondingly large facet on mandibular condyle
  - $C_1^1$  enlarged and rounded in cross section, especially in males
  - ventrally flattened auditory bulla
  - diastemal crest on mandible low and rounded
  - reduced maxillary fossa
  - thickened, heavy premaxilla
- Node 28. *Megatylopus* and *Titanotylopus* share
- reduced  $P_1$
  - reduced  $P_3$
- Node 29. *Megatylopus* is distinguished by
- reduced  $P^3$
  - cheek teeth higher crowned than *Titanotylopus*
- Node 30. *Titanotylopus* is derived relative to *Megatylopus* in having
- $P_1$  absent
  - $P_3$  more reduced than in *Megatylopus*
  - larger body size than *Megatylopus*
- Node 31. *Megacamelus*, *Gigantocamelus*, and *Camelus* share
- metapodials shorter in relation to basal length of the skull
  - cheek teeth more hypsodont than *Megatylopus* or *Titanotylopus*
- Node 32. *Megacamelus* and *Gigantocamelus* share
- spatulate lower incisors
  - splayed  $C_1$
- Node 33. *Megacamelus* is primitive in all characters relative to *Gigantocamelus* except for
- $I^3$  enlarged and caniniform
- Node 34. *Gigantocamelus* is distinguished by
- short, blunt chin with a shortened ramal symphysis
  - greater size than *Megacamelus*
  - lower incisors arrayed almost transversely
  - $I^3$  absent or vestigial
- Node 35. *Camelus* is distinguished by
- reduced paroccipital process
  - metapodials subequal in length and shorter than the basal length of skull
  - maxillary fossa reduced or absent
  - zygomatic arch straight in lateral view
  - retracted nasals
  - center of suspensory ligament scar raised

### *Aepycamelus* Macdonald, 1956

TYPE-SPECIES.—*Aepycamelus giraffinus* Macdonald, 1956:198 (= *Alticamelus giraffinus* Matthew and Cook, 1909:402).

*Aepycamelus* is the geologically oldest of the giant camels, ranging from the Barstovian through the early Hemphillian and occurring through the southern and western United States (Figure 2). When Marsh (1894:274) described *Procamelus altus* from Oregon, he based it solely upon an isolated calcaneum. Cope (1894:869) took almost instant exception to the designation of such an undiagnostic element as a type specimen. Matthew (1901:429), under the impression that the type of *P. altus* was more extensive, described *Alticamelus* from northeastern Colorado and named *A. altus* the genotypic species. When he became aware of the indeterminate nature of the type of *A. altus*, Matthew renamed the Colorado material *A. giraffinus* (Matthew and Cook, 1909:402). Macdonald (1956:198) maintained that *Alticamelus* was a nomen vanum and proposed *Aepycamelus* as a replacement name, with *A. giraffinus* as the new genotypic species.

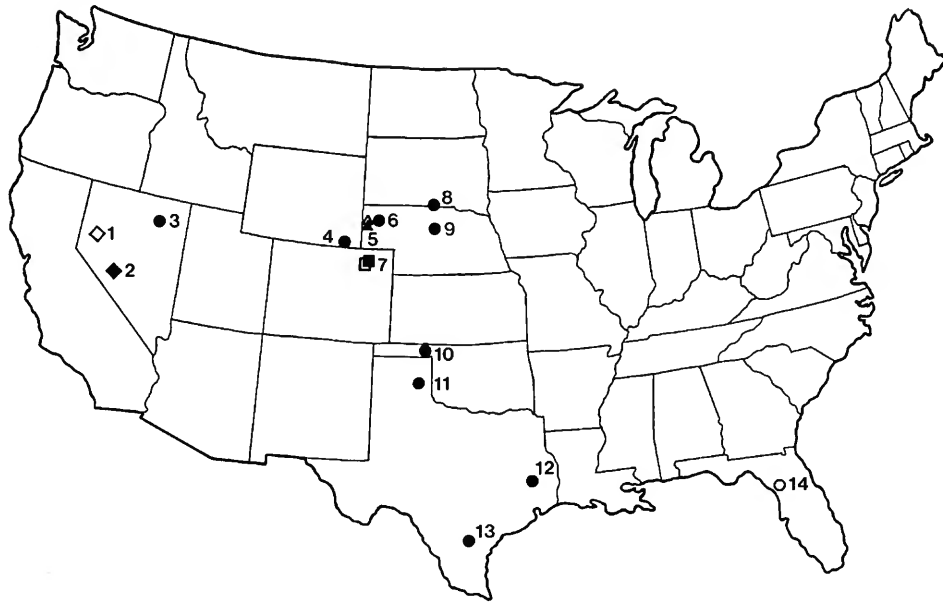
All of the species of *Aepycamelus* are noted for extremely elongate, slender limbs and cervical vertebrae. The teeth are quite brachyodont, and the dental formula is  $I_3^1-3C_1^1P_4^1M_3^3$ . When present,  $I^{1,2}$  are reduced to stumps as in the type of *A. giraffinus*.  $P_2^2$  is always present and less reduced than in *Procamelus*.

Matthew and Cook (1909:402) described *Alticamelus procerus* from the Snake Creek beds of Nebraska, and later Matthew (1924:187) described a second species, *Alticamelus priscus*, from the Sheep Creek beds. Matthew (1924:187) also referred material from Snake Creek to *Alticamelus leptocolon* described by him from the Pawnee Creek area of Colorado. Davidson (1923:399) described *Alticamelus alexandrae* from Barstow, California, but Macdonald (1949:190) referred this form to *Hesperocamelus*. Henshaw (1942:153) named a species from Tonopah, Nevada, *Alticamelus? stocki*. Macdonald (1956:199) described *Aepycamelus bradyi* from the Nightin-

gale Road fauna, Truckee Formation, Nevada. Leidy (1886:12) described *Auchenia major* from Mixson, Florida; Leidy and Lucas (1896:53) later changed the name to *Procamelus major*. Although Simpson (1930:196) referred this material to *Megatylopus major*, it is more likely an advanced species of *Aepycamelus* (pers. comm., Beryl E. Taylor, 1973).

*Aepycamelus bradyi* is the largest of the species, followed by *A. giraffinus*. *Aepycamelus procerus* is smaller than *A. giraffinus* and has completely lost  $I^{1,2}$ . *Aepycamelus stocki* is smaller than *A. procerus* but larger than *A. leptocolon* and retains  $I^{1,2}$ . *Aepycamelus priscus* is the smallest of the lot. In *A. bradyi* and *A. stocki* the premolars are a bit more reduced than in the other species of *Aepycamelus* but are not so reduced as in *Procamelus*. *Aepycamelus bradyi* has as well an almost complete internal crescent on  $P^3$ . Although such a project is not within the scope of this paper, it may be seen that the genus *Aepycamelus*, often difficult to distinguish from *Procamelus*, would benefit considerably from a revision.

A number of specimens have been referred simply to *Aepycamelus*. Hesse (1936:66) referred two partial jaws from Beaver Quarry, Oklahoma, to ?*Alticamelus*. Savage (1941:701) referred a series of metapodials and phalanges from the Optima fauna of Oklahoma, but the dimensions of these specimens are more characteristic of *Hemiauchenia*. Macdonald (1966:12) described an associated partial skeleton and jaws from the Camp Creek fauna of Nevada. Skinner, Skinner, and Gooris (1968:432) reported a partial radius-ulna of *Aepycamelus* from Turtle Butte, South Dakota. Webb (1969:147) referred two partial metapodials from Burge Quarry, Nebraska, to *Aepycamelus* sp. Patton (1969:149) referred limb elements from the Cold Spring fauna and the Lapara Creek fauna of Texas to *Aepycamelus* sp. Forsten (1970:48) referred an astragalus and some teeth from the Trail Creek fauna of Wyoming to ?*Alticamelus*; Cassiliano (1980:55) changed the reference to *Aepycamelus* sp. Galusha (1975:54) listed *Aepycamelus* cf. *A. priscus* in a preliminary faunal list from the Box Butte Formation of Nebraska.



Key			
◇	<i>A. bradyi</i>	5	Sheep Creek/Snake Creek, Nebraska
◆	<i>A. stocki</i>	6	Box Butte, Nebraska
□	<i>A. leptocolon</i>	7	Pawnee Creek, Colorado
■	<i>A. giraffinus</i>	8	Turtle Butte, South Dakota
△	<i>A. procerus</i>	9	Burge, Nebraska
▲	<i>A. priscus</i>	10	Beaver Quarry, Oklahoma
○	<i>A. major</i>	11	Clarendon, Texas
●	<i>A. sp.</i>	12	Cold Spring, Texas
1	Nightingale Road, Nevada	13	Lapara Creek, Texas
2	Tonopah, Nevada	14	Mixson, Florida
3	Camp Creek, Nevada		
4	Trail Creek, Wyoming		

FIGURE 2.—Geographic distribution of *Aepycamelus*.

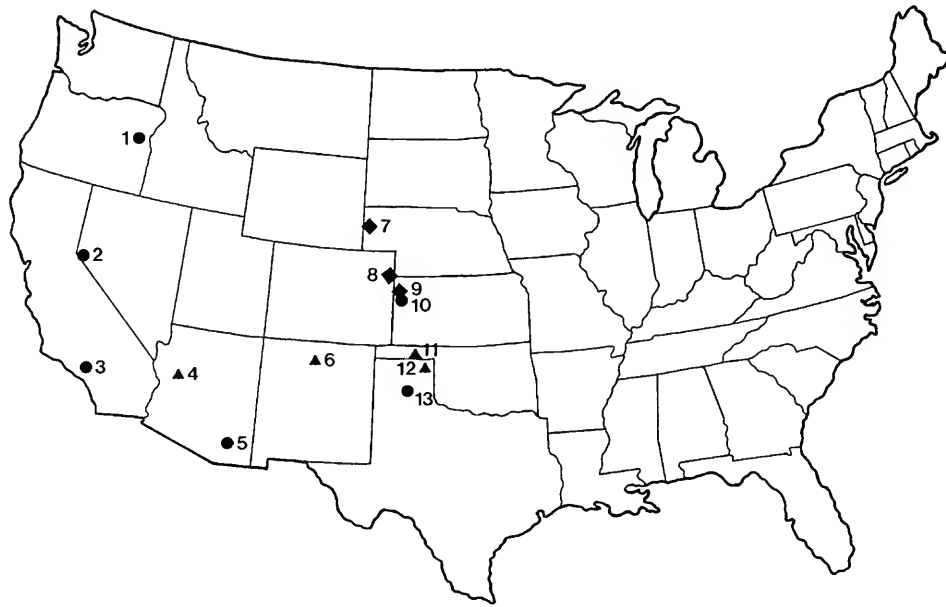
### *Megatylopus* Matthew and Cook, 1909

TYPE-SPECIES.—*Pliauchenia gigas* Matthew and Cook, 1909:396.

*Megatylopus* is the geologically oldest of the giant Camelini. It ranges from the late Clarendonian into early Blancan throughout the western United States (Figure 3). *Megatylopus* was originally proposed as a subgenus of *Pliauchenia* and was later elevated to generic rank. The type provenience of *M. gigas*, the genotypic species, is the late Hemphillian ZX Bar fauna, Snake Creek

Formation, Nebraska (Skinner, Skinner, and Gooris, 1977:360). *Megatylopus* shares with *Titanotylopus* a tendency to reduce the third and fourth premolars. Its teeth are higher crowned than those of *Titanotylopus*, but both genera are more brachyodont than *Megacamelus*, *Gigantocamelus*, and *Camelus*. The limbs of *Megatylopus*, particularly the metapodials, are not shortened in relation to the basal length of the skull.

