Seed predation and defleshing in the agouti-dispersed palm *Astrocaryum standleyanum*

Patrick A. Jansen*,†,‡,1, Kelly Elschot*, P. Johannes Verkerk‡,§ and S. Joseph Wright†

* Community and Conservation Ecology, University of Groningen, PO Box 14, NL-9750 AA Haren, the Netherlands
† Smithsonian Tropical Research Institute, Apartado 0843-03092, Balboa, Ancón, Panamá, República de Panamá
‡ Center for Ecosystem Studies, Wageningen University, PO Box 47, 6700 AA Wageningen, the Netherlands
§ Laboratory of Nematology, Wageningen University, PO Box 8123, 6700 ES Wageningen, the Netherlands
(*Accepted 5 June 2010*)

Abstract: The agouti (*Dasyprocta punctata*) meticulously defleshes *Astrocaryum standleyanum* palm seeds before scatter hoarding. On Barro Colorado Island, Panama, we experimentally tested three hypotheses on how this behaviour could reduce seed predation to the mutual benefit of the tree and the rodent. The first and established hypothesis – that defleshing reduces seed predation by bruchid beetles by intercepting larvae – was rejected. Experiments in which manually defleshed seeds or entire fruits were incubated at different times showed that defleshing reduced bruchid infestation before fruit fall but not after fruit fall. The second hypothesis – that defleshing reduces cache pilferage by making seeds less conspicuous – was supported. An experiment in which intact fruits and manually defleshed seeds were placed in mimicked agouti caches and followed showed that seeds with flesh were pilfered at higher rates than defleshed seeds. The third hypothesis – that defleshing reduces post-dispersal infestation of cached seeds – was rejected. An experiment in which intact fruits and manually defleshed seeds were placed in mammal exclosures and later collected to assess infestation showed that burial reduced seed infestation but defleshing did not. Thus, seed defleshing reduced palm seed predation, but in a different way than previously believed. We also found that (1) bruchid beetles can be pre-dispersal rather than post-dispersal seed predators, (2) seed infestation by scolytid beetles may control bruchid larvae, and (3) scolytids rather than bruchids are the main invertebrate seed predators of this palm.

Key Words: *Astrocaryum standleyanum*, Barro Colorado Island, Bruchidae, cache pilferage, *Dasyprocta punctata*, scatter-hoarding, Panama, Scolytidae, seed handling, seed predation

INTRODUCTION

Post-dispersal seed predation is a major source of offspring mortality in trees (Hulme 1998, Janzen 1971a). Many large-seeded tree species escape seed predation through seed dispersal by scatter-hoarding animals (Jansen & Forget 2001). These animals hoard seeds in shallow single-seeded caches in the soil surface, scattered across the home range, as food supplies for periods of food scarcity (Vander Wall 1990). When the scatter hoarder fails to recover some of the cached seeds, scatter-hoarding reduces seed predation in several ways. The dispersal of seeds away from the parent tree potentially reduces post-dispersal infestation by parent-associated predators

and pathogens, the scattering of seeds across a wide area reduces the likelihood of density-responsive seed predators finding and killing the seeds, and the burial of seeds enhances seed survival and seedling establishment (Jansen *et al.* 2008a, Jansen & Forget 2001, Vander Wall 1990). A potential advantage to the plant that has received less attention is a reduction of seed predation rates as a result of the rodents' tendency to carefully husk and deflesh seeds before caching them (Jansen *et al.* 2006).

A prime example of seed-defleshing behaviour is seen in the Central American agouti *Dasyprocta punctata* Grey. This large Neotropical forest rodent meticulously peels off and discards or eats all exocarp and mesocarp material before scatter-hoarding seeds such as those of the fleshy-fruited palm *Astrocaryum standleyanum* L. H. Bailey (Arecaceae) (Smythe 1989, pers. obs.). Smythe (1989) argued that agoutis deflesh *A. standleyanum* before

---

caching to remove the eggs and larvae of bruchid beetles that are contained in the pulp before those can penetrate the endocarp and attack the seed. Thus, agoutis would reduce seed predation to the mutual benefit of the animal and its food tree. However, recent studies suggest that agoutis prefer to consume bruchid larvae rather than seeds, possibly because larvae have a higher nutritional value than endosperm (Gálvez & Jansen 2007, Silvius 2002). Thus, it might not be in the interest of agoutis to intercept bruchid larvae.

We performed a series of manipulative experiments to evaluate three alternative, but comparable hypotheses on how seed-defleshing behaviour may reduce seed predation to the mutual benefit of hoarder and tree. The first and established hypothesis – the 'interception hypothesis' – was that defleshing reduces post-dispersal seed predation by bruchid beetles by removing bruchid beetle larvae (or eggs) present in the material around seeds before these larvae can enter the seed (Smythe 1978). The second hypothesis – the 'theft-prevention hypothesis' – was that defleshing reduces post-dispersal seed predation by lowering the likelihood that seed-eating mammals recover cached seeds. Odour enhances cache detection by rodents and other macrosmatic mammals (Murie 1977, Stapanian & Smith 1978, Vander Wall 1991, Vander Wall *et al.* 2003), and decaying fruit flesh and other soft material has a stronger odour than the seeds themselves. Defleshing might reduce cues that other animals use to find caches, and increase the recovery advantage for the cache owner. Although a proportion of seed pilfered from caches by rodents may be re-cached, every instance of seed handling comes with a risk of seed consumption (Vander Wall 1995). The third hypothesis – the 'conservation hypothesis' – was that defleshing reduces seed predation by reducing seed infestation by microbes and invertebrates that first arrive after seeds are cached. Fruit flesh is an excellent medium for microbes (Janzen 1977). By removing exocarp and mesocarp before caching seeds, rodents might reduce microbial infestation and increase seed storage life (Vander Wall 1990). In each of these three hypotheses, the scatter hoarder and the plant share a common advantage – seed defleshing reduces post-dispersal seed predation – but the mechanisms differ. Note that we do not consider explanations unrelated to seed predation, such as defleshing facilitating the transport and caching of seeds by reducing seed weight and volume, or the material around the seed being removed to be consumed as food. However, these do not explain why seed defleshing is done meticulously and exhaustively.

