AN ANNOTATED CHECKLIST AND KEY TO THE REPTILES OF MEXICO EXCLUSIVE OF THE SNAKES

BY

HOBART M. SMITH AND EDWARD H. TAYLOR
The scientific publications of the National Museum include two series, known, respectively, as Proceedings and Bulletin.

The Proceedings series, begun in 1878, is intended primarily as a medium for the publication of original papers, based on the collections of the National Museum, that set forth newly acquired facts in biology, anthropology, and geology, with descriptions of new forms and revisions of limited groups. Copies of each paper, in pamphlet form, are distributed as published to libraries and scientific organizations and to specialists and others interested in the different subjects. The dates at which these separate papers are published are recorded in the table of contents of each of the volumes.

The series of Bulletins, the first of which was issued in 1875, contains separate publications comprising monographs of large zoological groups and other general systematic treatises (occasionally in several volumes), faunal works, reports of expeditions, catalogs of type specimens, special collections, and other material of similar nature. The majority of the volumes are octavo in size, but a quarto size has been adopted in a few instances in which large plates were regarded as indispensable. In the Bulletin series appear volumes under the heading Contributions from the United States National Herbarium, in octavo form, published by the National Museum since 1902, which contain papers relating to the botanical collections of the Museum.

The present work forms No. 199 of the Bulletin series.

Alexander Wetmore,
Secretary, Smithsonian Institution.
CONTENTS

Introduction .............................................................................. 1
Itinerary and gazetteer of localities for the Walter Rathbone Bacon
Expedition, 1938–1941 ................................................................ 4
Class Reptilia ........................................................................... 12
  Subclass Anapsida ................................................................. 12
    Order Testudines ................................................................. 12
      Suborder Atheca ............................................................... 13
        Family Dermochelyidae .................................................. 13
          Genus Dermochelys ...................................................... 13
      Suborder Carettoidea ........................................................ 14
        Family Cheloniidae ......................................................... 14
          Genus Lepidochelys ...................................................... 14
          Genus Caretta .............................................................. 15
          Genus Eretmochelys ...................................................... 16
          Genus Chelonia ............................................................ 17
      Suborder Trionychoidae ..................................................... 18
        Family Trionychidae ........................................................ 18
          Genus Amyda .............................................................. 18
      Suborder Cryptodira .......................................................... 18
        Family Dermatemyidae .................................................. 19
          Genus Dermatemys ..................................................... 19
        Family Chelydridae ........................................................ 20
          Genus Chelydra ........................................................... 20
        Family Kinosternidae .................................................... 21
          Genus Kinosternon ....................................................... 21
          Genus Claudius ............................................................ 26
          Genus Staurotypus ....................................................... 27
        Family Testudinidae ......................................................... 27
          Genus Gopherus ........................................................... 28
        Family Emyidae .............................................................. 28
          Genus Geoemyda ........................................................ 29
          Genus Pseudemys ........................................................ 31
          Genus Chrysemys ........................................................ 33
          Genus Terrapene ........................................................ 34
          Genus Malaclemys ....................................................... 36
          Genus Clemmys ........................................................... 37
  Subclass Lepidosauria ............................................................ 37
    Order Squamata .................................................................. 37
      Suborder Amphisbaenia .................................................... 37
        Family Bipedidae .......................................................... 37
          Genus Bipes ............................................................... 38
<table>
<thead>
<tr>
<th>Class</th>
<th>Subclass</th>
<th>Order</th>
<th>Page</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reptilia</td>
<td>Lepidosauria</td>
<td>Squamata</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Family Gekkonidae</td>
<td></td>
<td></td>
<td>40</td>
</tr>
<tr>
<td>Genus Coleonyz</td>
<td></td>
<td></td>
<td>41</td>
</tr>
<tr>
<td>Genus Gonatodes</td>
<td></td>
<td></td>
<td>45</td>
</tr>
<tr>
<td>Genus Phylloactylus</td>
<td></td>
<td></td>
<td>46</td>
</tr>
<tr>
<td>Genus Thecadactylus</td>
<td></td>
<td></td>
<td>49</td>
</tr>
<tr>
<td>Genus Hemidactylus</td>
<td></td>
<td></td>
<td>49</td>
</tr>
<tr>
<td>Genus Aristelliger</td>
<td></td>
<td></td>
<td>51</td>
</tr>
<tr>
<td>Genus Peropus</td>
<td></td>
<td></td>
<td>51</td>
</tr>
<tr>
<td>Genus Sphaerodactylus</td>
<td></td>
<td></td>
<td>52</td>
</tr>
<tr>
<td>Family Iguanidae</td>
<td></td>
<td></td>
<td>53</td>
</tr>
<tr>
<td>Genus Anolis</td>
<td></td>
<td></td>
<td>55</td>
</tr>
<tr>
<td>Genus Corythophanes</td>
<td></td>
<td></td>
<td>68</td>
</tr>
<tr>
<td>Genus Laemancus</td>
<td></td>
<td></td>
<td>69</td>
</tr>
<tr>
<td>Genus Basiliscus</td>
<td></td>
<td></td>
<td>71</td>
</tr>
<tr>
<td>Genus Iguana</td>
<td></td>
<td></td>
<td>72</td>
</tr>
<tr>
<td>Genus Clenosaurosa</td>
<td></td>
<td></td>
<td>73</td>
</tr>
<tr>
<td>Genus Enyaliosaurus</td>
<td></td>
<td></td>
<td>75</td>
</tr>
<tr>
<td>Genus Dipsoasaurus</td>
<td></td>
<td></td>
<td>77</td>
</tr>
<tr>
<td>Genus Sauromalus</td>
<td></td>
<td></td>
<td>78</td>
</tr>
<tr>
<td>Genus Holbrookia</td>
<td></td>
<td></td>
<td>81</td>
</tr>
<tr>
<td>Genus Callisaurus</td>
<td></td>
<td></td>
<td>85</td>
</tr>
<tr>
<td>Genus Uma</td>
<td></td>
<td></td>
<td>89</td>
</tr>
<tr>
<td>Genus Petrosaurus</td>
<td></td>
<td></td>
<td>90</td>
</tr>
<tr>
<td>Genus Streptosaurus</td>
<td></td>
<td></td>
<td>91</td>
</tr>
<tr>
<td>Genus Crotaphytus</td>
<td></td>
<td></td>
<td>91</td>
</tr>
<tr>
<td>Genus Gambelia</td>
<td></td>
<td></td>
<td>93</td>
</tr>
<tr>
<td>Genus Phrynosoma</td>
<td></td>
<td></td>
<td>94</td>
</tr>
<tr>
<td>Genus Sceloporus</td>
<td></td>
<td></td>
<td>105</td>
</tr>
<tr>
<td>Genus Sator</td>
<td></td>
<td></td>
<td>130</td>
</tr>
<tr>
<td>Genus Urosaurus</td>
<td></td>
<td></td>
<td>140</td>
</tr>
<tr>
<td>Genus Uta</td>
<td></td>
<td></td>
<td>147</td>
</tr>
<tr>
<td>Family Xantusiidae</td>
<td></td>
<td></td>
<td>151</td>
</tr>
<tr>
<td>Genus Lepidophyosma</td>
<td></td>
<td></td>
<td>151</td>
</tr>
<tr>
<td>Genus Gaigia</td>
<td></td>
<td></td>
<td>153</td>
</tr>
<tr>
<td>Genus Xantusi</td>
<td></td>
<td></td>
<td>154</td>
</tr>
<tr>
<td>Family Scincidae</td>
<td></td>
<td></td>
<td>155</td>
</tr>
<tr>
<td>Genus Mabuya</td>
<td></td>
<td></td>
<td>155</td>
</tr>
<tr>
<td>Genus Scincella</td>
<td></td>
<td></td>
<td>156</td>
</tr>
<tr>
<td>Genus Buenees</td>
<td></td>
<td></td>
<td>160</td>
</tr>
<tr>
<td>Family Anelytropsidae</td>
<td></td>
<td></td>
<td>170</td>
</tr>
<tr>
<td>Genus Anelytropsia</td>
<td></td>
<td></td>
<td>170</td>
</tr>
<tr>
<td>Family Teiididae</td>
<td></td>
<td></td>
<td>170</td>
</tr>
<tr>
<td>Genus Ameiva</td>
<td></td>
<td></td>
<td>170</td>
</tr>
<tr>
<td>Genus Cnemidophorus</td>
<td></td>
<td></td>
<td>174</td>
</tr>
<tr>
<td>Genus Gymnophthalmus</td>
<td></td>
<td></td>
<td>192</td>
</tr>
<tr>
<td>Family Helodermidae</td>
<td></td>
<td></td>
<td>192</td>
</tr>
<tr>
<td>Genus Heloderma</td>
<td></td>
<td></td>
<td>192</td>
</tr>
</tbody>
</table>
**CONTENTS**

Class Reptilia—Continued
Subclass Lepidosauria—Continued
Order Squamata—Continued
Suborder Sauria—Continued
  
  **Family Anguidae**…………………………………………………………… 194
  Genus *Celestus*…………………………………………………………… 194
  Genus *Abronia*…………………………………………………………… 196
  Genus *Barisia*…………………………………………………………… 198
  Genus *Gerrhonotus*……………………………………………………… 203
  Genus *Elgaria*…………………………………………………………… 205
  
  **Family Xenosauridae**………………………………………………….. 207
  Genus *Xenosaurus*……………………………………………………… 207
  
  **Family Anniellidae**………………………………………………….. 208
  Genus *Anniella*………………………………………………………… 209

Suborder Serpentes.1

Subclass Archosauromorpha—Continued
Order Loricata………………………………………………………………… 209
Family Crocodylidae………………………………………………………… 210
  Genus *Crocodylus*……………………………………………………… 210
Family Alligatoridae……………………………………………………….. 211
  Genus *Caiman*………………………………………………………… 212

Species inquirendae………………………………………………………… 212
State lists…………………………………………………………………… 215
Index……………………………………………………………………… 231

---

AN ANNOTATED CHECKLIST AND KEY TO
THE REPTILES OF MEXICO, EXCLUSIVE
OF THE SNAKES

By Hobart M. Smith and Edward H. Taylor

INTRODUCTION

This volume constitutes the last of a series of checklists and keys to the herpetological fauna of Mexico. The first, "An Annotated Checklist and Key to the Snakes of Mexico" (U. S. Nat. Mus. Bull. 187), was published on October 5, 1945. The second, "An Annotated Checklist and Key to the Amphibia of Mexico" (U. S. Nat. Mus. Bull. 194), appeared on June 17, 1948. In this, the third volume, the turtles, amphisbaenians, lizards, and crocodilians are treated. The preparatory work that preceded these checklists was pursued as time permitted from 1932 to the present, a period of about 17 years. It has involved a number of expeditions to Mexico at our own expense and the collecting of more than 50,000 herpetological specimens.

Except in rare instances the materials have all passed under the scrutiny of one or the other of us, and the detailed results of these studies are embodied in more than a hundred short or longer reports. Our own ideas have often changed with the acquisition of materials not originally available.

It was hoped that the long-awaited work on turtles by Stejneger and Hartweg would see the light before the appearance of the turtle checklist, but since there is still no certainty of the completion of this work in the immediate future it does not seem wise to delay on this account.

We have refrained from anticipating certain forms not now recognized that are present even in our own collection; the specimens representing them are referred without comment to the recognized form they most closely resemble. A number of such forms belong to Anolis, no adequate revision of which has yet been made, although the senior author is contemplating such a work. Certain other groups, especially Cnemidophorus and Uta, merit reinvestigation.

There are many other unsolved problems in Mexican herpetology; much more exploration remains to be performed. Probably no single

---

1 From 1938 to 1941 I was financed by the Smithsonian Institution on several expeditions, through tenure of a Walter Rathbone Bacon Traveling Scholarship.—H. M. S.
problem of distribution is more demanding of explanation than the extreme dearth of Plethodontidae in the Guerrero "island" and in the high western plateau, and the apparent replacement of that family in the latter region, largely by the Ambystomidae.

The treatment of forms in this volume differs little from that of the two preceding volumes, except that the name of the collector of each type is added. This name often is as important perhaps as the name of the author of a species. Unfortunately, the collector is frequently unknown or uncertain. This is true of the collections from Mexico studied by Wiegmann, in which case we have given Deppe credit for obtaining most of the reptiles. The other two men constituting the collecting party, Count Von Sack and Schiede, were, we believe, engaged primarily in the collection of orchids. However, it is not at all improbable that some forms accredited to Deppe were actually taken by one of the others.

We refrain here from considering the implications that may be deduced from a perusal of the combined data of the three volumes, such as an investigation of faunal origins, lines of migration, relative age of taxonomic groups, and faunal areas of ecological relationships. However, we are fully aware that such consideration would doubtless yield valuable information.

The number of forms recognized in the herpetofauna of Mexico, including those forms described since the first of these checklists was published, is relatively large. The following table represents the numbers in the various groups:

<table>
<thead>
<tr>
<th>Order or suborder</th>
<th>Families</th>
<th>Genera</th>
<th>Species and subspecies</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gymnophiona</td>
<td>1</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Caudata</td>
<td>4</td>
<td>16</td>
<td>65</td>
</tr>
<tr>
<td>Salientia</td>
<td>7</td>
<td>25</td>
<td>163</td>
</tr>
<tr>
<td>Testudines</td>
<td>8</td>
<td>18</td>
<td>49</td>
</tr>
<tr>
<td>Amphisbaenia</td>
<td>1</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>Sauria</td>
<td>10</td>
<td>47</td>
<td>394</td>
</tr>
<tr>
<td>Serpentes</td>
<td>8</td>
<td>80</td>
<td>486</td>
</tr>
<tr>
<td>Loricata</td>
<td>2</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td><strong>41</strong></td>
<td><strong>191</strong></td>
<td><strong>1,165</strong></td>
</tr>
</tbody>
</table>

That there will in the future be considerable shifting of species to the subspecific category, and changing forms we regard as subspecies to the species category, is to be anticipated. On occasion we have found even ourselves at variance on certain problems of this sort. We both feel, however, that the currently and deservedly popular trend toward recognition of subspecies where they exist has brought
with it, almost inevitably, occasional overzealous reduction of species to subspecies. Such changes are made without due demand for facts, but instead with speculations that often merely serve as an excuse to tamper with an accepted arrangement that may be equally as plausible.

One of the most exasperating tasks has been the unraveling of certain purely nomenclatorial knots. Several names, for example, are of controversial orthography. Worthy of special mention are *Agkistrodon* vs. *Ancistrodon*, *Cnemidophorus tessellatus* vs. *C. tesselatus*, *Corythophanes* vs. *Corythophanes*, and *Kinosternon* vs. *Cinosternon*. Opinion 36 of the International Commission on Zoological Nomenclature interprets the permission given by Article 19 to correct any “error of transcription” to mean also “error of transliteration,” whether the original author was aware of his error or not. By such procedure, the names *Ancistrodon*, *tesselatus*, *Corythophanes*, and *Cinosternon* would be recommended. On the other hand, Moore, Weller, and Knight (Journ. Paleont., vol. 16, 1942, pp. 250–261) maintain with excellent reason that only in very clear-cut and exceptional cases is any modification of the original orthography of a generic name justified. Their view is strengthened by Blackwelder, Knight, and Sabrosky (Science, vol. 108, 1948, pp. 37–38). Maintenance of original form would perpetuate *Agkistrodon*, *tesselatus*, *Kinosternon*, and *Corythophanes*. Since the proper procedure is not universally or even generally agreed upon, we see little advantage in deviation from currently accepted form until some means of real standardization is provided by the International Commission on Zoological Nomenclature.

For the first time in these checklists we here adopt the practice of placing a comma between scientific name and author in citations of all references except the original.

We wish to acknowledge the assistance of W. Leslie Burger in the laborious task of checking and rechecking various points in the manuscript, and in criticizing certain parts, and of Dr. Doris M. Cochran, zoologist, division of reptiles and amphibians, United States National Museum, who devoted much time to curatorial duties connected with the voluminous collection and in making available for study the National Museum herpetological collections. We are grateful likewise for the corrections and improvements suggested by Dr. D. F. Hoffmeister, Karl P. Schmidt, and Dr. Frederick Shannon. The Graduate Research Boards of the University of Illinois and the University of Kansas have generously provided financial support for research and clerical work involved in completion of the present project, and the Walter Rathbone Bacon Scholarship of the Smithsonian Institution furnished a substantial sum for travel and research in Mexico.
ITINERARY AND GAZETTEER OF LOCALITIES FOR THE WALTER RATHBONE BACON EXPEDITION, 1938-1941

As previously stated, the senior author was enabled to collect in and study material from Mexico by tenure of a Walter Rathbone Bacon Traveling Scholarship from 1938 to 1941. The collection obtained during those years has formed an important addition to other material from Mexico and merits an elaboration of the routes traversed by the expedition and the localities at which specimens were secured. Such an itinerary was, as a matter of fact, to have accompanied a study of the lizards of the Bacon collection, as stated in our introduction to the summary of the amphibians (Proc. U. S. Nat. Mus., vol. 95, 1945, p. 521). It now seems unlikely that a complete survey of the Bacon lizards can appear for a number of years, for a satisfactory completion of such a study involves revision of a number of very sketchily understood genera, such as Anolis and Cnemidophorus. In the meantime a need for an account of the Bacon itinerary and localities continues to exist. We have, accordingly, taken this opportunity to present this information in this the final checklist.

The expedition personnel consisted of the senior author and his wife. They received hearty support in field work from a number of individuals, through whose combined efforts about 22,000 specimens were secured during a period of two years. As closely as can now be determined (the lizards not yet having been fully studied), 500 species and subspecies (146 amphibians, 160 lizards, 1 amphibiaenian, 170 snakes, 20 turtles, 3 crocodilians) were secured.

Among those who contributed to the success of the expedition are Dr. Alexander Wetmore, who saw to it that the expedition was properly planned and could function smoothly; Dr. Doris M. Cochran, who must have nearly equaled the expedition personnel in time expended for the collection, inasmuch as the laborious task of cataloging the specimens rested with her; Dr. Linton P. Satterthwaite, who provided the facilities for our stay at Piedras Negras in the midst of a little-known, remote, and austere area; Mr. and Mrs. Dyfrig McH. Forbes, who provided for nearly two years a base of operations in Veracruz and who were a constant source of companionship, inspiration, and material; Eizi Matuda, who very generously provided facilities for a two-month stay at his finca and who sympathetically aided us in every possible way to sample the herpetofauna of the area as thoroughly as possible; Thomas MacDougall, who in Tehuan tepec secured for us numerous specimens and arranged for our travel into areas otherwise difficult of access; Ernest Rateike, of Palenque, Chiapas, who accepted us as a guest in his home for a month; and a
host of local residents, far too numerous to mention, who materially augmented our collections and made our visits pleasant and successful.

Inasmuch as we traveled chiefly by car, our routes are best followed on a road map of Mexico. We entered Mexico on October 5, 1938, at Ciudad Juárez, Chihuahua, and followed the Chihuahua–El Paso highway southward to Vado (October 9). We then turned back to Ahumada and followed a poor trail through Carrizal and Rancho Nuevo to Progreso (October 10). We camped beside the Río Santa María about a mile southwest of Progreso for 5 days, interrupted only by one hurried round trip to Casas Grandes for the benefit of a snake-bitten cowboy. At this camp we were aided greatly by 10 or 15 cowboys who brought in material as fast as it could be preserved. Our specimen containers full, we determined to go to the border to ship specimens and likewise to obtain higher wheels for the car (a half-ton panel truck), with which we had encountered numerous difficulties because of deep ruts and high centers. We left the Progreso camp on October 15, passing through Casas Grandes and Ascención, and reached Las Palomas the same day. After two days in Columbus and Deming, N. Mex., we retraced our steps (October 20) from Las Palomas to Casas Grandes. As the weather had become cool, we continued southward along the road through San Buenaventura and Carmen to the main El Paso–Chihuahua highway, thence southward to Ciudad Chihuahua. We left there on October 27 for Torreón, arriving October 29. We collected in the vicinity of Torreón October 30 and 31, and on November 1 started for Ciudad Durango. We could get no farther than Pedriceña, however, and, after exploring with little success a side road to Nazas, returned to Torreón on November 4. We continued to San Pedro the next day and collected in the vicinity of that town until November 10. After a side trip to Parras off the Torreón–Saltillo highway, we made an attempt to find Jaral, Coahuila, a locality made famous zoologically by Heller and Barber. This turned out to be an abandoned ranch near Hipólito, and, as no likely habitat for the montane species recorded from Jaral appeared to be nearer than 20 miles or so, we continued to Saltillo (November 14). We collected in the vicinity of Saltillo, Arteaga, and Mount Zapalinaámé until November 17, when we left for Monterrey.

On November 18 we arrived at Hacienda La Clementina (near Forlón), and we remained there until November 28, when we drove to Laredo to ship another lot of specimens. We returned to La Clementina on December 3 and left December 5 for Mexico City. We stayed at Huichihuayán for several days, leaving December 9. Our route, with brief stops at or near various towns, then led through Mexico City to Tehuacán, Puebla, thence through Orizaba to Potrero Viejo, Veracruz, where we arrived on December 11.
Potrero Viejo remained our headquarters for over a month. We made numerous short trips during that time; especially noteworthy were those to Palma Sola (on the Veracruz–Orizaba highway), to Tezonapa, Veracruz, and Cosolapa, Oaxaca, and to Cuautlapan, Orizaba, Córdoba, Acultzingo, and other localities along the Córdoba–Tehuacán road. We left Potrero Viejo on January 16, 1939, and after a brief stop in Mexico City made headquarters in Cuernavaca, Morelos, for nearly two weeks, with side trips to Puente de Ixtla (Morelos), Cacahuamilpa (Guerrero), and Zempoala (Morelos and México).

Leaving Cuernavaca February 2, we continued toward Acapulco, reaching there February 5. We collected in the vicinity of Acapulco, with side trips to Coyuca, until February 11. Our route, with frequent stops, then led northward again to Mexico City, where we arrived on March 1. After two days in search of axolotls and other ambystomids in the vicinity of Mexico City (with side trips to Texcoco, Zumpango, and Chimalhuacán), we started (March 4) on the road to Guadalajara. Our first deviation from this route was on March 9 and 10, to Pátzcuaro. On March 11 we reached Uruapan on another side trip and continued southward to Apatzingán, returning to Uruapan on March 19. An attempt was made to find *Crotalus poly-STICTUS* in the marshes of the eastern end of Lake Chapala, but without success, since most of the marshes have been drained and are now under cultivation. We reached Guadalajara on March 24 and returned to Potrero Viejo on March 26. On March 30 we left Potrero Viejo enroute to Laredo with another shipment of specimens, arriving at Laredo on April 3, 1939.

After nearly a month in the United States we left Laredo on April 29, 1939, and arrived in Potrero Viejo on May 2. From there we went to Veracruz by train and secured passage for Álvaro Obregón, arriving on May 13. A river boat was then taken, via Ciudad del Carmen, to Tenosique, Tabasco, the head of navigation on the Río Usumacinta. Our final destination, Piedras Negras, Guatemala, was reached on May 21 after two days by mule. We collected in the immediate vicinity of Piedras Negras until June 23, when we left for Tenosique.

From Tenosique (June 30) we continued downstream to Emiliano Zapata and there procured mules for a 2-day trip to San Juanito, a ranch half a mile from the village of Palenque. There we remained, with a side trip only to the ruins of Palenque several miles distant in the hills, until August 6, when we left for Álvaro Obregón and, immediately thereafter, Potrero Viejo, where we arrived on August 14. There we were joined by the junior author and with him left on August 18 for Mexico City, where we arrived, via the usual route through Tehuacán and Puebla, on August 22, after numerous brief
CHECKLIST OF REPTILES OF MEXICO

Geographic movements.

On August 29 we took the Acapulco road and followed it with only one deviation: to Tixtla, Guerrero, arriving at Acapulco on September 3. We returned to Mexico City by the same route, arriving on September 6. Again we collected in the immediate environs of Mexico City, chiefly investigating montane faunas. On September 10 we left for Guadalajara but turned back near Sahuayo. We then, on September 12, started northward on the Pachuca Road to El Chico National Park in Hidalgo. After one day there we returned, September 18, to Mexico City, where Dr. Taylor left us.

We then returned, September 19, to Potrero Viejo. After a few local trips we again returned to the Guadalajara road, leaving September 29. We took the side road to the Nevado de Toluca (October 2) and then retraced our steps to Mexico City, arriving October 3. After a few days spent in local trips, we turned northward to Laredo with another load of specimens, arriving at Laredo October 17. Our only side trip enroute was to Galeana, Nuevo León, via the road from Linares, Tamaulipas (October 13, 14).

After nearly two months in the United States, we again crossed the border at Laredo on December 10, 1939, and headed directly for Potrero Viejo, Veracruz, arriving December 14. After a few local trips near Potrero Viejo, we left (December 27) for Tehuantepec by road. We arrived on December 30 and remained until January 28, 1940. A number of side trips were taken, by truck to Cerro Arenal, by rail to Matías Romero, Oaxaca, by rail to Salina Cruz, and by rail to Tonala. Illness necessitated return to Potrero Viejo and ultimately to Mexico City. We remained there until March 15, when we returned to Potrero Viejo. On March 17 we started on a side trip, lasting until March 24, which led by highway through Tehuacán, Tecamachalco, thence northeastward on the Jalapa road, to Teziutlán (Puebla) from El Limón, Totalco, and eastward to Puerto Nacional.

Shortly thereafter we left for Tehuantepec by rail again, arriving April 1. We stayed only one day and traveled by rail to Acapetahua, whence we went by bus to Escuintla. From there pack animals were used to transport our equipment to La Esperanza, a finca about 5 miles northeast of Escuintla. We arrived there on April 4 and remained until June 5, 1940. Various short side trips were taken to nearby fincas.

On June 5 we returned to Tehuantepec and, after one day, to Potrero Viejo, arriving June 9. About one month later, on July 6, we crossed the border at Laredo with no deviations from the direct route from Potrero Viejo to the border.
We returned immediately to Mexico City by way of the Pan American Highway and remained there until August 7, except for a trip by rail to Guanajuato, Guanajuato, on July 19-21. On August 8 we again drove to Potrero Viejo. Within a few days we made a brief and final foray along the highway from Acultzingo to Tehuacán before packing all equipment that had been stored at Potrero Viejo. We left our headquarters there for the final time on August 8 and, with but a brief stop in Mexico City, traveled slowly northward along the Pan American Highway, with a heavy load, crossing the border at Laredo on August 24.

The following gazetteer includes only those localities represented by specimens in the Bacon collection that are not to be found on the National Geographic Society’s 1939 map of Mexico, Central America, and the West Indies, scale 1:5,702,400. Most of the localities are plotted on other, larger-scale maps, but inasmuch as these are not always readily available we include all missing from the Geographic Society’s map, which is readily available to all investigators. The localities are arranged alphabetically within the states, which are themselves alphabetically arranged.

CHIAPAS

Acacoyagua: 3 miles northwest of Escuintla.
Aguacate: 7 miles north-northeast by north of Palenque.
Belén: 20 miles southeast of Escuintla.
Cerro Obando: 3 miles northeast of La Esperanza (which see).
Colonia Hidalgo: 7 miles northeast of Acacoyagua (which see).
Colonia Soconusco: 12 miles northeast of La Esperanza (which see).
Cruz de la Piedra: 2 miles west of La Esperanza (which see).
Finca Juárez: 10 miles northeast of La Esperanza (which see).
Javarinero: 13 miles east of Palenque.
La Esperanza: 3 miles east of Acacoyagua (which see).
La Magnolia: 3 miles northwest of La Esperanza (which see).
Las Nubes: On Cerro Obando, 2 miles northeast of La Esperanza (which see).
Motozintla: 25 miles east-northeast by east of Escuintla.
Rancho Las Gradas: 2 miles west of La Esperanza (which see).
Salto de Agua: On Cerro Obando, 1 mile northeast of La Esperanza (which see).
San Juanito: 1 mile east-northeast of Palenque.
Santa Rosa: Near Comitán.

CHIHUAHUA

Carmen: 32 miles east-northeast by east of Galeana.
Carrizal: 10 miles west-southwest by south of Ahumada.
Ciudad Delicias: 5 miles south of Meoqui.
Ojo de Federico: A small ranch 8 miles southeast of Ascención.
Progreso: 35 miles northeast of Galeana, near Río Santa María.
Rancho Nuevo: A small ranch 10 miles west-northwest by west of Carrizal.
Samalayuca: 25 miles south of Ciudad Juárez.
CHECKLIST OF REPTILES OF MEXICO

COAHUILA

Hipólito: 25 miles north-northeast of General Cepeda.
Zapaliname: A mountain 3 miles south of Saltillo.

DISTRITO FEDERAL

Atzacoalco: 10 miles northeast of Mexico City.
Cañada de Contreras: 5 miles southeast of Mexico City.

DURANGO

La Goma: 12 miles west of Lerdo, on the south side of the Río Nazas.
La Loma: 12 miles west of Lerdo, on the north side of the Río Nazas.
Pedriceña: 25 miles east-southeast of Nazas.

GUATEMALA

Desempeña: 4 miles southeast of Piedras Negras.
Pozo de la Jicotea: 2 miles southeast of Piedras Negras.

GUERRERO

Agua Bendita: 13 miles south of Taxco.
Agua del Obispo: 8 miles north of Dos Caminos.
Cacahualnipa: 10 miles southeast of Tetecala, Morelos.
Coyuca: 7 miles northeast of Acapulco.
El Treinta: 20 miles north of Acapulco.
Garrapatas: 5 miles southwest of Dos Caminos.
Julian Blanco: 4 miles north of Dos Caminos.
Ocotito: 3 miles north of Dos Caminos.
Omitlame: 3 miles northwest of Chilpancingo.
Paso del Limonero: 9 miles northeast of Acapulco.
Tierra Colorada: 3 miles southwest of Garrapatas (which see).
Xaltianguis: 31 miles northeast of Acapulco.

HIDALGO

Barranca de los Horcones: 6 miles south of Durango (which see).
Chapulhuacán: 19 miles southwest of Tamazunchale, San Luis Potosí.
Colonia: 6 miles southeast of Pachuca.
Durango: 15 miles west-southwest by south of Jacala.
El Chico Parque Nacional: 15 miles northeast of Pachuca.
Maguey Verde: 7 miles south of Durango.
Tianguistengo: 10 miles north-northeast of Zacualtipán.

MÉXICO

Chalco: 13 miles northeast of Xochimilco.
Chimalhuacán: 12 miles east of Mexico City.
Lerma: 10 miles east of Toluca.
Nevado de Toluca: 32 miles south-southwest by south of Toluca.
Río Frío: 18 miles west-northwest of Texmelucán, Puebla.
Santa Magdalena: 15 miles east of Mexico City.
Zempoala: 6 miles west-southwest of Tres Cumbres, Morelos.
MICHOCÁN

Carapa(n): 23 miles north of Paracho.
El Temascal: 20 miles east of Morelia.
La Palma: 10 miles north of Sahuayo.
Puerto Hondo: 23 miles east of Morelia.
Rancho San José: 24 miles east of Morelia.
Tacícuaro: 5 miles east of Quiroga.

NUEVO LEÓN

Mamulique Pass: 45 miles north of Monterrey.
Santa Caterina: 7 miles west of Monterrey.