Additional species of *Megatylopus* are *M. cochranii* (Hibbard and Riggs, 1949:854) from Keefe Canyon, Kansas, and *M. matthewi* (Webb,



Key			
◆	<i>M. gigas</i>	6	Chamita Formation
▲	<i>M. matthewi</i>	7	Snake Creek, Nebraska
●	<i>M. sp.</i>	8	Wray area, Colorado
1	Little Valley, Juniper Creek, and Black Butte, Oregon	9	Edson Quarry, Kansas
2	Smiths Valley, Nevada	10	Found Quarry, Kansas
3	Kinsey Ranch, California	11	Optima, Oklahoma
4	Wikieup, Arizona	12	Coffee Ranch, Texas
5	Redington and Camel Canyon, Arizona	13	Axtel, Currie Ranch, and Christian Ranch, Texas

FIGURE 3.—Geographic distribution of *Megatylopus*.

1965:42) from the Coffee Ranch fauna of Texas. *Megatylopus cochrani* was originally described as *Pliauchenia cochrani* but was transferred to *Megatylopus* by Webb (1965:42). Although these workers have commented on the similarity of *M. cochrani* to *Camelops*, Corner and Voorhies (pers. comm., 1984) believe that *M. cochrani* may be more closely related to *Titanotylopus*. *Megatylopus matthewi* is distinguished from *M. gigas* by the complete internal crescent on P<sup>3</sup>, the greater reduction of P<sub>3</sub>, and a deeper maxillary fossa. In addition to other specimens of *M. matthewi*, Dalquest (1980:110) described five thoracic vertebrae preserved in articulation. Based on their structure, he proposed a large dorsal hump, py-

ramidal in profile rather than rounded. Table I lists additional occurrences of *Megatylopus*.

A fourth species of *Megatylopus*, *M. major*, was originally described by Leidy (1886:12) as *Auchenia major* and based upon an isolated astragalus from Mixson, Florida. Leidy and Lucas (1896:53) transferred the species to *Procamelus* and assigned to it a composite dentition, also from Mixson, which they believed had come from a single individual. Simpson (1930:196) changed the identification to ?*Megatylopus major*, noting that the species was nearly as large as *M. gigas*. He also distinguished *M. major* as having broader cheek teeth than *M. gigas* and a complete internal crescent on P<sup>3</sup>. Subsequent excavation

TABLE 1.—Additional occurrences of *Megatylopus*.

Species	Locality	Reference
<i>M. gigas</i>	Wray area, Colorado	Cook, 1922:11
	Edson Quarry, Kansas	Harrison, 1983:8
<i>M. matthewi</i>	Chamita Formation, New Mexico	MacFadden, 1977:791
	Optima, Oklahoma	Schultz, 1977:75
	Wikieup, Arizona	MacFadden, Johnson, and Opdyke, 1979:357
	Ocote, Mexico	Dalquest and Mooser, 1980:18
<i>M. cochrani</i>	White Bluffs, Washington	Gustafson, 1978:48
<i>M. sp. or ?M.</i>	Smiths Valley, Nevada	Macdonald, 1959:885
	Little Valley, Oregon	Shotwell, 1970:98
	Juniper Creek, Oregon	Shotwell, 1970:98
	Black Butte, Oregon	Shotwell, 1970:96
	Kinsey Ranch, California	Miller and Downs, 1974:11
	Axtel, Texas	Schultz, 1977:89
	Currie Ranch, Texas	Schultz, 1977:89
	Christian Ranch, Texas	Schultz, 1977:89
	Redington, Arizona	Lindsay, 1978:270
	Camel Canyon, Arizona	Lindsay, 1978:270
	Found Quarry, Kansas	Bennett, 1979:12

of the Mixson bone bed produced a much larger sample of this camel, currently housed in the Frick Collection of the Department of Vertebrate Paleontology, American Museum of Natural History. Skulls and mandibles bearing complete dentitions as well as diagnostically elongate metapodials and cervical vertebrae indicate that *M. major* should be transferred to *Aepyocamelus major* (pers. comm., Beryl E. Taylor, 1973).

#### *Titanotylopus* Barbour and Schultz, 1934

TYPE-SPECIES.—*Titanotylopus nebraskensis* Barbour and Schultz, 1934:291.

Barbour and Schultz (1934:291) described *Titanotylopus nebraskensis* based on a single mandible from a Pleistocene gravel pit near Red Cloud, Nebraska (Figure 4). Other than a proximal phalanx from another Pleistocene gravel pit, no more specimens have been referred to *T. nebraskensis*. Based upon this material, Webb (1965) and Breyer (1976) felt that the differences between *Gigantocamelus* and *Titanotylopus* warranted distinction only at the specific level. Cor-

ner and Voorhies (pers. comm., 1984) have since identified additional material, which is referable to *T. nebraskensis* and which they believe validates the generic independence of these two taxa. I agree with Corner and Voorhies that the name *Titanotylopus* should be applied to a single species, *T. nebraskensis*, as yet known only from early Irvingtonian localities in Nebraska.

The type mandible of *Titanotylopus* is very large, 662 mm long, with a dental formula of  $I_3C_1P_2M_3$ . The mandibular symphysis is long and extends well beyond the large canines. The non-spatulate incisors are arrayed in an arc. There is no indication of  $P_1$ .  $P_3$  is broken, but its position is indicated by alveoli for its two roots.  $P_4$ , like the molars, has sustained some breakage. All of the cheek teeth are quite brachyodont.

In their study of *Gigantocamelus spatulus* from Keefe Canyon, Hibbard and Riggs (1949) classified as female those jaws with small  $C_1$  and no  $P_1$ , whereas jaws classified as male had large  $C_1$ , and  $P_1$  was present. Webb (1965:36) felt that the type of *T. nebraskensis* fell within the range of variation assigned to females and hence attached

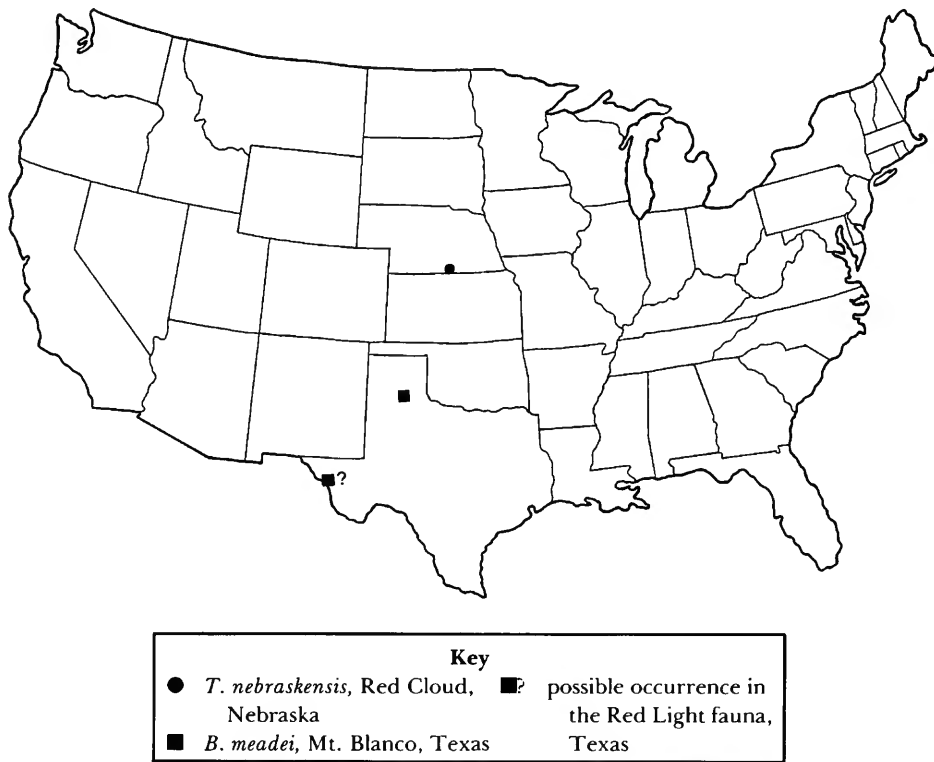


FIGURE 4.—Type-localities of *Titanotylopus nebraskensis* and *Blancocamelus meadei*.

little significance to the missing  $P_1$ . His synonymy of *Titanotylopus* and *Gigantocamelus* has been followed by most authors except Hibbard (Skinner et al., 1972:114). Considerations of  $P_1$  aside, the highly derived chin and greater degree of hypsodonty in *G. spatulus* as well as the much shorter distance between  $C_1$  and  $P_3$  in *T. nebraskensis* must weigh heavily against the congenerity of these two species.

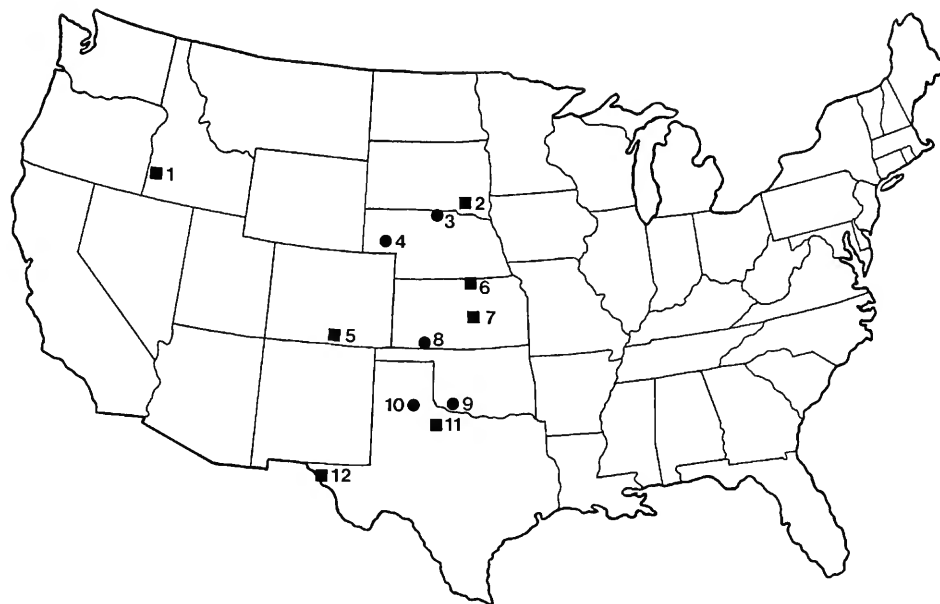
#### *Gigantocamelus* Barbour and Schultz, 1939

**TYPE-SPECIES.**—*Gigantocamelus fricki* Barbour and Schultz, 1939:17 (= *Pliauchenia spatula* Cope, 1893:70; = *Gigantocamelus spatulus* Meade, 1945:531; = *Titanotylopus spatulus* (Meade) fide Webb, 1965:36).

*Gigantocamelus* is known from Blancan localities throughout the central and western United States (Figure 5). The giant camel from the Blanco beds of Texas was first described as

*Pliauchenia spatula* by Cope (1893:70). Barbour and Schultz (1939:17) subsequently described a large sample of giant camel from Lisco, Nebraska, as *Gigantocamelus fricki*. Meade (1945:531) recognized these camels as the same species and united them under the name *Gigantocamelus spatulus*. Hibbard and Riggs (1949:844) followed Meade when they described a third large sample of this giant camel from Keefe Canyon, Kansas.