STUDY SITE AND SPECIES

Our study sites were Barro Colorado Island (BCI) and the nearby Pipeline Road (PLR) in the Parque Nacional Soberania, Panama (9°10′N, 79°51′W). BCI is a former hilltop that was isolated from the mainland by the formation of Gatun Lake to complete the Panama Canal in 1914. BCI and PLR support semi-deciduous tropical moist forest. Mean annual rainfall is 2600 mm, and there is a distinct 4-mo dry season (Leigh 1998). BCI and PLR have a fairly complete fauna with mammal densities similar to much more remote sites (Wright *et al.* 1994). One important component absent today is the white-lipped peccary (*Tayassu pecari*), which has relevance to our study because these animals are known to feed on cached palm seeds (Kiltie 1981).

The Central American agouti is a terrestrial, diurnal caviomorph rodent weighing 2–4 kg that is common in tropical forests from Central America and the West Indies to northern South America. Agoutis are of great ecological importance as seed predators and seed dispersers. They scatter-hoard seeds of several large-seeded plant species during periods of high fruit abundance, and retrieve them during periods of scarcity. They carry seeds in their mouths, one at a time, and bury them individually in widely scattered caches. Because cached seeds may remain unrecovered, agoutis function as seed dispersers of their food plants (Smythe 1989, Smythe *et al.* 1982). Before caching, agoutis routinely spend time defleshing seeds (Smythe 1989), even if the seeds are already free of fruit flesh (P. A. Jansen *et al.*, unpubl. data).

Astrocaryum standleyanum is a monoecious, arbores-cent cocosoid palm, occurs from Nicaragua to Colombia (Croat 1978), and is among the ten most abundant tree species in central Panama (Pyke *et al.* 2001). On BCI, adults produce up to eight pendulous infructescences, each up to 150 cm long with up to 800 fruits that ripen mainly during April–June (De Steven *et al.* 1987, Jansen *et al.* 2008b). Fruits are ovoid drupes that consist of one 20 × 15-mm seed (rarely two or three), enclosed in three layers: a hard endocarp (shell) that is 1.5–3 mm thick; a sweet, fleshy mesocarp (flesh) that is 4–5 mm thick; and finally a harder, bright orange exocarp (skin) that is 0.5 mm thick. In the following, we loosely use 'seed' to denote either the actual seed or the stone: the seed plus endocarp. Flesh and seeds are consumed by a variety of frugivorous and granivorous mammals and are one of the most important food sources for the agouti on BCI (Glanz *et al.* 1982, Hoch & Adler 1997, Kays 1999, Smythe *et al.* 1982). In turn, *A. standleyanum* is believed to depend strongly on scatter hoarding by agoutis that may cache seeds of *A. standleyanum* up to >100 m away from the mother tree (P. A. Jansen *et al.*, unpubl. data), which significantly increases the chances of survival and germination (Smythe 1989).

The invertebrate seed predators of *A. standleyanum* include a bruchid beetle (Coleoptera: Bruchidae) and a scolytid beetle (Coleoptera: Scolytidae). The bruchid beetle – identified as *Pachymerus bactris* (L.) in this study

(M. N. Garcia, pers. comm.) – caused high proportions of *A. standleyanum* seed predation (up to 75%) in previous studies (Smythe 1989, Wright *et al.* 2000). *Pachymerus bactris* adults are ~12 mm long, and are believed – with all other pachymerine bruchids – to oviposit on fruits after they fall to the ground (Johnson & Romero 2004, Smythe 1989). The larvae bore through the endocarp into the seed on which they feed. The beetles pupate and hatch inside the endocarp, spend the dry season inside as adults, and emerge at the onset of the rainy season (P. A. Jansen, pers. obs.). Emerging adults leave unmistakable 5-mm diameter emergence holes (Smythe 1989).

The scolytid beetle, *Coccotrypes* sp., was identified as a major post-dispersal seed predator in the current study. Species of *Coccotrypes* are post-dispersal seed predators that specialize on palms and can infest nearly 100% of a seed crop (Hammond *et al.* 1999, Janzen 1972, Notman & Villegas 2005, Silva & Tabarelli 2001). Adult females (*c.* 1 mm long) bore their way through the endocarp into the seed in which they lay multiple eggs and form a new colony that feeds on the seed. In other plant species, seeds with low levels of scolytid infestation still germinated, but higher levels killed seeds (Curran & Webb 2000, Russo 2005). The entrance holes, that can amount to dozens per seed, can be easily counted as an estimate of infestation.