OAXACA

Cajón de Piedra: 12 miles south-southwest of Salina Cruz.
Cerro Arenal: 16 miles west of Tehuantepec.
Cerro de Chipehua: 16 miles southeast of Salina Cruz.
Cerro de Huamelula: 34 miles west-southwest by south of Tehuantepec.
Cerro Guengola: 10 miles west-northwest by west of Tehuantepec.
Cosolapa: 8 miles northwest of Acatlán.
Coyol: between San Antonio and Las Cruces.
El Limón: 25 miles southeast of Tehuantepec.
Escurano: 15 miles west-northeast of Tehuantepec.
La Concepción: 32 miles west of Tehuantepec.
Lachiguiri: 29 miles northeast of Tehuantepec.
La Gloria: 8 miles southeast of Chimalapa.
Las Pilas: 4 miles northwest of Tehuantepec.
Las Vacas: 40 miles west of Tehuantepec.
Llano Ocotal: 20 miles west-southwest by west of Tehuantepec.
Matías Romero: 34 miles north of San Gerónimo.
Mixtequilla: 4 miles northwest of Tehuantepec.
Palmar: 37 miles west-northwest by west of Tehuantepec.
Pixixi: 6 miles south of Tehuantepec.
Portillo Guayabo: 16 miles west of Tehuantepec.
Portillo Las Vacas: 40 miles west of Tehuantepec.
Portillo Los Nanches: 32 miles northwest of Tehuantepec.
Rincón San Pedro: 16 miles northeast of Tehuantepec.
Río Grande: 20 miles west-southwest by south of Tehuantepec.
Río Hondo: 41 miles west-northwest by west of Tehuantepec.
San Francisco Guichina: 61 miles west-northwest by west of Tehuantepec.
San José Manteca: 61 miles west-northwest by west of Tehuantepec.
San Mateo del Mar: 14 miles east-northeast by east of Salina Cruz.
San Pedro Quiechapa: 10 miles west of Yautepec.
Tenango: 24 miles west-southwest by west of Tehuantepec.
Tres Cruces: 43 miles west-northwest by west of Tehuantepec.
Yerba Santa: 10 miles west-northwest by west of Tehuantepec.

PUEBLA

Alchichica: 10 miles southwest of Perote, Veracruz.
Cacaloapam: 8 miles southeast of Tlacotepec.
El Seco: 27 miles northeast of Tepenca.
La Virgin: 2 miles north of Cacaloapam.
CHECKLIST OF REPTILES OF MEXICO

Ozumbilla: 4 miles east-southeast of Morelos.
Pájaro Verde: 300 feet west of Puebla–Veracruz state line, about 7 miles south-east of Morelos.
Tecamachalco: 15 miles northwest of Tlacotepec.

SAN LUIS POTOSÍ

Pujal: 20 miles north of Tancanhuitz.

TABASCO

El Retiro: 13 miles south of Tenosique.
Los Rieles: 10 miles south of Tenosique.
Santo Tomás: 12 miles southeast of Tenosique.

TAMAULIPAS

Ciudad Mante (=Villa Juárez): 18 miles east-northeast by north of Antiguo Morlos.
Forlón: 18 miles east-southeast of Llera.
Hacienda La Clementina: 13 miles east-southeast of Llera.

TLAXCALA

Apizaco: 8 miles north-northwest by north of Tlaxcala.

VERACRUZ

Acultzingo: 9 miles east-southeast by east of Morelos, Puebla.
Atoyac: 4 miles east-northeast of Potrero Viejo (which see).
Cerro Gordo: 20 miles east-southeast of Jalapa.
Cuautlapan: 4 miles southwest of Fortín (which see).
Cruz Blanca: 8 miles northeast of Perote.
El Limón Totalco: 10 miles southwest of Perote.
El Maguey: 8 miles east-southeast by east of Potrero Viejo (which see).
Encero: 8 miles east-southeast of Jalapa.
Fortín: 5 miles east of Córdoba.
Matacabestre: 7 miles north-northwest by north of Joaquín.
Mata de Caña: 18 miles east-southeast of Jalapa.
Medellín: 10 miles south of Veracruz.
Metlac: 1 mile west of Fortín (which see).
Ojo de Agua: 2 miles east of Paraje Nuevo (which see), near Potrero Viejo.
Pan de Olla: 8 miles south of Teziutlán, Puebla.
Paraje Nuevo: 2 miles east of Peñuela (which see).
Paso del Macho: 5 miles northeast of Atoyac.
Peñuela: 4 miles southeast of Córdoba.
Potrero Viejo: 2 miles east of Paraje Nuevo (which see).
Presidio: 22 miles southwest of Córdoba.
San Juan de La Punta: 15 miles east-southeast by south of Córdoba.
San José de Gracia: 13 miles southeast of Córdoba.
Palma Sola: 9 miles east-southeast by south of Córdoba.
Tequeyutepec: 10 miles northeast of Jalapa.
Tezonapa: 8 miles northwest of Acatlán.
Toxtlacuaya: 17 miles northwest of Jalapa.
Xuchil: 1 mile north of Potrero Viejo (which see).
Class REPTILIA Laurenti

Reptilia Laurenti, Specimen medicum exhibens synopsin reptilium, 1768, p. 19.

Subclasses.—Three subclasses are represented by living members. All occur in Mexico.

KEY TO MEXICAN SUBCLASSES, ORDERS, AND SUBORDERS OF REPTILIA

1. Provided with a shell encasing body above and below, consisting of plastron and carapace; turtles...subclass Anapsida...order Testudines (p. 12) 5
2. Anal slit longitudinal; skull diapsid...subclass Archosauiria.
   - order Loricata (p. 209)
   - Anal slit transverse; skull modified diapsid; lower (quadratejugal-quadrate) arch interrupted...subclass Lepidosauria...order Squamata (p. 37)
3. Two limbs only, the forelegs; body ringed with grooves.
   - subclass Amphisbaenia (p. 37)
   - Four limbs or none; body not ringed with grooves
4. Limbs present or, if not, movable eyelids present; lizards.
   - subclass Sauria (p. 39)
   - Limbs absent; movable eyelids absent; snakes...subclass Serpentes
5. No epidermal scutes on shell
6. Epidermal scutes present
   - subclass Atheca (p. 13)
   - Limbs with distinct digits, 3 with claws...subclass Trionychoidea (p. 18)
7. Limbs oar-shaped, with 1 to 3 claws...subclass Carettidea (p. 14)
   - Limbs not oar-shaped, at least foreleg with 4 or 5 claws.
   - subclass Cryptodira (p. 18)

Subclass ANAPSIDA Williston


Orders.—A single living order exists, the Testudines.

Order TESTUDINES Batsch

Chelonia Macartney, in Ross, Transl. Cuvier’s Leçons d’anatomie comparée...vol. 1, 1802, tab. 3.
Testudinata Oppel, Die Ordnungen, Familien und Gattungen der Reptilien...1811, p. 3.

Suborders.—Five living suborders of Testudines are recognized; four occur in Mexico, and the fifth (Pleurodira) is restricted to southern portions of both hemispheres.
Suborder ATHECA Cope


Families.—One.

Family DERMOCHELYIDAE 4 Fitzinger

Dermatochelydae Fitzinger, Systema reptilium, 1843, p. 30.

Genera.—One.

Range.—Tropical oceans of the world.

Genus DERMOCHELYS Blainville


Genotype.—Testudo coriacea Linnaeus.

Range.—World-wide, in tropical oceans and occasionally into temperate waters.

Species.—As many as three species (or subspecies) may be valid. The species has been recorded in Mexico only from the western coast, although Atlantic records are to be expected.

DERMOCHELYS CORIACEA 4 (Linnaeus)


Type.—Unknown.

Type locality.—Palermo, Sicily, by present restriction.

Range.—Pacific coast and probably the tropical Atlantic coast. Recorded definitely from Sonora: Guaymas; Baja California: Los Coronados Islands.

4 Actually the first family name proposed for this group was Sphargidae Gray (Ann. Philos., vol. 10, 1825, p. 212), based upon the genus Sphargis Merrem, 1820, a junior synonym of Dermochelys Blainville, 1816; both genera are based upon the same type, Testudo coriacea Linnaeus. Many taxonomists retain the oldest family name regardless of the status of its type. We believe there is a certain degree of error counted by such a procedure, however, inasmuch as a family name based upon a generic name later found to be a junior homonym of an earlier name in another family would, very unfortunately, be preserved; and what if the senior homonym had also served as a type for its family? It seems to us that only currently valid generic names should be available as types for family names, and that all synonyms and homonyms should be regarded unavailable for that purpose.

4 A common arrangement restricts coriacea to the Atlantic, schlegelii to the Pacific, either as species or subspecies. We are unable to find that anything more than geographic probability has led to such arrangement.
Suborder CARETTOIDEA\(^6\) Fitzinger

Carettoidea Fitzinger, Neue Classification der Reptilien . . . , 1826, p. 5.

_Families._—One.

Family CHELONIIDAE Gray


_Genera._—Four.

_Range._—Tropical oceans, world-wide.

**KEY TO GENERA OF THE FAMILY CHELONIIDAE**

1. One pair of prefrontal scutes; costal scutes 4; horny cutting edge of lower jaw coarsely dentate, that of upper jaw strongly ribbed vertically; bony alveolar ridge of upper jaw with a low but regularly raised auxiliary ridge behind anterior ridge, which is very strong and terminates anteriorly in a tooth at posterolateral corner of premaxillary pit.________Chelonia (p. 17)

   Two pairs of prefrontals; costal scutes 4–9; horny cutting edge of lower jaw smooth or feebly denticulate, that of upper jaw without markedly elevated vertical ribbing on its inner surface; bony alveolar surface of upper jaw smooth or with a single ridge; this ridge not terminating anteriorly in a sharp tooth._________________________ 2

2. Costal scutes 4 pairs; dorsal scutes usually conspicuously imbricate; bony alveolar surface of upper jaw with a sharp-crested ridge.

   _Eretmochelys_ (p. 16)

   Costal scutes 5 pairs or more; dorsal scutes not conspicuously imbricate; bony alveolar surface of upper jaw smooth or with a rounded ridge________ 3

3. Four enlarged inframarginal scutes on bridge; dorsal color gray to olive-green; maxillaries not in contact, separated by prevomer.________Lepidochelys (p. 14)

   Three enlarged inframarginal scutes on bridge; dorsal color brown or reddish brown; maxillaries in contact between prevomer and premaxillaries.

   _Caretta_ (p. 15)

Genus LEPIDOCHELYS Fitzinger

_Lepidochelys_ Fitzinger, Systema reptilium, fasc. 1, 1843, p. 30.

_Cauvana_ Gray, Catalogue of the tortoises . . . in the British Museum, 1844, p. 52 (type, _Chelonia olivacea_ Eschscholtz).

_Genotype._—_Chelonia olivacea_ Eschscholtz.

_Species._—Two are generally recognized.

_Range._—Tropical oceans, world-wide.

**KEY TO SPECIES OF LEPIDOCHELYS**

1. Inframarginal scutes without pores; usually 5 pairs of costal scutes; color usually gray; bony alveolar surface of upper jaw with a conspicuous ridge.

   _kempfi_ (p. 15)

   Each inframarginal scute usually with a pore at its posterior border; usually more than 5 pairs of costal scutes; color olive; bony alveolar surface of upper jaw with a low (not conspicuous) ridge_________________________olivacea (p. 15)

---

\(^6\) Deraniyagala (Spolia Zeylanica, vol. 24, 1945, p. 98) suggests that this group may be of pleurodirous origin.

LEPIDOCHELYS KEMPII (Garman)


Type.—Several cotypes, Mus. Comp. Zool.

Type locality.—Gulf of Mexico, here restricted to Key West, Fla.

Range.—Atlantic Ocean and Gulf of Mexico. In Mexico, the entire Atlantic coast. The only exact record known is from Quintana Roo (Isla de Mujeres).

LEPIDOCHELYS OLIVACEA (Eschscholtz)


Type.—Unknown.

Type locality.—Manila Bay, Philippine Islands.

Range.—Indian and Pacific Oceans. In Mexico, the entire Pacific coast. Reported only from the states of Chiapas, Oaxaca, Guerrero, Colima, and Sonora (Tiburón Island).

Genus CARETTA Rafinesque


Caouana Cocteau, in Ramón de la Sagra, Historia física, política y natural de la Isla de Cuba, vol. 4, Rept., 1838, p. 31 (type, Testudo cephalo Schneider).

Eremonia Gray, Hand list of shield reptiles of the British Museum, 1873, p. 91 (type, Caouana elongata Gray).

Genotype.—Caretta nasuta Rafinesque=Caretta caretta caretta (Linnaeus).

Species.—One, with two races, is generally recognized.

Range.—Tropical ocean, world-wide.

KEY TO SPECIES OF CARETTA

1. Neural bones usually 9 or more; if fewer, the costal bones usually interrupting the neural series by contact with each other at one or more places.

   caretta gigas (p. 16)

   Neural bones 7 or 8, the series rarely interrupted by median contact of costal bones

   carettar caretta (p. 16)
CARETTA CARETTA CARETTA (Linnaeus)

Testudo caretta Linnaeus, Systema naturae, ed. 10, vol. 1, 1756, p. 197.


Testudo cephalo Schneider, Algemeine Naturgeschichte der Schildkröten . . ., 1783, pp. 303-308 (type unknown; type locality, Charleston, S. C., by present restriction).


Type.—Unknown.

Type locality.—“About the American Islands,” here restricted to the Bermuda Islands.

Range.—Western Atlantic Ocean; the entire Atlantic coast of Mexico. Reported in Mexico only from Yucatán.

CARETTA CARETTA GIGAS Deraniyagala


Type.—Presumably in Colombo Museum, Ceylon.

Type locality.—Ceylon.

Range.—Indian and Pacific Oceans and possibly eastern Atlantic Ocean. In Mexico, the entire Pacific coast. Reported only from Sinaloa and Baja California.

Genus ERETMOCHELYS Fitzinger

Eretmochelys Fitzinger, Systema reptilium, fasc. 1, 1843, p. 30.

Genotype.—Testudo imbricata Linnaeus.

Species.—One is generally recognized, but two species (or subspecies) are distinguished by some authorities.

ERETMOCHELYS IMBRICATA ¹ (Linnaeus)

Testudo imbricata Linnaeus, Systema naturae, ed. 12, 1766, p. 350.

Eretmochelys imbricata, Fitzinger, Systema reptilium, fasc. 1, 1843, p. 30.—

Chelonia lachrymata Cuvier, Le règne animal, ed. 2, vol. 2, 1829, p. 13 (no type or type locality designated; Bermuda Islands by present restriction).


Chelonia pseudo-carettta Lesson, op. cit., p. 302 (type and type locality as in the preceding).

¹ The Atlantic Ocean specimens are frequently regarded as Eretmochelys imbricata (or E. i. imbricata), the Pacific Ocean ones as Eretmochelys imbricata bissa (or E. i. squamata). We are unable to determine characters for such separation.
Caretta bissa Rüppell, Neue Wirbelthiere Abyssiniens, Amphibiens, 1835, p. 4, pl. 2 (type unknown; type locality, Red Sea).


Type.—Unknown.

Type locality.—American seas, here restricted to the Bermuda Islands.

Range.—Tropical oceans, world-wide; both coasts of Mexico. Reported from Baja California, Sonora, Oaxaca, Campeche, and Quintana Roo (Isla de Mujeres).

Genus CHELONIA Brongniart

Mydas Cocteau, in Ramón de la Sagra, Historia física, política y natural de la Isla de Cuba, vol. 4, 1838, p. 22.
Euchelonia Tschudi, Untersuchungen über die Fauna Peruana . . ., 1846, p. 22.
Megemys Gistel, Naturgeschichte des Thierreichs, 1848, p. 8 (type of all above, Testudo mydas Linnaeus).

Genotype.—Testudo mydas Linnaeus.

Species.—Possibly four forms (species or subspecies) are recognizable. None of these are well enough known to us to diagnose properly.

Range.—Tropical oceans, world-wide.

CHELONIA MYDAS (Linnaeus)

Testudo mydas Linnaeus, Systema naturae, ed. 10, 1758, p. 197.
Testudo viridis Schneider, Algemeine Naturgeschichte der Schildkröten . . ., 1782, pp. 299–303 (type unknown; type locality, Charleston, S. C., by present restriction).
Chelonia virgata Schweigger, Prodromi monographiæ cheloniorum . . ., 1814, p. 21 (type unknown; type locality, “Seas of Torrid Zone,” here restricted to the Bermuda Islands).

*A common arrangement restricts mydas to the Atlantic Ocean, agassizii to the Pacific, either as species or subspecies. We are unable to determine characters justifying such an arrangement.
Type.—Unknown.
Type locality.—Ascension Island.
Range.—Tropical oceans, world-wide, both coasts of Mexico. Reported from Veracruz, Yucatán, Quintana Roo (Isla de Mujeres), Oaxaca, Guerrero, Sonora (Tiburón Island), and Clarion Island.

Suborder TRIONYCHOIDEA Fitzinger
Trionychoidea Fitzinger, Neue Classification der Reptilien . . ., 1826, p. 7.

Families.—Two, one of which (Carettochelyidae) is restricted to New Guinea.

Family TRIONYCHIDAE Gray

Genera.—Seven, only one of which occurs in the Americas.
Range.—Asia, Africa, North America.

Genus AMYDA Schweigger
Genotype.—Amyda javanica Geoffroy [=Amyda cartilaginea (Bood- daert)].
Species.—About 21, of which 4 are American; two or three subspecies are recognized of one species (A. spinifera). One occurs in Mexico.10
Range.—Asia, North America.

AMYDA EMORYI (Agassiz)
Type locality.—Rio Grande River, near Brownsville, Tex.
Range.—Southern Oklahoma and Arizona southward into northern Mexico. Recorded from Coahuila: Hacienda Los Borregos (near Juárez), San Juan, Cuatro Ciéncas, Hacienda La Gacha; Nuevo León: Rodriguez; Tamaulipas: Matamoros, Río Purificación north of Ciudad Victoria; Baja California?.

Suborder CRYPTODIRA Cope

Families.—Six, all but one of which (Platysternidae, southern Asia) occur in Mexico.

CHECKLIST OF REPTILES OF MEXICO

KEY TO MEXICAN FAMILIES OF CRYPTODIRA

1. Abdominal scutes in contact with marginals or separated from them only by membrane; inguinal scute short, less than half length of bridge; 12 plastral scutes

All plastral scutes separated from marginals by a series of inframarginals, or inguinal scute very long, half length of bridge or longer; 12 or fewer plastral scutes

2. Four or 5 inframarginals

Two or 3 (rarely 1) inframarginals

3. Bridge very narrow, covered chiefly by the displaced abdominal scutes, which are widely separated from each other medially

Bridge broad, abdominal scutes in normal position

4. Digits with no webs whatever; rear foot stump-shaped, plantar surface as broad as long

Digits with at least some vestige of webs; rear foot more elongate, plantar surface longer than broad

Family DERMATEMYIDAE Gray

Dermatemyidae Gray, Suppl. to the catalogue of the shield reptiles . . , 1870, p. 49.

Genera.—One.

Range.—Central Mexico to Honduras.

Genus DERMATEMYS Gray


Genotype.—Dermatemys mawii Gray.

Range.—Atlantic coast from central Veracruz to Honduras.

Species.—One recognized.

DERMATEMYS MAWII Gray


Emys Berardi Duméril and Bibron, in Duméril and Duméril, Catalogue méthodique de la collection des reptiles, livr. 1, 1851, p. 11 (type locality, environs of Veracruz, Mexico; type in Mus. Hist. Nat. Paris).


Limnochelone micrura Werner, Zool. Anz., vol. 24, 1901, p. 298 (type locality, Mexico, here restricted to Alvarado, Veracruz; type unknown).

Type.—Brit. Mus.

Type locality.—Unknown, here restricted to Alvarado, Veracruz.

Range.—Rivers of the Atlantic coast from central Veracruz to Guatemala, probably excluding the northern part of the Yucatán Peninsula. Recorded from the states of Veracruz, Oaxaca, Tabasco, Yucatán, and Campeche.
Family CHELYDRIDAE Swainson


Genera.—Two, one of which (Macrochelys) is restricted to the United States. A supposed New Guinea genus, Devisia, is an erroneously labeled Chelydra serpentina.

Range.—North America south to northern South America.

Genus CHELYDRA Schweigger


Chelonura Fleming, Philosophy of zoology . . . , vol. 2, 1870, p. 64 (type, Testudo serpentina Linnaeus).


Saurochelys Latreille, Familles naturelles du règne animal . . . , 1825, p. 92 (type as above).

Chelidura Schweigger, Atlantic Journ., 1832, p. 64 (type as above).


Chelonura Holbrook, North American herpetology, vol. 1, 1836, p. 139 (type as above).

Genotype.—Testudo serpentina Linnaeus.

Range.—Southeastern Canada, United States east of the Rocky Mountains, south to Ecuador.

Species.—Three species or subspecies; one is definitely recorded from Mexico, another probably occurs there, and the third (acutirostris) is restricted to Panama and South America.

KEY TO MEXICAN SPECIES OF CHELYDRA

1. Bridge about one-seventh length of plastron; 4 chin barbels; second vertebral shield 32 to 34 percent length of carapace; height of skull at quadrate 44 to 48 percent length of skull to condyle; width of palatine bone 28 to 30 percent length of skull------------------------------rossignonii (p. 20)

Bridge about one-ninth length of plastron; 2 chin barbels; second vertebral shield less than 30 percent length of carapace; height of skull at quadrate 38 to 43 percent length of skull to condyle; width of palatine bone 19 to 25 percent length of skull----------------------------serpentina (p. 21)

CHELYDRA ROSSIGNONII (Bocourt)


Type locality.—Panzós, Río Polochic, Guatemala.

Range.—Atlantic slopes of Guatemala and presumably adjacent Mexico southward to Costa Rica. Not yet reported definitely from Mexico.
CHECKLIST OF REPTILES OF MEXICO

CHELYDRA SERPENTINA (Linnaeus)


Type.—Unknown.
Type locality.—“Warmer region,” here restricted to New Orleans, La.

Range.—North America east of the Rocky Mountains; in Mexico, Atlantic slopes south to the Yucatán Peninsula; recorded only from the states of Veracruz and Campeche.

Family KINOSTERNIDAE Agassiz


Genera.—Three.
Range.—North America, south to northern South America.

KEY TO GENERA OF KINOSTERNIDAE

1. Plastron very small, cruciform, with 7–9 scutes; bridge narrow, its length (parallel to body axis) exceeded by its width; inframarginals little longer than broad

Plastron larger, not cruciform, with 10 or 11 scutes; bridge broader, its length much exceeding its width; inframarginals about three times as long as broad, or longer

..........................Kinosternon (p. 21)

2. A ligament between inframarginals and plastron

A suture between inframarginals and plastron

..........................Claudius (p. 26)

..........................Staurotypus (p. 27)

Genus KINOSTERNON Spix

Kinosternon Spix, Ranae et testudinis brasiliensis species novae, 1825, p. 17.
Thyrosterum Agassiz, op. cit., p. 429 (type, Kinosternum integrum LeConte).

Genotype.—Kinosternon longicaudatum Spix=K. scorpioides Linnaeus.

Range.—United States, except the northwestern quarter, south to northern South America.

Species.—About 22 species and subspecies, 12 of which are known to occur in Mexico.

KEY TO MEXICAN SPECIES OF KINOSTERNON

1. Plastron rounded behind, not incised or indented; stridulation organs (on concealed surfaces of shank and thigh) in males poorly developed or absent; plastron large, completely closing shell in adults

..........................2
Plastron incised or indented behind; stridulation organs in males well developed or not; plastron at least somewhat smaller, not completely closing shell even in adults. 7
2. Carapace tricarinate; axillary and inguinal scutes (if present) widely separated. 3
Carapace flat or unicarinate; axillary and inguinal scutes narrowly in contact or narrowly separated. 5
3. Axillary scute usually absent; posterior edge of abdominals convex. abaxillare (p. 22)
Axillary scute present; posterior edge of abdominals straight. 4
4. Anterior lobe of plastron longer than immovable portion of plastron. cruentatum cruentatum (p. 23)
Anterior lobe of plastron as long as or shorter than immovable portion of plastron. cruentatum consors (p. 24)
5. Anterior lobe of plastron longer than fixed portion. 6
Anterior lobe of plastron not longer than fixed portion. acutum (p. 23)
6. Gular more than half length of anterior lobe of plastron. creaseri (p. 23)
Gular less than half length of anterior lobe of plastron. leucostomum (p. 25)
7. Carapace tricarinate; stridulation organs absent; plastron slightly smaller than opening of carapace. integrum (p. 25)
Carapace flat or unicarinate; stridulation organs well developed in males; plastron distinctly smaller than opening of carapace. 8
8. Ninth marginal about as high as tenth, much higher than eighth. 9
Ninth marginal much lower than tenth, about height of eighth. 10
9. Gular more than half length of anterior lobe of plastron (60–64 percent); length of anterior lobe less than combined length of gular plus length of interfemoral suture plus width of nuchal (83–93 percent). flavescens stejnegeri (p. 24)
Gular half or less than half length of anterior lobe of plastron (30–50 percent); length of anterior lobe greater than combined length of gular plus length of interfemoral suture plus width of nuchal (128–196 percent). flavescens flavescens (p. 24)
10. Anterior vertebral very narrow, widely separated from second marginal on each side; gular less than half length of anterior lobe; posterior marginal (eleventh) little lower than penultimate (tenth). herrerae (p. 24)
Anterior vertebral broad, in contact with second marginal on each side; gular variable in length; posterior marginal distinctly lower than penultimate. 11
11. Carapace depressed; entire shell twice as broad as deep; head light, dark-spotted; sutures between scutes of carapace not distinctly black-bordered, those between plastral scutes not or narrowly marked with brown. sonoriense (p. 26)
Carapace peaked, arched as viewed directly from front; shell less than twice as broad as deep; head dark, light-spotted, sutures on carapace narrowly but distinctly black-edged, those on plastron broadly marked with brown. hirtipes (p. 25)

**KINOSTERNON ABAXILLARE** Baur


*Type.*—U.S.N.M. No. 7518; C. H. Berendt collector.

*Type locality.*—Tuxtla Gutiérrez, Chiapas, Mexico.

*Range.*—Plateau of Chiapas. Recorded only from the type locality.
KINOSTERNON ACUTUM Gray


Kinosternon Effeldtii Peters, Monatsb. Berlin Akad. Wiss., 1873, p. 603, pl. 5, figs. 1-3 (type locality, "presumably" Veracruz, Mexico, here restricted to Cosamaloapam; type Berl. Mus.).

Type.—Brit. Mus.

Type locality.—Unknown, here restricted to Cosamaloapam, Veracruz.

Range.—Atlantic coast from central Veracruz southward to British Honduras, excluding the northern part of the Yucatán Peninsula. Recorded from the states of Veracruz, Tabasco, and Campeche.11

KINOSTERNON CREASERI Hartweg


Type locality.—One mile south of the Hacienda, Chichen Itzá, Yucatán.

Range.—Known only from the general region of the type locality, and Quintana Roo (Vivienda de Platanal).

KINOSTERNON CRUENTATUM CRUENTATUM Duméril and Bibron


Type locality.—"Amér. septentr.," here restricted to San Mateo del Mar, Oaxaca.

Range.—Atlantic and Pacific drainages in Oaxaca, Tabasco, Campeche, and Chiapas. In Central America, Guatemala.

**KINOSTERNON CRUENTATUM CONSORS Stejneger**


*Type.*—U.S.N.M. No. 13912.

*Type locality.*—Cozumel Island, Quintana Roo.

*Range.*—The type locality and northern Yucatán (Progreso and Telchac Puerto).

**KINOSTERNON FLAVESCENS FLAVESCENS (Agassiz)**


*Type.*—Unknown.

*Type locality.*—Texas and Arizona, here restricted to Waco, Tex.

*Range.*—Northern Mexico from Coahuila to Tamaulipas; recorded in Coahuila (Jaral). In the United States from Arizona and Texas north to Colorado and Kansas.

**KINOSTERNON FLAVESCENS STEJNEGERI Hartweg**


*Type locality.*—Llano, Sonora (midway between Nogales and Hermosillo).

*Range.*—Known only from Sonora (type locality) and Durango (Pedriceña); perhaps also from southern Coahuila (said to intergrade with *flavescens flavescens* at Jaral, Coahuila).

**KINOSTERNON HERRERAI Stejneger**


*Type.*—U.S.N.M. No. 61249; Alfonso Herrera, donor.

*Type locality.*—Xochimilco, Valley of Mexico, Distrito Federal, Mexico, in errore; here restricted to La Laja, Veracruz.

*Range.*—Known only from the type locality.

---

13 A record from La Majada, near Apatzingán, Michoacán (Schmidt and Shannon, Fieldiana, vol. 31, 1947, p. 69), is referable to *K. integrum*. 
**KINOSTERNON HIRTIPE Wagler**

*Cinosternon hirtipes Wagler*, Natürliches System der Amphibien, . . ., 1830, pl. 5, figs. 29, 30.


*Type.*—Munich Museum.

*Type locality.*—Mexico, here restricted to Mazatlán, Sinaloa.

*Range.*—Western Texas and southern Arizona southward through the main Mexican Plateau, from Chihuahua to México; recorded from the states of Chihuahua, Sinaloa, Michoacán, Colima, Guanajuato, San Luis Potosí, Hidalgo, México, and from Distrito Federal and the Tres Marías Islands.

**KINOSTERNON INTEGRUM LeConte**


*Cinosternum integrum*, BoulenGer, Catalogue of the chelonians, rhynchocephalians and crocodiles in the British Museum, 1889, p. 42.


*Type.*—Acad. Nat. Sci. Philadelphia; Mr. Pease collector.

*Type locality.*—Mexico, here restricted to Acapulco, Guerrero.

*Range.*—The plateau of Mexico from Sonora to Oaxaca east to Veracruz. Known from Tres Marías Islands and the states of Sonora, Sinaloa, Nayarit, Colima, Michoacán, Guerrero, Oaxaca, Morelos, Guanajuato, Aguascalientes, Jalisco, San Luis Potosí, Puebla, and Veracruz.

**KINOSTERNON LEUCOSTOMUM Duméril and Bibron**


*Cinosternum brevigulare* Günther, Biologia Centrali-Americana, Reptilia and Batrachia, 1885, pp. 17–18 (type locality, Playa Vicente, Mexico, Sallé collector; type in Brit. Mus. Nat. Hist.).


*Type locality.*—“N. Orléans; Mexique; Rio-Sumasinta (Amér.

Thyrosterum sonoriense, Agassiz, Contributions to the natural history of the United States, vol. 1, 1857, p. 428; vol. 2, pl. 5, figs. 8-11 (Kinosternum sonoriense LeConte).


? Cinosternon punctatum Gray, Catalogue of the shield reptiles in the British Museum, 1855, p. 45, pl. 20c, figs. 5, 6 (type in Brit. Mus.; "North America," here restricted to Tucson, Ariz.).


Type locality.—Tucson, Ariz.

Range.—Sonora, Chihuahua, and Durango (Durango); in the United States: southeastern California, southern Arizona, New Mexico to western Texas.

Genus CLAUDIUS Cope


Genotype.—Claudius angustatus Cope.

Range.—Restricted to the Atlantic coast from Veracruz to British Honduras.

Species.—One.

CLAUDIUS ANGUSTATUS Cope


Type.—U.S.N.M. Nos. 6518, 6525.

Type locality.—Tabasco.

Range.—Veracruz southward to British Honduras. Recorded in Mexico from the states of Veracruz, Tabasco, and Campeche.
Genus STAUROTYPUS Wagler


*Genotype.*—*Terrapene triporcata* Wiegmann.

*Range.*—Atlantic slopes of southern Mexico south to Guatemala.

*Species.*—Two.