*Gigantocamelus* is a much more hypsodont camel than *Titanotylopus*. Its chin is blunt with the ramal symphysis extending only a few centimeters beyond the large, splayed canines. The spatulate lower incisors are arrayed almost transversely. In *Titanotylopus* the chin is long,  $C_1$  is large but not greatly splayed, and the nonspatulate incisors are arrayed in an arc.  $P_1$  is present in *Gigantocamelus* and absent in *Titanotylopus*. As indicated in the preceding section, many workers have followed Webb (1965) and Breyer (1976)



Key	
●	<i>G. spatulus</i>
■	<i>G. sp.</i>
1	Grand View, Idaho
2	Delmont, South Dakota
3	Sand Draw, Nebraska
4	Lisco, Nebraska
5	Donnelly Ranch, Colorado
6	White Rock, Kansas
7	Kentuck, Kansas
8	Keefe Canyon, Kansas
9	Holloman, Oklahoma
10	Mt. Blanco, Texas
11	Gilliland, Texas
12	Hudspeth, Texas

FIGURE 5.—Geographic distribution of *Gigantocamelus*.

in synonymizing *Gigantocamelus* and *Titanotylopus*. I believe that the above characters, in addition to the new data from Corner and Voorhies, support the generic validity of *Gigantocamelus*.

Several other workers have made reference to *Gigantocamelus* in one or more of its previous taxonomic incarnations. Dumble (1894:559), Wortman (1898:128), Gidley (1903:627), Matthew (1901:423; 1909:120), and Merriam (1917:435) noted the presence of *Pliauchenia spatula* at Mt. Blanco. Matthew (1899:75) listed material from Goodnight, Texas, as *P. spatula*, but this is more likely *Megatylopus*. From the Saw Rock fauna of Kansas, Hibbard (1953:407) referred a toe bone to *Gigantocamelus* cf. *G. spatulus*. Although I have not seen this specimen, the measurements fall within the range of *Megaca-*

*melus*. If indeed the toe can be identified as *Gigantocamelus*, it would be the earliest occurrence of this genus. From the Gilliland fauna of Texas, Hibbard and Dalquest (1962:86) referred a distal radius, some distal metapodials, an astragalus, and three phalanges to ?*Gigantocamelus*, and later added a cervical vertebra and changed the identification to ?*Titanotylopus*, reflecting Webb's synonymy. Strain (1966:50) referred two distal metapodials from the Hudspeth fauna of Texas to *Gigantocamelus* sp. Semken (1966:164) referred a partial calcaneum from the Kentuck fauna of Kansas to *Gigantocamelus* sp. A number of foot and limb bones from the Grand View fauna of Idaho were referred to *Titanotylopus* sp. by Shotwell (1970:96). Hibbard (Skinner et al., 1972:114) referred some material from the Sand

Draw fauna of Nebraska to *G. spatulus*. Martin and Harksen (1974:14) referred a partial mandible from the Delmont fauna of South Dakota to *Titanotylopus*. Dalquest (1975:42) described additional specimens of *T. spatulus* from Mt. Blanco. Eshelman (1975:47) referred a proximal ulna from the White Rock fauna of Kansas to *Gigantocamelus* sp. Hager (1975:14) referred some tooth fragments and foot bones from the Donnelly Ranch fauna of Colorado to *Gigantocamelus* sp. Dalquest (1977:260) described a radius of *T. spatulus* from the Holloman fauna of Oklahoma. Corner and Voorhies (pers. comm., 1984) regard the specimens from the Gilliland and Holloman faunas as generically indeterminate between *Gigantocamelus* and *Titanotylopus*. Ferrusquia-Villafranca (1978:255) listed *Gigantocamelus mexicanus* and *G. magnus* from the Mexican faunal assemblage. Dalquest (1974:196–197) noted that these two species were based

upon the same type material, *G. mexicanus* being the senior synonym, and identified them as *Camelops*.

### *Megacamelus* Frick, 1929

TYPE-SPECIES.—*Pliauchenia merriami* Frick, 1921:358 (= *Megacamelus merriami*, new combination).

*Megacamelus* is presently known only from the late Hemphillian of the southwestern United States (Figure 6). Frick (1921:358) described *Pliauchenia merriami* from Mt. Eden, California, and later (1929:107) named *Megacamelus blicki* from Keams Canyon, Arizona. Study indicates that these camels are conspecific. The Mt. Eden camel is clearly not *Pliauchenia* and although Webb (1965:36) referred it to *Titanotylopus*, it does not belong to that genus for reasons discussed below. Therefore, a new taxonomic com-



FIGURE 6.—Geographic distribution of *Megacamelus*.



ination, *Megacamelus merriami*, is proposed for the giant camels from Mt. Eden and Keams Canyon.

***Megacamelus merriami* (Frick, 1921), new combination**

FIGURES 7-16

*Pliauchenia merriami* Frick, 1921:358.

*Megacamelus blicki* Frick, 1929:107.

**HOLOTYPE.**—UCMP 23483, anterior portion of the upper jaws bearing right and left  $I^3$ ,  $C^1$ ,  $P^1$ ; anterior portion of the lower jaws bearing right  $I_{2,3}$ ,  $C_1$ ,  $P_1$ , partial  $P_3$  and left  $I_{2,3}$ ,  $C_1$ ,  $P_1$ ; distal humerus, proximal radius-ulna, distal radius-ulna, scaphoid, lunar, cuneiform, pisiform, distal tibia, astragalus, calcaneum, navicular, cuboid, ectocuneiform, proximal metatarsus, 2 distal metapodials, 6 proximal phalanges, 5 medial phalanges, 7 distal phalanges, and 8 sesamoids.

**REFERRED SPECIMENS.**—From Mt. Eden, California: UCMP 23416, left  $P^3$ ; UCMP 23433, right partial upper molar; UCMP 23783, left  $I_2$ ; UCMP 23790, right  $I_3$ ; UCMP 23791, left  $I_3$ ; UCMP 23435, left  $M_{2(?)}$ ; UCMP 23789, left  $M_{1(?)}$ .

From Keams Canyon, Arizona (all of the following are F:AM numbers): skulls, 23201, 23202, 23202A, 23203, 23207; partial skulls, 23203A, 23203B, 23203C, 23204, 23205, 23205A, 23208, 23209, 104395; partial maxillae, 23206, 23210; mandibles, 23216, 23218, 23220, 23230, 23231, 23232, 23233, 23235, 23239A; rami, 23217A, 23217B, 23219, 23221, 23222, 23223A, 23223B, 23225, 23226, 23227, 23228, 23229, 23233A, 23234, 23240B; partial rami, 23224, 23317, 23320, 104274; incisors, 104329, 104330, 104333, 104334, 104335; canine, 104383; associated postcrania, 23312 (astragalus, calcaneum, tarsal fragments, and metatarsus); atlas, 104291, 104292, 104293, 104294, 104295; axis, 104284, 104299; cervical vertebrae, 104281, 104282, 104283, 104285, 104286, 104287, 104288, 104289, 104290, 104297, 104298; thoracic vertebrae, 104286, 104296, 104305; lumbar vertebrae, 104277, 104300, 104301, 104304; sacra, 104280,

104302; scapulae, 23245, 23246, 23247, 23248, 23249, 23250, 23251, 23275, 23276, 104278, 104279, 104408, 104607; humeri, 23254, 23255, 23256, 23257, 23258, 104405, 104406, 104407; radius-ulnae, 23260, 23261, 23261A, 23262, 23263, 23264, 23265, 23266, 23267, 23268, 23269, 23270, 104400, 104600; scaphoids, 104311, 104351, 104352, 104376, 104377, 104603; lunars, 104309, 104323, 104324, 104349, 104350, 104361; cuneiforms, 104325, 104332, 104389, 104605; pisiforms, 104312, 104345, 104356, 104370, 104371, 104372; trapezoids, 104316, 104341, 104359; magna, 104339, 104360, 104390; unciforms, 104307, 104308, 104342, 104343, 104344, 104346, 104364; metacarpi, 23277, 23278, 23279, 23280, 23281, 23282, 23283, 23284, 23285, 23301, 104401; pelvis, 104303, 104396; femora, 23290, 23291, 23292, 23293, 23294, 23295, 104403, 104404; patellae, 104358, 104394; tibiae, 23271, 23296, 23297, 104399; distal fibula, 104347, 104375; astragali, 104229, 104230, 104231, 104232, 104233, 104234, 104235, 104236, 104272, 104273, 104602; calcanea, 104237, 104238, 104239, 104240, 104601; naviculars, 104310, 104331, 104373, 104374; cuboids, 104306, 104336, 104353, 104354, 104355, 104362, 104363; ectocuneiforms, 104313, 104322, 104378, 104379, 104380, 104381, 104382, 104384, 104385, 104386, 104392, 104393; metatarsi, 23302, 23303, 23304, 23305, 23306, 23307, 23308, 23313; partial metapodials, 23314, 23316, 104326, 104327, 104328, 104397, 104398, 104402; sesamoids, 104315, 104317, 104318, 104319, 104320, 104321, 104337, 104338, 104348, 104357, 104365, 104366, 104367, 104368, 104369, 104387, 104388, 104606; proximal phalanges, 104241, 104242, 104243, 104244, 104245, 104246, 104247, 104248, 104249, 104250, 104251, 104252, 104253, 104254, 104255, 104256, 104257, 104258, 104259, 104260, 104261, 104262, 104263, 104265, 104266, 104267, 104268, 104269; medial phalanges, 104210, 104211, 104212, 104213, 104214, 104215, 104216, 104217,

TABLE 2.—Measurements (cm) of the skull and upper dentition of *Megacamelus merriami*, new combination, from Keams Canyon (O.R. = observed range,  $\bar{X}$  = sample mean, s.d. = standard deviation).

Element	No.	O.R.	$\bar{X}$	s.d.
Length from premaxilla to occipital crest	3	69.30–82.05	73.65	
Length from premaxilla to occipital condyles	3	61.67–71.35	65.54	
Width of muzzle across I <sup>3</sup>	3	6.68–8.25	7.68	
Width of muzzle across C <sup>1</sup>	3	7.69–8.59	8.16	
Minimum width at post-orbital constriction	4	6.71–9.65	8.45	
Maximum width across zygomatic arches	2	26.40–27.34	26.87	
Width of occipital condyles	7	9.25–11.55	10.14	0.76
Diastema I <sup>3</sup> -C <sup>1</sup>	2	2.10–2.30	2.20	
Diastema C <sup>1</sup> -P <sup>1</sup>	2	3.08–3.27	3.17	
Diastema P <sup>1</sup> -P <sup>3</sup>	3	7.10–8.83	7.81	
I <sup>3</sup> length	2	2.20–3.19	2.69	
I <sup>3</sup> width	2	1.22–1.73	1.47	
C <sup>1</sup> length	3	2.88–3.46	3.15	
C <sup>1</sup> width	3	1.82–2.33	2.11	
P <sup>1</sup> length	4	1.94–2.52	2.20	
P <sup>1</sup> width	4	1.38–1.74	1.54	
P <sup>3</sup> length	6	1.84–3.05	2.62	0.44
P <sup>3</sup> width	7	1.88–2.03	1.95	0.06
P <sup>4</sup> length	7	2.85–3.25	3.03	0.18
P <sup>4</sup> width	6	2.75–3.31	3.04	0.22
M <sup>1</sup> length	6	4.30–4.75	4.54	0.15
M <sup>1</sup> width	6	3.80–4.45	4.11	0.27
M <sup>2</sup> length	8	4.89–5.66	5.42	0.35
M <sup>2</sup> width	6	3.82–4.75	4.46	0.34
M <sup>3</sup> length	7	5.40–6.13	5.69	0.28
M <sup>3</sup> width	6	3.86–4.39	4.20	0.19
I <sup>3</sup> -M <sup>3</sup> length	3	37.77–41.10	39.06	
P <sup>3</sup> -M <sup>3</sup> length	5	18.96–20.03	19.41	
dP <sup>2</sup> length	2	1.39–1.50	1.44	
dP <sup>2</sup> width	2	0.42–0.82	0.62	
dP <sup>3</sup> length	3	3.29–3.60	3.43	
dP <sup>3</sup> width	3	2.45–2.65	2.55	
dP <sup>4</sup> length	3	3.72–4.15	3.86	
dP <sup>4</sup> width	3	3.00–3.20	3.07	
dP <sup>2</sup> -M <sup>2</sup> length	1	16.00		
dP <sup>3</sup> -M <sup>2</sup> length	2	14.88–15.96	15.42	

104218, 104219, 104220, 104221, 104222, 104223, 104224, 104225, 104270, 104604; distal phalanges, 104226, 104227, 104228, 104271, 104314, 104340, 104391.