METHODS

Interception hypothesis

We performed three experiments to determine the timing of oviposition by bruchids and evaluate the interception hypothesis, i.e. that seed defleshing reduces post-dispersal seed predation by bruchids. Experiment 1 evaluated the interception hypothesis for ripe seeds that were still on the tree. In April 2006, we collected 440 ripe *A. standleyanum* fruits from the crowns of eight individual palms at PLR with a 10-m pole saw. Ripe fruits are distinguished by soft exocarp and mesocarp and hard, filled endosperm. We randomized these fruits, left half intact, and removed the exocarp and mesocarp from the remainder with a pocket knife. This treatment mimicked seed defleshing by agoutis; the difference from seeds defleshed by agoutis was indistinguishable to the human eye. We incubated all seeds for 4 mo at ambient temperature and humidity in screen-covered boxes inside a screened shade house to exclude adult bruchids. We then inspected the seeds for exit holes of bruchids, opened the endocarps with a hammer and recorded the presence/absence of bruchid larvae.

Experiment 2 evaluated the interception hypothesis for freshly fallen seeds. In June 2005, we collected 466 fruits fallen overnight by returning on consecutive mornings to 5-m^2 pieces of shade cloth suspended below mature infructescences of five individual *A. standleyanum* along

PLR. We immediately placed these fruits in 1.3-cm wire mesh exclosures at the base of the palms to expose seeds to *P. bactris* but exclude rodents. The exclosures were 50–80 cm diameter wide and 25–30 cm tall, were staked into the ground with 3–5 40-cm-long iron bars, and had a closed top of the same wire mesh. After 0, 1, 2, 4 and 8 d of exposure, we transported 20% of the fruits from each exclosure to the laboratory, where we manually removed the exocarp and mesocarp to simulate seed-defleshing by agoutis (as above) and incubated the defleshed seeds for 3 mo, after which we recorded bruchid infestation as in Experiment 1.

Experiment 3 evaluated whether and to what extent *P. bactris* oviposits after fruit fall. In June 2004, we collected 850 freshly fallen *A. standleyanum* fruits from the ground below 20 individual palms on BCI and mixed them. We incubated 50 randomly chosen fruits in an air-conditioned laboratory (20–25 °C) in five styrofoam containers provided with cotton-stoppered tubes of water. We simultaneously placed 50 randomly chosen fruits in each of 16 mammal exclosures (as above) in the forest. The mammal exclosures were ~50 cm in diameter and were separated by >100 m. We removed 10 randomly chosen fruits from each exclosure 5, 10, 20, 40 and 80 d later and incubated them for 12 mo, after which we recorded bruchid infestation as in Experiment 1.

Theft-prevention hypothesis

Experiment 4 evaluated the theft-prevention hypothesis, i.e. whether seed defleshing reduced the pilferage of cached seeds. In June 2005, we collected 50 ripe *A. standleyanum* fruits from below fruiting individuals on BCI, and removed the exocarp and mesocarp from 25 of them, mimicking seed defleshing by agoutis (as above). Then, we experimentally cached 25 seeds with fruit flesh and 25 without, at 2.5 cm depth in similar-diameter holes, mimicking agouti-made caches. Caches were placed along 2.5 km of trails on BCI at 50-m intervals, spaced out far enough for cache fates to be considered independent (i.e. no animal can use the presence of a cached seed as cue for finding a seed cached ≥50 m further away). Caches were 0.5 m away from a sapling in a fixed direction, and marked with numbered flagging tape attached to the sapling at eye level. We monitored their disappearance, which was evident from empty holes, 1, 2, 4, 8, 16 and 32 d after caching.

Conservation hypothesis

Experiment 5 evaluated the conservation hypothesis, i.e. whether defleshing reduces seed loss to microbes and invertebrates attacking seeds after dispersal. To disentangle any effects of defleshing from those of burial, our experiment mimicked both in a factorial design. In

May 2006, we selected eight reproductive *A. standleyanum* in each of 12 forest plots (six on BCI and six along PLR), placed one mammal exclosure (as in experiment 2) at each palm (96 in total), and placed 25 *A. standleyanum* seeds in each exclosure (2400 in total, from a single randomized batch, collected from a small number of abundantly fruiting *A. standleyanum* at PLR). The 25 seeds in each exclosure were either defleshed or not defleshed, as in experiment 1, and buried or not buried. The buried defleshed seeds mimicked agouti caches not pilfered by mammals, except for the fact that they were clumped rather than scattered. After 4 mo, all seeds were collected and externally and internally analysed for infestation by microbes and insects, as in experiment 1.

Scolytid versus bruchid beetles

Results from the above experiments showed that the percentage of seeds that were infested by bruchids was much higher than the percentage of seeds that had bruchid larvae or exit holes under field conditions, possibly due to post-dispersal seed infestation by scolytid beetles. Therefore, we conducted a sixth experiment to evaluate whether bruchid presence in endocarps in the field is negatively related to scolytid infestation. We collected 10–40 freshly fallen ripe fruits with seed traps (as in experiment 2) at five palms in each of 12 forest plots (as in experiment 5) and placed them in mammal exclosures (as in experiment 2) located near the base of each palm (60 exclosures in total) in June 2005. After 11 mo, we collected 1331 seeds from 55 exclosures (five exclosures had disappeared) and recorded the number of scolytid entry holes (1 mm diameter) per seed – each hole represents an independent colonization of the seed by a female – as well as the presence of a bruchid larva or bruchid emergence hole (5–6 mm diameter).