**KEY TO SPECIES OF STAUROTYPUS**

1. Suture between humerals longest; abdominals much broader than long; length of bridge 5½ or more times in length of plastron.---------*salvinii* (p. 27)

Suture between pectorals longest; abdominals as broad as long; length of bridge less than 5½ times in length of plastron.---------*triporcatu*s (p. 27)

**STAUROTYPUS TRIPORCATU*S (Wiegmann)


*Staurotypus triporcatu*s, Wagler, *Natürliches System der Amphibien* . . ., 1830, pl. 5, figs. 44-45.

*Staurotypus salvinii*, Bocourt (nee Gray), *Mission scientifique au Mexique* . . ., *Études sur les reptiles*, livr. 1, 1870, pp. 22-23, pl. 5, fig. 3.


*Type.*—Presumably in the Berlin Mus.; Deppe collector.

*Type locality.*—Rio Alvarado, Veracruz.

*Range.*—Atlantic slopes from Veracruz to Guatemala and British Honduras (recorded in Mexico only from the states of Veracruz and Tabasco).

**STAUROTYPUS SALVINII* Gray


*Staurotypus marmoratus* Fischer, *Archiv für Naturg.*, 1872, p. 265, pl. 10 (type locality, "Mexico," here restricted to Santa Efégenia, Oaxaca; St. Petersburg Mus.).


*Staurotypus biporcatu*s Siebenrock, *Zool. Jahrb.*, Suppl. 10, No. 3, 1909, p. 438 (type locality restricted here to that of *S. salvinii* Gray; type here restricted, as lectotype, to that of *S. salvinii* Gray).

*Type.*—Brit. Mus.

*Type locality.*—Huamuchil, Guatemala.

*Range.*—Pacific slopes from the Isthmus of Tehuantepec to Guatemala (in Mexico recorded only from Oaxaca).

**Family TESTUDINIDAE** Gray


861316—50—3
Genera.—About four, of which one occurs in Mexico.

Range.—World-wide in tropical and subtropical areas, except the East Indies and Australia.

Genus Gopherus Rafinesque

Gopherus Rafinesque, Atlantic Journ., vol. 1, 1832, p. 64.
Genotype.—Testudo polyphemus Daudin.
Range.—Southern United States and northern Mexico.
Species.—Three recognized, two of which occur in Mexico.

KEY TO MEXICAN SPECIES OF Gopherus

1. Carapace dome-shaped, rounded on top; carapace length about twice shell height (47–56 percent) — berlandieri (p. 28)
   Carapace with flat-topped contour; carapace length more than twice shell height (36–48 percent) — agassizii (p. 28)

Gopherus berlandieri (Agassiz)

Testudo berlandieri, Boulenger, Catalogue of the chelonians, rhynchocephalians and crocodiles in the British Museum, 1889, p. 156.

Type.—U.S.N.M. No. 60, two specimens.
Type locality.—Lower Rio Grande, Tex., here restricted to Brownsville.

Range.—Nuevo León, Tamaulipas, and Coahuila; in United States: Southern Texas.

Gopherus agassizii (Cooper)

Testudo agassizii, Boulenger, Catalogue of the chelonians, rhynchocephalians and crocodiles in the British Museum, 1889, p. 156.

Type.—Unknown.
Type locality.—Mountains near Fort Mojave, Calif.

Range.—Baja California, Sonora including Tiburón Island; in the United States, southeastern California and southwestern Arizona north into Nevada and Utah.

Family Emyidae

Genera.—About 25, of which 6 occur in Mexico.
Range.—World-wide except Australia and East Indies.

KEY TO MEXICAN GENERA OF EMYIDAE

1. Plastron hinged, movable; no inframarginals. .................................. Terrapene (p. 31)
   Plastron fixed, immovable; inframarginals present .................................. 2

2. Alveolar surface of upper jaw with a ridge either smooth or toothed, parallel to labial cutting edge ................................................................. 5
   Alveolar surface of upper jaw ridgeless, smooth ........................................ 3

3. A strong interrupted median keel on carapace, with a marked tubercle at the end of each vertebral; a low dorsal crest of enlarged compressed scales on tail; bridge much shorter than posterior lobe of plastron.
   Malaclemys (p. 36)
   Dorsal keel on carapace absent or continuous, not tuberculate; no crest of scales on tail ................................................................. 4

4. Bridge much shorter than posterior lobe of plastron. .................. Clemmys (p. 37)
   Bridge longer than posterior lobe of plastron .................................. Geoemyda (p. 29)

5. Rear of carapace perfectly smoothly rounded, edges of all plates even, no notches except a median rear one; a terminal notch flanked on either side by a projection on upper jaw, and cutting edge of lower jaw smooth.
   Chrysemys (p. 33)
   Rear of carapace at least somewhat serrate; no terminal notch flanked on either side by a projection in upper jaw, or if so cutting edge of lower jaw serrate .................................. Pseudemys (p. 31)

Genus GEOEMYDA Gray


Genotype.—Testudo spengleri Gmelin.
Range.—India to Japan and the Malay Archipelago; the coasts of central Mexico south into South America.
Species.—Nineteen species and subspecies are known, nine in the Americas, four in Mexico.

KEY TO MEXICAN SPECIES OF GEOEMYDA

1. Bridge yellow ........................................................................ areolata (p. 30)
   Bridge black, uniform or with a few yellow markings ......................... 2

2. Upper jaw hooked or smooth; plastron dark with a yellow margin. rubida (p. 30)
   Upper jaw notched; plastron yellow, with a broad dark median stripe .. 3

3. Carapace almost uniform brown, without distinct markings, its entire border strongly bowed, nowhere straight; sides of head in front of eye with usually 2, sometimes 3, bright horizontal lines .................................. pulcherrima incisa (p. 30)
   Carapace olive-brown, with bright, yellow-red concentric or hieroglyphic pattern of lines on each scale; young with dark, light-centered spots on costals; only lateral border of carapace strongly bowed; sides of head in front of eye always with 3 lines ........ pulcherrima pulcherrima (p. 30)
Geoemyda areolata (Duméril and Bibron)

*Emys areolata* Duméril and Bibron, in Duméril and Duméril, Catalogue méthodique de la collection des reptiles . . ., 1851, p. 10.


**Type locality.**—El Petén, Guatemala, here restricted to La Libertad.

**Range.**—Coast of central Veracruz south to northern Guatemala and British Honduras. Recorded in Mexico from Veracruz, Tabasco, Campeche, Yucatán, and Quintana Roo (Cobá, Cozumel Island).

Geoemyda pulcherrima pulcherrima (Gray)

*Emys pulcherrima* Gray, Catalogue of the shield reptiles in the British Museum, vol. 1, 1855, p. 25, pl. 25, fig. 1.—Bocourt, Mission scientifique au Mexique, Études sur les reptiles, livr. 1, 1870, pp. 15-16, pl. 7, figs. 1, 1a, 1b (color).


**Type locality.**—Mexico, here restricted to Presidio de Mazatlán, Sinaloa.

**Range.**—Pacific slopes from southern Sonora to Guerrero and probably western Oaxaca. Recorded from the states of Sonora, Sinaloa, Nayarit, Colima, and Guerrero.

Geoemyda pulcherrima incisa (Bocourt)


**Type locality.**—La Unión, El Salvador.

**Range.**—Pacific slopes of the Isthmus of Tehuantepec southward to at least El Salvador. Recorded from Oaxaca and Chiapas.

Geoemyda rubida (Cope)


**Type.**—Unknown.

**Type locality.**—Juchitán, Oaxaca.

**Range.**—The Pacific coast from Colima to the Isthmus of Tehuantepec (recorded from Colima, Michoacán, Guerrero, and Oaxaca

---

18 The exact location of San Juan del Río (type locality of *Rhinoclemmys mexicana*) is uncertain; it probably is one of the two cities of that name in the state of Oaxaca.
CHECKLIST OF REPTILES OF MEXICO

Genus PSEUDEMYS Gray


Type.—Testudo concinna LeConte.

Species.—About 8, and a total of about 22 species and subspecies, of which 8 forms occur in Mexico.

Range.—The United States east of the Rocky Mountains, Baja California, both coasts of Mexico from the Río Grande on the east and southern Sonora on the west southward to Argentina.

KEY TO MEXICAN SPECIES OF PSEUDEMYS

1. Ridge on alveolar surface of upper jaw distinctly toothed; temporal stripe absent or not expanded----------------------------------------------- 2
   Ridge on alveolar surface of upper jaw not toothed; temporal stripe or spot much expanded-------------------------------------------- 3

2. A strong notch at apex of upper jaw, flanked on each side by a toothlike projection; teeth on alveolar ridge long, numerous; nostrils terminal; snout not pointed, obtuse.............................................floridana texana (p. 33)
   No notch at apex of upper jaw, and no subterminal toothlike projections; teeth on alveolar ridge smaller, less numerous; nostrils pierced below and posterior to tip of snout; snout pointed, acuminate............................................................umbra (p. 33)

3. A light, round temporal spot, completely isolated from orbital markings— 4
   Supratemporal light spot elongate, reaching orbit.................................................. 5

4. Temporal light spot completely isolated, not continuous with lines either anteriorly or posteriorly; bridge with longitudinal black stripes.
   scripta gaigeae (p. 33)
   Temporal light spot continuous toward rear with longitudinal light stripe; bridge black spotted......................................................... scripta nebulosa (p. 32)

5. Markings solid, intensely black, replacing or obscuring a linear juvenile pattern, and including a broad area along middle of plastron, lengthwise along middle of bridge, and on posterolateral corners of the marginal scutes above and below; these markings evident in bony plates as well as in epidermal scutes.................................................................scripta hiltoni (p. 32)
   Markings not as described.............................................................................................................. 6

6. Plastral markings always present, either smudged, or, if in lines, the lines forming ocelli; supratemporal stripe, if visible, reaching eye.
   scripta elegans (p. 32)
   Plastral markings, if visible, not smudged, but in lines that tend to be arranged longitudinally, not ocellate; supratemporal stripe reaching eye or not—— 7

7. Dark ocellus of each costal scute nearest latter's posteroverentral corner.
   scripta cataspila (p. 32)
   Dark ocelli nearest posterodorsal corners of costals............scripta ornata (p. 31)

PSEUDEMYS SCRIPTA ORNATA (Gray)

Emys ornata Gray, Synopsis reptilium, 1831, p. 30.—BOCOURT, Mission scientifique au Mexique ..., Études sur les reptiles, livr. 1, 1870, pl. 3, fig. 1, la.


Emys venusta Gray, Catalogue of the shield reptiles ..., vol. 1, 1855, p. 24, pl. 24a (type locality, Mexico, Honduras and America, here restricted to Honduras; types in Brit. Mus.).

Type.—Brit. Mus., two cotypes.
Type locality.—Mazatlán, Sinaloa.

Range.—Pacific slopes of Mexico southward from southern Sinaloa, and both Atlantic and Pacific slopes (presumably) of Central America to Panama and possibly northern South America; Atlantic slopes of Mexico east of the Isthmus of Tehuantepec (recorded in Mexico from the states of Sinaloa, Jalisco, Oaxaca, Chiapas, Tabasco, Campeche, and Yucatán, and from Cozumel Island).

**Pseudemys scripta nebulosa** (Van Denburgh)


Type.—Destroyed.

Type locality.—Los Dolores, Baja California, Mexico.

Range.—The southern half of Baja California.

**Pseudemys scripta hiltoni** Carr

**Pseudemys scripta hiltoni** Carr, Amer. Mus. Nov., No. 1181, 1942, pp. 1–4, fig. 1–3.


Type locality.—Guirocoba, 28 miles southeast of Álamos, Sonora, Mexico.

Range.—Known only from the type locality; presumably occurs in northern Sinaloa as well as in Sonora.

**Pseudemys scripta cataspila** (Günther)

*Emys cataspila* Günther, Biologia Centrali-Americana, Reptilia and Batrachia, 1885, p. 4, pl. 6, fig. B.


Type.—Brit. Mus., seven cotypes.

Type locality.—Mexico, here restricted to Alvarado, Veracruz.

Range.—Atlantic slopes of Mexico from the northern border to the Isthmus of Tehuantepec (recorded from the states of Tamaulipas and Veracruz).

**Pseudemys scripta elegans** (Wied)


Type.—Unknown.
Type locality.—Fox River at New Harmony, Ind.
Range.—Northeastern Mexico: Coahuila, Nuevo León, and Tamaulipas. Widely distributed in central and southern United States.

**PSEUDEMYS SCRIPTA GAIGEA** Hartweg


_Type locality._—Rio Grande River, Boquillas, Brewster County, Tex.
_Range._—Northern central Mexico (recorded from Coahuila and Durango).

**PSEUDEMYS UMBRA** (Bocourt)


*Emys umbra* Bocourt, _in O’Shaughnessy_, Zool. Rec., vol. 13, 1876 (1878), Rept., p. 6 (substitute name for *Emys grayi* Bocourt, preoccupied by *Emys grayi* Günther = *Clemmys caspica* caspica [Gmelin]).—Günther, _Biologia Centrali-Americana_, Reptilia and Batrachia, 1885, pp. x, 5, pls. 4, 5, 6 (fig. A).

_Type locality._—Río Nagualate, Guatemala.
_Range._—Pacific slopes of the Isthmus of Tehuantepec, southeastward to Guatemala. Recorded definitely only from the state of Oaxaca; other records from Chiapas may refer either to this or to P. s. ornata.

**PSEUDEMYS FLORIDANA TEXANA** Baur


*Pseudemys floridana texana,* Carr, Copeia, 1938, p. 108.

_Type locality._—San Antonio, Tex.
_Range._—Central Texas and northeastern Mexico. Recorded only from Nuevo León.

**Genus CHRYSEMYS** Gray

*Chrysemys* Gray, Catalogue of the tortoises . . ., 1844, p. 27.

_Genotype._—*Testudo picta* Schneider.
_Range._—Eastern three-fourths of the United States, extreme northern Mexico.
_Species._—One, with four subspecies. One race occurs in Mexico.

17 Schmidt and Owens (Publ. Field Mus. Nat. Hist., zool. ser., vol. 29, 1944, p. 101) regard this as a species distinct from that to which *elegans* belongs, but the distinguishing criterion mentioned appears to be one that in reality separates males from females of either form. We have not examined the specimens but regard all *Pseudemys* reported by Schmidt and Owens as *P. s. elegans.*
CHRYSEMYS PICTA BELLII (Gray)

Emys bellii Gray, Synopsis reptilium . . . , 1831, p. 31.  


Type.—Lost; originally in Brit. Mus.  
Type locality.—Unknown, here restricted to Manhattan, Kans.  
Range.—Western Illinois to Washington and British Columbia, south on the east of the Cascade Mountains to northern Chihuahua and southern Texas. Recorded only from the state of Chihuahua: Río Santa María, near Progreso.

Genus TERRAPENE Merrem


Type.—Testudo clausa Gmelin=Terrapene carolina (Linnaeus).  
Range.—North America east of the Rocky Mountains and Arizona, south to Yucatán and Nayarit.  
Species.—Eleven species and subspecies are recognized at present, six in Mexico.

KEY TO MEXICAN SPECIES OF TERRAPENE

1. Plastron with a conspicuous and well-defined pattern of yellow lines on a dark background (or dark lines on a yellow background).----------ornata (p. 36)  
   Plastron without a lined pattern, either uniform or suffused here and there with dark color.------------------------------------------ 2

2. Pattern of carapace consisting of small, round or ovoid yellow dots 1.5-2 mm. in diameter, separated from each other by an average distance of 4 mm.  
   klauberi (p. 35)

Pattern not so.------------------------------------------ 3

3. Width of carapace 65-68 percent length of carapace, average 66 percent; height of shell 40-45 percent length of carapace; 4 rear claws; carapace horn-colored, without dark borders at sutures.----------coahuila (p. 36)  
   Width of carapace 68-79 percent length of carapace, average about 73 percent; height of shell 45-58 percent length of carapace, average 51 percent; rear claws 3 or 4; carapace either marked with yellow or horn-colored, but sutures always broadly margined with dark color.------------------------------------------ 4

4. No notch at apex of upper jaws; 4 rear claws.----------nelsoni (p. 35)  
   Apex of upper jaw notched; rear claws 3 or 4.------------------------------------------ 5

5. Rear claws always (so far as known).----------mexicana yucatana (p. 35)  
   Rear claws generally 3 (81 percent of 32 specimens).  
   mexicana mexicana (p. 34)

TERRAPENE MEXICANA MEXICANA (Gray)

Cistudo carolina var. mexicana, Bouleneger, Catalogue of the chelonians, rhynchocephalians and crocodiles in the British Museum, 1889, p. 118.


Terrapene yucatana, Ditmars (nee Boulenger), Zoologica, vol. 17, 1934, figs. 29, 30.

Type.—Brit. Mus. Nat. Hist.; two female cotypes; collector unknown.

Type locality.—Mexico, restricted by Müller to Tampico, Tamaulipas.

Range.—Atlantic slopes from central Tamaulipas and eastern Nuevo León to northern Veracruz. Known only from San Luis Potosí, Veracruz, and Tamaulipas.

TERRAPENE MEXICANA YUCATANA (Boulenger)


Type locality.—Northern Yucatán, here restricted to Chichen Itzá.

Range.—Northern part of the peninsula of Yucatán. Recorded only from Yucatán and Quintana Roo.

TERRAPENE NELSONI Stejneger


Type.—U.S.N.M. No. 46252; E. W. Nelson and E. A. Goldman collectors.

Type locality.—Pedro Pablo, Tepic [=Nayarit], Mexico, 2,500 feet altitude.

Range.—Known only from the type locality.

TERRAPENE KLAUBERI Bogert


18 Schmidt and Owens (Publ. Field Mus. Nat. Hist., zool. ser., vol. 29, 1944, p. 103) regard T. goldmani as a distinct species “of lowland San Luis Potost,” implying that its closest relative, T. mexicana, is restricted to other areas, presumably highland Mexico inasmuch as lowland San Luis Potost is scarcely distinguishable faunistically from northern Veracruz, southern Tamaulipas, and eastern Nuevo León. The entire area, as a matter of fact, is small. However, in spite of a few records of occurrence of T. mexicana on the plateau, notably by Duqué (loc. cit.), we do not regard any of them as reliable, and believe the species is restricted to the approximate range here stated. There can, therefore, be no geographical isolation of two populations to consider.
**Type.**—Amer. Mus. Nat. Hist. No. 63751; John W. Hilton collector.

**Type locality.**—Rancho Guirocoba, about 18 miles southeast of Álamos, Sonora.

**Range.**—Known only from the type locality and Sierra de Batuec, 8 kilometers northeast of Matapé, Sonora.

**TERRAPENE COAHUILA** Schmidt and Owens


**Type.**—Chicago Nat. Hist. Mus. No. 41234; Ernest G. Marsh collector.

**Type locality.**—Quatro Citéneas, Coahuila, Mexico.

**Range.**—Known only from the type locality.

**TERRAPENE ORNATA** (Agassiz)


**Type locality.**—The upper Missouri and Iowa, here restricted to Council Bluffs, Iowa.

**Range.**—Widespread in the United States between the Rocky Mountains and the Mississippi River reaching east to Indiana and west to Arizona; extreme northern Mexico. Reported only from Sonora and Chihuahua.

**Genus MALACLEMYS** Gray

*Malaclemys* Gray, Catalogue of the tortoises . . . , 1844, p. 28.

**Genotype.**—*Testudo concentrica* Shaw.

**Range.**—Atlantic and Gulf coasts of the United States and Mexico.

**Species.**—One, of five distinct races, only one of which has been found in Mexico.

**MALACLEMYS TERRAPIN LITTORALIS** Hay


**Type.**—U.S.N.M. No. 33913; W. P. Hay collector.

**Type locality.**—Rockport, Tex.

**Range.**—Coast of Texas southward possibly to the Yucatán Peninsula. Recorded only from “Mexico” (Strauch).
Genus CLEMYS Ritgen


Genotype.—Testudo punctata Schoepff [= Clemmys guttata (Schneider)].

Range.—Southern Europe, northwestern Africa, southern China and Japan, extreme western and the eastern third of the United States.

Species.—Fifteen species and subspecies are recognized, five in North America, one of which enters Mexico.

CLEMMYS MARMORATA PALLIDA Seeliger

Clemmys marmorata pallida Seeliger, Copeia, 1945, pp. 158-159.


Type locality.—Lower Coyote Creek, near Alamitos, Orange County, Calif.

Range.—Central California near the mouth of the Sacramento River south to northwestern Baja California. Recorded only from northern Baja California.

Subclass LEPIDOSAURIA Romer

Lepidosauria Romer, Vertebrate paleontology, ed. 2, 1945, p. 595.

Orders.—Two orders are represented by living members; one (Rhynchocephalia) is restricted to certain islands off North Island, New Zealand, and the other is world-wide in distribution.

Order SQUAMATA Oppel


Suborders.—Three living suborders are recognized: the Sauria, Serpentes, and Amphisbaenia.

Suborder AMPHISBAENIA Gray

Amphisbaenia Gray, Catalogue of the tortoises, crocodilians and amphisbaenians in British Museum, 1844, p. 68 (as an order).

Families.—Two, only one of which occurs in Mexico. When the anatomy of the forms of amphisbaenians is better known, it is probable that several other families will be recognized.

Range.—West Indies, tropical parts of South America north to Arizona, Florida, and Baja California; Spain, Turkey through Africa.

Family BIPEDIDAE Stejneger


Genera.—One. For the present we prefer to refer all Mexican forms to the single genus Bipes despite the differences in digital structure.
Studies on the cranial anatomy may reveal differences not now apparent.

Range.—The southern tip of Baja California, Arizona, the Rio Balsas Valley, and southern Guerrero.

Genus BIPES Latreille


Genotype.—*Bipes canaliculatus* Bonnaterre.

Range.—Baja California, Guerrero, southeastern Arizona.

Species.—Three.

KEY TO SPECIES OF BIPES

1. Digits 5.------------------------------------------------------------- 2
   Digits 3, clawed--------------------------------------------- tridactylus (p. 38)
2. All digits equally developed, all generally with claws; 6 preanal pores.  
   canaliculatus (p. 38)
   One digit smaller and lacking claw; 2 preanal pores.--------biporus (p. 38)

**BIPES TRIDACTYLUS** (Dugès)


Type.—Alfredo Dugès Mus., Guanajuato, Guanajuato, Mexico.

Type locality.—Tecpan de Galeana, Guerrero.

Range.—Known only from type locality (only the type specimen known).

**BIPES BIPORUS** (Cope)


Type.—U.S.N.M. No. 8568, cotypes (?), T. H. Street collector, and No. 12599, 12 specimens, L. Belding collector.

Type locality.—Cape San Lucas, Baja California.

Range.—The cape region of southern Baja California. Reported from La Paz, Cape San Lucas.

**BIPES CANALICULATUS** Bonnaterre


**CHECKLIST OF REPTILES OF MEXICO**


*Lacerta lumbrocoides* Shaw, Naturalists’ Miscellany, vol. 6, 1795, pl. and text No. 212 (type locality, Mexico, here restricted to Mexcala, Guerrero; type originally in British Museum).


*Chamaesaura propus* Schneider, Historia amphibiorm naturalis et literaria, fasc. 2, 1801, p. 211 (here restricted to Mexcala, Guerrero).

_Type._ Not known.
_Type locality._—Mexico, here restricted to Mexcala, Guerrero.
_Range._—Known in the basin of Río Balsas, Guerrero. Reported from Mexcala, Tеваузиапan, and Balsas. 19

**Suborder Sauria Macartney**

Sauria Macartney, in Ross’s translation Cuvier’s Leçons d’anatomie comparée, vol. 1, 1802, table 3.

**Families.**—Nineteen, of which 10 occur in Mexico.

**KEY TO MEXICAN FAMILIES OF SAURIA**

1. Neither forelegs nor hind legs evident externally
   - Both forelegs and hind legs visible externally
   - 2

2. No eye opening; eye scarcely or not visible below lateral head scales; one unpaired median scale between frontal and rostral. **Anelytropsidae** (p. 170)
   - Eye opening present, although small; a pair of scales on dorsal surface of head between frontal and rostral
     - **Anniellidae** (p. 208)
   - 3

3. No movable eyelids; eyes permanently open
   - Eyelids present, movable
   - 4

4. All scales around middle of body uniform, flat, cycloid, smooth, in about 13 rows
   - **Teiidae** 20 (p. 170)
   - Not so
   - 5

5. Head covered with large, flat plates; ventral scales large, quadrangular, abruptly differentiated from granular lateral scales. **Xantusiidae** (p. 151)
   - Dorsal surface of head covered with granular scales; ventral scales smaller, rounded, not abruptly differentiated from granular lateral scales
   - **Geckkonidae** (p. 40)

6. All scales around middle of body perfectly smooth (sometimes finely striated), cycloid, more or less equal in size (those in the middorsal row or pair or rows may be widened)
   - Not so
   - 7

7. Only one scale in contact with frontal anteriorly; five supraoculars.
   - **Anguidae** 21 (p. 194)

---

20 *Gymnophtalmus* only.
21 *Diploglossus* only.
Two or three scales in contact with frontal anteriorly; three or four supraco-
oeulars.........................................................Scincidae (p. 155)
8. A granular fold along sides of body, abruptly differentiated from much
enlarged dorsal and ventral scales...............................................Anguidae (p. 194)
Not so.................................................................................. 9
9. Ventral scales large, quadrangular, in 8 longitudinal series; dorsals granular.
Tetiidae (p. 170)
Ventral scales smaller, in more numerous rows.................................. 10
10. Ventral scales quadrangular; scales on top of head small, more or less uniform,
tubercular; dorsal scales granular with numerous enlarged tubercles..... 11
Ventral scales pointed or rounded, not quadrangular; body and head scales
as described or not....................................................................... 12
11. Digits on hind leg of nearly equal length; enlarged tubercles covering most of
dorsal surface of body; a series of about four large, paired postmentals on
midline of chin, immediately back of mental.................................. Helodermidae (p. 192)
Digits on hind leg varying greatly in length, fourth toe three times as long
as fifth; enlarged tubercles on body much less numerous, not occupying as
great an area as granules; no enlarged postmentals.......................... Xenosauridae (p. 207)
12. Head and most of body, except belly, covered with very minute granules; no
parietal “eye”; no keels or tubercles along ventral surfaces of digits.
Gekkonidae (p. 40)
Head covered with larger scales; a parietal “eye” usually visible; at least
one tubercle, or several keels, on most or all of lamellae on ventral surfaces
of digits................................................................................. Iguanidae (p. 53)

Family GEKKONIDAE Stejneger

1, 1885, p. 3.

Subfamilies.—Studies by Noble (Amer. Mus. Nov., No. 4, 1921,
pp. 1–16) and by M. A. Smith (Rec. Indian Mus., 1933, p. 16) indicate
that neither the Eublepharidae nor the Uroplataidae are worthy
of either family or subfamily distinction.

Genera.—About 75 genera are commonly recognized.

Range.—World-wide, in tropical and semitropical areas.

Remarks.—Eight genera of this family occur in Mexican territory.
Three of these have probably been introduced accidentally from other
parts of the world. Peropus, with a single representative, arrived on
the west coast of Mexico probably from the Philippines. Hemidac-
tylus, with three species, has a somewhat more complicated history.
One species reached the west coast, probably from the Philippines
(frenatus), another came across the Atlantic to the east coast from
Europe or Africa (turcicus), and a third (mabouia) probably came
from somewhere in the West Indies or, not improbably, from Madag-
scar or South Africa where it also occurs. Aristelliger undoubtedly
was imported from the West Indies. A fourth genus, Gonatodes, has
only recently been reported in Mexico, and may possibly have been transported by man from Central America.

It would appear from collections that all these genera have very limited or discontinuous distribution in Mexico.

The genera Phyllodactylus, Sphaerodactylus, and Thecadactylus very probably reached Mexico without the aid of man.

**KEY TO MEXICAN GENERA OF GEKKONIDAE**

1. Eyelid fully developed, body covered with small cycloid somewhat imbricating scales, with or without somewhat larger tubercles on body or tail; a few preanal pores present; digits cylindrical, never distinctly widened.

   **Coleonyx** (p. 41)

   Eyelid rudimentary, not capable of covering eye........................................ 2

2. Under surface of digits unwidened, covered with flat smooth scales, lacking leaflike lamellae; no preanal or femoral pores; diminutive size.

   **Gonatodes** (p. 45)

   A part of under surface of digits with widened lamellae present; femoral pores present or absent; size variable.......................................................... 3

3. A single widened terminal lamella on digits with the claw lateral to it; no femoral pores; diminutive in size................................. **Sphaerodactylus** (p. 52)

   Digits with one pair or several pairs of widened lamellae.......................... 4

4. A single terminal pair of lamellae, the claw between them; size larger; preanal and femoral pores absent in Mexican forms............ **Phyllodactylus** (p. 46)

   Part of under surface of digits widened with a single or double series of lamellae, usually not present under terminal joint; pores present or absent........... 5

5. Subdigital lamellae undivided, in a single row................................. **Aristelliger** (p. 51)

   Subdigital lamellae in two rows, divided medially.................................. 6

6. Tail with enlarged, sometimes spiny tubercles, forming longitudinal rows or transverse whorls; body with at least some enlarged tubercles, which may be flat or trihedral; femoral or preanal pores present.... **Hemidactylus** (p. 49)

   Tail and body lacking spines or enlarged tubercles; pores present or absent... 7

7. Tail with flat ventral surface, serrated lateral edge, and a median row of enlarged scales on ventral surface; a long, doubly curved series of femoral pores; a fold of skin on posterior side of thigh and leg....... **Peropus** (p. 51)

   Tail rounded, tapering, covered with imbricate scales, a little larger on under surface; no femoral or preanal pores; no skin fold on hind leg.

   **Thecadactylus** (p. 49)

---

**Genus COLEONYX Gray**


**Genotype.**—*Coleonyx elegans* Gray.

**Range.**—The genus extends from Southwestern United States to Panama.

**Species.**—Five, with a total of 12 forms. Four species and 10 forms are known or are to be expected in Mexico.
1. Dorsal surface covered with small granular scales, without larger tubercular scales ......................................................... 2
  Dorsal surface with larger tubercules intermingled with the small granular scales ......................................................... 9

2. Two light and 3 dark bands between level of arm and leg insertions. 

   fasciatus (p. 43) 
   Usually 4 or parts of 4 dark bands between levels of arm and leg insertions, or the bands broken up into spots ........................................................................ 3

3. Preanal pores small, usually 4 in number (rarely more or less), the pore-bearing scales separated medially; maximum snout-vent measurement about 59 mm. ......................................................... brevis (p. 43) 
   Preanal pores larger, usually 6 to 10, the series continuous, not separated medially; maximum snout-vent measurement 77 mm ........................................................................ 4

4. Usually 4 or less postmental scales touching mental variegatus slevini (p. 45) 
   Usually 5 or more postmental scales touching mental ........................................................................................................ 5

5. Preanal pores in males usually 8 or more variegatus bogerti (p. 44) 
   Preanal pores in males usually 7 or less ................................................................................................................................. 6

6. Dark transverse body bars in the adults considerably wider than the light interspaces .............................................................. 7
   Dark transverse body bars in adults about equal to or narrower than the light interspaces; or bars obsolete and replaced by spotting ........................................................................ 8

7. A middorsal light longitudinal line usually splitting the dorsal body bars in the adults variegatus sonoriensis (p. 45) 
   No middorsal light longitudinal line; adults with longitudinal edges of the dark body bars even, with narrow uniform interspaces. variegatus peninsularis (p. 44)

8. Dark body bands in the adults unicolor; top of head unicolor; nuchal light loop narrow and clear variegatus abbotti (p. 44) 
   Dark body bands in the adults with light centers producing a double-barred effect, or bars obsolete and replaced by spotting; top of head spotted; nuchal light loop irregular or obsolete variegatus variegatus (p. 44)

9. Enlarged supranasals 23 separated by a median circular scale. 

   elegans nemoralis (p. 43) 
   Enlarged supranasals in contact. .......................................................... elegans elegans (p. 42)

**COLEONYX ELEGANS ELEGANS** Gray


**Type.**—Brit. Mus. Nat. Hist., male spec.; Mr. Dyson collector.

---

23 Adapted from Klauber, op. cit., pp. 205-206.

23 We believe the scale above nostril should be designated supranasal rather than prenasal; and those scales following the mental should be postmental rather than gular, since the latter term refers to throat or gullet, rather than chin.
Type locality.—Belize, British Honduras.