TABLE 3.—Measurements (cm) of the mandible and lower dentition of *Megacamelus merriami*, new combination, from Keams Canyon (O.R. = observed range,  $\bar{X}$  = sample mean, s.d. = standard deviation).

Element	No.	O.R.	$\bar{X}$	s.d.
Maximum length of ramal symphysis	5	12.81–17.10	14.89	
width of mandibular condyle	8	4.35–5.93	5.36	0.51
Diastema I <sup>3</sup> -C <sub>1</sub>	4	0.63–1.06	0.88	
Diastema C <sub>1</sub> -P <sub>1</sub>	5	3.36–4.38	3.90	
Diastema P <sub>1</sub> -P <sub>3</sub>	6	7.26–8.23	7.55	0.41
C <sub>1</sub> length	6	3.19–3.88	3.48	0.25
C <sub>1</sub> width	6	2.19–3.17	2.58	0.41
P <sub>1</sub> length	6	2.23–2.63	2.36	0.18
P <sub>1</sub> width	6	1.23–1.89	1.49	0.29
P <sub>3</sub> length	8	1.87–2.38	2.08	0.16
P <sub>3</sub> width	7	1.16–1.35	1.24	0.06
P <sub>4</sub> length	10	2.72–3.26	2.96	0.18
P <sub>4</sub> width	9	1.72–2.07	1.88	0.12
M <sub>1</sub> length	12	4.05–5.05	4.41	0.33
M <sub>1</sub> width	12	2.32–2.90	2.64	0.19
M <sub>2</sub> length	12	4.86–5.44	5.09	0.28
M <sub>2</sub> width	14	2.27–3.53	2.96	0.38
M <sub>3</sub> length	11	5.98–6.84	6.49	0.26
M <sub>3</sub> width	9	2.61–3.43	3.04	0.29
C <sub>1</sub> -M <sub>3</sub> length	5	36.05–39.41	38.09	
P <sub>3</sub> -M <sub>3</sub> length	6	20.34–21.74	20.71	0.51
dP <sub>2</sub> length	3	1.06–1.31	1.18	
dP <sub>2</sub> width	3	0.12–0.13	0.13	
dP <sub>3</sub> length	3	2.05–2.29	2.18	
dP <sub>3</sub> width	3	1.20–1.27	1.23	
dP <sub>4</sub> length	2	4.21–5.30	4.75	
dP <sub>4</sub> width at posterior cusp	2	2.06–2.16	2.11	

DESCRIPTION.—The extensive sample from Keams Canyon contains several fine skulls and mandibles as well as a wealth of postcrania (Figures 7–11). With one exception (F:AM 23312), none of the material is associated. The Keams Canyon camel compares well with that described by Frick (1921) from Mt. Eden. Both exhibit very large, caniniform I<sup>3</sup>, C<sup>1</sup>, and P<sup>1</sup>, a massive premaxilla, and heavy anterior ramus with large C<sub>1</sub> and large, caniniform P<sub>1</sub>. The size and proportions of the postcrania from both localities are comparable.

The skull of *M. merriami* is long with a flattened dorsal profile and a deep, massive rostrum. The occipital crest is a broad fan that extends

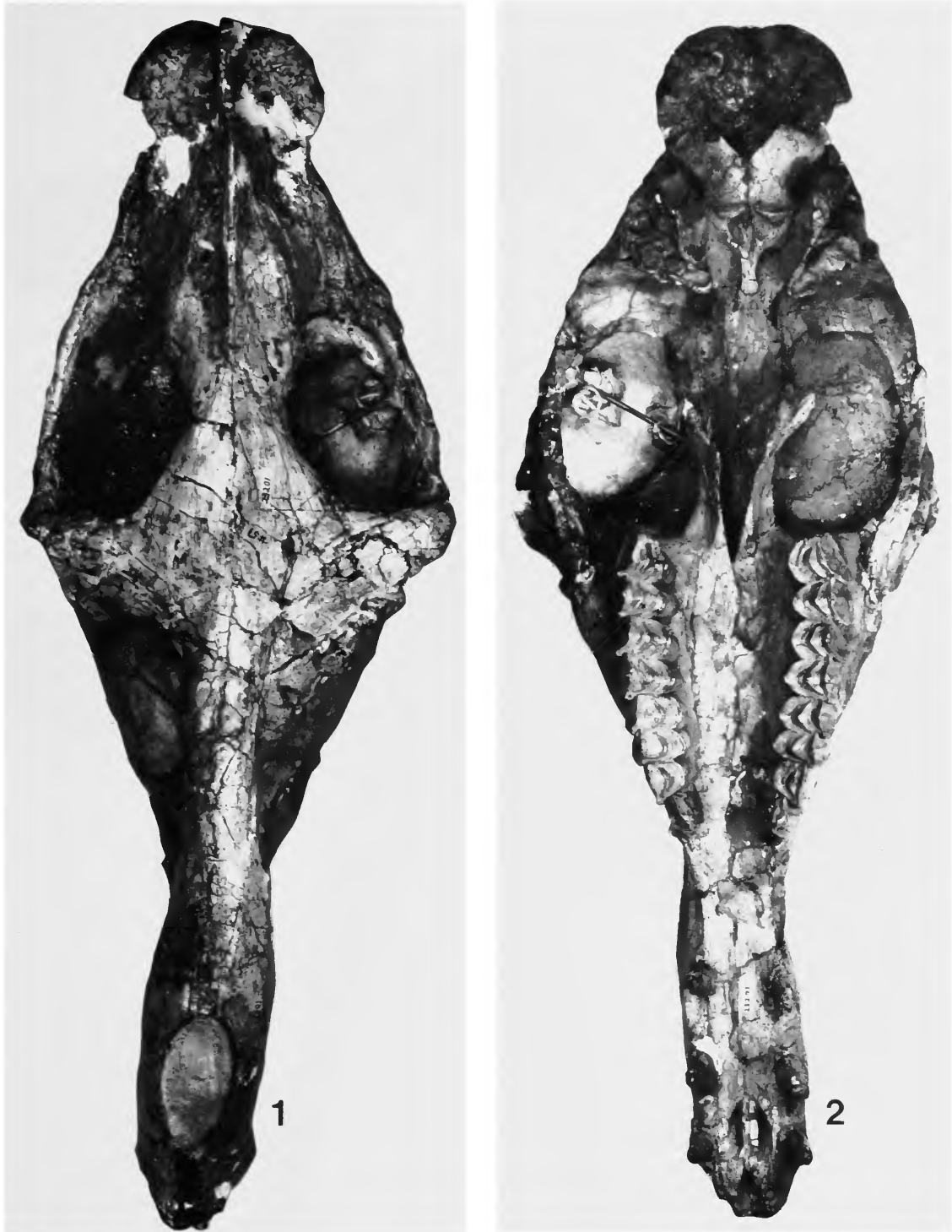


FIGURE 7.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona, F:AM 23201, skull: 1, dorsal view; 2, occlusal view. ( $\times \frac{1}{4}$ .)



FIGURE 8.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona: 1, F:AM 23201, skull, lateral view; 2, 3, F:AM 23232, mandible, occlusal and lateral views. ( $\times \frac{1}{4}$ .)

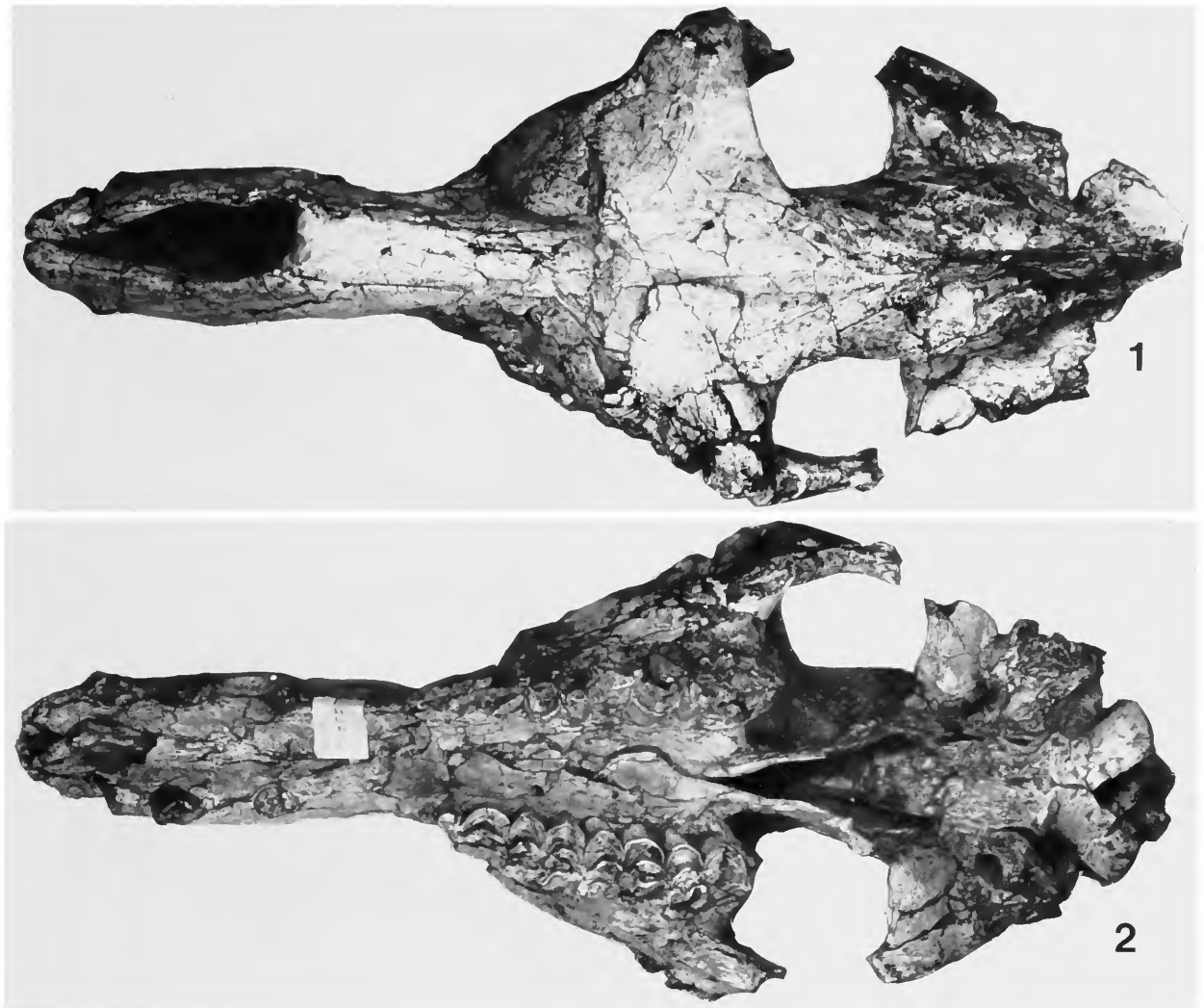


FIGURE 9.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona, F:AM 23202, skull: 1, dorsal view; 2, occlusal view. ( $\times \frac{1}{4}$ .)

well posterior to the occipital condyles. The sagittal crest is likewise well developed. The orbit is circular in outline followed by a strong postorbital bar. A triangular lacrimal vacuity is present in most of the Keams Canyon *M. merriami*, but it is a highly variable feature. In one specimen, F:AM 23202, it is present on the left side of the skull, but reduced to a slit on the right side (Figure 9). Another skull, F:AM 23202A, bears well-developed lacrimal vacuities on both sides (Figure 10). Although Meade (1945:531) re-

ported the presence of a lacrimal vacuity in *M. spatulus* from Mt. Blanco, Hibbard and Riggs (1949:846) reported its absence and, moreover, suggested that the opening in the Mt. Blanco skulls could represent an artifact of preservation.