Analysis

All analyses were conducted in R 2.8.0 (R Foundation for Statistical Computing, Vienna, Austria) and its contributed libraries MASS and survival. We used Generalized Linear Modelling with binomial errors to test for differences in the proportion of seeds infested (experiments 1, 3 and 5). We used Generalized Linear Mixed Modelling with binomial errors and random factors for hierarchical levels to test for differences in the proportion of seeds infested in experiments where seeds were grouped within trees and plots (experiments 2 and 6). We used Kaplan–Meier survival analysis and the log-rank test to test for a difference in cache survival time (experiment 4).

RESULTS

Interception hypothesis

In experiment 1, which evaluated whether seed defleshing affected bruchid infestation of seeds still ripening on trees, bruchid presence was significantly lower in defleshed seeds (16%) than in non-defleshed seeds (24%; GLM with binomial errors: Wald = 4.61, df = 2, P = 0.032). In experiment 2, which evaluated the interception hypothesis for freshly fallen seeds, the proportion of seeds infested by bruchids was independent of how long after fruit fall seeds were defleshed (Figure 1a; GLMM with palms as random factor: $\chi^2_1 = 0.0259$, P = 0.87). Bruchids had infested up to 75% of the seeds defleshed on the same day as fruit fall. In experiment 3, which evaluated bruchid oviposition after fruit fall, we found that the proportion of seeds infested by bruchids (mean = 0.36, SD = 0.18, n = 85 samples) did not increase with the duration of exposure in the forest (GLM with binomial errors, duration log-transformed: $t_{63} = 1.02$, P = 0.31).

Theft-prevention hypothesis

In experiment 4, which evaluated whether seed defleshing reduced the pilferage of cached seeds, we found that caches with defleshed seeds had significantly longer life times (half-life > 32 d) than did caches with non-defleshed seeds (half-life < 4 d; Figure 1b; Log-rank test on Kaplan–Meier survival estimates: $\chi^2 = 17.9$, df = 1, P < 0.001).

Conservation hypothesis

In experiment 5, which evaluated whether defleshing reduces seed loss to microbes and invertebrates attacking seeds after dispersal, we found that only 12 (0.6%) of the 2037 seeds that were recovered (some exclosures had been raided by mammals) contained bruchid larvae, while only 16 (0.8%) had fungus. In contrast, 381 (19%) were infested by *Coccotrypes* bark beetles. The fraction infested was significantly lower for buried seeds (16%) than for unburied seeds (21%; Figure 1c; GLM with binomial errors: $F_{1,94} = 6.66$, P < 0.001), but there was no significant difference between defleshed and untreated seeds ($F_{1,93} = 0.11$, P = 0.74), nor was there evidence for interaction between burial and defleshing ($F_{1,92} = 0.09$, P = 0.77). Thus, burial enhanced seed conservation but defleshing did not.

Scolytids versus bruchids

The above experiments showed a discrepancy between the percentage of seeds that were infested by bruchids (65% and 25% in experiments 2 and 3) and the percentage of
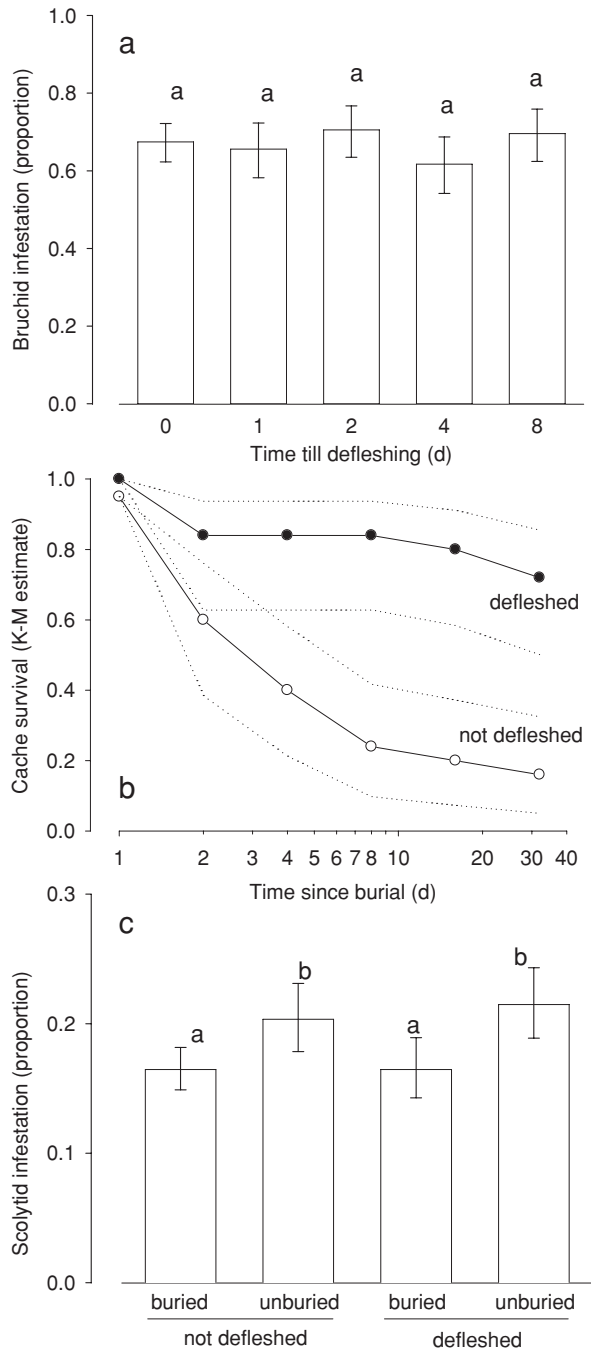
Figure 2. Relationship between bruchid beetle infestation and scolytid beetle infestation of *Astrocaryum standleyanum* seeds after 11 mo on the forest surface. Significantly fewer bruchids developed as seeds showed more scolytid beetle entrance holes. Symbol size scales with the number or observations over which the proportion was calculated. Line shown is the fitted weighted logistic regression model.