Range.—Atlantic slopes from central Veracruz through the Yucatán Peninsula, including Honduras and northern Guatemala; on Pacific slopes in Chiapas and undoubtedly adjacent Guatemala. Reported from Veracruz: Jalapa, Potrero Viejo, San Lorenzo, Orizaba, Tezonapa, Zempoala, Presidio, San Juan Cuatotolapam; Oaxaca: San Cristóbal, Cosolapa; Tabasco: Teapa, Tenosique; Campeche: Tuxpeña Camp, Apasote, Encarnación; Yucatán: Chichen Itzá, Mayapán, Puz, Gongora Caves, Ziz, Xyyc Cave; Chiapas: La Esperanza, Piedra Parada; Quintana Roo: Mujeres Island.

**COLEONYX ELEGANS NEMORALIS** Klauber


Type.—EHT-HMS No. 10509; Hobart M. Smith collector.

Type locality.—Hacienda Paso del Río, Colima.

Range.—Colima to southeastern Oaxaca (probably also Michoacán). Probably confined to the coastal area of these states. Reported from Guerrero: Cocoyul, Agua del Obispo, 4 to 5 miles north of Acapulco; Michoacán: vicinity of Jorullo (?); Oaxaca: (said to be intergrades with *elegans*) Tehuantepec, Mixtequilla, Tapanatepec, etc.

**COLEONYX FASCIATUS** (Boulenger) 14

*Eublepharis fasciatus* Boulenger, Catalogue of the lizards in the British Museum, vol. 1, 1885, p. 423.—Güntner, Biologia Centrali-Americana, Reptilia and Batrachia, 1893, p. 84, pl. 31, fig. A.


Type locality.—Ventanas, Durango.

Range.—Known from western Durango, and Sinaloa. The only known specific locality other than the type locality is *Sinaloa*: 10 miles south of Presidio.

**COLEONYX BREVIS** Stejneger


Type.—U.S.N.M. No. 13627; Mr. Marnock collector.

Type locality.—Helotes, Bexar County, Tex.

Range.—The Rio Grande Valley, from northern New Mexico to southern Texas, and southward through eastern Chihuahua and

---

14 Bogert and Oliver (Bull. Amer. Mus. Nat. Hist., vol. 83, 1915, p. 330) suggest the possibility of intergradation of this form with *variegatus*. There is no evidence whatsoever that such is the case. See Klauber, loc. cit.

861316—50—4
western Tamaulipas to southern Coahuila and extreme eastern Durango. Reported from Tamaulipas: Mier; Nuevo León: 5 miles south of Sabinas Hidalgo, near China, Ciénega de Flores, Mamulique Pass; Coahuila: Monclova, Saltillo; Durango: 6 miles northeast of Pedriceña.

**COLEONYX VARIEGATUS VARIEGATUS** (Baird)


**Type.**—U.S.N.M. No. 3217 (now lost).

**Type locality.**—Colorado Desert (*fide* Klauber, *loc. cit.*), here restricted to Winterhaven, Calif.

**Range.**—Southern Nevada south through eastern California and western Arizona to northeastern Baja California and northwestern Arizona. Reported from *Baja California*: San Felipe, Colorado River Delta, Colorado Desert; *Sonora*: Paso McDougall, Sierra Blanca, Punta Peñasco, etc.

**COLEONYX VARIEGATUS ABBOTTII** Klauber


**Type.**—No. 34790, collection of L. M. Klauber; William Moore collector.

**Type locality.**—Proctor Valley, between Jamul and Upper Otay Reservoir, San Diego County, Calif.

**Range.**—Southwestern California and northwestern Baja California. Reported from *Baja California*: Ensenada, 65 miles southeast of Tecate, San José (lat. 31°), ? Calmalfi, Isla de Cedros.

**COLEONYX VARIEGATUS BOGERTII** Klauber


**Type.**—L. M. Klauber No. 32486; Lee W. Arnold collector.

**Type locality.**—Xavier, Pima County, Ariz.

**Range.**—Southeastern Arizona and presumably adjacent Sonora and New Mexico. Not yet recorded from Mexico.

**COLEONYX VARIEGATUS PENINSULARIS** Klauber


**Type.**—Mus. Comp. Zool. No. 37210; Miguel L. Cornejo collector.
Type locality.—La Paz, Baja California, Mexico.
Range.—Eastern side of southern Baja California. Reported from Baja California: San José del Cabo, between Loreto and Comondú.

COLEONYX VARIEGATUS SLEVINI Klauber


Type.—California Acad. Sci. No. 51697; J. R. Slevin collector.
Type locality.—“South Santa Inez Island” (lat. 27° W.), Gulf of California (Baja California coast).
Range.—Isla Santa Inez and Isla de San Marcos, Gulf of California.

COLEONYX VARIEGATUS SONORIENSIS Klauber


Type locality.—Five miles southeast of Hermosillo, Sonora.
Range.—Known only from central western Sonora. Reported from La Posa 10 miles northwest of Guaymas, Tepoca Bay, Sierra Álamo (30 miles west of Caborca), Isla de Tiburón.

Genus GONATODES Fitzinger

Gonatodes Fitzinger, Systema reptilium, 1843, pp. 91-92.

Genotype.—Gymnodactylus albogularis Duméril and Bibron =Gonatodes albogularis (Duméril and Bibron).
Range.—Tropical America; West Indies. Species in South Asia, Africa, and Malaya formerly regarded as members of the genus are now placed in Cnemaspis.
Species.—About 10, of which only one occurs in Mexico.

GONATODES FUSCUS (Hallowell)

Gymnodactylus fuscus, A. Duméril, Arch. Mus., vol. 8, 1856, p. 477.—Bocourt, Mission scientifique au Mexique ..., Études sur les reptiles, livr. 2, 1873, pp. 48, 49, pl. x, fig. 5, 5a, 5b, 5c.
Type.—Acad. Nat. Sci. Philadelphia?
Type locality.—Nicaragua, here restricted to Rama.
Range.—Huixtla, Chiapas (only definite Mexican record), southward in Central America to northwestern South America; Jamaica (probably introduced); Florida (probably introduced).

Genus PHYLLODACTYLUS Gray

Phylloactylus Gray, Spicilegia zoologica, 1830, p. 3.
Discodactylus Fitzinger, Systema reptilium, 1843, p. 95 (type, Phylloactylus pulcher Gray).

Genotype.—Phylloactylus pulcher Gray.

Range.—California to and including most of South America; Galápagos Islands, southern Asia, Africa, Madagascar, Australia, and cetera.

Species.—About 48 species are known; 9 are known from Mexico.25

KEY TO MEXICAN SPECIES OF PHYLLODACTYLUS

1. Dorsal squamation consisting of minute rounded scales without admixture of larger tubercles; no enlarged tubercles on tail; small, length 52 mm. unctus (p. 47)

Dorsal squamation with an admixture of larger tubercles; larger tubercles present or absent on tail. 2

2. Tail lacking any trace of enlarged tubercles on dorsal surface. 3
Tail with smaller or larger tubercles intermixed with dorsal lepidosis. 4

3. Scales on head between middle of orbits 15–17; on dorsal and lateral surface 14 to 16 rows of large, very distinct trihedral tubercles; smaller, 44 + mm. bordai (p. 47)

Scales between orbits 19–24; dorsal tubercles low, small, rather indistinct, arranged in 8 to 12 rows; larger, 90 mm. decampi (p. 47)

4. Two rows of tubercles on dorsal surface of tail (greatly reduced or absent in homolepidurus). 5
More than two rows of enlarged tubercles, at least on basal third of tail. 6

5. Scales between middle of orbits 22–26; tubercles on back and tail large, well defined. muralis (p. 47)

Scales between middle of orbits 19–24; tubercles on back much smaller than in muralis, the caudal tubercles reduced so that they can be discerned only with difficulty; 67 mm. homolepidurus27 (p. 48)

25 Phylloactylus pulcher Gray, Spicilegia Zoologica, from "Tropical America," may be found to occur in Mexico. It is distinct from all others listed here by having 20 to 22 longitudinal series of dorsal tubercles (rather than 14 to 16), and larger ventral scales, in 22 longitudinal and 53 transverse series (rather than 25 to 30 and 65, respectively).

We likewise are unable to place Phylloactylus mentalis Werner (Jahrb. Hamburg. Wiss. Anstalten, vol. 27, No. 2, 1910, p. 5; Zool. Mus. Hamburg; L. von Poppinghausen coll.; Central America or Mexico). Mosauer (Copeia, 1936, p. 144) includes a few notes on the type.

26 Not counting scales on vestigial eyelids.

27 Bogert and Oliver (Bull. Amer. Mus. Nat. Hist., vol. 83, 1945) note that at least nine species of Phylloactylus have been described from western Mexico in recent years and that certain of these are allopatric and their characters strongly suggest that a more accurate "indication of the biological conditions that exist" may be obtained by considering them as subspecies or races of the same species. They proceed to place homolepidurus as a subspecies under tuberculatus. Since the latter name cannot be fixed at the present time, we do not follow this proposal. Unfortunately, these authors have not examined most of the described forms and have overlooked the fact that at least four of the species may occur in the same general locality. They state that "the lizards of this genus exhibit considerable variation." These forms are probably no more variable than other lizards. For many years most North American species of Phylloactylus were classified in museums as "tuberculatus," and examination of such material might create the erroneous idea that "tuberculatus" was an extremely variable species.
6. Four distinct longitudinal rows of enlarged tubercles at least on basal fourth of tail, usually in rather regular transverse rows

7. Scales between orbits 12–16; dorsal and posterior part of thigh with numerous large conical tubercles; color darker above, cream color below; 80 mm.

8. Scales between orbits 25–30; 95 mm

PHYLODACTYLUS UNCTUS (Cope)

Phyllodactylus (Diplodactylus) unctus, BOCOURT, Mission scientifique au Mexique

Type.—U.S.N.M. No. 5304.
Type locality.—Cape San Lucas, Baja California.
Range.—Southern Baja California. Reported from Baja California: San José del Cabo; Isla Partida near Isla Espíritu Santo, Isla Gallina, and Isla San Francisco.

PHYLODACTYLUS DELCAMPI Mosauer

Phyllodactylus delcampi MosaUER, Copeia, 1936, pp. 141–146.
Type.—Mus. Comp. Zool. No. 41238; W. Mosauer collector.
Type locality.—Tierra Colorada, Guerrero.
Range.—Known only from the type locality.

PHYLODACTYLUS BORDAI Taylor

Type.—EHT–HMS No. 27732; E. H. Taylor collector.
Type locality.—Six miles north of Taxco, Guerrero.
Range.—Northern Guerrero; known only from the type locality.

PHYLODACTYLUS MURALIS Taylor

Type.—EHT–HMS No. 10902; H. M. Smith collector.
Type locality.—Totolapam, Oaxaca.
Range.—Oaxaca. Reported from Totolapam, Tehuantepec, San Gerónimo, Salina Cruz.
PHYLLODACTYLUS HOMOLEPIDURUS Smith


**Type.**—EHT–HMS No. 10853 (originally EHT field No. 146); E. H. Taylor collector.

**Type locality.**—Five miles southwest of Hermosillo, Sonora.

**Range.**—Southern Sonora and adjacent Chihuahua. Reported from Sonora: La Posa, 5 miles southwest of Hermosillo, Guirocoba; Chihuahua: Batopilas.

PHYLLODACTYLUS MAGNATUBERCULATUS Taylor


**Type.**—EHT–HMS No. 10995; H. M. Smith collector.

**Type locality.**—Acapulco, Guerrero.

**Range.**—Known only from the type locality.

PHYLLODACTYLUS MAGNUS Taylor


**Type.**—EHT–HMS No. 21783; E. H. Taylor collector.

**Type locality.**—Tierra Colorada, Guerrero.

**Range.**—Guanajuato, Oaxaca, and Chiapas. Reported from Guerrero: Garrapatas, El Ocotito, Tierra Colorada, Agua del Obispo; Oaxaca: Tehuantepec, Oaxaca, Tres Cruces, Cerro Arenal, Cajón de Piedra, Escurana; Chiapas: Tonala.

PHYLLODACTYLUS LANEI 28 Smith


**Type.**—EHT–HMS No. 10942 (originally EHT field No. 146); E. H. Taylor and H. M. Smith collectors.

**Type locality.**—Near Tierra Colorada, Guerrero, Mexico.

**Range.**—Sinaloa and (?) western Durango 29 southward to central-southern Guerrero, probably confined to the coastal areas up to an elevation of about 3,500 feet. Recorded from the states of Sinaloa (including the Tres Marías Islands), ? Durango, Nayarit, ? Jalisco, Colima, Michoacán, and Guerrero.

28 The suggestion of common identity of *P. lanei* and *P. tuberculatus* by Mosauer is, we believe, untenable (Mosauer, Copeia, 1936, p. 194).

29 A specimen of *Phyllodactylus* collected at Durango, Mexico by Dr. T. H. Webb (U. S. N. M. No. 3208) and figured by Baird (Reptiles of the Boundary, U. S. and Mexican Boundary Surveys, 1859, p. 12, pl. 23, figs. 1-8) may or may not belong to this species, since Durango is considerably higher than this species is known to range.
PHYLLODACTYLUS TUBERCULOSUS Wiegmann


Type.—? Zool. Mus. Berlin; collector unknown.
Type locality.—"Californien."
Range.—? Extreme southern part of California, south through Baja California.

Genus THECADACTYLUS Gray


Genotype.—Gecko rapicauda Houttuyn.
Range.—Southern Mexico to northern South America (Peru); Antilles; islands north of Australia in Torres Strait.
Species.—Two.

THECADACTYLUS RAPICAUDUS (Houttuyn)


Type.—Unknown.
Type locality.—American Islands, here restricted to Chichen Itzá, Yucatán.
Range.—Southward from Yucatán through Central America to northwestern South America; Lesser Antilles. Reported from Yucatán: Chichen Itzá; Puz Cave, Oxkutzcab; Chakxix Cave, Tekax. The Guadalajara reference of Cope (U. S. Nat. Mus. Bull. 32, 1887, p. 28) is very doubtful.

Genus HEMIDACTYLUS Oken

Tachybates Fitzinger, Systema reptilium, 1843, p. 105 (type, "Hemidactylus mabuya Cuvier").
Phoépus Fitzinger, Systema reptilium, 1843, p. 105 (type, "Hemidactylus javanicus Cuvier").
Genotype.—*Gekko tuberculosis* Daudin [=*Hemidactylus mabouia* Moreau de Jonnès].

Range.—Widespread in Africa, southern Eurasia, tropical America, Polynesia.

Species.—About 60 known. Three are found in Mexico.

KEY TO MEXICAN SPECIES OF *HEMIDACTYLUS*

1. Preanal pores 4 to 10, forming a short angular series; no femoral pores; dorsal scales minute granules intermixed with large tubercles usually larger than interspaces between them; inner digits well developed. turcicus (p. 50)
   Preanal and femoral pores forming a continuous series  
2. Small granules of the dorsal surface with only a few relatively small, convex tubercles, these usually on posterior part of body; pores 15–18 on each side, not separated mesially. frenatus (p. 50)

Small granules of the dorsal surface mixed with very numerous trihedral tubercles, irregularly scattered; pores from 15 to 30 on each side. mabouia (p. 50)

*HEMIDACTYLUS MABOUIA* (Moreau de Jonnès)


*Type locality.*—"Antilles," here restricted to St. Vincent, Lesser Antilles.

*Range.*—In Mexico known only from "Veracruz" (probably city). Widespread in the West Indies, and the northern and eastern coast of South America; Madagascar and Africa. The American localities are probably due to relatively recent introductions from Africa, or the West Indies.

*HEMIDACTYLUS FRENATUS* Schlegel


*Type.*—Leyden Mus.

*Type locality.*—Java.

*Range.*—Guerrero, in the region about Acapulco. Widespread in southern Asia, Pacific Islands, etc. Reported from Guerrero: Acapulco, Tierra Colorada.

*HEMIDACTYLUS TURCICUS TURCICUS* (Linnaeus)


*Hemidactylus turcicus turcicus*, Loveridge, Copeia, 1941, p. 247.

*Type.—Unknown.*

*Type locality*—"Habitat in Oriente," here restricted to Cairo, Egypt.

*Range.*—Tamaulipas south to Yucatán, distribution very probably discontinuous. Found in the West Indies, Florida, North Africa, and southwestern Asia. Reported from Tamaulipas: 12 miles east of Llera; Veracruz: no specific locality; Tabasco: Álvaro Obregón, Emiliano Zapata; Campeche: Ciudad del Carmen; Yucatán: Progreso. The Mexican records are due to modern introductions, probably from northern Africa.

**Genus ARISTELLIGER** Cope


*Genotype.*—*Aristelliger lar* Cope.

*Species.*—Six, all West Indian except the one Yucatán form.

*Range.*—West Indies and the peninsula of Yucatán.

**ARISTELLIGER GEORGEENSIS** (Bocourt)

*Idiodactylus georgeensis* Bocourt, Mission scientifique au Mexique..., Études sur les reptiles, livr. 2, 1873, pp. 41-42, pl. 10, fig. 1.


*Type locality.*—St. George Island, near Belize, British Honduras.

*Range.*—The eastern edge of the Yucatán Peninsula. Recorded in Mexico only from Cozumel Island (Quintana Roo).

**Genus PEROPUS** Wiegmann


*Dactyloperus* Fitzinger, Systema reptilium, 1843, p. 103 (type, *Peropus peronii* Duméril and Bibron= *Peropus mutilatus* Wiegmann).


*Chalinocnemis* Dugès, La Naturaleza, vol. 6, 1883, p. 312 (type, *Hemidactylus navarri* Dugès= *Peropus mutilatus* Wiegmann).

*Spasmocnemis* Dugès, loc. cit. (type as for preceding).

*Genotype.*—*Hemidactylus mutilatus* Wiegmann.

---

1 We are inclined to maintain *Peropus* and *Gebyra* based on the different character of the subdigital lamellae.
Range.—Widespread in southern Asia from Ceylon to the Philippines; south through the Malay Archipelago to New Guinea; Seychelles; western Mexico.

Species.—One form occurs in Mexico, probably introduced from the Philippines by man; about 10 others are known.

**PEROPUS MUTILATUS** (Wiegmann)


_Hemidactylus Navarri Dusès_, La Naturaleza, vol. 6, 1883, pp. 309–312, pl. 7a (type locality, San Blas, Nayarit; 2 types in Alfredo Dugès Museum, Guanajuato, Guanajuato.)


_Type locality._—Manila, Philippine Islands.

Range.—Seaports of Nayarit and Sinaloa. Widespread in southeastern Asia, Philippine Islands, Dutch Indies, to New Guinea and certain Pacific Islands. Reported from _Nayarit_: San Blas, Tepic; _Sinaloa_: Presidio, Mazatlán.

**Genus SPHAERODACTYLUS** Wagler


_Genotype._—Sphaerodactylus spurator_ Cuvier.

Range.—West Indies, southern Mexico, Central America.

Species.—About 38 forms. Three are known or expected from Mexico.\(^{32}\)

**KEY TO MEXICAN SPECIES OF SPHAERODACTYLUS**

1. Dorsal scales granular, keeled.----------------------------- *lineolatus* (p. 52)
   Dorsal scales imbricate, smooth.-------------------------------- 2

2. A single nuchal band; two distinct spots at base of tail, light spots at elbow and knee usually discernible.----------------------------- *glaucus* *glaucus* (p. 53)
   A cream-colored nuchal band bordered by black bands of equal width anteriorly and posteriorly; two narrow longitudinal light lines from collar to eye; indistinct narrow light bands on back; spots on tail base absent or obsolete; no light spots at knee and elbow in young or adult.

   *glaucus torquatus* (p. 53)

**SPHAERODACTYLUS LINEOLATUS** Lichtenstein


_Type._—Zool. Mus. Berlin, three cotypes.

\(^{32}\) Until new material is discovered in Mexico, the Mexican reports of _Sphaerodactylus fantasticus_, _antiracnus_, _punctatissimus_ (cinereus), and _spurator_, are to be regarded as uncertain. The type locality of _antiracnus_ is "Mexico," but possibly in error (see p. 213). It would not be impossible for a part of these forms to occur, as recent introductions in Mexico. Banana fruits serve as a ready means of transportation.
Type locality.—Veraguas, Panamá.

Range.—The eastern edge of the Yucatán Peninsula southeastward to Colombia. No definite records from Mexico.\(^3\)

**SPHAERODACTYLUS GLAUCUS GLAUCUS Cope**


*Type.*—U.S.N.M. No. 6572, three cotypes (now one, No. 13570, in Mus. Comp. Zool.); Arthur Schott collector.

*Type locality.*—Near Mérida, Yucatán.

*Range.*—Known from southern Veracruz to Guatemala and British Honduras, and possibly to Costa Rica. Reported from Oaxaca: Tehuantepec, Salina Cruz, Guengola Mountain; Tabasco: Teapa; Campeche: Tuxpeña Camp, Ciudad del Carmen, Balchacaj, Apazote; Veracruz: Pérez, Coatzacoalcos River; Yucatán: Mérida.

**SPHAERODACTYLUS GLAUCUS TORQUATUS Strauch**


*Type.*—Mus. Petrograd No. 3268, three specimens; Hr. Salmin collector.

*Type locality.*—“Mazatlan, Mexico” (= Mazatlán, Oaxaca?).

*Range.*—Uncertain. Known from the type locality and Cajón de Piedra, Oaxaca.\(^3\)

Family IGUANIDAE Gray


*Genera.*—About 65 genera are commonly recognized; 21 occur in Mexico.

---

\(^3\) The species is included here on the strength of its occurrence in British Honduras (Schmidt, Publ. Field Mus. Nat. Hist., zool. ser., vol. 22, 1941, p. 489), for although it has been cited from "Mexico" by numerous authors, no definite record of its occurrence in that country is known to us. Its existence in at least Quintana Roo is virtually a certainty.

\(^3\) It now seems most probable that this name was applied to a unicolor intergrade between *S. g. glaucus* and *S. g. torquatus* such as those known to occur in the vicinity of Tehuantepec, Oaxaca. The entire description, of both scutellation and coloration, fits glaucus, not lineolatus, although Barbour (op. cit., p. 238) places Peters's name in the synonymy of the latter species. See Smith, loc. cit., and Taylor, loc. cit., for discussion.—H. M. S.

\(^3\) The subspecies does not occur in the vicinity of the city of Tehuantepec, but only, so far as we are aware, near the coast southwest of Salina Cruz, where a village by the name of Mazatlán does exist.
**Range.**—Extreme southwestern Canada and extreme southeastern New York, southward throughout the Americas; Bermuda Islands; West Indies; Revillagigedos Islands; Galápagos Islands; Madagascar (two genera); Fiji and Tonga Islands (one genus).

**KEY TO MEXICAN GENERA OF IGUANIDAE**

1. No femoral pores
   
   | Femoral pores present, conspicuous in males, less well defined in females |
   |-------------------------|-----------------|---------------------|
   | 2                      | 5               |
   
2. Ventral lamellae on next to last phalanx of digits expanded laterally, forming a padlike structure; posterior part of head not produced
   
   | Ventral lamellae on digits not expanded to produce a pad; posterior part of head more or less produced |
   |---------------------------------|-----------------|
   | 3                      | 4               |

3. Infraocular lamellae with a single, tubercular keel
   
   | Infraocular lamellae with several sharp keels |
   |----------------------------------------|----------------|
   | 4                      | 5               |

4. Toes with a lateral fringe
   
<table>
<thead>
<tr>
<th>No fringe on toes</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
</tr>
</tbody>
</table>

5. A single, median, dorsal, longitudinal series of enlarged scales
   
   | Scales in vertebral row not conspicuously if any larger than adjacent scales |
   |------------------------------------------------|---------------|
   | 6                      | 9               |

6. Tail with whorls of much-enlarged, spiny scales, usually separated from each other by whorls of small scales
   
   | Scales on dorsal and lateral surfaces of tail (except the vertebral row) subequal in size, small |
   |------------------------------------------------|---------------|
   | 7                      | 8               |

7. A group of much enlarged, spinose, protuberant scales on shank and sometimes on thigh; basal caudal region (or entire tail in short-tailed species) highly modified, sometimes flattened and widened, with whorls of greatly enlarged scales separated from each other by no more than 1 row of small scales; total length less than 450 mm
   
<table>
<thead>
<tr>
<th>Enyaliosaurus</th>
</tr>
</thead>
<tbody>
<tr>
<td>8</td>
</tr>
</tbody>
</table>

8. A large, circular shield below tympanum; a well-developed gular appendage
   
<table>
<thead>
<tr>
<th>Phrynosoma</th>
</tr>
</thead>
<tbody>
<tr>
<td>9</td>
</tr>
</tbody>
</table>

9. Head bearing bony spines posteriorly, or elevated somewhat in a projecting ridge
   
<table>
<thead>
<tr>
<th>Holbrookia</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
</tr>
</tbody>
</table>

10. Superciliary scales not imbricate, but juxtaposed, like other scales on head; rostral subdivided, with a median suture meeting lip
   
<table>
<thead>
<tr>
<th>Saurophalus</th>
</tr>
</thead>
<tbody>
<tr>
<td>11</td>
</tr>
</tbody>
</table>

11. Sutures between supralabials diagonal, not vertical; mental very small, bordered posteriorly by a median postmental
   
<table>
<thead>
<tr>
<th>Uma</th>
</tr>
</thead>
<tbody>
<tr>
<td>12</td>
</tr>
</tbody>
</table>

12. No ear opening
   
<table>
<thead>
<tr>
<th>Uma</th>
</tr>
</thead>
<tbody>
<tr>
<td>13</td>
</tr>
</tbody>
</table>

13. Interparietal much smaller than ear opening, not more than twice as large as any other dorsal head scale; several enlarged auricular lobules; a patch of enlarged scales with projecting points on rear of femur
Interparietal nearly as large as ear opening, or larger, much larger than any other dorsal head scale; no enlarged auricular lobules; no patch of enlarged scales on rear of femur.  

Callisaurus (p. 85)

14. No complete transverse gular fold, or if so body and tail compressed and dorsal scales nearly uniform, large, no more than 75 from interparietal to rear of thighs.

A complete transverse gular fold, marked by granular or reduced scales, much smaller than those preceding or following fold; dorsal scales very small, or at least not uniformly large and as few as 75 from interparietal to rear of thighs (except Uta squamata); body and tail flattened, never compressed even slightly.

Sator (p. 139)

15. A gular fold usually present; lateral scales granular; body and tail somewhat compressed.

Gular fold never present; lateral scales not granular in those species with compressed tail.

Sceloporus (p. 105)

16. Interparietal very large, about as large as or larger than ear opening.  

Interparietal very small, a great deal smaller than ear opening.

17. Two scales bordering mental posteriorly between infralabials; head very broad, neck narrow.

Three or more scales bordering mental posteriorly between infralabials; head elongate, neck not so sharply defined.

Crotaphytus (p. 91)

18. Dorsal scales all small, granular, smooth, very little larger than lateral scales; enlarged supraoculars in more than one row; one or more blackish bars across shoulders; size frequently over 75 mm. snout to vent.

Dorsal scales, at least toward middle of back, larger than lateral scales, keeled; enlarged supraoculars in one row; no blackish bars across shoulders; size less than 75 mm. snout to vent.

19. Caudal scales smaller than ventrals on body, not mucronate; three or four black bands across body.

Caudal scales large, mucronate, much larger than ventrals on body; a single narrow black band across shoulders.

Petrosaurus (p. 90)

20. A longitudinal dorsolateral line or dermal fold, usually with some enlarged scales on it; usually a second, lateral fold; no small, rounded blue blotch behind axilla; frontal divided or not.

Urosaurus (p. 140)

No longitudinal dorsolateral dermal fold; often a small, rounded, dark blue blotch behind axilla; frontal always divided.

Uta (p. 147)

Genus ANOLIS Daudin


Dactyloa Wagler, Naturliches System der Amphibien, 1830, p. 148 (type, Anolis viridis Wied).

Tropidopilus Fitzinger, Systema reptilium, 1843, p. 66 (type, Anolis fuscoauratus d’Orbigny, monotype).

Trachypilus Fitzinger, op. cit., p. 67 (type, Anolis sagraei Duméral and Bibron).


Clenocercus Fitzinger, op. cit., pp. 17, 68 (type, Dactyloa bullaris Wagler = Anolis carolinensis Linnaeus).

Gastrorropis Fitzinger, loc. cit. (type, Dactyloa nebulosa Wiegmann).

Genotype.—*Anolis bullaris* Daudin (=*Anolis carolinensis* Voigt).

Species.—Perhaps 350. Thirty-five forms are listed here, but two are not known from any definite locality, four are known only from “Mexico,” and the Mexican range of another is unknown.

Range.—Southeastern United States to Brazil; West Indies.