The upper dentition consists of  $I^3$ ,  $C^1$ ,  $P^{1,3,4}$ ,  $M^{1-3}$ .  $I^3$  is always present and, as mentioned above, is large and caniniform.  $C^1$  is large and deviates slightly from a vertical orientation. Presumably, when Barbour and Schultz (1939:24) stated that the canines of the Keams Canyon

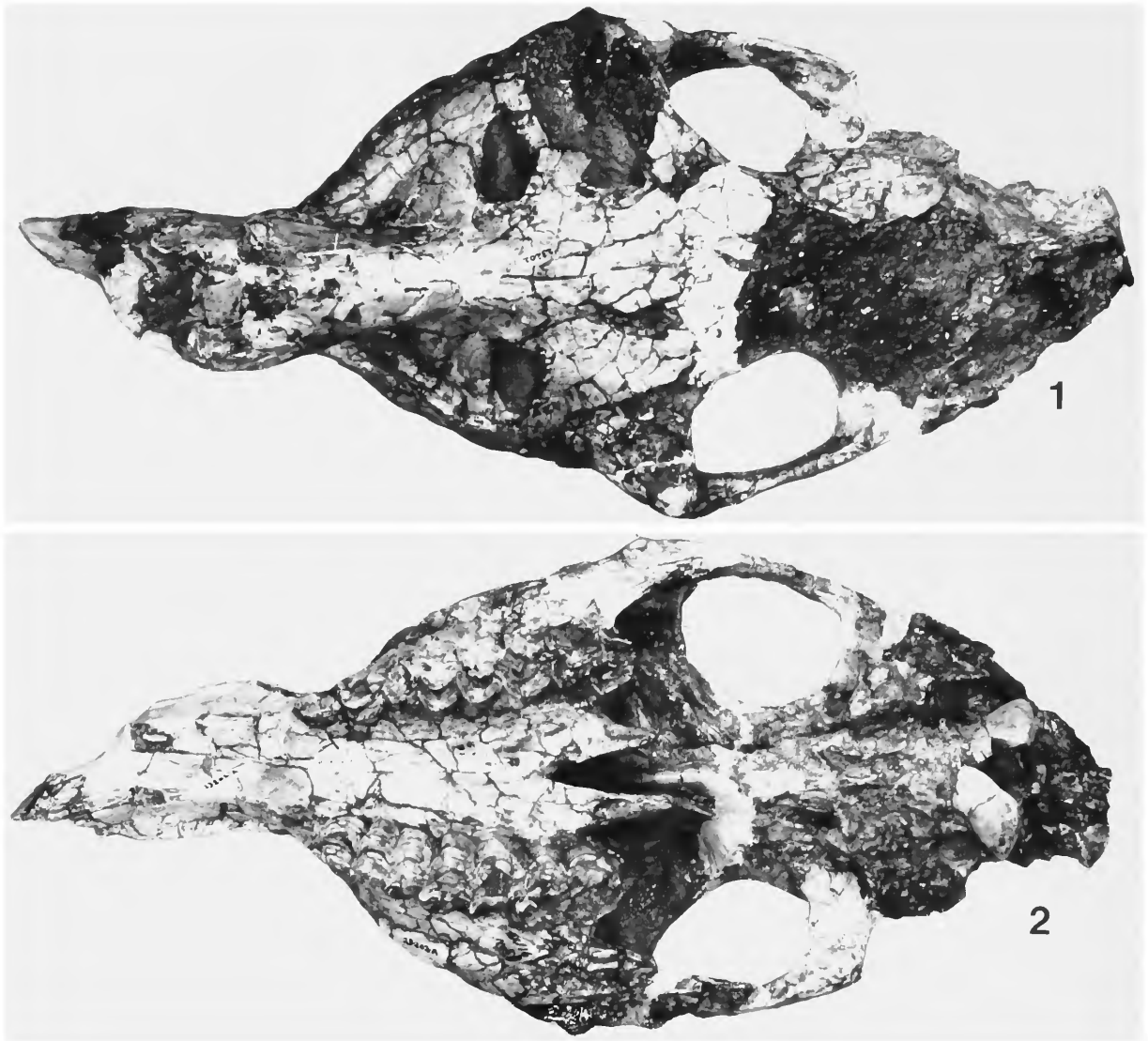


FIGURE 10.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona, F:AM 23202A, skull: 1, dorsal view; 2, occlusal view. ( $\times \frac{1}{4}$ .) Note the large lacrimal vacuities.

camel were not enlarged, it was in comparison to the extreme development observed in *Gigantocamelus spatulus* and *Titanotylopus nebraskensis*.  $P^1$  is caniniform and only slightly smaller than  $I^3$ .  $P^3$  is large with an incomplete internal crescent.  $P^4$  is not much longer than  $P^3$  but much wider due to its complete internal crescent. Only the parastyle shows much development on  $P^4$ , but a strong parastyle and mesostyle are present on

each molar. The teeth are higher crowned than those of *Megatylopus gigas* or *Titanotylopus nebraskensis*.

The mandible is long and massive but still smaller than that of *Gigantocamelus spatulus* or *Titanotylopus nebraskensis*. The mandibular proportions of *M. merriami* agree well with those of *G. spatulus*; however, *M. merriami* does not display the degree of variation in the symphyseal

TABLE 4.—Measurements (cm) of the postcrania of *Megacamelus merriami*, new combination, from Keams Canyon (O.R.= observed range,  $\bar{X}$  = sample mean, s.d. = standard deviation).

Element	No.	O.R.	$\bar{X}$	s.d.	Element	No.	O.R.	$\bar{X}$	s.d.
Atlas					Tibia				
length of centrum	2	6.75–6.80	6.77		length	1	67.00		
posterior height	1	9.36			distal width	2	10.95–11.45	11.20	
Scapula					Astragalus				
maximum length	2	61.53–64.06	62.79		length (tibial to tarsal surface)				
maximum width	1	38.35			medial	7	8.19–8.98	8.57	0.34
glenoid fossa					lateral	7	8.85–9.80	9.47	0.30
antero-	3	9.37–10.56	9.87		distal width	7	6.03–6.65	6.39	0.21
posterior					Calcaneum				
transverse	3	9.17–10.61	9.66		length	4	17.68–19.56	18.64	
Humerus					maximum antero-	4	7.21–8.30	7.90	
length	1	53.27			posterior				
distal width across					Metatarsus				
trochlea	5	10.60–11.70	11.14		length	5	48.37–50.58	49.44	
Radius-ulna					proximal width	2	8.06–9.37	8.71	
maximum length	4	72.65–85.70	80.33		distal width	4	10.01–10.75	10.35	
articular length	4	66.03–75.50	71.22		Proximal phalanx				
proximal width	6	10.19–11.52	10.87	0.43	length	21	11.05–13.80	12.41	0.84
distal width	5	10.40–12.62	11.39		proximal width	21	4.55–5.73	5.08	0.35
Metacarpus					distal width	22	3.82–5.14	4.42	0.41
length	7	48.39–54.29	51.08	2.07	Medial phalanx				
proximal width	8	8.11–11.64	9.12	1.10	length	18	7.22–8.47	7.76	0.34
distal width	7	11.38–12.30	11.88	0.33	proximal width	18	3.74–4.99	4.19	0.27
Femur					distal width	17	3.26–4.21	3.79	0.25
length	1	60.89			Distal phalanx				
proximal width	1	16.07			length	6	3.16–4.16	3.71	0.34
distal width	2	13.98–15.34	14.66		width of articular	6	2.45–2.80	2.60	0.13
width of patellar surface	3	5.07–5.79	5.46		surface				

region observed in *G. spatulus* by Meade (1945:532) or Hibbard and Riggs (1949:847). Meade, in the Mt. Blanco sample, and Hibbard and Riggs, in the Keefe Canyon sample, found jaws with widely splayed and canines and transversely arrayed incisors as well as jaws with more vertically oriented canines and more conventionally arrayed incisors. Hibbard and Riggs (1949), Webb (1965), and Breyer (1976) have attributed such variation to sexual dimorphism. In the Keams Canyon sample the incisors are procumbent and arrayed in a more shallow arc than in

*Titanotylopus*. The canines are very large, but only slightly splayed. No specimen displays the degree of canine flare and incisor-row bluntness characteristic of *G. spatulus* (Cope, 1893, pl. 21; Barbour and Schultz, 1939, fig. 9; Meade, 1945, pl. 54; Hibbard and Riggs, 1949, fig. 8). A groove is present between the median incisors on the ventral symphyseal surface, as noted by Cope (1893:71) and Meade (1945:532).

The lower dentition consists of  $I_{1-3}$ ,  $C_1$ ,  $P_{1,3,4}$ ,  $M_{1-3}$ . The incisors have spatulate crowns that with much wear assume a more rounded, peg-



FIGURE 11.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona: 1, 4, F:AM 23216, mandible, occlusal and lateral views; 2, 3, F:AM 23239A, partial ramus bearing deciduous  $P_{2-4}$  and  $M_1$  emerging, occlusal and lateral views. ( $\times \frac{1}{4}$ .)

like appearance. The canines bear the strong anterointernal and posterior enamel ridges typical of *G. spatulus*.  $P_1$  is present in all specimens and usually well developed. An exception is F:AM 23218, one of the smallest individuals, in which  $P_1$  is correspondingly small. The cheek tooth series is quite similar to that of *M. spatulus*. Meade (1945:533) reports an anteroexternal

style or "llama buttress" on  $M_{2,3}$  of one specimen and notes the presence of this feature in figures of *M. spatulus* from Lisco. No indication of a "llama buttress" is present in *M. merriami*.