Figure 1. Effect of defleshing on seed predation by bruchid beetles, pilferage of cached seeds and seed predation by scolytid beetles in the palm *Astrocaryum standleyanum*. Infestation by the bruchid beetle *Pachymerus bactris* for seeds that were defleshed at different times after fruit fall (a). Defleshing did not affect infestation, regardless of the timing. The beetle larvae had already entered the endocarp at the time of fruit fall; Kaplan–Meier survival estimates for experimental caches containing defleshed or non-defleshed seeds (b). Defleshing significantly reduced cache pilferage rates; Infestation by scolytid beetles for seeds that were subject to experimental burial and defleshing (c). Infestation was reduced by burial but not by removal of fruit pulp. Different letters within panels (a) and (c) denote significant differences. Dotted lines in (b) are the 95% CI of the Kaplan–Meier estimates.
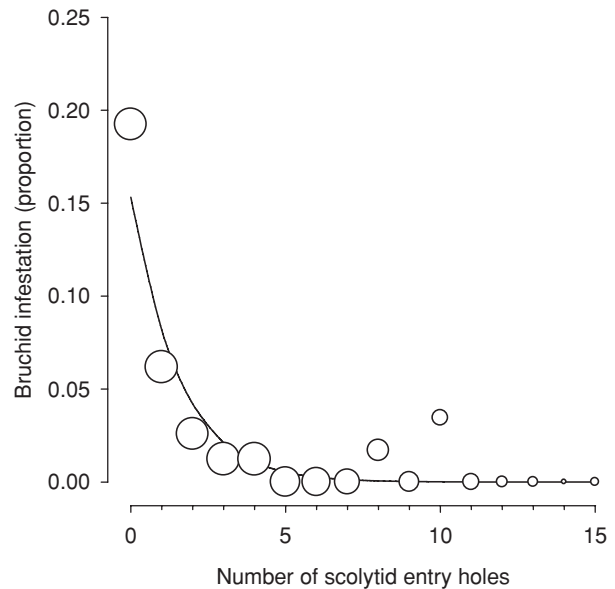
seeds that had bruchid larvae or exit holes under field conditions (0.5% in experiment 5). Do scolytids reduce bruchid survival? In an additional experiment to evaluate the relationship between bruchid and scolytid infestation of seeds in the field, the proportion of seeds with bruchid larvae or emergence holes was indeed significantly lower for seeds with scolytid entrance holes (0.017) than for seeds without (0.192; GLMM: $t_{1274} = -6.34$, P < 0.001). The proportion of seeds with bruchids also decreased significantly with the number of scolytid entrance holes (Figure 2; $t_{1274} = -2.46$, P = 0.014).

DISCUSSION

The interaction between *A. standleyanum* and the Central-American agouti has become a classic example of a plant–rodent interaction in which animal and plant both benefit (Leigh 1998, Leigh & Rubinoff 2005, Moore 2001, Vander Wall 1990), particularly because the defleshing of palm seeds before caching was assumed to intercept an important insect seed predator, to the advantage of the agouti as well as the palm (Smythe 1989). Our study, however, shows that agoutis do not in fact intercept bruchid larvae. We found that the bruchid beetle infesting *A. standleyanum* seeds – now identified as *Pachymerus bactris* – attacks seeds in the canopy, before they leave the mother tree, and that the larvae have already entered the seed when the fruits fall to the ground. *Pachymerus*

bactris does not infest seeds after seed fall. Monkeys and squirrels feeding on unripe fruit might still intercept beetle larvae, but fruit defleshing by agoutis after fruit fall no longer affects infestation. Also, prolonged exposure of palm seeds on the ground below the palm does not increase the fraction infested by bruchids. Thus, the original interception hypothesis is rejected.

The alternative hypothesis that seed defleshing by agoutis reduces predation of cached seeds by other insects and microbes (conservation hypothesis) also was not supported. Post-dispersal seed predation by invertebrates (i.e. scolytid beetles) was important. However, the infestation of seeds by scolytids was reduced by burial, as in Silva & Tabarelli (2001), but not by defleshing. Although we cannot rule out the possibility that agoutis reduce cues for seed finding by scolytid beetles further than we did, for example because they scatter and deflesh seeds better, we consider it unlikely that a strong effect was masked by any such bias. Fungal infestation was unimportant during the 4 mo of this experiment. This seems to be due to the fact that *A. standleyanum* seeds germinate slowly and have very thick endocarps and thus seem well-protected against fungi, at least until the germination pores open. In general, rodents create their long-term food reserves with well-protected, slow-germinating seeds with a long storage life (Jansen *et al.* 2006), for which fungal infestation may be less important.