KEY TO MEXICAN SPECIES OF ANOLIS

1. Tail strongly compressed.......................................................... 2
   Tail subcylindrical, no more than slightly compressed......................... 4
2. Two continuous rows of scales along middorsal line on tail...*barkeri* (p. 58)
   One row of scales along middorsal line of tail.................................. 3
3. Ventrafis keeled........................................................................... 2
   Ventrafis smooth........................................................................... 3
4. Dorsal scales as large as or larger than ventrals, and both strongly keeled. 5
   Dorsal scales smaller than ventrals, or one or both sets of scales smooth; or,
   if body scales as above, occipital much larger than ear opening............... 8
5. Supraorbital semicircles in contact or separated by a single series of scales. *megapholidotus* (p. 59)
   Supraorbital semicircles separated by 2 or 3 series of scales........... 6
6. Dorsal and ventral scales subequal in size................................. *metallicus* (p. 59)
   Dorsal scales distinctly larger than ventrals................................... 7
7. Tibia 75 percent to 85 percent length of head (to anterior border of ear); adult
   size smaller, about 36 mm.; dewlap brilliant red with a large purple spot
   at base in life.................................................................................. 2
   Tibia 90 percent length of head, or longer; adult size larger, about 50 mm.;
   dewlap red, without a purple spot.................................................. *tropidonotus* (p. 60)
8. Ventrafis smooth 30
   Ventrafis keeled........................................................................... 25
9. Supraorbital semicircles in contact and a single row of only 4 or fewer large
   supraoculars................................................................................... 10
   Supraorbital semicircles separated or, if in contact, supraoculars numerous (8
   or more enlarged)............................................................................ 14
10. Two rows of loreals; female with a large gular “pouch”; all scales on body
    flat and granular, small and equal on back and sides, larger on belly.
    *impetigosus* (p. 60)
    Four to six rows of loreals; females (not known in *schmidti*) lacking a large
    gular pouch; dorsal scales keeled, at least faintly larger than lateral scales. 11
11. Ventrafis and dorsals subequal in size; enlarged supraoculars three-fourths
    width of supraocular area............................................................. *schmidti* (p. 60)
    Ventrafis distinctly larger than (at least 1 1/2 times as large as) dorsals; enlarged
    supraoculars narrower, half width of supraocular area or less............... 12
12. Two gulars in contact with mental between chin shields; 9 or fewer scales
    across snout between second canthals (counting as the first canthal that
    which borders the superciliaries).................................................... 13
    Three or (generally) 4 gulars in contact with mental between chin shields;
    10 to 12 scales across snout between second canthals......................... *gadovii* (p. 61)
13. Six scales between nasals; 5 scales in contact with rostral between supralabials;
    ventrals larger........................................................................... *dunni* (p. 61)

30 In six species (*cymbopha, luciventris, limifrons rodriguezi, milleri, schimidtii, and schmidti*) the ventrals may
appear either smooth or keeled; these are keyed out under both categories. All others are uniformly and
plainly either smooth or keeled.
Seven or 8 scales between nasals; 6 scales in contact with rostral between supralabials; ventrals smaller.................................................. taylori (p. 61)
14. All dorsals and laterals equal in size, minute, granular; tibia 85 percent length of head (to anterior border of ear); scales of supraorbital semicircles enlarged; head scales feebly keeled............................................. damulus (p. 61)
At least 2 or more vertebral rows of slightly enlarged scales, or, if all dorsal scales uniform, tibia less than 70 percent length of head, and either scales of supraorbital semicircles not enlarged, or head scales perfectly smooth... 15
15. Scales of supraorbital semicircles, at least in the frontal and frontoparietal area, not or scarcely larger than scales between; tail compressed, sub-triangular in section with a serrate crest; tibia much shorter than head (70 percent or less) all dorsal and lateral scales equal in size.

pentaprion (p. 61)
Scales of supraorbital semicircles distinctly larger than scales between; tail not or only slightly compressed, with little or no crest.............................................. 16
16. Supraorbital semicircles either in contact or separated by a single series of scales; postanals enlarged in males (males not known in utowanae, cymbops, bekeri, and baccatus)........................................................................ 17
Supraorbital semicircles separated by 2 or 3 rows of scales; postanals not enlarged in males (not known in gäntherii).......................................................... 22
17. Interparietal smaller than ear opening.................................................. 18
Interparietal larger than ear opening...................................................... 19
18. Three loreal rows.............................................................................. utowanae (p. 62)
Seven loreal rows.................................................................................. cymbops (p. 62)
19. All dorsal and lateral scales equal in size, minute, granular, smooth; tibia only 60 to 65 percent head length; all dorsal head scales smooth.

beckeri (p. 62)
At least a few vertebral scales slightly enlarged, keeled; tibia longer in proportion to head length; dorsal head scales smooth or not................. 20
20. Upper head scales tricarinate......................................................... baccatus (p. 62)
Upper head scales smooth, slightly rugose, or feebly uncarinate; postanals enlarged in males............................................................... 21
21. One series of 3 enlarged supraoculars; male lacking first digit on all four feet37; dewlap red.............................................................. liogaster (p. 62)
Two series of 7 to 9 enlarged supraoculars; males with 5 digits on each foot; dewlap yellow................................................................. laeviventris (p. 62)
22. Tibia distinctly longer than head (15.5 mm. and 13.5 mm. respectively in the type and only known specimen)................................. schiedii (p. 63)
Head (snout to anterior border of ear opening) at least a little longer than tibia.................................................................................. 23
23. Interparietal plate twice as long as ear opening, much larger than adjacent, very small head scales.................................................... guntherii (p. 63)
Interparietal plate not more than 1½ times as long as ear opening, not more than 4 times as large as adjacent scales.................................................. 24
24. Ventral scales only slightly larger than dorsals; one large scale in anterolateral corner of supraocular area, in contact with superciliaries and separated from enlarged supraoculars by one row of small scales........... milleri (p. 64)
Ventral scales much larger than dorsals............................................. limifrons rodriguezii (p. 64)
25. Head shorter than tibia........................................................................ 26
Head longer than tibia............................................................................ 27

37 Materia recently examined demonstrates that this species like others of Anolis possesses five digits on both limbs in both sexes.
26. Ventrals very strongly keeled; dorsal scales smooth, no larger than laterals; head broad. \textit{capito} (p. 65)
    Ventrals very weakly keeled or smooth; dorsal scales keeled, somewhat larger than laterals; head longer. \textit{schiedii} (p. 63)
27. Lamellae under second and third phalanges of fourth toe, about 27; interparietal scarcely enlarged, much smaller than ear opening. \textit{petersii} (p. 65)
    Lamellae under second and third phalanges of fourth toe, 23 or less; interparietal variable. 28
28. Two or 3 rows of scales between supraorbital semicircles. 29
    Supraorbital semicircles in contact or separated from each other by no more than 1 row of scales. \textit{biporactus} (p. 65)
29. Ventrals very strongly keeled; 22 or 23 lamellae under phalanges 2 and 3 of fourth toe. \textit{schmidti} (p. 60)
    Ventrals very weakly keeled; not over 16 or 17 lamellae. 24
30. A single row of not more than 5 large supraoculars. 31
    Two or more rows of 7 or more enlarged supraoculars. 35
31. Ventrals very weakly keeled; postanals enlarged and dewlap yellow in males; supraorbital semicircles broadly in contact. \textit{nebuloides} (p. 66)
    Ventrals very strongly keeled. 32
32. Males with enlarged postanals and dewlap more or less unicolor. 33
    Males without enlarged postanals and dewlap with a conspicuous central dark blue spot. 34
33. Dorsal scales very slightly smaller than ventrals. \textit{nebulosus} (p. 65)
    Dorsal scales markedly smaller than ventrals. \textit{nebuloides} (p. 66)
34. Supraorbital semicircles in contact with each other and with interparietal. \textit{kidderi} (p. 66)
    Supraorbital semicircles separated from each other (generally) and from interparietal (invariably). \textit{ustus} (p. 66)
35. Interparietal little if any larger than ear opening; dewlap in males without a central blue spot. 36
    Interparietal nearly or fully twice as long as ear opening; dewlap in males with a central blue spot or not. 38
36. Ventral scales indistinctly keeled; dorsal scales granular except two vertebral rows of abruptly enlarged scales. \textit{cymbops} (p. 62)
    Ventral scales strongly keeled; dorsal scales uniform in size in both vertebral and paravertebral areas. 37
37. Dorsals very small, much smaller than ventrals; 17–19 lamellae under phalanges 2 and 3 of fourth toe; 6–8 rows of loreals. \textit{lemurinus bourgeaui} (p. 66)
    Dorsals rather large, distinctly smaller than ventrals; 15 or 16 lamellae; 4–6 rows of loreals. \textit{cummingii} (p. 67)
38. Dorsals very slightly smaller than ventrals, in about 14 rows; a purple spot in center of gular fan in males; postanals not enlarged. \textit{heliactin} (p. 67)
    Dorsals smaller; a purple spot or not; postanals enlarged or not. 39
39. Both dorsals and ventrals smaller; no purple spot in center of dewlap; postanals enlarged in males. \textit{laeviventris} (p. 62)
    Dorsals and ventrals larger; a purple spot in center of dewlap; postanals not enlarged in males. \textit{sericeus} (p. 67)

\textbf{ANOLIS BARKERI} Schmidt

\textit{Anolis barkeri} Schmidt, Publ. Field Mus. Nat. Hist., zool. ser., vol. 24, 1939, pp. 7–9, fig. 2.

\textit{Type}.—Brit. Mus. Nat. Hist. No. 36.6.6.12; R. Wright Barker collector.
Type locality.—Cascajal, upper Uzpanapa River, Veracruz.

Range.—Known only from the type locality.

**ANOLIS SAGREI MAYENSIS** Smith and Burger


*Type*—E. H. Taylor No. 11523; H. M. Smith collector.

*Type locality*.—Panlao, Campeche (a small island at mouth of Río Mamantel, Laguna de Términos).

*Range*.—The Atlantic coast from central Mexico to northern South America. Recorded in Mexico from the states of Yucatán: Progreso, Mérida, Chichen Itzá; Campeche: Balchacaj, Ciudad del Carmen, Panlao; Tabasco: San Juan Bautista; Quintana Roo: Cozumel Island.

**ANOLIS COZUMELAE** Smith


*Type*.—Chicago Nat. Hist. Mus. No. 751; male; C. F. Millspaugh collector.

*Type locality*.—Cozumel Island, Yucatán Peninsula.

*Range*.—Cozumel Island, Quintana Roo.

**ANOLIS METALLICUS** Bocourt


*Type locality*.—Mexico.

*Range*.—Unknown.

**ANOLIS MEGAPHOLIDOTUS** Smith


*Type*.—EHT-HMS No. 11149 (originally EHT-HMS field No. 1509); E. H. Taylor and H. M. Smith collectors.

*Type locality*.—Between Rincón and Cajones (about 40-45 km. south of Chilpancingo), Guerrero.

*Range*.—Known only from the type locality; presumably ranges throughout the Sierra Madre del Sur in Guerrero.

---

13 Barbour (loc. cit.) synonymizes this with *A. h. uniformis*, and Stuart (loc. cit.) suggests synonymy with *A. tropidonotus*. Inasmuch as both the original description and the subsequent illustration indicate the possession by the type of relatively small dorsal scales equal in size to the ventrals, while in both *tropidonotus* and *uniformis* the dorsals are much larger, we regard *metallicus* as distinct from either.
ANOLIS HUMILIS UNIFORMIS Cope


Type.—U.S.N.M., Nos. 6774, 24734–48, 24750 (Guatemala), 24859 (Yucatan), cotypes; Mus. Comp. Zool. No. 10933 (Guatemala).

Type locality.—Yucatan and Guatemala, here restricted to 2 miles north of Santa Teresa, Guatemala.

Range.—The southern portion of the Yucatan Peninsula, from Chiapas and probably Tabasco to El Petén and British Honduras. Known in Mexico from Chiapas: Usumacinta River (across from Piedras Negras, El Petén, Guatemala), Tlacotalpa, Mineral de Santa Fé; Tabasco: Teapa.

ANOLIS TROPIDONOTUS Peters


Type.—Zool. Mus. Berlin, 6 specimens; Dr. Hille collector.

Type locality.—Huanuco (sic) = probably, Huatusco, Veracruz.

Range.—Atlantic slopes from Veracruz to Nicaragua, including the Yucatán Peninsula. Recorded from the states of Veracruz, Tabasco, Oaxaca, Campeche, Yucatán, and Quintana Roo (Cobá).

ANOLIS IMPETIGOSUS Cope


Type locality.—Unknown.

Range.—Unknown.

ANOLIS SCHMIDTI Smith


Type locality.—Manzanillo, Colima.

Range.—Known only from the type locality.
ANOLIS GADOVII Boulenger


_Type locality._—Tierra Colorada, Guerrero.
_Range._—Known only from the type locality.

ANOLIS DUNNI Smith

Anolis dunni Smith, Copeia, 1936, p. 9.—Smith and Spieler, Copeia, 1945, pp. 165-168.

_Type._—EHT-HMS No. 1506; E. H. Taylor collector.
_Type locality._—Between Rincón and Cajones, Guerrero (a place now called Agua del Obispo).
_Range._—Known only from the type locality.

ANOLIS TAYLORI Smith and Spieler

Anolis taylori Smith and Spieler, Copeia, 1945, pp. 165-168.

_Type._—U.S.N.M. (H. M. Smith field No. 10085); E. H. Taylor collector.
_Type locality._—One mile north of Acapulco, Guerrero.
_Range._—Known only from the type locality.

ANOLIS DAMULUS Cope


_Type locality._—Unknown.
_Range._—Unknown.

ANOLIS PENTAPRION Cope


_Type._—Formerly in U.S.N.M., now lost; A. Schott collector.
_Type locality._—Truando River, Colombia.
_Range._—Atlantic slopes from the Isthmus of Tehuantepec to Colombia. Known in Mexico only from Chiapas: Palenque.
ANOLIS UТОWANAE Barbour

Anolis utowanae Barbour, Copeia, 1932, p. 11.

Type.—Mus. Comp. Zool. No. 31035; Thomas Barbour collector (purchased from Indian).

Type locality.—Ten miles north of Mazatlán, Sinaloa.

Range.—Coastal Sinaloa, known only from the type locality.

ANOLIS CYMBOPS Cope


Type locality.—“Veracruz,” Mexico.

Range.—Known only from the type locality.

ANOLIS BECKERI Boulenger


Type.—Royal Belg. Mus., two cotypes; A. Boucard, collector.

Type locality.—Yucatán.

Range.—Yucatán Peninsula, including British Honduras. Recorded in Mexico only from Yucatán: Chichen Itzá.

ANOLIS BACCATUS Bocourt


Type.—Mus. Hist. Nat. Paris; Auguste Sallé collector.

Type locality.—Mexico.


ANOLIS LIOGASTER Boulenger


Type locality.—Omilteme, Guerrero (7,600 feet).

Range.—Known only from the type locality.29

ANOLIS LAEVIVENTRIS (Wiegmann)

Dactyloa (Anolis) laeviventris Wiegmann, Herpetologia Mexicana, 1834, p. 47.


Anolis wiegmanni Fitzinger, Systema reptilium, 1843, p. 67 (substitute name for Dactyloa laeviventris Wiegmann).


Type locality.—Mexico, here restricted to Jalapa, Veracruz.

Range.—Atlantic slopes from Central Veracruz to the Isthmus of Tehuantepec. Recorded from the states of Veracruz: Jalapa, Orizaba, Mirador, Actopam, Jicaltepec, Xico; and Tabasco.41

ANOLIS SCHIEIID (Wiegmann)


Type locality.—Mexico (by inference).

Range.—Unknown.43

ANOLIS GUNTERII Bocourt

Anolis guntherii Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 2, 1873, pp. 61–62, pl. 15, fig. 15.

40 The original type series included two, a male and a female, in the Brit. Mus. Nat. Hist., from Cobán, Guatemala; one, U.S.N.M. No. 12206, from Arriba, Costa Rica, and one, U.S.N.M. No. 6117, now lost, from Jalapa, Veracruz. Dunn (Proc. New England Zool. Club, vol. 12, 1930, p. 18), in discussing A. intermedius of Central America, mentions "the type of nannodes, U.S.N.M. No. 12206, Costa Rica," stating that it is the same as intermedius. We regard this action as constituting a restriction of the name to intermedius and rendering U.S.N.M. No. 12206 the lectotype of nannodes. The Mexican specimen obviously represents laeviventris, while the Guatemalan species represented by the Cobán specimens remains unnamed. Stuart (Misc. Publ. Mus. Zool. Univ. Michigan, No. 69, 1948, p. 50) restricts the name nannodes to the Guatemalan species, inasmuch as a name is available for each of the other two species (now laeviventris of Mexico, intermedius of Costa Rica) represented in the cotypes. This procedure certainly is the most desirable, but in view of the fact that Dunn previously restricted the name to the Costa Rican species, Stuart's course cannot, apparently, be maintained. The name Anolis stuarti Smith, new name, is proposed for the Guatemalan species represented by the two cotypes of nannodes in the British Museum of Natural History, from Cobán, Alta Verapaz. For a description of the types, see Boulenger, Cat. Liz. Brit. Mus., vol. 2, 1885, p. 78. The Anolis nannodes (naut.) from numerous localities in Mexico is the species laeviventris.—H. M. S.

41 In various papers (first in Proc. Acad. Nat. Sci. Philadelphia, 1886, p. 123) Cope records this species from Yucatán and from "Tehuantepec"; Dufré (La Naturaleza, ser. 2, vol. 2, 1896, p. 479) records it from Tangancicuaro (Michoacán), Guadalajara (Jalisco), and Cerro de los Amoles (state?)'; Moquard (Bull. Soc. Phîlom. Paris. ser. 9, vol. 1, 1899, pp. 155–156) records it from Guadalajara, Jalisco; and Peters (Monatsh. Berl. Acad., 1899, p. 874) records it from Matamoros, Puebla. We believe all these records refer to other species with smooth ventrals. We are unable at this time to place them accurately.

42 Bocourt possessed for his description the type of schiedii and a series of specimens from Cobán, Guatemala; all the latter are, however, apparently referable to A. cobanensis Stuart.

43 Cope's record (U. S. Nat. Mus. Bull. 32, 1887, p. 31) for Jalapa, Veracruz, is highly questionable.

Type.—Mus. Milano.

Type locality.—Mexico.

Range.—Unknown.

ANOLIS MILLERI Smith, new species


Type.—U.S.N.M. No. 120957; Walter S. Miller collector.

Type locality.—Quetzaltepec, Oaxaca.

Range.—Known only from the type locality.

ANOLIS LIMIFRONS RODRIGUEZII Bocourt


Type locality.—Panzós, Alta Verapaz, Guatemala.

Range.—Atlantic slopes from the Isthmus of Tehuantepec to Honduras, including the peninsula of Yucatán except its semiarid northwestern coast. Recorded in Mexico from Oaxaca (no locality); Chiapas: Palenque; Campeche: Balchacaj, Champotón, Tres Brazos, Encarnación; Yucatán: Citilpech, Chichen Itzá, Mérida; and Quintana Roo: Cozumel Island.

44 Diagnosis.—As in the accompanying key (p. 57). Differs from schiedii in two important respects: The head length from snout to ear (15.8 mm.) is distinctly greater than tibia length (13.6 mm.), and the interparietal is much larger, its length equaling the diameter of ear opening (the tibia, 15.5 mm., is longer than head, 13.5 mm., and length of the interparietal is little more than half the greatest diameter of the ear opening, in schiedii). A. schiedii is unique, in its group of the genus, in its very long tibia.

Description.—A lengthy description of the type was published in Smith and Laufe (loc. cit.) and needs no repetition.—H. M. S.

46 Spoiled rodriguezii by error.

46 The type, according to Bocourt’s figure and description, does not differ from the form most authors in the past have called aureolus, except perhaps in size of the interparietal. It appears probable to us that the deceptive marking on the type of rodriguezii (a broad vertebral light stripe), which Bocourt apparently thought was constant, may have been responsible for his failure to consider rodriguezii and the type of his rubigenosus as conspecific.

Although no other record indicates the occurrence of rodriguezii in Oaxaca, the known distribution elsewhere strongly implies occurrence on Atlantic slopes of that state near or at the Isthmus of Tehuantepec.
ANOLIS CAPITO Peters


**Type.**—Zool. Mus. Ber. No. 4086, two cotypes; C. Hoffman collector.

**Type locality.**—Costa Rica, here restricted to Palmar.

**Range.**—Tabasco to Panama. Recorded in Mexico only from “Tabasco.”

ANOLIS PETERSII Bocourt


**Type locality.**—Alta Verapaz, Guatemala, here restricted to Finca Samac.

**Range.**—Atlantic slopes from central Veracruz to Alta Verapaz in Guatemala. Recorded in Mexico only from Veracruz: Cuautlapan, Mirador, Orizaba; *San Luis Potosí*: Xilitla.

ANOLIS BIPORCATUS (Wiegmann)

*Dactyloa* biporcat Wiegmann, Herpetologia Mexicana, 1834, p. 47.


**Type.**—Zool. Mus. Berlin; F. Deppe collector.

**Type locality.**—Mexico, here restricted to Piedra Parada, Chiapas.

**Range.**—Chiapas to Panama. Known in Mexico only from Chiapas (Piedra Parada).

ANOLIS NEBULOSUS (Wiegmann)

*Dactyloa* nebulosa Wiegmann, Herpetologia Mexicana, 1834, p. 47.


**Type.**—Zool. Mus. Berlin, two cotypes; F. Deppe collector.
Type locality.—Mexico (by inference), here restricted to Mazatlán, Sinaloa.

Range.—Western Mexico, from Sinaloa to the Isthmus of Tehuantepec. Known from the states of Morelos, Puebla, Nayarit, Colima, Oaxaca, Guerrero, Michoacán, Sinaloa, Jalisco, Durango, and the Tres Marías Islands.47

**ANOLIS NEBULOIDES** Bocourt


Type locality.—Putla, Oaxaca.

Range.—Western Mexico from Chihuahua to Oaxaca. Recorded from the states of Chihuahua, Sonora, Colima, Guerrero, Puebla, and Oaxaca.48

**ANOLIS KIDDERI** Ruthven


Type.—Mus. Zool. Univ. Michigan No. 72851; Edwin P. Creaser collector.

Type locality.—Mérida, Yucatán.

Range.—The western part of the Yucatán Peninsula. Recorded from Yucatán and Campeche.

**ANOLIS USTUS** Cope


Type locality.—Belize, British Honduras.

Range.—The Yucatán Peninsula and the area at its base. Recorded in Mexico from Yucatán and Campeche.

**ANOLIS LEMURINUS BOURGEAEI** Bocourt


47 Records from the states of Veracruz and Yucatán are probably erroneous, and that from Chihuahua is said to be referable to *A. nebuloides*. Virtually all records for both species are, for that matter, open to question. The two forms have never been adequately characterized, and very likely more than two species are involved.

48 Undoubtedly some records of *nebuloides* belong here, and perhaps *vice versa*. A record for Veracruz is probably in error.
Anolis biporcatus, Bocourt⁴⁶ (née Wiegmann), Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 2, 1873, pp. 98–99, pl. 15, fig. 8.—Boulenger, op. cit., pp. 88–89.


Type.—Mus. Hist. Nat. Paris, one female (Orizaba, collected by Bourgeau); Zool. Mus. Berlin, one male (Huatusco, collector?).

Type locality.—Huatusco and Orizaba, Veracruz, here restricted to Huatusco.

Range.—Atlantic slopes from Veracruz to Guatemala and British Honduras. Recorded in Mexico from the states of Veracruz, Tabasco, Campeche, and Yucatán.

ANOLIS CUMINGII Peters


Type.—Zool. Mus. Berlin; Herr Cuming collector.

Type locality.—Mexico.

Range.—Uncertain; possibly the upper Balsas Basin. Recorded only from La Paz (Puebla?).

ANOLIS HELIACTIN Cope


Type locality.—Mexico.

Range.—Recorded only from “Oaxaca.”

ANOLIS SERICEUS Hallowell


⁴⁶ And of other authors prior to Schmidt, 1941; see synonymy of A. biporcatus.

⁵⁰ According to Stuart (op. cit., 1948, p. 51) the type and several of the paratypes of A. u. veraepacis Barbour are referable to A. bourgeaei while the remaining paratypes are A. sericeus.
Anolis jacobij Bocourt, op. cit., p. 74, pl. 13, fig. 8 (Mus. Hist. Nat. Paris; Veracruz, here restricted to Veracruz).

Type.—Acad. Nat. Sci. Philadelphia, now lost; Mr. Pease donor. 
Type locality.—El Encero de Jalapa, Veracruz.
Range.—Atlantic slopes from Tamaulipas and Pacific slopes from the Isthmus of Tehuantepec south to Nicaragua. Recorded in Mexico from the states of Tamaulipas, San Luis Potosi, Veracruz, Oaxaca, Tabasco, and Chiapas.

Genus CORYTHOPHANES Boie


Genotype.—Agama cristata Merrem.
Species.—Three.
Range.—Central Veracruz and the Pacific coast of Chiapas southeastward to Costa Rica.

KEY TO SPECIES OF CORYTHOPHANES

1. Nuchal crest interrupted, not continuous with dorsal crest.  hernandezii (p. 68) Nuchal crest continuous with dorsal, although sometimes low where the two meet. 
2. Parietal crests joining very near extremity of helmet; upper head scales keeled or rugose. percarinatus (p. 69) Parietal crests joining about halfway from orbits to extremity of helmet; upper head scales smooth cristatus (p. 69)

CORYTHOPHANES HERNANDEZII (Wiegmann)


Corythophanes mexicanus Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 3, 1874, pp. 122-123, pl. 17, fig. 1 (substitute name for Chamaeleopsis hernandezii Wiegmann, adopting Hernández's pre-Linnaean name Chamaeleo Mexicanus).52

51 The author of this generic name, and of the specific name as well, has been accepted as Gray by some authors, as Wiegmann by others. Wiegmann published his own description in 1831, in great detail; Gray published but a few words, also in 1831, but attributed both names to "Wiedemann, MSS" (in error; = Wiegmann). Even though Gray's description may have preceded Wiegmann's, it seems obvious that Wiegmann was ethically the author, having provided the only reasonable description, and his manuscript having been acknowledged by Gray as the source of the latter's information. It is true that Gray stated specimens were in the British Museum (Boulenger cites one purchased by Gray), so that Gray could well have drawn up his description from them and not from Wiegmann's manuscript. The correct authorship thus remains debatable.
52 Wiegmann cited Chamaeleo Mexicanus of Hernández in both of his papers (1831 and 1834) but did not adopt the name.
Type.—Zool. Mus. Berlin; F. Deppe collector.

Type locality.—Mexico, here restricted to Jalapa, Veracruz.

Range.—Atlantic slopes from central Veracruz south and east to Guatemala. Recorded in Mexico from various localities in the states of Veracruz, San Luis Potosí (Xilitla), Oaxaca, Chiapas, Yucatán, and Quintana Roo (Cobá).

CORYTHOPHANES CRISTATUS (Merrem)

Agama cristata Merrem, Tentamen systematis amphibiorum, 1821, p. 50.


Type.—Unknown.

Type locality.—Ceylon (in error), here restricted to Orizaba, Veracruz.

Range.—Atlantic slopes from central Veracruz to Costa Rica. Reported in Mexico from the states of Veracruz, Campeche, Yucatán, and Chiapas (Usumacinta River across from Piedras Negras, Petén, Guatemala).

CORYTHOPHANES PERCARINATUS Duméris


Type locality.—Escuintla, Guatemala.

Range.—Pacific slopes of Guatemala and Chiapas, and Atlantic slopes in Alta Verapaz, Guatemala. Known in Mexico only from Colonia Hidalgo, 8 kilometers north of La Esperanza, Chiapas.

Genus LAEMANCTUS Wiegmann


Genotype.—Laemanctus longipes Wiegmann.

Range.—Lowlands from San Luis Potosí south to and including Guatemala, and British Honduras; ? Colima.

Species.—Four, all occurring in Mexico.

KEY TO MEXICAN SPECIES OF LAEMANCTUS

1. Scales on upper surface of snout much larger than those on occipital region... 2
   Scales on upper surface of snout not distinctly larger than those on occipital region; no dorsal crest; part of scales on body bicarinate or tricarinate; head yellowish above, the posterior contour black; body above dark purple or

---

13 While Boie designated Merrem’s species as type of his genus Corythophanes, he did not actually print the combination. That it was his intent so to spell the name is indicated in his article in vol. 21 (1828) of Isis von Oken, p. 363, where he (Bole) spells the name Corytophanes. Kaup (Isis von Oken, vol. 21, 1828, p. 1147) does likewise.
dark green, with blackish cross bands; a light lateral streak; lilac below; 690
mm. — deborreii (p. 71)

2. A series of projecting triangular scales on the posterior contour of the head; a
feeble dorsal crest. — U.S.'NM. Type
No. 308; Arthur Schott collector.

3. Scales around middle of body, 57-61; reddish or purplish above with dark brown
bands across body, most distinct in the vertebral region; a dark brown band
from eye to tympanum, sometimes continued along side of back; a white
streak from below the eye to forelimb, and from axilla to groin; a white spot
on each side of base of tail; lower surfaces pink; 730 mm.

longipes (p. 71)

LAEMANCTUS ALTICORONATUS Cope

Proc. Amer. Philos. Soc., 1869, pl. 11.—BoulenGer, Catalogue of the

Type. — U.S.N.M. No. 308; Arthur Schott collector.

Type locality. — Mérida, Yucatán.

Range. — Yucatán. Reported from Chichen Itzá and Mérida.

LAEMANCTUS SERRATUS Cope

pl. 21, fig. 4.—Bocourt, Mission scientifique au Mexique . . . , Études sur
les reptiles, livr. 3, 1874, pp. 114-116, pl. 17, fig. 4 (non Wiegmann, fide
Cope loc. cit.).


Type. — Mus. Leyden.

Type locality. — “Orizaba Valley, Mexico.”

Range. — Lowlands from extreme eastern Guanajuato to Yucatán.
Reported from Yucatán: Chichen Itzá; Oaxaca: “Oaxaca” (no specific
locality), Tlacolutla; Veracruz: Boca del Río, Jicaltepec, Misantla,
Orizaba; Guanajuato: Huasteca Potosina; Hidalgo: Zacualtipan; Cam-
peche: Champotón, Oxpemul; San Luis Potosi: Xilitla; Tamaulipas:
near Gómez Farias.

Mrs. Helen T. Gaige suggests the possibility that serratus and alticoronatus may be identical since spec-
imens from Yucatán and Campeche seem to show the characters of both (Carnegie Inst. Washington Publ.
No. 457, 1936, p. 296).
LAEMANCTUS DEBORREI Boulenger


Type.—Museum de Bruxelles, two adults; M. Linden collector (?).

Type locality.—Tabasco.

Range.—Atlantic slopes from Tabasco to British Honduras. Reported from Tabasco: La Venta; Oaxaca: Mountains near Santo Domingo (U. S. Nat. Mus.).

LAEMANCTUS LONGIPES Wiegmann


Type.—Zool. Mus. Berlin; F. Deppe collector.

Type locality.—Jalapa, Veracruz.


Genus BASILISCUS Laurenti


Oedicoryphus Wagler, Natürliches System der Amphibien, 1830, p. 148 (type, Basiliscus vittatus Wiegmann).


Genotype.—Basiliscus americanus Laurenti=(Basiliscus basiliscus (Linnaeus)).

Range.—Lowlands of Mexico south through Jalisco and Tamaulipas, through Central America to northwestern South America.

Species.—Five; one occurs in Mexico.

BASILISCUS VITTATUS Wiegmann


Type.—Zool. Mus. Berlin; F. Deppe collector.

Type locality.—Mexico, here restricted to Veracruz, Veracruz.

Range.—The coasts from Jalisco and southern Tamaulipas, south to Nicaragua. Reported from numerous localities in the states of Tamaulipas, Veracruz, Tabasco, Campeche, Yucatán, Quintana Roo, Chiapas, Oaxaca, Guerrero, Michoacán, Colima, and Jalisco.

Genus IGUANA Laurenti

Iguana Laurenti, Specimen medicum exhibens synopsin reptilium, 1768, p. 47.

Hypsilophus Wagler, Natürliches System der Amphibien, 1830, p. 147 (type, Lacerta iguana Linnaeus).

Genotype.—Lacerta iguana Linnaeus.

Range.—Mexico on the Pacific coast from Sinaloa to Chiapas, on the eastern coast north to central Veracruz. Absent on the higher parts of the plateau. Lowlands of Central America, and central and northern South America.

Species.—Two, one with two subspecies. Extralimital are Iguana iguana iguana and Iguana delicatissima of South America.

IGUANA IGUANA RHINOLOPHA Wiegmann


Type.—Zool. Mus. Berlin; F. Deppe collector.

Type locality.—Mexico, by inference here restricted to Córdoba, Veracruz.

Range.—Veracruz and Sinaloa southward along both coasts to Panama. Reported from Sinaloa: Mazatlán, Presidio; Nayarit: San Blas; Colima: Manzanillo, Tecomán, Colima; Jalisco: ?San Blas (possibly Nayarit); Michoacán: Hacienda California, Chacán River Sierra Madre, Jorullo; Guerrero: Pie de la Cuesta, Acapulco; Puebla: Chiautla; Oaxaca: Tehuantepec, Río Coatzacoalcos, ?Tetela, Agua Fría, Tequesistlán, Totolapan; Veracruz: Córdoba, Vega de Alatorre, vicinity of Cuatotolapam, Matacabestro, Puente Nacional; Tabasco: Tlacotalpan, Tenosique; Campeche: Ciudad del Carmen; Quintana Roo: Cozumel Island; Chiapas: La Esperanza.
Genus CTENOSAURA Wiegmann


Genotype.—_Ctenosaura cycluroides_ Wiegmann [= _C. acanthura_ (Shaw)].