The limbs and feet of *M. merriami*, especially the metapodials and phalanges, do not exhibit the shortening in relation to the basal length of the skull seen in *G. spatulus*. Hence, Barbour and



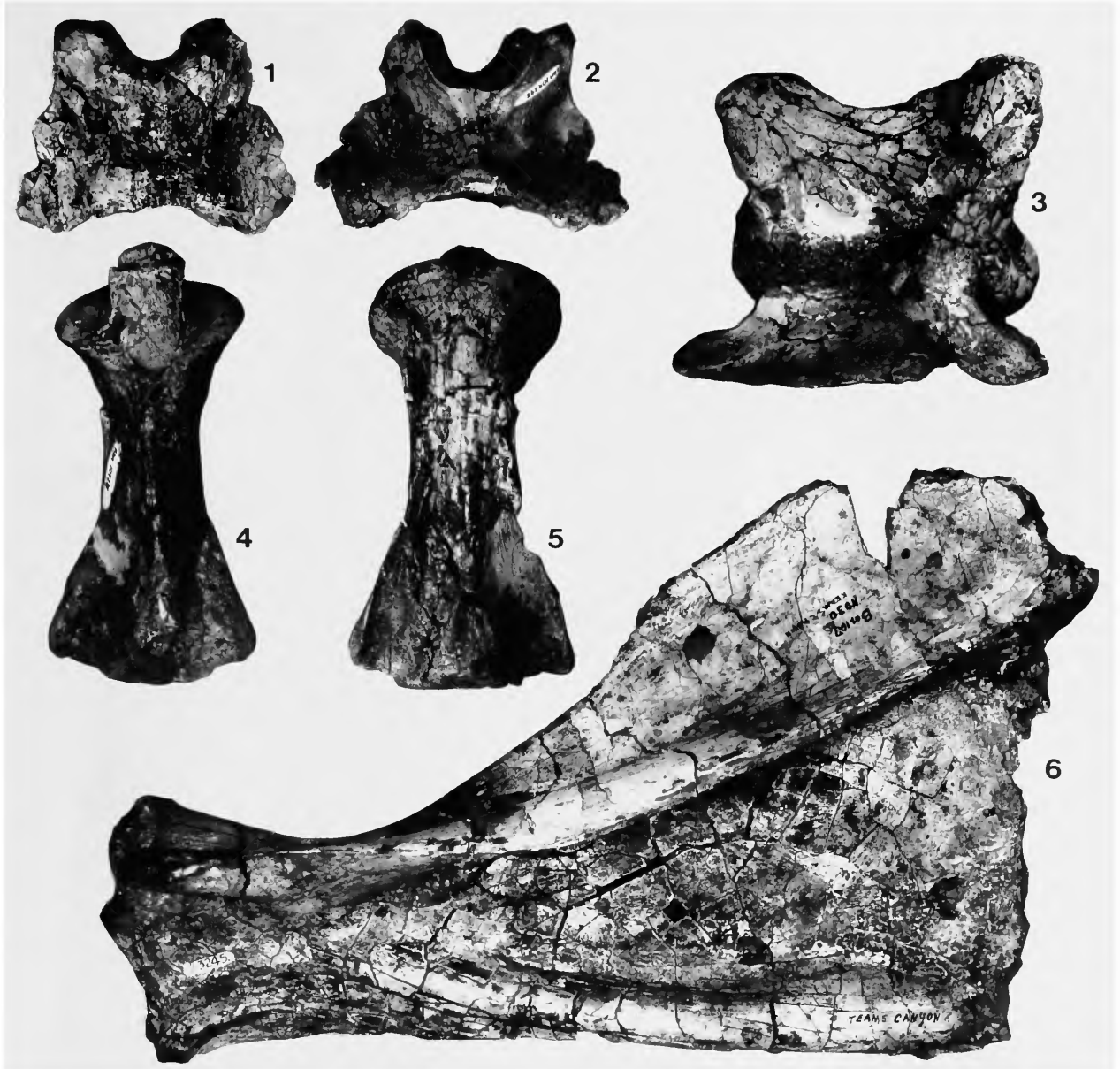


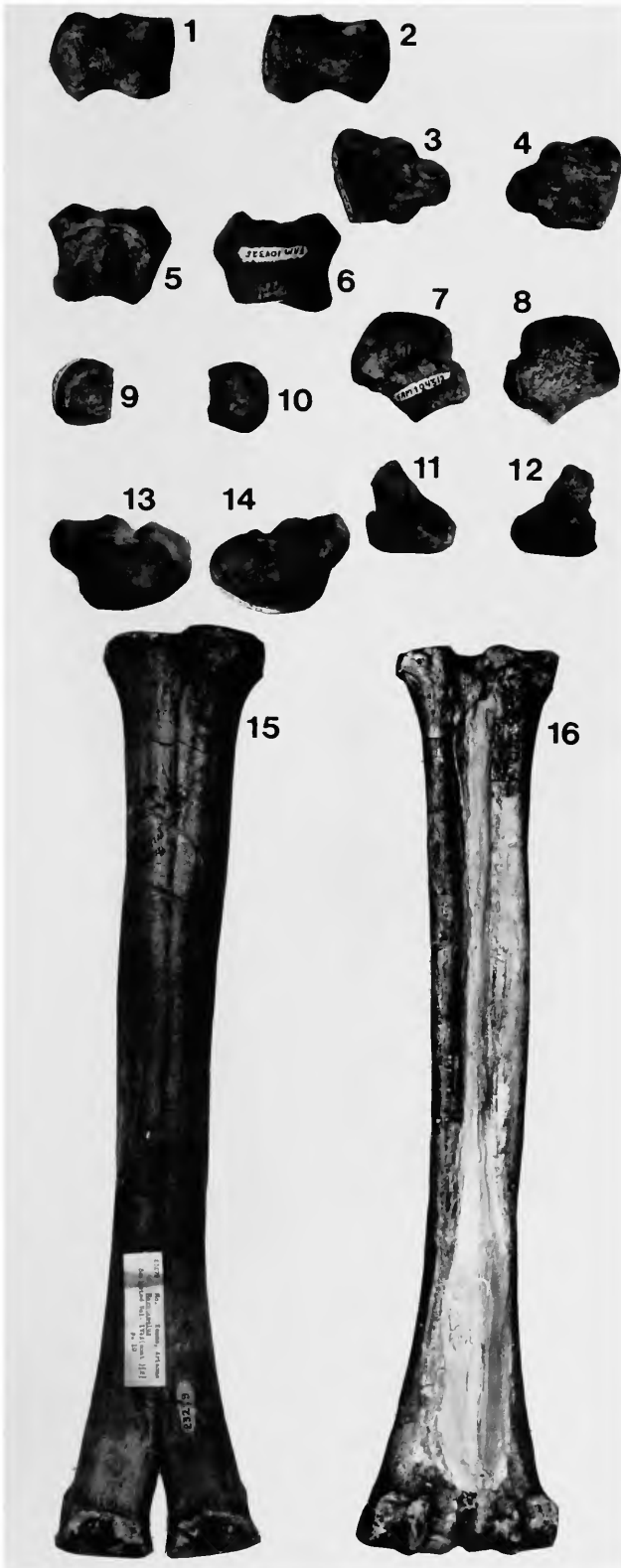
FIGURE 12.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona: 1, 2, F:AM 104293, atlas, dorsal and ventral views; 3, F:AM 104281, sixth cervical vertebra, lateral view; 4, 5, F:AM 104284, axis, dorsal and ventral views; 6, F:AM 23245, left scapula, lateral view. ( $\times \frac{1}{4}$ .)

Schultz (1939:24) remarked that "the skeletal elements appear to be more massive in the Nebraska form" (= *G. spatulus*). The limbs of *M. merriami* are, however, shorter and stockier than those of *Megatylopus*.

DISCUSSION.—*Megacamelus merriami* is most closely related to *Gigantocamelus spatulus* but differs from it in the presence of the large, canini-form  $I^3$ , smaller size, less shortened limbs, and lower-crowned teeth. Breyer (1983:305) re-



FIGURE 13.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona: 1, 2, F:AM 23256, humerus, anterior and posterior views; 3, 4, F:AM 23262, radius-ulna, anterior and posterior views. ( $\times \frac{1}{4}$ .)



ferred the Keams Canyon material to *Titanotylopus nebraskensis* on the basis of the projection of the mandibular symphysis beyond the canines. This condition is primitive for camelines and hence not a valid criterion. Moreover, several characters such as the presence of  $P_1$  in all specimens, greater degree of hypsodonty, and the greater distance between  $C_1$  and  $P_3$  preclude the referral of the Keams Canyon camel to *Titanotylopus*.

#### *Camelus* Linnaeus, 1758

TYPE-SPECIES.—*Camelus dromedarius* Linnaeus, 1758:65.

*Camelus* is the smallest of the giant camels. The two extant species, *G. dromedarius* (monogibbose) and *C. bactrianus* (digibbose), range throughout most of the arid and semi-arid regions of the Old World. *Camelus bactrianus* is native to Chinese Turkestan and Mongolia, where small wild populations still exist (Walker, 1964:1374). Both *C. bactrianus* and *C. dromedarius* have been domesticated for several thousand years, and the original native range of the latter species can no longer be determined.

*Camelus* has an extensive fossil record in the Pleistocene of the Old World and has been found in association with human artifacts and remains (Gauthier-Pilters and Dagg, 1981:5). Camels migrated from North America via Beringea near the end of the Tertiary, probably during the late Ruscinian. *Camelus* (*Paracamelus*) Schlosser (1903) occurs in several late Pliocene localities in the People's Republic of China. As yet no fossil material of *Camelus* or *Paracamelus* has been recovered from North America.

FIGURE 14.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona: 1, 2, F:AM 104311, scaphoid, medial and lateral views; 3, 4, F:AM 104323, lunar, medial and lateral views; 5, 6, F:AM 104325, cuneiform, medial and lateral views; 7, 8, F:AM 104312, pisiform, medial and lateral views; 9, 10, F:AM 104359, trapezoid, posteromedial and anterolateral views; 11, 12, F:AM 104360, magnum, proximal and distal views; 13, 14, F:AM 104343, unciform, proximal and distal views; 15, 16, F:AM 23279, metacarpus, anterior and posterior views. ( $\times \frac{1}{4}$ .)



FIGURE 15.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona: 1, 2, F:AM 23293, femur, anterior and posterior views; 3, 4, F:AM 23296, tibia, anterior and posterior views. ( $\times \frac{1}{4}$ .)