We did find clear support for the hypothesis that defleshing by agoutis reduces cache robbery by mammalian food competitors (theft-prevention hypothesis). Experimentally cached seeds were pilfered at much lower rates when defleshed before caching. Our study site lacks white-lipped peccaries, which are known to forage for buried palm seeds (Kiltie 1981), but there is no reason to believe that the observed difference in cache survival was due to their absence. Previous studies suggest that olfaction plays an important role in cache pilferage (Stapanian & Smith 1978, Vander Wall *et al.* 2003). Thus, we believe that seed defleshing reduces the olfactory cue for food competitors finding and stealing the buried food.

Bruchid versus scolytid beetles

An additional finding from our study is that bruchid beetles were neither an important enemy for *A. standleyanum* nor an important food competitor for agoutis. Infestation percentages for seeds incubated in the laboratory were important, but bruchids appeared unimportant under field conditions. This was due to low survival of bruchid larvae under field conditions, not to the defleshing of the seeds by agoutis. We found that the major invertebrate seed predators of *A. standleyanum* are *Coccotrypes* scolytid beetles, not bruchid beetles. On BCI,

up to 78% of the *A. standleyanum* seeds in the soil show signs of infestation by scolytid beetles (H. Sharrott & P. A. Jansen, unpubl. data). We found that bruchid presence in seeds was negatively correlated with the number of scolytid entrance holes, suggesting that scolytid beetles are responsible for the low survival of bruchid beetles; they either eat or outcompete bruchid larvae in palm seeds.

Bruchids also seem relatively unimportant to *A. standleyanum* and agoutis because seeds rarely contain more than one bruchid larva, which consumes only a limited portion of the endosperm, after which the seed can still germinate and establish (C. X. Garzon-Lopez & P. A. Jansen, unpubl. data). This contrasts with the sympatric palm *Attalea butyracea*, where a single larva will typically consume the entire seed (Gálvez & Jansen 2007). Additionally, recent studies in *A. butyracea* suggest that agoutis may consider bruchid larvae better food than palm endosperm (Gálvez & Jansen 2007, Silvius 2002). The fact that bruchid larvae spend a long time inside the endocarps – live larvae were present in endocarps after 4 and even 11 mo – implies that bruchid larvae are available to agoutis during August–March, when fruit abundance is relatively low and the rodents feed on cached *A. standleyanum* seeds. Thus, the selective pressure to prevent bruchid infestation might be negative for agoutis (Silvius 2002), and limited for *A. standleyanum*.

Smythe (1989) based his conclusion that agoutis intercept bruchid larvae on a comparison of results from three experiments. In one experiment 100 fruits were collected the second morning after they had fallen and stored on trays. Seven days later, exocarp and mesocarp were removed and the defleshed seeds were incubated in jars. After 4 mo, 34 bruchid beetles had emerged. In contrast, in two other experiments, no bruchid exit holes were found in 125 seeds that had been defleshed immediately and left buried in germination bags in the field for several months. Smythe concluded that the larvae entered the endocarps during the 8 d following seed fall unless timely defleshing removed them. Our results suggest that differences in bruchid emergence in Smythe (1989) were due not to the defleshing treatment but rather to the different environments in which the seeds were placed; with and without protection of bruchid larvae from scolytid beetles.

Bruchids as pre-dispersal seed predators

Johnson & Romero (2004) distinguish three oviposition guilds in bruchid beetles: guild A includes species that lay eggs on intact fruits when still on the plant, guild B includes species that lay eggs directly on mature seeds when still on the plant, and guild C includes species that lay eggs on fruits or seeds that have fallen to the ground. All palm bruchids (subfamily Pachymerinae)

were believed to be in guild C, infesting endocarps only after fruit have fallen to the ground (Johnson & Romero 2004). Removal of the exocarp and mesocarp facilitates subsequent oviposition and enhances seed predation by guild C species, for example in *Speciomerus giganteus* attacking *Attalea butyracea* in Costa Rica and Panama (Janzen 1971b, Wright 1983), in *P. cardo* attacking *A. maripa* in Brazil (Silvius & Fragoso 2002), and in *Caryoborus serripes* attacking *Astrocaryum chambira* in Peru (Delgado *et al.* 1997). Exocarps protect seeds against guild C species, and fruit handling by mammals gives bruchid beetles access to the endocarps (Silvius 2005).

Our study is the first to identify a palm bruchid belonging to guild A, infesting endocarps while still on the parent plant, hence acting as a pre-dispersal seed predator (cf. Johnson & Romero 2004). This implies that *P. bactris* does not require fruit handling by mammals to oviposit successfully. Forget (1991) reported that an unidentified *Pachymerus* infested *A. paramaca* while still in French Guiana, and unidentified bruchids from guild A have also been observed in *Attalea maripa* in Brazil (J. Fragoso, pers. comm.). Pre-dispersal seed infestation by bruchids is more important than is currently appreciated.

Implications

Seed defleshing has become standard preparation for propagating *A. standleyanum* in nurseries (Potvin *et al.* 2003) because it was believed to prevent seed infestation by bruchids. Our study suggests that this tedious process will in fact not reduce seed predation by microbes or invertebrates. The only demonstrated effect of defleshing – reduced loss of buried seeds to mammals – is irrelevant in protected nurseries. Whether seed defleshing enhances survival and germination ratios in other ways remains to be shown.