Species.—Five, all but one of which (_C. bakeri_ of Utilla Island, Honduras) occur in Mexico.

Range.—Baja California, Sonora, and perhaps southern Arizona on the west, and Tamaulipas on the east, south along both coasts to Panama.

KEY TO MEXICAN SPECIES OF _CTENOSAURA_

1. Small scales between whorls of enlarged scales reduced to 1 row (and no more than 1 row) between at least a few whors (generally near middle of tail), often on entire distal half or more of tail.................................................. 2
2. Fifth whorl of enlarged scales on tail................................................................................................. 3

2. Fifth whorl of enlarged scales preceded by more than a single row of small scales................................................. _acanthura_ (p. 74)

3. Three rows of small scales, complete or incomplete, preceding each of the proximal 5 whors of enlarged caudal scales._________________________ _pectinata_ (p. 75)

3. Small scales reduced to only two rows preceding one or more of the proximal five caudal whors........................................................................... _similis_ (p. 73)

**CTENOSAURA SIMILIS SIMILIS** (Gray)

_Iguana (Ctenosaura) similis_ GRAY, _in_ Griffith, Animal Kingdom, by the Baron Cuvier, vol. 9, Synopsis . . ., 1831, p. 38.

_Cyclura (Ctenosaura) similis_, WIEGMANN, Herpetologia Mexicana, 1834, p. 42.


_Type._—Lost.

_Type locality._—Restricted (by Bailey) to Tela, Honduras.

_Range._—Coastal areas from the Isthmus of Tehuantepec on the

---

51 We are not totally in agreement with Bailey's treatment of the species in this genus, nor with some of the more recent records no doubt influenced by his work. The four forms we recognize are more or less completely allopatric; at least such suggestions as of _C. acanthura_ in Chihuahua and _C. similis_ in Guerrero appear to us wholly unwarranted. The only extensive overlapping of ranges of which we are aware occurs on the Atlantic coast and involves _C. acanthura_ and _C. similis_.

55 Bocourt (loc. cit.) states that the types are from "Guatemala" and "La Unión," El Salvador. Barbour and Loveridge (Bull. Mus. Comp. Zool., vol. 69, 1929, p. 247), however, state that a cotype in the Mus. Comp. Zool. is from "Mexico." The type locality is here restricted to La Unión.
Pacific and central Veracruz on the Atlantic south to Panama. Recorded in Mexico from the states of Veracruz, Tabasco, Campeche, Yucatán, Quintana Roo (including Mujeres and Cozumel Islands), Oaxaca, and Chiapas (Tonalá, Palenque).\textsuperscript{57}

**CTENOSAURA ACANTHURA (Shaw)**


*Iguana (Clatosaura) armata* Gray, in Griffith, Animal Kingdom, by the Baron Cuvier, vol. 9, Synopsis . . . 1831, p. 38 (type lost; no type locality cited, here restricted to Tampico, Tamaulipas).

*Cyclus shawii* Gray, loc. cit. (synonymic name for *Iguana (Clatosaura) acanthura* Gray).

*Iguana (Clatosaura) lanceolata* Gray, loc. cit. (type lost; no type locality cited, here restricted to Tampico, Tamaulipas).

*Iguana (Clatosaura) bellii* Gray, loc. cit. (type lost; no type locality cited, here restricted to Tampico, Tamaulipas).

*Cyclus articulata* Wiegmann, Herpetologia Mexicana, 1834, p. 43 (types unknown; type locality, Mexico).

*Cyclus denticulata* Wiegmann, ibid., pp. 43–44 (substitute name for *Clatosaura cycluroides* Wiegmann).

*Cyclus semicolorata* Fitzinger, Systema reptilium, 1843, p. 56 (synonymic name for *Cyclus denticulata* Wiegmann).\textsuperscript{58}


**Type locality.**—Not given by Shaw. Boulenger (Catalogue of the lizards in the British Museum, vol. 2, 1885, p. 196) gives “California,” which probably means Baja California. Since the species does not occur there we accept Bailey’s restriction to Tampico, Tamaulipas.

**Range.**—Atlantic slopes from central Tamaulipas south to the Isthmus of Tehuantepec. Reliably reported only from the states of Tamaulipas, San Luis Potosí (Pujal), Veracruz, and Oaxaca.\textsuperscript{59}

\textsuperscript{57} Burt and Myers (Stanford Univ. Publ., biol. ser., vol. 8, 1942, p. 290) record the species, in error, from Acapulco, Guerrero.

\textsuperscript{58} Fitzinger (loc. cit.) attributes this name to Wiegmann, as do also Lichtenstein and Martens (Nomenclator . . . Berolimensis, 1856, p. 8): we are, however, unable to locate Wiegmann’s use of the name and attribute it to Fitzinger, as did Bailey, Proc. U. S. Nat. Mus., vol. 73, 1928, p. 10.

\textsuperscript{59} Reexamination of specimens in the U. S. Nat. Mus, referred by Bailey (loc. cit.) to *C. acanthura* reveals that two other species were confused with *C. acanthura*. One specimen from Mirador, Veracruz, is a *C. similis*; and all from Uriapan, Michoacán; Tehuantepec, Oaxaca (except Nos. 26341, 55498); Isabel Island; Tres Marías Islands; Culcatlán, Oaxaca; Tlapa and Balsas, Guerrero, are *C. pectinata*. That all other west-coast specimens in his list, from other museums, are also *C. pectinata* we believe is virtually a certainty.
Cyclura pectinata Wiegmann, Herpetologia Mexicana, 1834, p. 42, pl. 2.


Type.—Mus. Zool. Berlin. No. 574; F. Deppe collector.

Type locality.—Restricted to Colima, Colima.

Range.—Pacific coast from southern Sinaloa southward to the Isthmus of Tehuantepec. Recorded from Isabel and the Tres Marías Islands and the states of Sinaloa, Durango (Ciudad, Ventanas), Nayarit, Jalisco, Colima, Michoacán, Morelos, Guerrero, Puebla, and Oaxaca. A record for the state of Guanajuato is unacceptable.


Ctenosaura insulana Dickerson, op. cit., pp. 462–463 (U.S.N.M. No. 64430; C. H. Townsend collector; Cerralvo Islands, Baja California).

Type.—U.S.N.M. No. 529, four cotypes; John Xantus collector.

Type locality.—Cape San Lucas, Baja California.

Range.—The southern two-thirds of Baja California, central Sonora (including San Pedro Nolasco Island) south to northern Sinaloa, and southwestern Chihuahua. Recorded only from the states cited. It is possible that the species extends as far north as southern Arizona.

Genus ENYALIOSAURUS Gray

Enyaliosaurus Gray, Catalogue of the specimens of lizards in the collection of the British Museum, 1845, p. 192.

861316—50—6

Genotype.—Cyclura quinquecarinata Gray.

Range.—Michoacán to the Isthmus of Tehuantepec, the Peninsula of Yucatán, and northern Guatemala.

Species.—Five, all but one of which (C. palearis of Guatemala) occur in Mexico.

KEY TO MEXICAN SPECIES OF ENYALIOSAURUS

1. Whorls of enlarged caudal scales not interspersed with small scales. defensor (p. 77)

2. Dorsal and lateral caudal scales of nearly uniform character in each whorl. erythromelas (p. 77)

3. Median row of dorsal scales extending to or almost to sacrum, noticeably raised, of medium height; tail twice length of body. quinquecarinata (p. 76)

Median row of dorsal scales extending only two-thirds distance to sacrum and not noticeably raised; tail about 1½ times body length. clarki (p. 76)

ENYALIOSAURUS QUINQUECARINATUS (Gray)


Type locality.—Unknown (restricted to Tehuantepec, Oaxaca, by Bailey, loc. cit.).


ENYALIOSAURUS CLARKI (Bailey)


Ctenosaura clarki BAILEY, Proc. U. S. Nat. Mus., vol. 73, art. 12, 1928, pp. 44-46, pl. 27.

Type.—Mus. Comp. Zool. No. 22454; Hans Gadow collector.

Type locality.—Ovopeo, Michoacán, elevation 1,000 feet.

Range.—Known definitely only from the type locality; a record for Cuautla, Morelos, probably belongs here (Dugès, La Naturaleza, ser. 2, vol. 2, 1896, p. 480).
ENYALIOSAURUS ERYTHROMELAS (Boulenger)


Ctenosaura (Cachryx) annectens Werner, Jahrb. Hamburg Wiss. Anst., pt. 2, 1911, p. 25 (locality unknown; P. Phol collector, Hamburg Museum; "Mexico").

_Type locality._—Unknown; here restricted to Balchacaj, Campeche.
_Range._—Known only from Balchacaj, Campeche.

ENYALIOSAURUS DEFENSOR Cope


_Type._—U. S. N. M. No. 12282; male, adult, and half grown, three cotypes; Arthur C. V. Schott collector.
_Type locality._—Yucatán, here restricted to Chichen Itzá.
_Range._—Northern part of the Yucatán Peninsula. Reported only from Chichen Itzá, Yucatán.

Genus DIPSOSAURUS Hallowell


_Genotype._—Crotaphytus dorsalis Baird and Girard.
_Range._—Sonora, Baja California, and some coastal islands of the Gulf. In United States: southern and central Arizona, Colorado and Mojave Deserts east to the Colorado River, southern Nevada, and Utah.
_Species._—Three, with five forms recognized.

KEY TO MEXICAN SPECIES OF DIPSOSAURUS

1. Two or more scale rows between nostril and rostral (62–83 percent); frequently with longitudinal dark lines on the body________dorsalis dorsalis (p. 78)
   One scale row between nostril and rostral (75 to 80 percent); dark lateral lines usually lacking_________________________ 2

2. Dorsal pattern consisting of large, round, white spots, and no longitudinal lines; brown reticulation on throat enclosing round or oval cream spots.
   dorsalis sonoriensis (p. 79)
   Dorsal pattern consisting of short longitudinal streaks, and few if any light spots, which are not dark-edged; throat either striped with dark longitudinal streaks or without streaks, in which case large rounded spots are present.  3
3. Gular region without longitudinal dark streaks; large rounded spots on lower jaw and side of neck; central gular region suffused with dark brown; femoral pores average 19.1........................................catalinensis (p. 78)
Gular region with longitudinal dark streaks, without definite rounded light spots laterally, sometimes suffused with dark centrally....................4
4. Femoral pores fewer, average 18.46..........................dorsalis lucasensis (p. 78)
Femoral pores more numerous, average 21.8.........................carmenensis (p. 78)

DIPSOSAURUS CARMENENSIS Van Denburgh


Type.—California Acad. Sci. No. 50504; Joseph R. Slevin collector.
Type locality.—Carmen Island, Baja California.
Range.—Carmen Island and Los Coronados Islands, Baja California.

DIPSOSAURUS CATALINENSIS Van Denburgh


Type.—California Acad. Sci. No. 50505; Joseph R. Slevin collector.
Type locality.—Santa Catalina Island, Baja California.
Range.—Known only from the type locality.

DIPSOSAURUS DORSALIS LUCASENSIS Van Denburgh


Type.—California Acad. Sci. No. 46090.
Type locality.—“San José del Cabo, Baja California, Mexico.”
Range.—Southern tip of Baja California, southward from Santa Margarita Island; Cerralbo Island. Reported from Miraflores, San José del Cabo, Santa Anita, Agua Caliente, Eureka, etc.

DIPSOSAURUS DORSALIS DORSALIS (Baird and Girard)

Dipsosaurus dorsalis, Hallowell, ibid., vol. 7, 1854, p. 92.

Type.—U.S.N.M. No. ; John Le Conte collector.
Type locality.—“Desert of Colorado, Cal.,” here restricted to Winter-haven (=Fort Yuma), Imperial County.
Range.—Extreme southwestern Utah and eastern California southward into northwestern Sonora, and eastern Baja California south to Magdalena Island; recorded on the latter island and on those of San Luis, Ángel de la Guarda, San Marcos, Monserrate, and San José. Reported from Baja California: Volcano Lake, San Felipe Bay, 13 miles northwest of El Mejor, El Cajón Cañon, San Ignacio, 600 feet, etc.; Sonora: Punta Peñasco.
**DIPSOSAURUS DORSALIS SONORIENSIS** Allen


**Type locality.**—Hermosillo, Sonora.

**Range.**—Gulf slope of Sonora, except extreme northwestern part, and southward into northern Sinaloa. Reported from Sonora: 5 miles southwest of Hermosillo, Miramar, La Posas, Guaymas, Rancho Costa Rica, Bocoit; Sinaloa: Ahome.

**Genus SAUROMALUS** Duméril


**Genotype.**—*Sauromalus ater* Duméril.

**Species.**—Seven, one with three subspecies.

**Range.**—Southern Utah and southern Nevada south through southern California, western Sonora and the Gulf slopes of Baja California.

**KEY TO SPECIES OF SAUROMALUS**

1. One or more transverse bands dorsally, across body or rump.  
   No transverse body bands present.  
   2. Largest nuchal scales equally as large as or larger than frontal plates.  
   hispidus (adult) (p. 80)  
   Largest nuchal scales smaller than frontal plates.  
   3. Dorsal pattern of large, irregular, dark brown or black blotches on a yellowish ground color.  
   varius (p. 81)  
   Dorsal pattern of small, dark brown or black spots on a gray ground color.  
   klauberi (p. 80)  
   4. Ventral scale rows usually less than 130.  
   Ventral scale rows usually more than 130.  
   5. Dorsal scales in a head length usually less than 20.  
   hispidus (juvenile) (p. 80)  
   Dorsal scales in a head length usually more than 20.  
   slevini (p. 80)  
   6. Transverse body bands with light centers and dark brown or black borders giving a double-banded effect.  
   Transverse bands unicolor.  
   7. Scales around middle of upper foreleg (humeral scales) usually fewer than 50.  
   Humeral scales more than 50.  
   obesus obesus (p. 81)  
   8. No reddish suffusion on dorsal and ventral areas in adult males; maximum adult length averaging somewhat less than in *o. tumidus*; average scale counts lesser.  
   More or less brilliant reddish suffusion on dorsal and ventral areas in adult males; maximum adult length averaging more than in *o. townsendi*; average scale counts greater.  
   obesus townsendi (p. 81)  
   obesus tumidus (p. 81)  
   9. Ventral scale rows 151 or more.  
   Ventral scale rows usually less than 151.  

---

SAUROMALUS HISPIDUS Stejneger


Type.—U.S.N.M. No. 8563; Thomas H. Streets collector.

Type locality.—Ángel de la Guarda Island, Baja California.

Range.—Ángel de la Guarda, Smith, Pond, Nuevo Amor, Granite, Mejía, and South San Lorenzo Islands, Gulf of California, Baja California.

SAUROMALUS KLAUBERI Shaw


Type.—L. M. Klauber No. 6859.

Type locality.—Santa Catalina Island, Baja California.

Range.—The type locality.

SAUROMALUS SLEVINI Van Denburgh


Type.—California Acad. Sci. No. 50503, adult male; Joseph R. Slevin collector.

Type locality.—South end of Monserrate Island, Baja California.

Range.—Monserrate, Carmen, and Coronados Islands, Baja California.

SAUROMALUS AUSTRALIS Shaw


Type.—L. M. Klauber No. 30170; Robert S. Hoard collector.

Type locality.—San Franciscuito Bay, Baja California.

Range.—The mainland of central and southern Baja California on the Gulf coast.

SAUROMALUS ATER Duméril


Type.—Mus. Hist. Nat. Paris; M. Jaurès collector? ("qui a été donné . . . par M. Jaurès, lieutenant à bord de la frégate la Danaïde").

Type locality.—Unknown; presumably one of the islands in the southern part of the Gulf of California, Baja California, here restricted to Espíritu Santo Island.

Range.—Espíritu Santo, Isla Partida, San Francisco, San Diego, Santa Cruz, and San Marcos Islands, Gulf of California, Baja California.
SAUROMALUS VARIUS Dickerson


**Type.**—U.S.N.M. No. 64441; C. H. Townsend collector.

**Type locality.**—San Esteban Island, Gulf of California.

**Range.**—The type locality.

SAUROMALUS OBESUS TOWSENDEI Dickerson


**Type.**—U.S.N.M. No. 64442; C. H. Townsend collector.

**Type locality.**—Tiburón Island, Gulf of California.

**Range.**—Tiburón Island, Sonora, and adjacent coast of Sonora at least as far south as Guaymas and east to the vicinity of Hermosillo.

SAUROMALUS OBESUS TUMIDUS Shaw


**Type.**—L. M. Klauber No. 27323; L. M. Klauber collector.

**Type locality.**—Telegraph Pass, Gila Mountains, Yuma County, Ariz.

**Range.**—Extreme northwestern Sonora, southwestern and south central Arizona. Intergrades only, with *S. o. townsendi*, are recorded from Mexico; typical specimens undoubtedly occur in extreme northwestern Sonora and extreme northeastern Baja California, along the Colorado River.

SAUROMALUS OBESUS OBESUS (Baird)


**Type.**—U.S.N.M. No. 4172; G. H. Thomas collector (?)..

**Type locality.**—Fort Yuma, Calif.

**Range.**—Southeastern California, southern Nevada, southern Utah, western Arizona, and extreme northern Baja California. Although known from numerous localities within a few hundred feet of the Baja California line, none have actually been taken in that state.

Genus HOLBROOKIA Girard


Genotype.—Holbrookia maculata Girard.

Range.—Northern states of Mexico south to San Luis Potosí and Durango. In the United States: Texas, New Mexico, and Arizona.

Species.—Four, with twelve forms, of which ten are known to occur or are to be expected in Mexico.

KEY TO MEXICAN SPECIES OF HOLBROOKIA

1. Tail flat, with broad, black, ventral bands; lateroventral black marks placed far posteriorly, continued dorsally above lateral fold, and slanting anteroventrally on belly.................................................. \textit{texana} (p. 85)
Tail rounded; no ventral subcaudal marks save small spots in one subspecies; lateroventral marks placed farther anteriorly, not extending upon dorsal surface, and slanting posteroverventrally on belly.......................... 2
2. Several black spots under tail; dorsal pattern of large, well-defined spots equally sharp-edged on all sides; no light dots in pattern. \textit{maculata lacerata} (p. 83)
   No black spots under tail; pattern not as described.......................... 3
3. Dorsal body scales distinctly keeled, rather small.......................... 11
   Dorsal body scales not keeled (although sometimes pointed and somewhat convex), larger, flat.................................................. 4
4. Tail generally shorter than the snout-vent measurement, usually not over 51 percent of total length (the higher proportions are of males)........ 5
Tail generally as long or longer than snout-vent measurement, 50–58 percent of total length (lower proportions are of females)................. 7
5. Enlarged supraoculars in contact with frontals. \textit{maculata elegans} (p. 84)
   Enlarged supraoculars separated from frontals by a series of small scales. 6
6. Size greater, maximum 71 mm., average 59 mm. snout to vent; in males a large blue patch surrounding lateroventral black spots on belly. \textit{maculata dickersonae} (p. 83)
   Maximum size 61 mm., average 50 mm. snout to vent; in males a smaller blue patch, or none, partially surrounding lateroventral black spots on belly.................................................. \textit{maculata approximans} (p. 83)
7. Femoral pores usually less than 12.............................................. 8
   Femoral pores usually 12 or more............................................. 10
8. Upper labial region strongly flared, almost flat......................... \textit{bunkeri} (p. 84)
   Upper labial region less flared, obtuse........................................ 9
9. Scales on limbs distinctly keeled; upper labials strongly imbricate. \textit{maculata pulchra} (p. 84)
   Scales on limbs smooth; upper labials less strongly imbricate. \textit{maculata dickersonae} (p. 83)
10. Femoral pores about two-thirds as wide as pore scales are long in adult males, about one-half in adult females......................... \textit{maculata elegans} (p. 84)
   Femoral pores about one-half as wide as the pore scales are long, in adult males, about one-fourth in adult females. \textit{maculata thermophila} (p. 84)
11. Anterior gulars a transverse row of 4 subequal scales, the inner pair not separated by linear or rounded scales...................... \textit{propinqua piperata} (p. 85)
   Not so......................................................................................... \textit{propinqua propinqua} (p. 85)
HOLBROOKIA MACULATA APPROXIMANS Baird


Type.—Lost.

Type locality.—Lower Rio Grande of Tamaulipas (fide Baird, United States-Mexican boundary survey, Reptiles, 1859, p. 8).

Range.—Western Texas to eastern Arizona, and from southeastern Utah southward through Chihuahua and central Coahuila, and southward in the west to Jalisco. Reported from Chihuahua: 35 and 40 miles south of Moctezuma, between Churo and Chiricahui, Ojos del Diablo, Río Santa María, Ramos, Casas Grandes; Coahuila: ? Concordia, Sierra de Santa Rosa, Buena Vista, Hda. La Rosita; San Luis Potosi: 5 miles northeast of Cándido Navarro; Sonora: headwaters of San Pedro; Durango; El Oro; Jalisco: Colotlán.82

HOLBROOKIA MACULATA DICKERSONAE Schmidt


Type.—U.S.N.M. No. 2668A; Lt. D. N. Couch collector.

Type locality.—Castañuelas, Coahuila.

Range.—The northern portion of the central plateau, as far south as Guanajuato, and adjacent areas immediately northward in southern Coahuila and Durango. Reported from Coahuila: Castañuelas, Hipólito, Jaral, Agua Nueva, Jimulco, Saltillo, álamo de Parras, 10 miles east of Torreón; Durango: near Avilco, 5 miles north of Conejos, 25 miles north of Bermejillo, Gómez Palacio; San Luis Potosi: Jesús María; Zacatecas: La Colorada, Berriozábal; Guanajuato: San Felipe.

HOLBROOKIA MACULATA LACERATA Cope


81 The type locality for this name strongly indicates that the species involved is H. propinqua, since that is the only one of this group whose range reasonably may include the state of Tamaulipas. Nevertheless, since the description better fits H. maculata, and since Baird presumably well knew H. propinqua (having described it also), we retain the previous allocation of the name H. approximans pending further investigations.

82 A number of records for "Tamaulipas," without specific locality, are regarded erroneous either by misidentification or incorrect locality data.
Type.—U.S.N.M. No. 10160.
Type locality.—Erath County, Tex., west of the Upper Brazos; on the Guadalupe River in Kendall or Comal County. Here restricted to Erath County, Tex.
Range.—Central Texas plains region south to northern Coahuila. Recorded in Mexico only from Coahuila: Sabinas.

HOLBROOKIA MACULATA PULCHRA Schmidt


Type.—Amer. Mus. Nat. Hist. No. 14777; R. D. Camp collector.
Type locality.—Carr Canyon, Huachuca Mountains, Ariz., 5,200 feet.
Range.—Known from the general region of the type locality; to be expected in adjacent Chihuahua and Sonora.

HOLBROOKIA MACULATA THERMOPHILA Barbour


Type locality.—San José de Guaymas, Sonora.
Range.—Sonora; possibly in northern Sinaloa. Reported from Sonora: 8 miles south of Nogales, 53 miles south of Nogales, 8 miles south of Magdalena, Canoa, near Noria, 30 miles south of Noria, 5 miles southwest of Hermosillo, 54 miles south of Hermosillo, 10 miles north of Guaymas, Batamotal, Álamos, Guirocoba. ?Sinaloa: ?Bacubirito.

HOLBROOKIA MACULATA ELEGANS Bocourt

Holbrookia elegans Bocourt, Mission scientifique au Mexique . . . , Études sur les reptiles, livr. 3, 1874, pp. 164-165, pl. 17 bis, figs. 8, 8a.

Type locality.—Mazatlán, Sinaloa.
Range.—Sinaloa. Reported from Presidio de Mazatlán, Mazatlán, Escuinapa, ?Bacubirito, Rosario.

HOLBROOKIA BUNKERI Smith


Type locality.—15 miles south of Juárez, Chihuahua.

Range.—Known only from the type locality.

**HOLBROOKIA PROPINQUA PROPINQUA** Baird and Girard


Type.—Unknown.

Type locality.—Between Indianola and San Antonio, Tex., here restricted to 9 miles southwest of Somerset, Atascosa County.

Range.—Central and southern Texas, southward presumably into Tamaulipas. No definite record for Mexico is known.

**HOLBROOKIA PROPINQUA PIPERATA** Smith and Burger


Type locality.—Beach at Etiopa, 2 miles south of Tecolutla, Veracruz.

Range.—Presumably southern Tamaulipas and northern Veracruz; recorded only from the type locality.

**HOLBROOKIA TEXANA** (Troschel)


Type.—Not known.

Type locality.—New Braunfels, on the Guadalupe River, Tex.; latitude 28° N. (=New Braunfels, Comal County, Tex.).

Range.—Southeastern Arizona to central Texas and southward through the northern parts of all the border states in Mexico except Baja California. Reported from Tamaulipas, San Luis Potosí, Nuevo León, Coahuila, Chihuahua, Durango, and Sonora.

**Genus CALLISAURUS** Blainville


*Megadactylus* Fitzinger, Systema reptilium, 1843, p. 59 (type, *Callisaurus draconoides* Blainville).

Genotype.—Callisaurus draconoides Blainville.

Range.—Southwestern United States, southward from Nevada to Baja California, Sonora, and adjacent islands.

Species.—One species with 10 forms; only one (C. d. myurus Richard-ardson of Nevada) is extralimital.

KEY TO MEXICAN SPECIES OF CALLISAURUS

1. A fringe of small spinose scales along one side of toes somewhat as in Uma. draconoides crinitus (p. 87)

2. Males with two nearly vertical lateral black blotches usually followed by a third black spot; dorsal caudal bars never black, usually with strongly undulate or pointed posterior margins. draconoides draconoides (p. 86)

Males with two large oblique lateral black blotches usually not followed by a third black spot; dorsal bars black and undulate or not. 3

3. Dark dorsal bars on tail with strongly undulate or pointed posterior margins from base to beyond middle of tail; dorsal tail bars never black.

draconoides carmenensis (p. 87)

Dark dorsal bars on tail with nearly straight or slightly undulate posterior margins at least from middle to end of tail; dorsal tail bars often black. 4

4. Dorsal tail bars black in both sexes; usually more or less red in dorsal coloration.

draconoides splendidos (p. 87)

Dorsal tail bars black in males only; usually little or no red in dorsal coloration. 5

5. Lateroventral black bars in males ill defined, blurred or obsolete, usually united below. Lateroventral black bars in males well defined, intense, separate below.


Lamellae under fourth toe 30 or more. 7

7. Lamellae under fourth toe 30 to 33. draconoides brevipes (p. 88)

Lamellae under fourth toe 35–39. draconoides insitatus (p. 88)

8. Femoral pores generally 16 or fewer; hind leg generally 92 percent of body length, or more; tail generally 58 percent of total length, or more; interparietal and supraorbital semicircles separated completely.

draconoides gabbi (p. 87)

Femoral pores generally 17 or more; hind leg generally 91 percent of body length, or less; tail generally 56 percent of total length, or less; interparietal and supraorbital semicircles usually not separated completely.

draconoides ventralis (p. 88)

CALLISAURUS DRACONOIDES DRACONOIDES Blainville


Type.—Mus. Hist. Nat. Paris; M. Botta collector.

Type locality.—“California,” here restricted to Cape San Lucas, Baja California.
Range.—Cape region of Baja California; Isla Espíritu Santo.

**CALLISaurus dracoNoides carmenensiS Dickerson**


**Type locality.**—Carmen Island, Gulf of California, Baja California.

**Range.**—The southern two-thirds of Baja California, except the Pacific coast in the center of that area, and the Cape region, between lat. 29°30' N. and 24° N.; the islands of San José, Carmen, San Marcos, South Santa Inés, Coronados, and Santa Margarita.

**CALLISaurus dracoNoides SplenDiDUS Dickerson**


**Type locality.**—Isla Ángel de la Guarda, Gulf of California, Baja California.

**Range.**—Island of Ángel de la Guarda, Baja California.

**CALLISaurus dracoNoides criniTUSt Cope**


*Callisaurus draconoides crinitus*, Tevis, Copeia, 1944, pp. 7–12.


**Type.**—U. S. N. M. No. 14895.

**Type locality.**—"Lower California" [=Ballenas Bay, Baja California].

**Range.**—Along the Pacific shore of central Baja California from Millers Landing (lat. 28°30' N.) to Santo Domingo (lat. 25°30' N.).

**CALLISaurus dracoNoides gabbii Cope**


_Type._—Apparently none designated.
_Type locality._—“Northern Lower California,” here restricted to Gardners Laguna, Salton River.

_Range._—Southern Nevada, western Arizona, and southeastern California southward into extreme northwestern Sonora and northeastern Baja California. Reported from several localities in Baja California, and from Sonora; Colorado River, Shoal Point.

**CALLISaurus DRACONIODES VENTRALIS** (Hallowell)


_Type._—U. S. N. M. No. 2670; S. W. Woodhouse collector.
_Type locality._—New Mexico, here restricted to Tucson, Ariz.
_Range._—Southern Arizona except the western and eastern edges, southward into northern Sonora. Recorded in Mexico only from the latter state, from between Sonoyta and Puerto Peñasco.

**CALLISaurus DRACONIODES INUSITATUS** Dickerson


_Type locality._—Tiburón Island, Sonora.
_Range._—Tiburón Island, and adjacent Sonoran coast to Guaymas.

**CALLISaurus DRACONIODES BREVIPES** Bogert and Dorson


_Type locality._—Guirocoba, 18 miles southeast of Álamos, Sonora, Mexico, elevation approximately 1,485 feet.
Range.—Extreme southern Sonora (Álamos and Guirocoba) and northern Sinaloa (Río de Choix).

**CALLISaurus Draconoides Bogerti** Martín del Campo


**Type.**—Inst. Biol. México; Victoriano Rodríguez collector.

**Type locality.**—Isla de los Chivos, en el puerto de Mazatlán, Sinaloa.

**Range.**—Known only from the type locality and the vicinity of Mazatlán.

**Genus Uma Baird**


**Genotype.**—*Uma notata* Baird.

**Species.**—Four, one of which includes two subspecies. Three forms occur in Mexico.

**Range.**—Southeastern California and adjacent Arizona, Sonora and Baja California, and southwestern Coahuila.

**Key to Mexican Forms of Uma**

1. Two strongly enlarged auricular lobules; a black bar on rear of femur; dorsal pattern with 3 median rows of spots and black chevrons with reticulum outlining whitish dots or blotches. **exsul** (p. 89)
   - Four strongly enlarged auricular lobules; no black bar on femur. **notata notata** (p. 89)
   2. Ventrolateral blotch smaller; femoral pores 20–31, usually fewer than 26. **notata cowlesi** (p. 90)
      - Ventrolateral blotch larger; femoral pores 23–32, usually more than 26.

**Uma Exsul** Schmidt and Bogert


**Type.**—Amer. Mus. Nat. Hist. No. 67404; Karl P. Schmidt and C. M. Bogert collectors.

**Type locality.**—Dunes 12 miles north of San Pedro de Las Colonias, Coahuila.

**Range.**—Presumably restricted to the vicinity of the type locality, the northern edge of the dry Laguna de Mayran.

**Uma Notata Notata** Baird


*Uma notata notata*, Heifetz, Copeia, 1941, pp. 101–104, figs. 1c, 2c, 3.


**Type.**—U.S.N.M. No. 4124; Williamson and Heerman collectors.

**Type locality.**—“Mojave Desert,” in error; restricted by Heifetz
to the "Colorado Desert" and here restricted to the vicinity of Yuma, Ariz.