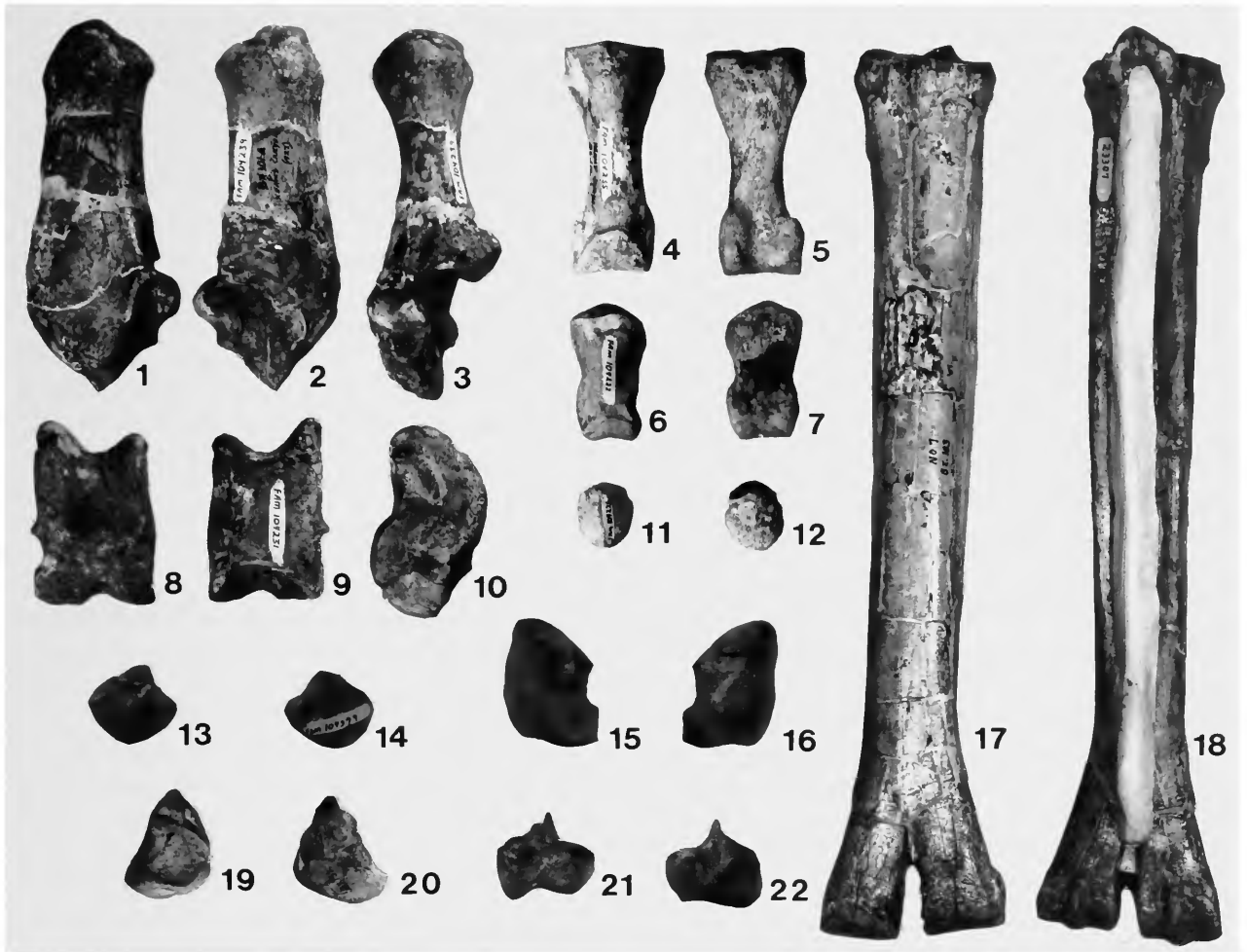


FIGURE 16.—*Megacamelus merriami*, new combination, from Keams Canyon, Arizona: 1-3, F:AM 104239, calcaneum, lateral, medial, and anterior views; 4, 5, F:AM 104255, proximal phalanx, anterior and posterior views; 6, 7, F:AM 104222, medial phalanx, anterior and posterior views; 8-10, F:AM 104231, astragalus, anterior, posterior and lateral views; 11, 12, F:AM 104226, distal phalanx, anterior and posterior views; 13, 14, F:AM 104379, ectocuneiform, proximal and distal views; 15, 16, F:AM 104366, cuboid, proximal and distal views; 17, 18, F:AM 23307, metatarsus, anterior and posterior views; 19, 20, F:AM 104310, navicular, proximal and distal views; 21, 22, F:AM 104375, distal fibula, medial and lateral views. ( $\times \frac{1}{4}$ .)

### *Blancocamelus* Dalquest, 1975

TYPE-SPECIES.—*Blancocamelus meadei* Dalquest, 1975:37.

This genus is represented solely by *B. meadei*, described by Dalquest (1975:37) from Mt. Blanco, Texas. Dalquest noted that although Meade (1945:538) was aware of the uniqueness of this camel, he mistakenly applied to it an

unpublished name, *Leptotylopus percelsus*, from a 1924 manuscript of W.D. Matthew. As used by Meade, the name was a nomen nudum. The taxon to which Matthew had applied the name in manuscript was subsequently identified as *Tanupolama* (= *Hemiauchenia*) *blancoensis*. Thus, the genus was left without a valid name until Dalquest (1975) proposed *Blancocamelus meadei* for it.

*Blancocamelus* is known only from postcranial elements. Its limbs are exceedingly long, but quite slender, evoking mental images of a giant *Hemiauchenia*. Indeed, the posterior surface of the proximal phalanx presents an asymmetrical, W-shaped scar for the attachment of the suspensory ligament that is quite like that of *Hemiauchenia* (Breyer, 1976, fig. 2). Although Meade (1945) and Kurtén and Anderson (1980:302) have speculated upon the possible affinities of *Blancocamelus* and the aepycamelines, I prefer for the present to group it with the lamines. With the exception of a possible occurrence in the Blacan Red Light fauna of Texas (Akersten, 1972:29), *Blancocamelus* is restricted to the type-locality (Figure 4).

### *Camelops* Leidy, 1854

**TYPE-SPECIES.**—*Camelops kansanus* Leidy, 1854:172.

*Camelops* is by far the best known of the lamines, giant or otherwise. It occurs from the late Blacan into the early Holocene in localities throughout the western United States (Kurtén and Anderson, 1980, fig. 15.4). Since its description by Leidy (1854:172), the genus *Camelops* has undergone a bewildering series of synonymies, referrals, and revisions. Much of this morass was clarified by Webb (1965), who followed Savage (1951) in recognizing five species: *C. kansanus* Leidy (1854:172), *C. hesternus* (Leidy, 1873:255), *C. huerfanensis* (Cragin, 1892:258), *C. sulcatus* (Cope, 1893:84), and *C. minidokae* Hay (1927:93). These five species plus *C. traviswhitei* Mooser and Dalquest (1975:341) were recognized by Kurtén and Anderson (1980).

*Camelops*, especially the later species, is very hypsodont with large lacrimal vacuities and marked maxillary fossae. The skull is long and does not display the rostral shortening characteristic of other lamines such as *Hemiauchenia*, *Lama*, and *Vicugna*. The mandible is long with a sharp diastemal crest and uninflected angular processes. The dental formula is  $I\frac{1}{2} C_1^1 P_2^2 M_3^3$ . In *Camelops*  $I^3$  and  $C_1^1$  are reduced, laterally compressed, and recurved rather than enlarged and

rounded in cross section as in the giant camelines.  $P_{1,2,3}^2$  are lost and  $P_4^3$  are reduced. The molars are relatively narrow with external styles less strongly developed than in the camelines.

The limbs of *Camelops* are sturdy and the metapodials less slender than those of the other lamines. The area of attachment for the suspensory ligament on the posterior surface of the proximal phalanx is distinctive (Breyer, 1974, fig. 2B). *Camelops hesternus*, *C. traviswhitei*, and *C. huerfanensis* are the only species considered within the scope of giant camels. More detailed descriptions of *Camelops* are given in Savage (1951) and Webb (1965).

### Summary

The trend toward gigantism in camelids is first evident in *Aepycamelus* in the late Clarendonian and continued throughout the rest of the Cenozoic (Figure 17). Eight camelid genera are treated as giant camels in this paper.

*Megacamelus merriami*, new combination, is proposed for the large, late Hemphillian camel from Mt. Eden and Keams Canyon. The giant camels from Mt. Blanco, Lisco, and Keefe Canyon are referred to *Gigantocamelus spatulus*. *Titanotylopus* is applied only to *T. nebraskensis*. *Megatylopus major* is transferred to *Aepycamelus*.

The Camelini were all very large camels, but

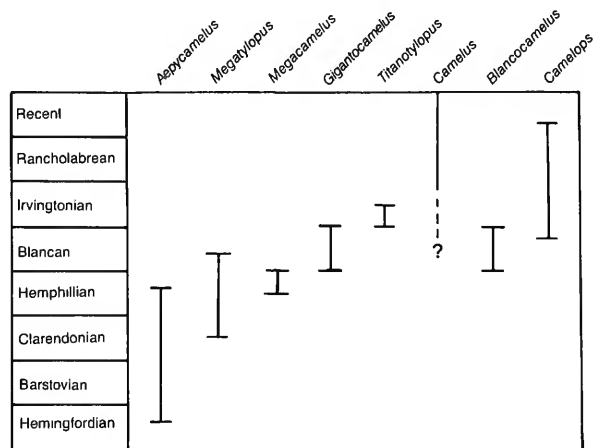


FIGURE 17.—Temporal distribution of the giant camels.

only two giants occur among the Lamini, *Camelops* (*C. hesternus*, *C. traviswhitei*, and *C. huerfaniensis*) and *Blancocamelus* (if, indeed, this genus belongs in the Lamini and not the Aepycamelinae). Most camels were considerably smaller. It

is intriguing that, in spite of over 40 million years of evolution in North America, and regardless of body size, camels became extinct in their place of origin following successful emigration to South America and Asia.

## Literature Cited

- Akersten, W.A.  
1972. Red Light Local Fauna (Blancan) of the Love Formation, Southeastern Hudspeth County, Texas. *Bulletin of the Texas Memorial Museum*, 20:1-53, figures 1-16.
- Barbour, E.H., and C.B. Schultz  
1934. A New Giant Camel, *Titanotylopus nebraskensis*, gen. et. sp. nov. *Bulletin of the Nebraska State Museum*, 1(36):291-294, figures 171, 172.  
1939. A New Giant Camel, *Gigantocamelus fricki*, gen. et sp. nov. *Bulletin of the Nebraska State Museum*, 2(2):17-27, figures 5-12.
- Bennett, D.K.  
1979. The Fossil Fauna from Lost and Found Quarries (Hemphillian: Latest Miocene), Wallace County, Kansas. *Occasional Papers of the University of Kansas Museum of Natural History*, 79:1-24, figures 1-6.
- Breyer, J.A.  
1974. Examination of Selected Postcranial Elements in Pleistocene Camelids. *University of Wyoming Contributions to Geology*, 13(2):75-85, figures 1-11.  
1976. *Titanotylopus* (= *Gigantocamelus*) from the Great Plains Cenozoic. *Journal of Paleontology*, 50(5):783-788, figures 1-3.  
1983. The Biostratigraphic Utility of Camel Metapodials. *Journal of Paleontology*, 57(2):302-307, figures 1-6.
- Cassiliano, M.  
1980. Stratigraphy and Vertebrate Paleontology of the Horse Creek-Trail Creek Area, Laramie County, Wyoming. *University of Wyoming Contributions to Geology*, 19(1):25-68, figures 1-43.
- Cook, H.J.  
1922. A Pliocene Fauna from Yuma County, Colorado with Notes on the Closely Related Snake Creek Beds from Nebraska. *Proceedings of the Colorado Museum of Natural History*, 4(2):1-29.
- Cope, E.D.  
1893. A Preliminary Report on the Vertebrate Paleontology of the Llano Estacado. *Geological Survey of Texas, 4th Annual Report*. 137 pages, 23 plates.  
1894. Marsh on Tertiary Artiodactyla. *American Naturalist*, 28:867-869.
- Cragin, F.W.  
1892. Observations on Llama Remains from Colorado and Kansas. *American Geologist*, 9:257-260.
- Dalquest, W.W.  
1974. The Mexican Camel Names, *Palauchenia mexicana* Del Castillo and *Palauchenia magna* Owen. *Journal of Paleontology*, 48(1):196-198.  
1975. Vertebrate Fossils from the Blanco Local Fauna of Texas. *Occasional Papers of the Texas Tech University Museum*, 30:1-52, figures 1-7.  
1977. Mammals of the Holloman Local Fauna, Pleistocene of Oklahoma. *The Southwestern Naturalist*, 22(2):255-268.  
1980. Camelidae from the Coffee Ranch Local Fauna (Hemphillian Age) of Texas. *Journal of Paleontology*, 54(1):109-117, figures 1-6.
- Dalquest, W.W., and O. Mooser  
1980. Late Hemphillian Mammals of the Ocote Local Fauna, Guanajuato, Mexico. *The Pearce-Sellards Series of the Texas Memorial Museum*, 32:1-25, figures 1-5.
- Davidson, P.  
1923. *Alticamelus alexandrae*, a New Camel from the Barstow Upper Miocene of the Mohave Desert. *University of California Publications, Bulletin of the Department of Geology*, 14:397-408, figures 1-16.
- Dumble, E.T.  
1894. The Cenozoic Deposits of Texas. *Journal of Geology*, 2:549-567.
- Eshelman, R.E.  
1975. Geology and Paleontology of the Early Pleistocene (Late Blancan) White Rock Fauna from North-Central Kansas. *University of Michigan Museum of Paleontology Papers on Paleontology*, 13:1-60, figures 1-6.
- Ferrusquía-Villafranca, I.  
1978. Distribution of Cenozoic Vertebrate Faunas in Middle America and Problems of Migration Between North and South America. In I. Ferrusquía-Villafranca, editor, *Conexiones Terrestres entre Norte y Sudamerica; Simposio Interdisciplinario sobre Paleogeografía Mesoamericana*. Mexico, *Universidad Nacional Autónoma, Instituto de Geología, Boletín*, 101:193-321.
- Forsten, A.  
1970. The Late Miocene Trail Creek Mammalian Fauna. *University of Wyoming Contributions to Geology*, 9(1):39-51, figures 1-6.
- Frick, C.  
1921. Extinct Vertebrate Faunas of the Badlands of Bautista Creek and San Timoteo Cañon, Southern California. *University of California Publications, Bulletin of the Department of Geology*, 12(5):277-