Mammal defaunation could interact with oviposition behaviour of beetles (guild) to have important implications for seed fate. The decline of frugivorous mammals in forest systems reduces access to endocarps for guild C bruchids, which cannot penetrate the exocarp, but not guild A bruchids. Defaunation may therefore differentially affect palm species depending on the oviposition guilds of the associated bruchids (Wright & Duber 2001, Wright *et al.* 2000).

ACKNOWLEDGEMENTS

LITERATURE CITED

CROAT, T. B. 1978. *Flora of Barro Colorado Island*. Stanford University Press, Stanford. 943 pp.

CURRAN, L. M. & WEBB, C. O. 2000. Experimental tests of the spatiotemporal scale of seed predation in mast-fruiting Dipterocarpaceae. *Ecological Monographs* 70:129–148.

DE STEVEN, D., WINDSOR, D. M., PUTZ, F. E. & DE LEON, B. 1987. Vegetative and reproductive phenologies of a palm assemblage in Panama. *Biotropica* 19:342–356.

DELGADO, C., COUTURIER, G. & DELOBEL, A. 1997. Oviposition of seed-beetle *Caryoborus serripes* (Sturm) (Coleoptera: Bruchidae) on palm (*Astrocaryum chambira*) fruits under natural conditions in Peru. *Annales de la Société Entomologique de France* 33:405–409.

FORGET, P. M. 1991. Scatterhoarding of *Astrocaryum paramaca* by *Proechimys* spp. in French Guiana: comparison with *Myoprocta exilis*. *Tropical Ecology* 32:155–167.

GÁLVEZ, D. & JANSEN, P. A. 2007. Bruchid beetle infestation and the value of *Attalea butyracea* endocarps for Neotropical rodents. *Journal of Tropical Ecology* 23:381–384.

GLANZ, W. E., THORINGTON, R. W., GIACALONE-MADDEN, J. & HEANEY, L. R. 1982. Seasonal food use and demographic trends in *Sciurus granatensis*. Pp. 239–252 in Leigh, E. G., Rand, A. S. & Windsor, D. M. (eds.). *The ecology of a tropical forest. Seasonal rhythms and long-term changes*. Smithsonian Institution Press, Washington DC.

HAMMOND, D. S., BROWN, V. K. & ZAGT, R. 1999. Spatial and temporal patterns of seed attack and germination in a large-seeded neotropical tree species. *Oecologia* 119:208–218.

HOCH, G. A. & ADLER, G. H. 1997. Removal of black palm (*Astrocaryum standleyanum*) seeds by spiny rats (*Proechimys semispinosus*). *Journal of Tropical Ecology* 13:51–58.

HULME, P. E. 1998. Post-dispersal seed predation: consequences for plant demography and evolution. *Perspectives in Plant Ecology, Evolution and Systematics* 1:32–46.

JANSEN, P. A., BONGERS, F. & PRINS, H. H. T. 2006. Tropical rodents change rapidly germinating seeds into long-term food supplies. *Oikos* 113;449–458.

JANSEN, P. A., VAN DER MEER, P. J. & BONGERS, F. 2008a. Is farther seed dispersal better? Spatial patterns of offspring mortality in three rainforest tree species with different dispersal abilities. *Ecography* 30:43–52.

JANSEN, P. A., BOHLMAN, S. A., GARZON-LOPEZ, C. X., OLFF, H., MULLER-LANDAU, H.-C. & WRIGHT, S. J. 2008b. Large-scale spatial variation in palm fruit abundance across a tropical moist forest estimated from high-resolution aerial photographs. *Ecography* 30:33–42.

JANSEN, P. A. & FORGET, P.-M. 2001. Scatterhoarding and tree regeneration. Pp. 275–288 in Bongers, F., Charles-Dominique, P.,

Forget, P.-M. & Théry, M. (eds.). *Nouragues: dynamics and plant–animal interactions in a Neotropical rainforest.* Kluwer Academic Publishers, Dordrecht.

JANZEN, D. H. 1971a. Seed predation by animals. *Annual Review of Ecology and Systematics* 2:465–492.

JANZEN, D. H. 1971b. The fate of *Scheelea rostrata* fruits beneath the parent tree: predispersal attack by bruchids. *Principes* 15:89–101.

JANZEN, D. H. 1972. Association of a rainforest palm and seed-eating beetles in Puerto Rico. *Ecology* 53:258–261.

JANZEN, D. H. 1977. Why fruits rot, seeds mold, and meat spoils. *American Naturalist* 111:691–713.

JOHNSON, C. D. & ROMERO, J. 2004. A review of evolution of oviposition guilds in the Bruchidae (Coleoptera). *Revista Brasileira de Entomologia* 48:401–408.

KAYS, R. W. 1999. Food preferences of kinkajous (*Potos flavus*): a frugivorous carnivore. *Journal of Mammalogy* 80:589–599.

KILTIE, R. A. 1981. Distribution of palm fruits on a rain forest floor: why white-lipped peccaries forage near objects. *Biotropica* 13:141–145.

LEIGH, E. G. 1998. *Tropical forest ecology.* Oxford University Press. 264 pp.

LEIGH, E. G. & RUBINOFF, I. 2005. Understanding and conserving tropical diversity: perspectives from Barro Colorado Island. Pp. 672 in Bermingham, E., Dick, C. W. & Moritz, C. (eds.). *Tropical rainforests: past, present, and future.* University of Chicago Press, Chicago.