Range.—Extreme southeastern Baja California, southwestern Arizona, and northeastern Baja California; possibly occurs in Sonora. Reported in Mexico only from Baja California.

**UMA NOTATA COWLESI** Heifetz

*Uma notata cowlesi* Heifetz, Copeia, 1941, pp. 104–106, fig. 5 (distribution).

*Type.*—California Acad. Sci. No. 53370; J. R. Slevin collector.

*Type locality.*—Shores of Tepoca Bay, Sonora.

*Range.*—The coast of northern Sonora, from Cerro Prieto to Tepoca Bay.

**Genus PETROSAURUS** Boulenger


*Genotype.*—*Uta thalassina* Cope.

*Range.*—Southern Baja California and certain adjacent islands in the Gulf of California.

*Species.*—Two.

**KEY TO SPECIES OF PETROSAURUS**

1. Two series of scales between subocular and labials; snout longer, less truncate; three distinct anterior dorsal cross bars; throat yellow, orange, or pale blue centrally—*thalassina* (p. 90)
   One series of scales between subocular and labials; snout shorter and more truncate; four distinct anterior dark bands; throat brownish marked with blackish centrally—*repens* (p. 90)

**PETROSAURUS REPENS** (Van Denburgh)


*Type.*—California Acad. Sci. No. 633; W. E. Bryant collector.

*Type locality.*—Comondú, Baja California.

*Range.*—The southern half of Baja California, exclusive of the Cape region; Isla Danzante.

**PETROSAURUS THALASSINUS** (Cope)


*Type.*—U.S.N.M. No. 5302; John Xantus collector.

*Type locality.*—Cape San Lucas, Baja California.
Range.—Cape region of Baja California and certain adjacent islands. Reported on the mainland from Cape San Lucas, La Paz, San José del Cabo, Miraflores, etc.; also Isla Partida, Isla Espíritu Santo.

Genus STREPTOSAURUS Mittleman


Genotype.—Uta mearnsi Stejneger.

Range.—Northern portion of Baja California and certain of the islands in the Gulf of California; in the United States: San Diego, Riverside, and Imperial Counties, Calif.

Species.—Two.

KEY TO FORMS OF STREPTOSAURUS

1: Throat with numerous round light spots; maximum length about 90 mm. mearnsi (p. 91)

Throat mottled, without definite rounded spots; larger maximum length, about 104 mm slevini (p. 91)

STREPTOSAURUS MEARNSI (Stejneger)


Type.—U.S.N.M. No. 21882; E. A. Mearns collector.

Type locality.—“Summit of Coast Range, United States and Mexico boundary line, California.”

Range. —Extreme southern California southward through the northern half of Baja California. Reported from Baja California: Ensenada, San Salado, San Matías, Parral, Matomi, Coyote Bay.

STREPTOSAURUS SLEVINI (Van Denburgh)


Type.—California Acad. Sci. No. 50506; Joseph R. Slevin collector.

Type locality.—Mejía Island, Gulf of California.

Range.—Mejía Island and Ángel de la Guarda Island, in the Gulf of California.

Genus CROTAPHYTUS Holbrook


Genotype.—Agama collaris Say.

63 The senior author has unintentionally referred Petrosaurus repens to this genus, owing to a lapsus; slevini was intended (Smith, Handbook of lizards, 1946, p. 173).
Range.—From Kansas south to Louisiana, west to the Pacific, as far north as central Oregon, and southward into northern Mexico. Absent from the northern Great Plains and some mountainous areas.

Species.—Four, with five forms recognized, all but one (C. c. collaris) occurring in Mexico.

KEY TO MEXICAN SPECIES OF CROTAPHYTUS

1. Black ring or rings about neck complete or incomplete; body pattern not reticulated. No black ring about neck, occasionally a black line present on shoulder; dorsal pattern boldly reticulated. 2

2. Collar formed of a single bar or ring; snout more elongate. 3

Collar formed of 2 rings, one or both incomplete. 3

3. Tail more distinctly compressed; 4 or 5 scale rows between suboculars and labials, and slightly enlarged scales on the middorsal line on tail. 3
dickersonae. 3

Tail not or scarcely discernibly compressed; minimum scale rows between suboculars and labials 2 (rarely 3); scale rows between supraorbital areas 2 (or rarely 3). collaris. 3

CROTAPHYTUS COLLARIS BAILEYI 34 Stejneger

Crotaphytus baileyi Stejneger, North Amer. Fauna, No. 3, 1890, p. 103, pl. 12, fig. 1.


Type.—U.S.N.M. No. 15821; C. Hart Merriam and Vernon Bailey collectors.

Type locality.—Painted Desert, Little Colorado River, Ariz.

Range.—Southern Idaho and Oregon southeastward through eastern California and northern Baja California to San Luis Potosí. Reported in Mexico from Baja California: San Salada, Trinidad, Las Palmas Canyon, Comondú, La Purísima, 6 miles and 33 miles north of Canipole, et cetera; Chihuahua: Chihuahua, Carmen, Progreso, San Buenaventura; Durango: 6 miles northeast of Pedriceña; Coahuila: Villa Acuña, Monclova, 4 miles west of Saltillo, Cuatro Ciéneegas, Pueblo Nuevo, Allende; Nuevo León: Pesquería Grande, Santa Catarina; Sonora: Hermosillo, Sierra Tule, Los Nogales, Río Grande 61 miles south of Los Nogales; San Luis Potosí: 30 miles north of Matehuala, Presa de Guadalupe.

34 With sufficient series it is probable that certain populations now associated with baileyi and collaris will be recognized as other subspecies. For instance, certain specimens from San Luis Potosí have three scale series between the supraocular regions. C. collaris collaris may occur.
CROTAPHYTUS INSULARIS Van Denburgh and Slevin


*Type.—* California Acad. Sci. No. 49151; J. R. Slevin collector.

*Type locality.—* Ángel de la Guarda Island, 7 miles north of Pond Island, Gulf of California, Baja California.

*Range.—* Restricted to type locality.

CROTAPHYTUS DICKERSONAE Schmidt


*Type.—* U.S.N.M. No. 64451; C. H. Townsend collector.

*Type locality.—* Tiburón Island, Sonora.

*Range.—* Known only from the type locality.

CROTAPHYTUS RETICULATUS Baird


*Type.—* Two cotypes, U.S.N.M. No. 2692, Ringgold Barracks, John H. Clark collector; No. 2731, Laredo, Tex., Arthur C. B. Schott collector.

*Type locality.—* Laredo and Ringgold Barracks, Tex., here restricted to Laredo.

*Range.—* Extreme southern Texas, along the Rio Grande, as far west as Eagle Pass, and southward in the northern parts of adjacent states in Mexico. Reported in Mexico from Tamaulipas: Mier; Nuevo León: 6 miles west of China, 4 miles west of China, 5 miles east of General Bravo, "Los Herrars"; Coahuila: 2 miles north of Nava.

Genus GAMBELIA Baird


*Genotype.—* *Gamhelia wislizenii* Baird and Girard.

*Range.—* Northern Idaho, Nevada, and Utah southward through southeastern California and Baja California, and into western Sonora and northern Coahuila.

*Species.—* One, with two subspecies. Only the typical form occurs in Mexico.

—This species has been reduced to subspecific status on the basis of specimens reported by Allen from Hermosillo and 40 miles north of Guaymas. Since we have not seen these specimens, and since he did not take into consideration some of the more important characters, we retain the specific status for this form pending a more considered revision.

Crotaphytus (Gambelia) wislizenii, Baird, United States and Mexican Boundary Survey, 1859, p. 7.

Gambelia wislizenii wislizenii, Smith, Handbook of lizards, 1946, pp. 159-164, pl. 30.


Crotaphytus copeii Yarrow, Proc. U. S. Nat. Mus., vol. 5, 1882, p. 441 (U.S.N.M. No. 12663; La Paz, Baja California).


**Type.**—U.S.N.M. No. 2770; H. Mollhausen collector.

**Type locality.**—Near Santa Fe, N. Mex.

**Range.**—Northern Idaho, Nevada, and Utah southward through southeastern California and all of Baja California except northwestern corner, western Chihuahua and northern Coahuila. Reported from the states of Baja California: Cedros and Magdalena Islands, numerous mainland localities; Sonora: Tiburón Island, numerous mainland localities; Chihuahua: 15 miles south of Juárez, Santa María, Progreso, Lake Santa María, Chihuahua, 11 miles south of Ahumada, 2 miles south of Moctezuma; Coahuila: Cuatro Ciénegas.

**Genus PHRYNOSOMA** Weigmann

Tapaja (or Tapaja, p. vi) Oken, Lehrbuch der Naturgeschichte, 1816, p. 295 (type, Lacerta orbicularis Linnaeus by present restriction; suppressed).


*Batrachosoma* Fitzinger, Systema reptilium, 1843, p. 79 (type, *Phrynosoma cornutum* Blainville).

*Tropidogaster* Fitzinger (nec Duméril and Bibron), loc. cit. (type, *Phrynosoma cornutum* Gray).

---

66 Cuvier (Le règne animal, vol. 2, 1817, p. 25) has been credited with this name by some authorities. He did not, however, use the name in latinized form anywhere in the first or 1817 edition of “Le Règne Animal”; he uses the name only as "les Tapayes."
CHECKLIST OF REPTILES OF MEXICO

95


Tapaya Girard (nee Fitzinger), in Wilkes, United States Exploring Expedition, Herpetology, 1858, p. 394 (type, Lacerta orbicularis Linnaeus).

Doliosaurus Girard, op. cit. p. 407 (type, Phrynosoma platyrhinos Girard).

Genotype.—Lacerta orbicularis Linnaeus.

Range.—Nebraska, Kansas, Oklahoma, and Texas westward to the Pacific, northward to British Columbia and Montana, and southward through Mexico to Guatemala.

Species.—Fifteen, with about 26 forms recognized; 21 occur in Mexico.

KEY TO MEXICAN SPECIES OF PHRYNOSOMA

1. Ventral abdominal scales smooth (a few scales may be keeled on breast of solare and a few specimens may show faint keeling on some of abdominal scales, but this form will have four occipital spines) 6
   Ventral abdominal scales distinctly keeled (rarely indistinctly keeled in cornutum) 2

2. Horns (or horny spines) present on back of skull 3
   No horns or horny spines on back of skull, the scales that form spines in other species are low rounded protuberances; two flaring temporal expansions on back of head separated by a medial notch; lower jaw with great vertical development, the chin shields separated from the infralabials by 4 or 5 scale rows; 76 mm. ditmarsi (p. 104)

3. Tail at least half length of body (usually more); large species with 2 lateral abdominal fringes of soft spines more or less clearly developed 4
   Tail shorter or but little longer than head; smaller species, with 1 series of lateral soft spines 5

4. Ventral scales large, 30 to 35 across widest part of belly; scales surrounding spines of upper lateral fringe very prominent; greater part of gular scales arranged in slightly serrate longitudinal series; 2 very prominent series of large scales widely separated on back; tail about as long as body; length, 170 mm. asio (p. 102)
   Ventral scales smaller, 55 to 70 across widest part of abdomen; 3 temporal and 2 occipital spines well developed; 3 brown bands across frontal area; 3 radiating lines below eye; rarely ventral keeling is indistinct; length, 140 mm. cornutum (p. 99)

5. Outer temporal part of skull prolonged backward into a very large “horn” several times size of small occipital spines; tail as long as head or slightly longer; 9-10 femoral pores taurus (p. 104)
   Outer temporal region not prolonged into a large horn; temporal spines about the same size as occipital spines; tail shorter than head; femoral pores about 10-10 braconnieri (p. 100)

6. No ear-opening present; tympanum superficial, covered with scales (very rarely unsealed in some Mexican specimens of modestum and some m'callii) 7
   Ear opening present, tympanum usually more or less deeply sunk 10

7. No lateral fringe of soft spines on sides of abdomen; occipital and temporal spines present but rather short; chin shields bordering labial series throughout; greater part of tail very slender and subcylindrical, not gradually tapering; venter white; chin usually speckled; 84 mm. modestum (p. 101)
   At least 1 series of lateral soft spines 8
8. Usually less than 12 femoral-preanal pores, the pore scales usually separated by 1 or more intervening scales.  

Pore series 17–23, the pores surrounded by several small scales and usually contiguous; temporal and occipital horns very elongate; 2 series of lateral fringes on abdomen; an indistinctly enlarged series of longitudinal gular scales on each side of chin, more evident posteriorly; a dark dorsal streak; immae late white on venter; tail much flattened; occasionally traces of abdominal keeling present.  

9. Pore series about 8 on each side; 3 enlarged temporals and 2 enlarged occipital spines, tending to curve laterally somewhat; small species, 72 mm. snout to vent; venter immaculate white.  

Pore series 7–12, the temporal and occipital horn much reduced; supraciliary spine short but heavy and prominent; a single series of scales in lateral fringe; venter usually punctate with dark; only rarely is the tympanum visible.  

10. Four much-elongated, flattened occipital horns, contiguous with a like temporal series, all directed more or less in same general plane; breast scales keeled; 1 distinct lateral fringe and a small secondary fringe indicated.  

11. Occipital horns at least twice as long as basal width; chin shields forming serrate series strongly visible from dorsal view of head; gulars with serrate series of enlarged scales.  

Occipital horns not twice as long as basal width; no serrate chin shields or specialized serrate series in gulars.  

12. Head not strongly notched or emarginate between occipital horns, distance between bases of horns one-half to rarely one and one-half times basal width of horn; occipital horns distinctly larger than supraciliary horns.  

13. Tail length a little less than half snout-to-vent length but distinctly longer than width of head; gular scales not convex; chest scales smooth.  

14. Occipital spines nearly erect, while temporal spines approach the horizontal, all relatively long; top of head conspicuously concave, supraciliary region elevated.  

Occipital spines on same plane as temporals, approaching horizontal, all shorter; top of head flat.  

15. Temporal horns not or barely extending backward as far as posterior tip of occipital horns.  

16. Occipital horns extending posteriorly farther than posterior temporals; preanal area with preanal pores not confined to a single row (usually several irregularly placed pores or 2 longitudinal rows of 2 to 4 scales).
Occipital and posterior temporal horns extending back an equal distance (or very nearly so); femoral pores 14–14, confined to single row. orbiculare dugesii (p. 98)

17. Fourth (or fifth) temporal horn (counting forward) sharply pointed and greatly exceeding fifth (or fourth) in length. orbiculare coronatum (p. 92)

Fourth and fifth temporal horns blunt, of nearly equal size, or fifth slightly larger. orbiculare coronatum (p. 92)

18. Nasal opening large; distance between inner edges less than 1/4 times maximum diameter of nostril. orbiculare cerroense (p. 103)

Nasal opening moderate, the distance between inner edges more than 1/4 diameters of nostril. orbiculare coronatum (p. 92)

19. Postrictial spine absent or rudimentary. orbiculare coronatum (p. 102)

Postrictial spine present. orbiculare coronatum (p. 102)

20. Frontal plates in adults smooth, convex. orbiculare coronatum (p. 103)

Frontal plates in adults rough, striated or rugose, and flat or peaked. orbiculare frontale (p. 103)

**PHYRINOSOMA ORBICULARE ORBICULARE** 67 (Linnaeus)

[Tapayxín, Lacertus orbicularis HERNÁNDEZ, Plantas y animales de la Nueva España, etc., 1651, C. xvi, p. 327, fig. (unnumbered).]

Lacerta orbicularis LINNAEUS, Systema naturae, ed. 12, vol. 1, 1789, p. 1062 (part, based on Hernández).

Agama orbiculara, Cuvier, Règne animal, vol. 2, 1817, p. 35.


Phrynosoma orbicularse orbicularse, SMITH, Trans. Kansas Acad. Sci., vol. 37, 1934, p. 290, pl. 11, fig. 2, pl. 12, fig. 5.

Phrynosoma wiegmanni GRAY, The zoology of Captain Beechey's voyage . . ., 1839, p. 96 (type unknown; type locality, "Mexico," here restricted to Mexico City, Distrito Federal).

**Type.** Presumably the unnumbered figure in Hernández, loc. cit.

**Type locality.**—Mexico (by inference), here restricted to México, D. F.

**Range.**—The central plateau of Mexico from Chihuahua and Nuevo León southward to Michoacán, Morelos, and Puebla. Reported from México: Toluca, Lerma, San Andrés, 11 miles south of Mexico (city), Teotihuacán, Zumpango; Puebla: Puebla, 15 kilometers northwest of San Martín, Matamoros near Tezutlán; Distrito Federal: México, Tlapam, Río San Juan de Dios, Tlalnepantla, between Navitas and Chalco; Hidalgo: Guerrero near Real del Mineral, Actopan, El Chico Parque Nacional, 2–3 miles west of Tulancingo, near Pachuca; Morelos: near Tres Marias (Km. 57); Chihuahua: south of Chihuahua City; Samachique, 21 miles south of Miñaca; Durango: Coyotes, El Salto, Ciudad; Jalisco: North of

---

67 We are unable to agree to the idea of the conspecificity of the Mexican orbicular group and the douglasii group. We feel that among other important distinguishing characters the posterior indentation of the skull, so marked in the latter group, has been overlooked in the past.
Rio Santiago, Guadalajara, Bolaños; Guanajuato: Guanajuato (city); San Luis Potosí: Jesús María, San Pedro; Querétaro: no specific locality; Nuevo León: Hacienda Pablillo; Tlaxcala: 13 kilometers northeast of Tlaxcala; Michoacán: “above 3000 feet” (Jorullo?); Zacatecas: Plateado, Sierra Madre.

PHRYNOSOMA ORBICULARE CORTEZII (Aug. Duméril and Bocourt)

Phrynosoma orbiculare, Wiegmann, Herpetologia Mexicana, 1834, p. 53, pl. 8, fig. 1.

Tapaya orbicularis var. A, Bocourt, *ibid.* (infra), livr. 4, 1874, pp. 223–224, pl. 11, fig. 2.

Tapaya Cortezii Aug. DUMÉRIL and BOCOURT, 68 Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 1, 1870, pl. 9, fig. 2, 2a–g.


Type.—Mus. Hist. Nat. Paris; Lucien Biart collector.

Type locality.—“Hacienda del Jasmin,” between “Orisaba and Cordoba,” Veracruz.

Range.—Eastern half of Puebla and adjoining parts of Veracruz. Reported from Veracruz: 15 miles east of San Marcos, 5 kilometers east of Las Vegas, Perote, Jalapa, Orizaba; Puebla: San Diego south of Tehuacán, Atzitzintla.

PHRYNOSOMA ORBICULARE DUGESII (Aug. Duméril and Bocourt)

Tapaya Dugesii 69 Aug. DUMÉRIL and BOCOURT, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 1, 1870, pl. 11, fig. 3, 3a–g.


Tapaya orbicularis var. B, Bocourt, *ibid.*, livr. 4, 1874, pp. 224–225, pl. 17, fig. 3, 3a–g.

Type.—Mus. Hist. Nat. Paris, two cotypes; Alfredo Dugès collector.

Type locality.—Colima.

Range.—Known only from Colima.

PHRYNOSOMA BOUCARDI (Aug. Duméril and Bocourt)

Tapaya Boucardii Aug. DUMÉRIL and BOCOURT, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 1, 1870, pl. 11, fig. 4, 4a–g.—Bocourt, *ibid.*, livr. 4, 1874, pp. 225–226.


Type.—Mus. Hist. Nat. Paris, two males, two females; A. Boucard collector.

Type locality.—“Plateau of Mexico,” here restricted to Zimapán, Hidalgo.

---

68 The name cortezii must be accredited to both authors of livr. 1. The name when it appears is Tapaya cortezii (nonis), and when quoted by Bocourt both names are used.

69 In 1870 the name was used as a full species, when associated with the figure. The name is dropped in the livr. 4 of the same work where it is regarded as a variety B of Tapaya orbicularis.
Range.—Central plateau region. Reported from Guanajuato: León; Hidalgo: Zimapán.

*Phrynosoma cornutum* (Harlan)


Type.—Unknown.

Type locality.—Great Plains east of the Rocky Mountains, here restricted to Fort Riley, Geary County, Kans.

Range.—Kansas to Texas, west to Arizona, south to Durango and San Luis Potosí. Reported from *Tamaulipas*: Alta Mira, Mier, Soto la Marina, Charco Escondido, Matamoros, Tampico, Marmolejo; *Nuevo León*: Monterrey, Pesquería Grande, 20 miles south of Nuevo Laredo, San Juan, 1 mile south of Nuevo Laredo; *San Luis Potosí*: Charco Largo; *Coahuila*: Castañuelos, Álamo de Parras, Monclova, San Pedro, Patos, Río Nazas, El Zacate, Buena Vista, Múzquiz, etcetera; *Durango*: 7 miles south of La Loma, 25 miles north of Bermejillo; *Chihuahua*: Batopilas, 5 miles north of Moctezuma, Progreso, Chihuahua (city), Huajuquilla (Jiménez), Sauz, Guzmán, etcetera; *Sonora*: Los Nogales.

*Phrynosoma m'callii* Hallowell


*Doliosaurus m'callii*, Girard, Wilkes United States Exploring Expedition, Herpetology, 1858, p. 408.


*Anota maccalli*,\(^{10}\) Bryant, Univ. California Publ. Zool., vol. 9, No. 1, 1911, pp. 5, 54, pl. 7.

Type.—Unknown.

Type locality.—Great Desert of the Colorado, between Vallecita and Camp Yuma, about 100 miles east of San Diego.

Range.—Northwestern Sonora and northeastern Baja California; in United States: Southeastern California and southwestern Arizona.

---

\(^{1}\) The senior author has expressed the opinion elsewhere that if the characters on which the genus *Anota* is based should prove constant this name should replace *Phrynosoma* for this species.
Reported from Sonora: East bank of Colorado River, 5 miles south of United States border. Baja California: “Yuhu Basin north of Petrified Forest.”

**PHRYNOSOMA DOUGLASSII HERNANDESI** (Girard)


*Type.—U.S.N.M. No. 197, a cotype.*

*Type locality.—New Mexico and “Sonora,” restricted to Santa Fe, N. Mex. (Cope, loc. cit., 1900).*

*Range.—Northern Sonora, Chihuahua; in United States: Colorado, southern New Mexico, southeastern and central Arizona, southwestern Texas. Reported from Sonora: “Mexican Boundary line”; Chihuahua: Ramos.*

**PHRYNOSOMA DOUGLASSII BRACHYCERCUM** Smith


*Type.—U.S.N.M. No. 23993; Edward Palmer collector.*

*Type locality.—“Durango.”*

*Range.—Known only from the type locality.*

**PHRYNOSOMA BRACONNIERI** Duméril and Bocourt


*Type.—Mus. Hist. Nat. Paris, adult; collected by Aide-Major Jacob; several specimens collected by Alfredo Dugès and A. Boucard.*

*Type locality.—Oaxaca.*

*Range.—Extreme southern edge of the central Mexican plateau, in semiarid portions of Puebla and Oaxaca (a specimen has been taken within a few hundred meters of the border of Veracruz, and very probably the species occurs in the mountains in the western part of that state). Reported from Puebla: Tehuacán, 14 miles northeast of Tehuacán, near Cacoloapam (Km. 226); Oaxaca: No specific locality, unless Oaxaca (city) is meant.
**PHRYNOSOMA MODESTUM** Girard


*Doliosaurus modestus*, Girard, Wilkes United States Exploring Expedition, Herpetology, 1858, p. 400.


**Type.**—U.S.N.M. (?), several ectotypes; General Churchill and “the party under Col. J. D. Graham” collectors.

**Type locality.**—The Río Grande west of San Antonio, Tex. (Churchill), and from between San Antonio and El Paso (Graham); here restricted to Las Cruces, N. Mex.

**Range.**—Western Texas to Arizona, southward to San Luis Potosí. Reported from *Nuevo León*: Santa Catarina, Pesquería Grande. *Chihuahua*: 11 miles south of Ahumada, Río Santa Marfa, Guzman, near Janos, Juárez, Chihuahua (city); *Coahuila*: 1.5 miles west of Saltillo, Músquiz, Jaral, Castañuelas, Palau, Álamo de Parras, San Pedro, Agua Nueva, Monclova, Buena Vista, Río Nazis, Cuatro Ciénegas, et cetera; *Sonora*: Nariz Temporal, Los Nogales, Sierra de la Nari; *Durango*: between Lerdo and La Goma, Pedriceña, 5 kilometers west of Torreón, 25 miles north of Bermejillo, 7 miles south of La Loma; *Zacatecas*: 3 miles west of La Colorada; *San Luis Potosí*: near Charco Largo (130 kilometers north of San Luis Potosí city), Hacienda La Parada.

---

**PHRYNOSOMA PLATYRHINOS PLATYRHINOS** Girard


*Doliosaurus platyrhinos*, Girard, in Wilkes, United States Exploring Expedition, Herpetology, 1858, p. 400.


*Phrynosoma calidiarum*, Ditmars, The reptile book, 1907, p. 157, pl. 46, figs. 10, 14, pl. 50, fig. 2.

**Type.**—U.S.N.M. No. 1897; H. Stansbury collector.

**Type locality.**—Great Salt Lake, Utah.

PHRYNOSOMA PLATYRHINOS GOODEI Stejneger

*Phrynosoma goodei* Stejneger, North Amer. Fauna, No. 7, 1893, p. 191 (footnote), pl. 2, figs. 3a–3e.


*Type.*—U.S.N.M. No. 8567a; T. H. Streets collector.

*Type locality.*—"Coast deserts of the state of Sonora, Mexico," here restricted to Puerto Libertad.

*Range.*—Known only from the gulf coast of Sonora. Reported from Puerto Libertad, 8 miles northwest of Alamo Muerto, west coast of Sonora.

PHRYNOSOMA ASIO Cope


*Batrachosoma asio*, BOCOURT, Mission scientifique au Mexique . . . , Études sur les reptiles, livr. 4, 1874, pl. 17, fig. 9, 9a–c.


*Type.*—U.S.N.M. (?) ; John Xantus collector.

*Type locality.*—Colima, Colima (by present restriction).

*Range.*—From Colima through coastal Michoacán, Guerrero, Oaxaca to Chiapas, and in the Balsas Basin. In Central America, recorded from Guatemala. Reported from *Oaxaca*: San Pedro, Tequesistlán, San Gerónimo, Ranchero Poso Río, vicinity of Tehuantepec, vicinity of Salina Cruz; *Guerrero*: Mexcala (on Balsas River); *Michoacán*: La Salada, Cofradía, San Salvador, Apatzingán; *Colima*: Tecoman, Colima (city); *Chiapas*: no definite locality.

PHRYNOSOMA CORONATUM CORONATUM (Blainville)


*Type locality.*—"California," here restricted to Cape San Lucas, Baja California.

*Range.*—Southern part of Baja California, north to about lat. 26° N.

PHRYNOSOMA CORONATUM JAMESI Schmidt


*Phrynosoma coronatum jamesi*, KLAUBER, Copeia, 1936, p. 110.—TEVIS, Copeia, 1944, pp. 13–15, map fig. 2.

*Type.*—U.S.N.M. No. 64450; H. Townsend collector.

*Type locality.*—Shore of San Bartolomé Bay, Baja California.
Range.—Central Baja California, between lat. 26° N. and lat. 28°30' N.

**PHRYNOSOMA CORONATUM FRONTALE** Van Denburgh


Type.—Stanford Univ. Mus. No. 93, adult male; C. H. Gilbert and W. W. Price collectors.

Type locality.—Bear Valley, San Benito County, Calif.

Range.—Discontinuous; central California east of the Sierra Nevada south to but not including extreme southwestern California; northwestern coast of Baja California except a small area adjacent to the United States border; Cedros Island.

**PHRYNOSOMA CORONATUM BLAINVILLII** Gray

*Phrynosoma blainvillii* Gray, The zoology of Captain Beechey’s voyage . . ., Reptiles, 1839, p. 96, pl. 29, fig. 1.


Type.—Brit. Mus. Nat. Hist., young specimen; de Blainville collector.

Type locality.—“California,” here restricted to San Diego.

Range.—Southern California and extreme northeastern Baja California.

**PHRYNOSOMA CERROENSE** Stejneger


Type.—U.S.N.M. No. 11977; L. Belding collector.

Type locality.—“Cerros” Island, Pacific coast of Baja California [=Cedros Island].

Range.—Known only from the type locality.
PHRYNOSOMA TAURUS Dugès

Phrynosoma taurus Dugès, MS. 1868, fide Duméril and Bocourt loc. cit., p. 234; and La Naturaleza, 1869, p. 143 (nomen nudum).—Aug. Duméril and Bocourt, Mission scientifique au Mexique . . . , Études sur les reptiles, livr. 1, 1870, pl. 12, figs. 8, 8a–g.—Dugès, La Naturaleza, 1873, pp. 302–305, figs. 1–4 (this description bears the date Nov. 23, 1870, but was not published until 1873; there were two specimens, one from "Cozcatlán," the other from Estado de Puebla).—Bocourt, Mission scientifique au Mexique . . . , Études sur les reptiles, livr. 4, 1874, pp. 234–235.—Boulenger, Catalogue of lizards in the British Museum, vol. 2, 1885, p. 249.—Smith and Neck, Anal. Esc. Nac. Cienc. Biol., vol. 3, 1943, pp. 208–210, pl. 3, figs. 1–2.


Range.—Puebla and Guerrero. Reported from Puebla: Zapatitlán, San Diego near Tehuacán, Cozcatlán (Dr. Bedel collector), Izúcar de Matamoros; Guerrero: 5 miles north of Chilpancingo, Amula 8,000 feet.

PHRYNOSOMA SOLARE Gray


Phrynosoma regale Girard, in Wilkes, United States Exploring Expedition, 1858, p. 406 (U. S. N. M. No. 161; Sierra de la Nariz, near Zuñi, Sonora).

Type.—Brit. Mus. Nat. Hist. (not listed in Boulenger, Catalogue of lizards in the British Museum, vol. 2, unless it is spec. a, σ [California], listed under Phrynosoma regale, p. 245); ? de Blainville collector.

Type locality.—"California," here restricted to Yuma, Ariz.

Range.—Southern Arizona southward into northeastern Baja California, western Sonora and into Sinaloa. Reported from Sonora: Santa Ana, Nogales, Hermosillo, 15 to 20 miles south and 45 miles southwest of Hermosillo, Tiburón Island, Empalme, 10 miles northwest of Guaymas, Alamos, Guirocoba; Baja California: Las Animas Bay; Sinaloa: Ahome, Sierra de Choix.

PHRYNOSOMA DITMarsi Stejneger


Type.—U.S.N.M. No. 36022; Mr. Eustace collector.

⁵¹ Whether the specimens mentioned above are really the types may be open to question. A description in manuscript was made by Dugès from a specimen in Mexico and presumably sent from Mexico to August Duméril in Paris. The Paris Museum had nine specimens of the same species. Drawings were made of one, and the drawings published under the name Phrynosoma taurus Dugès. Since the specimen figured is the first description or indication, we believe that the original of the drawing becomes the type and this, we believe, is a specimen in the Museum d'Histoire Naturelle de Paris. Thus the type specimen was presumably never seen by the author of the name. The specimen described by Dugès is in the Museum "Alfredo Dugès," Colegio del Estado de Guanajuato.
Type locality.—State of Sonora, not far from the boundary of Arizona.

Range.—Known only from the type locality.

Genus SCELOPORUS Wiegmann


Genotype.—Sceloporus torquatus Wiegmann.

Species.—In all, 104 forms are recognized at present, consisting of 54 species representing 15 groups. Eighty-nine forms of 52 species are known from Mexico.

Range.—Southern Canada on the west and southern New York on the east south throughout North and Central America to western Panama.