- 424, figures 1–165, plates 43–50.
1929. History of the Earth. Childs Frick Tertiary-Quaternary Explorations. *Natural History*, 29(1):106–108.
- Galusha, T.
1975. Stratigraphy of the Box Butte Formation, Nebraska. *Bulletin of the American Museum of Natural History*, 156(1):1–68, figures 1–16.
- Gauthier-Pilters, H., and A.I. Dagg
1981. *The Camel, Its Evolution, Ecology, Behavior, and Relationship to Man*. 208 pages. Chicago: University of Chicago Press.
- Gidley, J.W.
1903. The Fresh-water Tertiary of Northwestern Texas. American Museum Expeditions of 1899–1901. *Bulletin of the American Museum of Natural History*, 19:617–636, figures 1–4, plates 52–58.
- Gustafson, E.P.
1978. The Vertebrate Faunas of the Pliocene Ringold Formation, South-Central Washington. *Bulletin of the University of Oregon Museum of Natural History*, 23:1–62, figures 1–32.
- Hager, M.W.
1975. Late Pliocene and Pleistocene History of the Donnelly Ranch Vertebrate Site, Southeastern Colorado. *University of Wyoming Contributions to Geology, Special Paper*, 2:1–62, figures 1–33.
- Harrison, J.A.
1979. Revision of the Camelinae (Artiodactyla, Tylopoda) and Description of the New Genus *Alforjas*. *University of Kansas Paleontological Contributions*, 95:1–28, figures 1–3, plates 1–7.
1983. The Carnivora of the Edson Local Fauna (Late Hemphillian), Kansas. *Smithsonian Contributions to Paleobiology*, 54:1–42, figures 1–18.
- Hay, O.P.
1927. The Pleistocene of the Western Region of North America and Its Vertebrated Animals. *Publication of the Carnegie Institute of Washington*, 322B:1–346, figures 1–19, plates 1–11.
- Henshaw, P.C.
1942. A Tertiary Mammalian Fauna from the San Antonio Mountains Near Tonopah, Nevada. *Carnegie Institute of Washington, Contributions to Paleontology*, 530:77–168, figures 1–7, plates 1–11.
- Hesse, C.J.
1936. Lower Pliocene Vertebrate Fossils from the Ogallala Formation (Laverne Zone) of Beaver County, Oklahoma. *Carnegie Institute of Washington, Contributions to Paleontology*, 476:47–72, figures 1–10.
- Hibbard, C.W.
1953. The Saw Rock Canyon Fauna and Its Stratigraphic Significance. *Papers of the Michigan Academy of Science, Arts, and Letters*, 38:387–411, figures 1–5.
- Hibbard, C.W., and W.W. Dalquest
1962. Artiodactyls from the Seymour Formation of Knox County, Texas. *Papers of the Michigan Academy of Science, Arts, and Letters*, 47:83–99, figures 1–4.
1966. Fossils from the Seymour Formation of Knox and Baylor Counties, Texas, and Their Bearing on the Late Kansan Climate of That Region. *Contributions from the University of Michigan Museum of Paleontology*, 21(1):1–66, figures 1–8, plates 1–5.
- Hibbard, C.W., and E.S. Riggs
1949. Upper Pliocene Vertebrates from Keefe Canyon, Meade County, Kansas. *Bulletin of the Geological Society of America*, 60:829–880, figures 1–11, plates 1–5.
- Honey, J.G., and B.E. Taylor
1978. A Generic Revision of the Protolabidini (Mammalia, Camelidae), with a description of Two New Protolabidines. *Bulletin of the American Museum of Natural History*, 161(3):367–426, figures, 1–13.
- Kurtén, B., and E. Anderson
1980. *Pleistocene Mammals of North America*. 442 pages. New York: Columbia University Press.
- Leidy, J.
1854. Description of a Fossil Apparently Indicating an Extinct Species of the Camel Tribe. *Proceedings of the Academy of Natural Sciences of Philadelphia*, 1854:172–173.
1873. Contributions to the Extinct Vertebrate Fauna of the Western Territories. *United States Geological Survey of the Territories, Report*, 1:14–358, plates 1–37.
1886. Mastodon and Llama from Florida. *Proceedings of the Academy of Natural Sciences of Philadelphia*, 1886:11–12.
- Leidy, J., and F.A. Lucas
1896. Fossil Vertebrates from the Alachua Clays of Florida. *Transactions of the Wagner Free Institute of Science*, 4:1–61, plates 1–19.
- Lindsay, E.H.
1978. Late Cenozoic Vertebrate Faunas, Southeastern Arizona. *New Mexico Geological Society Guidebook for the 29th Field Conference*, 1978:269–275, figure 1.
- Linnaeus, C.
1758. *Anamalia*. In *Systema naturae . . . editio decima, reformata*, volume 1. London: British Museum (Natural History).
- Macdonald, J.R.
1949. A New Clarendonian Fauna from Northeastern Nevada. *University of California Publications, Bulletin of the Department of Geology*, 28(7):173–193, figures 1–11.
1956. A New Clarendonian Mammalian Fauna from the Truckee Formation of Western Nevada. *Journal of Paleontology*, 30(1):186–202, figures 1–13.

1959. The Middle Pliocene Mammalian Fauna from Smiths Valleys, Nevada. *Journal of Paleontology*, 33(5):872-887, figures 1-5.
1966. The Barstovian Camp Creek Fauna from Elko County, Nevada. *Los Angeles County Museum of Natural History Contributions in Science*, 92:1-18, figures 1-7.
- MacFadden, B.J.  
1977. Magnetic Polarity Stratigraphy of the Chamita Formation Stratotype (Mio-Pliocene) of North-Central New Mexico. *American Journal of Science*, 277:769-800, figures 1-10.
- MacFadden, B.J., N.M. Johnson, and N.D. Opdyke  
1979. Magnetic Polarity Stratigraphy of the Mio-Pliocene Mammal-Bearing Big Sandy Formation of Western Arizona. *Earth and Planetary Science Letters*, 44:349-364, figures 1-7.
- Marsh, O.C.  
1894. Description of Tertiary Artiodactyls. *American Journal of Science*, series 3, 48:259-274, figures 1-34.
- Martin, R.A., and J.C. Harksen  
1974. The Delmont Local Fauna, Blancan of South Dakota. *Bulletin of the New Jersey Academy of Science*, 19(1):11-17, figures 1-13.
- Matthew, W.D.  
1899. A Provisional Classification of the Fresh-water Tertiary of the West. *Bulletin of the American Museum of Natural History*, 12(3):19-75.  
1901. Fossil Mammals of the Tertiary of Northeastern Colorado. *Memoirs of the American Museum of Natural History*, 1(7):355-448, figures 1-34, plates 37-39.  
1909. Faunal Lists of the Tertiary Mammalia of the West. *Bulletin of the United States Geological Survey*, 361:91-138.  
1924. Third Contribution to the Snake Creek Fauna. *Bulletin of the American Museum of Natural History*, 50:59-210, figures 1-63.
- Matthew, W.D., and H.J. Cook  
1909. A Pliocene Fauna from Western Nebraska. *Bulletin of the American Museum of Natural History*, 26(27):361-414, figures 1-27.
- Meade, G.E.  
1945. The Blanco Fauna. *University of Texas Publication*, 4401:509-556, figures 1-4, plates 48-55.
- Merriam, J.C.  
1917. Relationships of Pliocene Mammalian Faunas from the Pacific Coast and Great Basin Provinces of North America. *University of California Publications, Bulletin of the Department of Geology*, 10(22):421-443, figure 1.
- Miller, W.E., and T. Downs  
1974. A Hemphillian Local Fauna Containing a New Genus of Antilocaprid from Southern California. *Los Angeles County Museum of Natural History Contributions in Science*, 258:1-35, figures 1-13.
- Mooser, O., and W.W. Dalquest  
1975. A New Species of Camel (Genus *Camelops*) from the Pleistocene of Aguascalientes, Mexico. *The Southwestern Naturalist*, 19(4):341-345, figures 1-2.
- Patton, T.H.  
1969. Miocene and Pliocene Artiodactyls, Texas Gulf Coastal Plain. *Bulletin of the Florida State Museum*, 14(2):115-226, figures 1-34.
- Savage, D.E.  
1941. Two New Middle Pliocene Carnivores from Oklahoma, with Notes on the Optima Fauna. *American Midland Naturalist*, 25(3):692-710, plates 1-4.  
1951. Late Cenozoic Vertebrates of the San Francisco Bay Region. *University of California Publications, Bulletin of the Department of Geological Sciences*, 28(10):215-314, figures 1-51.
- Schlosser, M.  
1903. Die Fossilen Säugethiere Chinas nebst einer Odontographie der recenten Antilopen. *Abhandlungen der Bayerischen Akademie der Wissenschaften*, 22(1):1-22.
- Schultz, G.E.  
1977. Guidebook for Field Conference on Late Cenozoic Biostratigraphy of the Texas Panhandle and Adjacent Oklahoma. *Special Publication of the Kilgore Research Center, West Texas State University*, 1:1-160, figures 1-35.
- Semken, H.A., Jr.  
1966. Stratigraphy and Paleontology of the McPherson *Equus* Beds (Sandahl Local Fauna), McPherson County, Kansas. *Contributions from the University of Michigan Museum of Paleontology*, 20(6):121-178, figures 1-7.
- Shotwell, J.A.  
1970. Pliocene Mammals of Southeast Oregon and Adjacent Idaho. *Bulletin of the University of Oregon Museum of Natural History*, 17:1-103, figures 1-43.
- Simpson, G.G.  
1930. Tertiary Land Mammals of Florida. *Bulletin of the American Museum of Natural History*, 59(3):149-211, figures 1-31.
- Skinner, M.F., and C.W. Hibbard (with the collaboration of E.D. Gutentag, G.R. Smith, J.G. Lundberg, J. Alan Holman, J. Alan Feduccia, and Pat Vickers Rich)  
1972. Early Pleistocene Preglacial and Glacial Rocks and Faunas of North-Central Nebraska. *Bulletin of the American Museum of Natural History*, 148(1):1-148, figures 1-60.
- Skinner, M.F., S.M. Skinner, and R.J. Gooris  
1968. Cenozoic Rocks and Faunas of Turtle Butte,

- South-Central South Dakota. *Bulletin of the American Museum of Natural History*, 138(7):331-436, figures 1-15, plates 20-25.
- Strain, W.S.  
1966. Blancan Mammalian Fauna and Pleistocene Formations, Hudspeth County, Texas. *Bulletin of the Texas Memorial Museum*, 10:1-55, figures 1-8, plates 1-13.
- Walker, E.P.  
1964. *Mammals of the World*. Volume 2. Baltimore: Johns Hopkins University Press.
- Webb, S.D.  
1965. The Osteology of *Camelops*. *Bulletin of the Los Angeles County Museum of Natural History*, 1:1-54, figures 1-22.
1969. The Burge and Minnechaduza Clarendonian Mammalian Faunas of North-Central Nebraska. *University of California Publications in Geological Sciences*, 78:1-191, figures 1-46.
- Wortman, J.L.  
1898. The Extinct Camelidae of North America and Some Associated Forms. *Bulletin of the American Museum of Natural History*, 10:93-142, figures 1-23, plate 11.







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