MOORE, D. 2001. Insects on palm flowers and fruits. Pp. 233–266 in Howard, F. W., Giblin-Davis, R., Moore, D. & Abad, R. (eds.). *Insects on palms.* CABI Publishing, Wallingford.

MURIE, J. O. 1977. Cues used for cache finding by agouti (*Dasyprocta punctata*). *Journal of Mammalogy* 58:95–96.

NOTMAN, E. M. & VILLEGAS, A. C. 2005. Patterns of seed predation by vertebrate versus invertebrate seed predators among different plant species, seasons and spatial distributions. Pp. 55–75 in Forget, P. M., Lambert, J. E., Hulme, P. E. & Vander Wall, S. B. (eds.). *Seed fate: predation, dispersal, and seedling establishment.* CABI, Wallingford.

POTVIN, C., CANSARI, R., HUTTON, J., CAISAMO, I. & PACHECO, B. 2003. Preparation for propagation: understanding germination of giwa (*Astrocaryum standleyanum*), wagara (*Sabal mauritiiformis*), and eba (*Socratea exorrhiza*) for future cultivation. *Biodiversity and Conservation* 12:2161–2171.

PYKE, C. R., CONDIT, R., AGUILAR, S. & LAO, S. 2001. Floristic composition across a climatic gradient in a Neotropical lowland forest. *Journal of Vegetation Science* 12:533–566.

RUSSO, S. E. 2005. Linking seed fate to natural dispersal patterns: factors affecting predation and scatter-hoarding of *Virola calophylla* seeds in Peru. *Journal of Tropical Ecology* 21:243–253.

SILVA, M. G. & TABARELLI, M. 2001. Seed dispersal, plant recruitment and spatial distribution of *Bactris acanthocarpa* Martius (Arecaceae) in a remnant of Atlantic forest in northeast Brazil. *Acta Oecologica* 22:259–268.

SILVIUS, K. M. 2002. Spatio-temporal patterns of palm endocarp use by three Amazonian forest mammals: granivory or 'grubivory'? *Journal of Tropical Ecology* 18:707–723.

SILVIUS, K. M. 2005. Frugivore-mediated interactions among bruchid beetles and palm fruits at Barro Colorado Island, Panama: implications for seed fate. Pp. 45–54 in Forget, P. M., Lambert, J. E., Hulme, P. E. & Vander Wall, S. B. (eds.). *Seed fate: predation, dispersal, and seedling establishment.* CABI Publishing, Wallingford.

SILVIUS, K. M. & FRAGOSO, J. M. V. 2002. Pulp handling by vertebrate seed dispersers increases palm seed predation by bruchid beetles in the northern Amazon. *Journal of Ecology* 90:1024–1032.

SMYTHE, N. 1978. The natural history of the Central American agouti (*Dasyprocta punctata*). *Smithsonian Contributions to Zoology* 257: 1–52.

SMYTHE, N. 1989. Seed survival in the palm *Astrocaryum standleyanum*: evidence for dependence upon its seed dispersers. *Biotropica* 21:50–56.

SMYTHE, N., GLANZ, W. E. & LEIGH, E. G. 1982. Population regulation in some terrestrial frugivores. Pp. 227–238 in Leigh, E. G., Rand, A. S. & Windsor, D. M. (eds.). *The ecology of a tropical rain forest.* Smithsonian Institution Press, Washington DC.

STAPANIAN, M. A. & SMITH, C. C. 1978. A model for seed scatterhoarding: coevolution of fox squirrels and black walnuts. *Ecology* 59:884–896.

VANDER WALL, S. B. 1990. *Food hoarding in animals.* University of Chicago Press, Chicago. 445 pp.

VANDER WALL, S. B. 1991. Mechanisms of cache recovery by yellow pine chipmunks. *Animal Behaviour* 41:851–864.

VANDER WALL, S. B. 1995. Dynamics of yellow pine chipmunk (*Tamias amoenus*) seed caches: Underground traffic in bitterbrush seeds. *Ecoscience* 2:261–266.

VANDER WALL, S. B., BECK, M. J., BRIGGS, J. S., ROTH, J. K., THAYER, T. C., HOLLANDER, J. L. & ARMSTRONG, J. M. 2003. Interspecific variation in the olfactory abilities of granivorous rodents. *Journal of Mammalogy* 84:487–496.

WRIGHT, S. J. 1983. The dispersion of eggs by a bruchid beetle among *Scheelea* palm seeds and the effect of distance to the parent palm. *Ecology* 64:1016–1021.

WRIGHT, S. J. & DUBER, H. C. 2001. Poachers and forest fragmentation alter seed dispersal, seed survival, and seedling recruitment in the palm *Attalea butyracea*, with implications for tropical tree diversity. *Biotropica* 33:583–595.

WRIGHT, S. J., GOMPPER, M. E. & DELEON, B. 1994. Are large predators keystone species in Neotropical forests? The evidence from Barro Colorado Island. *Oikos* 71:279–294.

WRIGHT, S. J., ZEBALLOS, H., DOMINGUEZ, I., GALLARDO, M. M., MORENO, M. C. & IBANEZ, R. 2000. Poachers alter mammal abundance, seed dispersal, and seed predation in a Neotropical forest. *Conservation Biology* 14:227–239.