KEY TO MEXICAN GROUPS OF SCELOPORUS

1. Postfemoral dermal pocket present ........................................... 2
   Postfemoral dermal pocket absent ........................................... 4
2. Postrostral scales absent; nasals and internasals in contact with rostral.  
   maculosus (p. 133)
   Postrostral scales present; internasals and nasals separated from rostral  3
3. Tail strongly compressed in males, rounded and light pink in females; femoral 
   pores 24 or more on each side ........................................... pyrocephalus (p. 138)
   Tail rounded in both sexes, not pink in females; femoral pores 20 or less.  
   variabilis (p. 129)
4. Lateral body scales not imbricate ........................................... 5
   Lateral body scales imbricate ............................................. 6
5. Preanal scales keeled in females; males with poorly developed postanals; no 
   distinct belly patches in males; tail over twice length of body.  
   utiformis (p. 135)
   Preanal scales smooth in females; males with well-developed postanals; 
   lateral belly patches distinct in males; tail less than twice length of body.  
   merriami (p. 132)
6. No postrostrals; nasals and internasals in contact with rostral ............ 7
   Postrostrals present; nasal and internasals separated from rostral ......... 8
7. Dorsal scales 50 or more; femoral pores 17 or more; preanal scales smooth in 
   females ...................................................... scalaris (p. 136)
   Dorsal scales 46 or less; femoral pores 16 or less; preanal scales keeled in females.  
   siniferus (p. 133)
8. Ventral scales, at least laterally, pointed, not notched; preanal scales keeled 
   in females; no lateral belly markings in males; femoral pore series widely 
   separated medially; two postrostrals ........................................ siniferus (p. 133)
   Ventral scales notched, or, if pointed, femoral pore series closely approxi-
   mated medially (separated by four scales or less); preanal scales smooth 
   in females ..................................................... 9
9. Males uniform white below; anterior section of frontal longitudinally divided; 
   femoral pore series widely separated medially ................................ chrysostictus (p. 133)
   Males with lateral belly patches distinct or, if not, anterior section of frontal 
   entire ..................................................... 10
10. Femoral pore series closely approximated medially; two postrostrals; lateral scale rows parallel or nearly so; scales on posterior surface of thigh granular; small species. \textit{scalaris} (p. 136)

Femoral pore series widely separated medially, or, if closely approximated, scales on posterior surface of thigh not granular; lateral scale rows strongly divergent; typically four postrostrals, sometimes three, rarely two. \textit{torquatus} (p. 119)

11. Scales on posterior surface of thigh granular. \textit{grammicus} (p. 119)

Scales on posterior surface of thigh not granular. \textit{grammicus} (p. 119)

12. Dorsal scales highly irregular in size. \textit{grammicus} (p. 119)

Dorsal scales subequal in size. \textit{grammicus} (p. 119)

13. Lateral nuchal scales much smaller than and well differentiated from dorsal nuchal scales; scales of lateral row of dorsal nuchals enlarged, strongly keeled and mucronate. \textit{grammicus} (p. 119)

Lateral nuchal scales not well differentiated from dorsal nuchal scales. \textit{grammicus} (p. 119)

14. Throat light blue, barred or mottled with white; lateral belly patches confluent medially; preanal scales large, about seven in a row from a line between femoral pore series to anus. \textit{grammicus} (p. 119)

Throat not as described or, if so, lateral belly patches not confluent medially and preanal scales smaller. \textit{grammicus} (p. 119)

15. Scales on posterior surface of thigh granular, femoral pore series widely separated medially, the folds defining the preanal area passing between the femoral pore series; throat mottled with blue and white, never with a distinct, blue spot medially or on either side posteriorly. \textit{graciosus} (p. 118)

Scales on posterior surface of thigh not granular or, if so, femoral pore series more closely approximated medially, the folds defining preanal area not or barely passing between femoral pore series. \textit{megalepidurus} (p. 121)

16. Dorsal, ventral, and lateral scales subequal in size; small species. \textit{megalepidurus} (p. 121)

Dorsal, ventral, and lateral scales distinctly differing in size; large or small species. \textit{megalepidurus} (p. 121)

17. A distinct, dark, light-bordered nuchal collar. \textit{torquatus} (p. 121)

No distinct, dark, light-bordered nuchal collar. \textit{torquatus} (p. 121)

18. Gular region mottled, not barred nor with a median or two lateral blue spots posteriorly. \textit{torquatus} (p. 121)

Gular region not mottled. \textit{torquatus} (p. 121)

19. Gular region barred in males and lacking a median posterior dark blue or black spot. \textit{spinosus} (p. 110)

Gular region not barred; or, if barred, with a median posterior dark blue or black spot. \textit{undulatus} (p. 117)

20. Gular region barred in males, with a median posterior dark blue or black spot. \textit{undulatus} (p. 117)

Gular region not barred. \textit{undulatus} (p. 117)

21. Supraoculars large, separated from superciliaries by no more than one complete and one incomplete row of small scales; and one or more of the posterior supraoculars in contact with median head scales. \textit{spinosus} (p. 110)

Supraoculars large or small; if any of the posterior supraoculars are in contact with median head scales, the supraoculars are separated from superciliaries by three or more complete or incomplete rows of small scales. \textit{undulatus} (p. 117)

22. Males lacking lateral belly patches. \textit{undulatus} (p. 117)

Males with lateral belly patches. \textit{undulatus} (p. 117)

23. A pair of dark blue or black spots on posterior part of gular region. \textit{undulatus} (p. 117)

No pair of dark spots on posterior part of gular region. \textit{undulatus} (p. 117)
24. Three or more rows of small scales between supraoculars and supracleiaries; or, if two, the scales of inner row considerably larger than those of outer row.  
One complete and one incomplete row of small, subequal scales between supraoculars and supracleiaries.  

25. One canthal, or, if two, the first forced above canthal ridge.  
Two canthals, the first not forced above canthal ridge.  

26. A broad, black nuchal collar complete about neck dorsally, or anterior section of frontal longitudinally divided.  
Neither true.  

27. Frontal ridges present.  
Frontal ridges absent.  

28. Adult males a uniform bright green above.  
Not so.  

**FORMOSUS GROUP**

*Species.*—Six, with a total of 11 forms. Extralimital are *lunaei*, *m. malachiticus*, and *m. smaragdinus*.  

*Range.*—Michoacán and Veracruz south to western Panama.  

**KEY TO MEXICAN SPECIES OF THE FORMOSUS GROUP OF SCELOPORUS**

1. Median frontonasal separated from lateral frontonasals.  
Median frontonasal in contact with lateral frontonasals.  

2. Dorsals about 31; internasals large, keeled, 3 from rostral to median frontonasal.  
Dorsals 37 to 45; internasals smaller, not keeled, rugose or not.  

3. A complete nuchal collar, sometimes narrowly interrupted medially.  
Nuchal collar incomplete.  

4. Anterior section of frontal usually longitudinally divided; canthals sharply ridged: no nuchal collar; ventrals a fourth, laterals not more than two-thirds, size of dorsals.  
Anterior section of frontal rarely longitudinally divided; canthals rounded; a nuchal collar or not; ventrals a half, laterals three-fourths size of dorsals.  

5. A broad, nearly or quite complete, nuchal collar.  
Collar, if present, restricted to sides of neck.  

6. Femoral pores 12-16; one or more supraoculars generally in contact with median head scales.  
Femoral pores 16-21; supraoculars not in contact with median head scales.  

7. Males without yellow on throat; dorsal scale rows black edged; dorsal head scales with a light blue center.  
Males with yellow or orange on throat; dorsal surface uniform blue; head scales not light spotted.  

**SCELOPORUS MALACHITICUS TAEIIOCNEMIS** Cope  


---

72a Material examined since this key was originally prepared places this species in the *torquatus* group.  

861316—50—8
Type.—U.S.N.M. No. 24768.
Type locality.—Guatemala, here restricted to Cobán.
Range.—Highlands of Chiapas and northern Guatemala in Alta Verapaz. Known in Mexico only from Chiapas: Pinobete, Mount Ovando, Teopisca.

SCLEPORUS MALACHITICUS SALVINI Günther


Type locality.—"Jalapa and Guatemala." Restricted (Smith, 1939, p. 40) to Jalapa, Veracruz.
Range.—Central western Veracruz to the Isthmus of Tehuantepec. Recorded only from the states of Veracruz and Oaxaca.

SCLEPORUS MALACHITICUS ACANTHINUS Bocourt


Type locality.—San Agustín, near Volcán de Atilán, Guatemala, 610 meters.
Range.—Pacific coast foothills, Chiapas to El Salvador. Known in Mexico only from the vicinity of Esquistla, Chiapas.

SCLEPORUS FORMOSUS FORMOSUS Wiegmann

_Sceloporus formosus_ Wiegmann, Herpetologia Mexicana, 1834, p. 50, pl. 7.
_Tropidolepis formosus_, Duménil and Bibron, Erpétologie générale, vol. 4, 1837, pp. 303–304.
**CHECKLIST OF REPTILES OF MEXICO**

**Type.**—? Zool. Mus. Berlin; F. Deppe collector.

**Type locality.**—Mexico, here restricted to Acultzingo, Veracruz.

**Range.**—Central Veracruz south in mountainous regions to the Isthmus of Tehuantepec. Recorded from various localities in the states of Veracruz, Puebla, and Oaxaca.

**SCHEL PORUS FORMOSUS SCITULUS Smith**


**Type.**—EHT-HMS No. 26962; R. C. Taylor and E. H. Taylor collectors.

**Type locality.**—Omitime, Guerrero.

**Range.**—Sierra Madre del Sur Mountains in Guerrero.

**SCHEL PORUS ASPER Boulenger**


**Type locality.**—La Cumbre de los Arrastrados, Jalisco.

**Range.**—Mountainous regions of western Mexico, from Nayarit to Guerrero. Recorded from Nayarit, Jalisco, Michoacán, and Guerrero.73

**SCHEL PORUS STEJNEGERI Smith**


**Type.**—U.S.N.M. No. 112635; E. H. Taylor, R. C. Taylor, and H. M. Smith collectors.

**Type locality.**—Tierra Colorada, Guerrero.

**Range.**—Guerrero, on southern low slopes of the Sierra Madre del Sur.

**SCHEL PORUS PREZYGUS Smith**74


**Type.**—U.S.N.M. No. 46881; E. W. Nelson and E. A. Goldman collectors.

**Type locality.**—Conjab, 5,300 feet (between San Bartolomé and Comitán), Chiapas.

**Range.**—Known only from the type locality in Chiapas.

---

73 The record from this state (Chilapa) (Ahl, Zool. Anz., vol. 106, 1934, p. 184) is open to doubt. The identification needs verification.

74 Group allocation of this species was not certain until after this account was in type. It belongs in the turguatus group.
Species.—Nine, with a total of 21 forms.
Range.—Guatemala northward to southern Nevada, south-western Colorado, southern Oklahoma, and central Texas.

KEY TO THE SPINOSUS GROUP OF SCELOPORUS

1. Femoral pores on both thighs total 7 or more .................................................. 3
Femoral pores on both thighs total 6 or less .................................................. 2

2. Ventral surfaces immaculate in adults of both sexes. horridus albiventris (p. 116)
Sides of belly blue, frequently dark-bordered, in males; gular region barred in males, sometimes in females. horridus oligoporus (p. 116)

3. Supraoculars completely in contact with superciliaries, or with one to four very small scales between; posterior superciliary greatly enlarged; venter immaculate in both sexes. edwardtaylori (p. 112)
Supraoculars separated from superciliaries by at least one row of small scales; posterior superciliary not enlarged; venter with distinct markings in males .......................................................... 4

4. Femoral pores on both thighs total 12 or less; 4 supraoculars, the fourth in contact with median head scales. horridus horridus (p. 116)
Femoral pores on both thighs total 13 or more; or, if 12, all supraoculars separated from median head scales .......................................................... 5

5. Femoral pores (one side) 17 or more .......................................................... 6
Femoral pores (one side) 16 or less .......................................................... 10

6. A black shoulder patch present, with a light posterior border ........................................ 10
No black shoulder patch; or, if present, without a light posterior border .................................. 7

7. Snout black; a broad, black bar across head at middle of supraocular region; chest and middle of belly orange in males; gular region in females white, with irregular, black marks; first canthal not in contact with lorilabials; ventral scales 44–54 .......................................................... 8
Snout not black, similar in color to rest of head; no black bar across head; chest and middle of belly not orange; gular region in both sexes bluish, with light lines following scale rows at anterior part of throat; first canthal in contact with lorilabials; ventral scales 38–42 magister lineatulus (p. 115)

8. Femoral pores usually (84 percent) 20 or less on each side; total pore counts usually (80 percent) 40 or less .......................................................... 9
Femoral pores usually (78 percent) 21 or more on each side; total pore counts usually (83 percent) 41 or more; lateral belly patches usually not confluent medially in males melanorhinus melanorhinus (p. 112)

9. Lateral nuchal pocket greatly reduced, with very few if any granules, and very little if any bare skin, shallow melanorhinus stuarti (p. 113)
Lateral nuchal pocket with numerous granules, considerable bare skin, and considerable depth melanorhinus calligaster (p. 113)

10. One or more supraoculars in contact with median head scales ........................................ 14
None of supraoculars in contact with median head scales ........................................ 11

11. Gular region barred .......................................................... 12
Gular region not barred .......................................................... 13

12. Dorsal scales usually more than 30; femoral pores usually more than 9 (one side); supraoculars usually 5 to 7 spinosus caeruleopunctatus (p. 116)
Dorsal scales usually 30 or less; femoral pores usually 9 or less (one side); supraoculars usually 4 spinosus spinosus (p. 116)
13. Belly patches in males continuous across chest; prefrontals never in contact; no
dorsolateral light lines in males; posterior surface of thigh mottled or with
a broad light line, dark-bordered. ........................................ 14

Belly patches small, confined to sides of belly in males; prefrontals usually in
contact; dorsolateral light lines present in males; posterior surface of thigh
nearly immaculate or with a narrow, short black line near insertion of hind
leg................................................................. olivaceus (p. 117)
14. Gular region barred......................................................... 15
Gular region not barred........................................................ 16
15. Femoral pores 13 or more (one side) .................................... orcutti licki (p. 114)
Femoral pores 12 or less (one side) ......................................... spinosus spinosus (p. 116)
16. Femoral pores (one side) 12 or less.................................... 17
Femoral pores (one side) 13 or more...................................... 21
17. Outer row of labiomental scales in contact with mental.

magister magister (p. 114)

Outer row of labiomental scales separated from mental.................. 18
18. First canthal in contact with lorilabials ................................ orcutti orcutti (p. 114)
First canthal separated from lorilabials.................................... 19
19. Lateral scales about one-half size of dorsal scales; median frontonasal not in
contact with frontal; lateral belly patches in males not passing over
chest................................................................. 20
Lateral scales considerably more than one-half size of dorsal scales; median
frontonasal usually in contact with frontal; lateral belly patches in males
passing over chest................................................................. 21
20. Dorsal markings absent or very dimly visible; a broad, light line on posterior
surface of thighs....................................................... lundelli gaigeae (p. 112)
Dorsal markings present and distinct at least in females, consisting of narrow,
dark cross bars; posterior surface of thigh mottled.

lundelli lundelli (p. 112)
21. Femoral pores (one side) 10 or less..................................... 22
Femoral pores (one side) 11 or more...................................... 23
22. First canthal rarely in contact with lorilabials; gular region never barred;
outer row of labiomentals rarely in contact with mental.

clarkii boulengeri (p. 113)

First canthal usually in contact with lorilabials; gular region barred, or outer
row of labiomentals sometimes in contact with mental (not in o. orcut-
ti)............................................................................ 23
23. First canthal separated from labiomentals; femoral pores 16 or less; lower fore-
legs distinctly barred; gular region never barred. clarkii clarkii (p. 113)
First canthal in contact with lorilabials, lower forelegs not barred dis-
distinctly................................................................. 24
24. Outer row of labiomentals in contact with mental; gular region not barred;
shoulder patch distinct................................................... 25
Outer row of labiomental scales separated from mental; gular region barred or
shoulder patch indistinct.................................................. 26
25. Dorsal scales 32 to 40; femoral pores 13–19; usually one or more scales in con-
tact with both subocular and supralabials; a distinct, light-bordered black
shoulder patch; dorsolateral light lines present, indistinct posteriorly, the
medial edge sometimes indistinct. orcutti licki (p. 114)
Dorsal scales 28 to 36; femoral pores 10–15; usually 2 complete rows of lorila-
bials below subocular; shoulder patch indistinct; no dorsolateral light
lines................................................................. orcutti orcutti (p. 114)
26. Supraoculars usually 5; femoral pores (one side) usually 15 or less.  

magister magister (p. 114)

Supraoculars usually 6 or 7; femoral pores usually 16 or more.  

27. No dark lines in adult males on sides of lateral scale rows; femoral pores (one side) usually less than 19; lateral belly patches confluent medially in adult males.  

magister rufidorsum (p. 115)

Dark lines on sides of lateral scale rows present in adult males; lateral belly patches not confluent medially in adult males.  

28. A distinct, narrow dorsal stripe about 1½ scale rows wide; females and young with 2 rows of dark spots, one on each side of middorsal stripe.  

magister monserratensis (p. 115)

A broad, light-colored dorsal area about 6 scale rows wide; dark spots on back absent or indistinct.  

magister zosteromus (p. 115)

SCELOPORUS LUNDELLI LUNDELLI Smith


Type.—Univ. Michigan Mus. Zool. No. 80674; C. L. Lundell collector.

Type locality.—Cohune Ridge (20 miles southeast of Benque Viejo), British Honduras.

Range.—The southern portion of the Yucatán Peninsula. Recorded in Mexico only from Campeche.

SCELOPORUS LUNDELLI GAIGEAE Smith


Type.—Chicago Nat. Hist. Mus. No. 31524; H. M. Smith collector.

Type locality.—Mérida, Yucatán.

Range.—Northern Yucatán; recorded only from Yucatán.

SCELOPORUS EDWARDTAYLORI Smith


Type.—EHT-HMS No. 8331 (EHT field No. 4221); E. H. Taylor and H. M. Smith collectors.

Type locality.—Ixtepec (= San Gerónimo), Oaxaca.

Range.—Central and southern Oaxaca.

SCELOPORUS MELANORHINUS MELANORHINUS Bocourt


Type.—Mus. Hist. Nat. Paris, three females, one male, cotypes; F. Sumichrast collector.
Type locality.—Isthmus of Tehuantepec, Oaxaca, here restricted to Tehuantepec (City).

Range.—Pacific slopes of Oaxaca.

**SCELOPORUS MELANORHINUS CALLIGASTER** Smith


Type.—U.S.N.M. No. 112201; H. M. Smith collector.

Type locality.—Acapulco, Guerrero.

Range.—Pacific slopes below about 3,500 feet, from Nayarit to Guerrero. Recorded from Nayarit, Jalisco, Colima, Michoacán, and Guerrero.

**SCELOPORUS MELANORHINUS STUARTI** Smith


Type locality.—Finca Canibal, 3,000 feet, Huehuetenango, Guatemala.

Range.—The upper valley of the Río Grijalva in Chiapas and Guatemala. Known in Mexico only from Chiapas: Piedra Parada, 12 miles north of Ocozucoutula.

**SCELOPORUS CLARKII CLARKII** Baird and Girard


Type.—U.S.N.M. No. 2940, three cotypes, of which two belong to *S. m. magister*, and one, with a white tag on its leg, is designated lectotype of *S. clarkii*; John H. Clark collector.

Type locality.—“Province of Sonora” (=southern Arizona), here restricted to Santa Rita Mountains.

Range.—Southern and central Arizona (except the western part of the State), southwestern New Mexico, and all of Sonora except the extreme western and southern parts. Recorded only from the states of Sonora and Chihuahua, and the islands of San Pedro Nolasco and Tiburón.

**SCELOPORUS CLARKII BOULENGERI** Stejneger

*Sceloporus boulengeri Stejneger*, North Amer. Fauna, No. 7, 1893, p. 180, pl. 1, figs. 5a-c.

Type.—U.S.N.M. No. 14079, three cotypes; A. Forrer collector.

Type locality.—Presidio (about 50 miles south from Mazatlán), Sinaloa.

Range.—Pacific slopes, southern Sonora south to northern Jalisco. Recorded from Sonora, Sinaloa, Nayarit, and Jalisco.

**SCELORUS ORCUTTI ORCUTTI** Stejneger

*Sceloporus orcutti Stejneger*, North Amer. Fauna, No. 7, 1893, p. 181, pl. 1, figs. 4a–c.


Type.—U.S.N.M. No. 16330; C. R. Orcutt collector.

Type locality.—Milquatay Valley, San Diego County, Calif.

Range.—California, from southern San Bernardino county southward into Baja California to the Sierra de la Gigante, and on some adjacent islands.

**SCELOPORUS ORCUTTI LICKI** Van Denburgh


Type.—Stanford Univ. Mus. No. 2987a, lectotype (one of two paratypes; type in California Acad. Sci. destroyed in 1906); Gustav Eisen, and Vaslet collectors.

Type locality.—Sierra San Lázaro, Baja California.

Range.—Cape region of Baja California, including Isla Espíritu Santo and Isla Partida.

**SCELOPORUS MAGISTER MAGISTER** Hallowell


Type.—U.S.N.M. No. 2967; A. L. Heermann collector.

Type locality.—Fort Yuma, Ariz.

Range.—Extreme western Texas to southern California (excluding western slopes), north to southern Nevada and southwestern Colorado, south to northern Durango and southern Sonora. Reported from Chihuahua, Coahuila, Durango, Sonora, and Baja California.
CHECKLIST OF REPTILES OF MEXICO

SCELOPORUS MAGISTER RUFIDORSUM Yarrow


_Type._—U.S.N.M. No. 11981; L. Belding collector.
_Type locality._—“San Quentin Bay,” Baja California. In Yarrow, U.S. Nat. Mus. Bull. 24, 1882, p. 64, the locality is listed “La Paz, California.”
_Range._—Northern Baja California, exclusive of the northeastern section, southward to include the Vizcaino Desert and Cedros Island; Coronados Islands.

SCELOPORUS MAGISTER LINEATULUS Dickerson


_Type._—U.S.N.M. No. 64263; C. H. Townsend collector.
_Type locality._—Santa Catalina Island, Gulf of California.
_Range._—Known only from Santa Catalina Island.

SCELOPORUS MAGISTER MONSERRATENSIS Van Denburgh and Slevin


_Type._—California Acad. Sci. No. 50509; Joseph R. Slevin collector.
_Type locality._—Monserrat Island, Gulf of California.
_Range._—Southern edge of the Vizcaino Desert in Baja California southward to the southern end of the Sierra de la Gigante; islands adjacent to the coast in the gulf except Santa Catalina.

SCELOPORUS MAGISTER ZOSTEROMUS Cope


_Type._—U.S.N.M. Nos. 5298 (23 specimens), 69472-69488; John Xantus collector.
_Type locality._—Cape San Lucas, Baja California.
_Range._—Cape region of Baja California.
Sceloporus horridus horridus Wiegmann, Herpetologia Mexicana, 1834, p. 50.


*Type.*—Zool. Mus. Berlin; F. Deppe collector.

*Type locality.*—Mexico, here restricted to Cuernavaca, Morelos.

*Range.*—Known in southern Morelos, Guerrero, and Oaxaca in the Balsas Basin, and southern Puebla.

Sceloporus horridus oligoporus Cope


*Type.*—U.S.N.M. Nos. 31386–31393, cotypes.

*Type locality.*—Colima, Colima.

*Range.*—Known from western Guerrero, central and southern Michoacán, Colima, and central Jalisco to Durango.

Sceloporus horridus albiventris Smith


*Type.*—EHT-HMS No. 8519; E. H. Taylor collector.

*Type locality.*—Tepic, Nayarit.

*Range.*—Known from coastal regions from northern Jalisco through Nayarit to Sinaloa.

Sceloporus spinosus spinosus Wiegmann

*Sceloporus spinosus* Wiegmann, Isis von Oken, vol. 21, 1828, p. 370; Herpetologia Mexicana, 1834, p. 50, pl. 7, fig. 3.


*Type.*—? Zool. Mus. Berlin; F. Deppe collector.

*Type locality.*—Mexico, here restricted to Puebla, Puebla.


Sceloporus spinosus caeruleopunctatus Smith


*Type.*—EHT-HMS No. 8467; H. M. Smith collector.
Type locality.—Slopes of Cerro de San Luis, about 15 miles north of Oaxaca, Oaxaca.

Range.—Known only from the highlands of Oaxaca.

SCELORPUS OLIACEUS Smith


Type.—EHT-HMS No. 2508; E. H. Taylor and John S. Wright collectors.

Type locality.—Arroyo Los Olmos, 3 miles southeast of Rio Grande City, Starr County, Tex.

Range.—Extreme south-central Oklahoma, southward through central Texas to southern Tamaulipas, central Nuevo León, southeastern Coahuila, and San Luis Potosí. Recorded from each state named.

UNDULATUS GROUP

Species.—Four, with a total of 13 forms. Extralimital are 9 forms all restricted to the United States and Mexico.

Range.—British Columbia and southern New York south to northern Baja California and central Zacatecas.

KEY TO MEXICAN SPECIES OF THE UNDULATUS GROUP OF SCELORPUS

1. Supraoculars large, entire; femoral pore series separated by 9 or more scales; gular patches small, widely separated.__________________________cautus (p. 117)
   Supraoculars smaller, divided; femoral pore series separated by 8 or fewer scales, or, gular patches large, often covering entire throat.______________2

2. Femoral pore series separated by 9 scales or more; scales on posterior surface of thigh abruptly differentiated from dorsal scales of same member, the median posterior scales not distinctly larger than adjacent lateral posterior scales.__________________________occidentalis biseriatus (p. 118)
   Femoral pore series separated by 8 scales or less; scales on posterior surface of thigh gradually merging with larger dorsals of same member, at least the median posterior scales distinctly larger than the adjacent lateral posterior scales.__________________________undulatus virgatus (p. 118)

3. Males without lateral belly patches; dorsolateral and lateral light stripes very clearly defined.__________________________undulatus consobrinus (p. 118)
   Males with lateral belly patches; dorsolateral and lateral light stripes poorly defined.__________________________undulatus virgatus (p. 118)

SCELORPUS CAUTUS Smith


Type.—EHT-HMS No. 13027; E. H. Taylor and H. M. Smith collectors.
Type locality.—Thirty miles north of El Salado, San Luis Potosí, in the state of Coahuila, Mexico.

Range.—Southern Coahuila, northern San Luis Potosí, and eastern Zacatecas.

**SCLEOPORUS OCCIDENTALIS BISERIATUS** Hallowell


Type locality.—“Borders of El Paso Creek and Tejon Valley,” Calif., here restricted to El Paso Creek.

Range.—Southern Idaho, southwest through Nevada and western Utah into California and the San Pedro Mártir range in Baja California.

**SCLEOPORUS UNDULATUS CONSObRINUS** Baird and Girard

*Sceloporus consobrinus* Baird and Girard, in Marcy and McClellan, Exploration of the Red River of Louisiana, . . ., 1853, pp. 224–226, 237, pl. 10, figs. 5–12.


Type.—Destroyed.

Type locality.—Beckham County, Okla., near the confluence of the north fork of the Red River and Suydam Creek.

Range.—Texas except eastern fourth, southwestern Oklahoma, extreme western and southern fourth of New Mexico, southeastern Arizona in vicinity of San Pedro River, south in Mexico to central Zacatecas, central Chihuahua on the west and central Tamaulipas on the east. Recorded only from Chihuahua, Coahuila, Durango, Nuevo León, and Zacatecas.

**SCLEOPORUS UNDULATUS VIRGATUS** Smith


Type locality.—Above Santa María mine, El Tigre Mountains, Sonora.

Range.—Southeastern Arizona and adjacent Mexico, at high elevations. Recorded in Mexico only from Sonora and Chihuahua.

**GRACIOSUS GROUP**

Species.—One, with three subspecies, only one of which occurs in Mexico.
Range.—Montana and Washington south to extreme northwestern Baja California.

**SCeloPorus graciosus vandenBurgianus** Cope


*Type.*—U.S.N.M. No. 21931; E. A. Mearns collector.

*Type locality.*—Summit of the coast range, San Diego County, Calif.

Range.—Mountains south from Ventura County, Calif., into northwestern Baja California.

**Grammicus Group**

*Species.*—Two, one of which includes three subspecies.

Range.—Extreme southern Texas and Chihuahua south to Oaxaca.

**Key to Species of the Grammicus Group of Sceloporus**

1. Dorsal scales unequal; a series of enlarged scales on each side of middorsal line, separated from each other by small, flat scales. *heterolepis* (p. 121)

   Dorsal scales more or less uniform in size. .......................... 2

2. Scales on sides of neck not abruptly differentiated from dorsal nuchal scales; no enlarged series of scales on sides of neck; dorsal scales 48–66.

   *grammicus grammicus* (p. 119)

   Scales on sides of neck abruptly differentiated from dorsal nuchal scales; two series of enlarged scales on sides of neck posterior to ear; dorsal scales 52–93. .......................... 3

3. Dorsal scales usually less than 70 (52–74) .......................... *grammicus disparilis* (p. 120)

   Dorsal scales usually 70 or more (68–93). *grammicus microlepidotus* (p. 120)

**Sceloporus grammicus grammicus** Wiegmann


Sceloporus rubriventris Günther, Biologia Centrali-Americana, Reptilia and Batrachia, 1890, p. 72, pl. 32, fig. C (Omitlteme, Guerrero; Brit. Mus. Nat. Hist.).

*Type.*—Zool. Mus. Berlin; F. Deppe collector.

*Type locality.*—Mexico, here restricted to Chilpancingo, Guerrero.

Range.—Southern Oaxaca (San Pedro Quiechapa, Ozolotepe) and the Sierra Madre del Sur in Guerrero (Omitlteme, Chilpancingo, Tamazulapan, Iguala).
Puebla,


Sceloporus microlepidotus Wiegmann, Herpetologia Mexicana, 1834, p. 51.—Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 3, 1874, pp. 194–195, pl. 18 bis, fig. 13, a–d.


Sceloporus microlepidopterus Herrera, La Naturaleza, ser. 2, vol. 1, 1890, p. 331 (lapsus calami).


**Type.—** Zool. Mus. Berlin; F. Deppe, collector.

**Type locality.—** Mexico, here restricted to México, D. F.

**Range.—** Southern part of the Mexican plateau from Jalisco, northern Guanajuato, and northern Hidalgo south to Oaxaca. Recorded from the states of Colima, Distrito Federal, Guanajuato, Hidalgo, Jalisco, Oaxaca (Reyes),55 Puebla, Veracruz, México, Morelos, Tlaxcala, and Michoacán.

**SCLELOPORUS GRAMMICUS DISPARILIS** Stejneger


**Type.—** U.S.N.M. No. 33041; William Lloyd collector.

**Type locality.—** Lomita Ranch, 6 miles north of Hidalgo, Tex.

**Range.—** Northern Hidalgo, Guanajuato, and southern Zacatecas northward to central Chihuahua and northern Coahuila and the southern tip of Texas. Recorded from Chihuahua, Coahuila, Durango, Guanajuato, Hidalgo, Nuevo León, San Luis Potosí, Tamaulipas, and Zacatecas.

---

55 Intergrades of *S. g. grammicus* and *S. g. microlepidotus* are from Cerro San Felipe and Cerro San Juan, north of Oaxaca City.
MEGALEPIDURUS


Type locality.—La Cumbre de los Arrastrados (to which the type locality is here restricted), Real Alto, “Ríocho (=Rancho?)” La Berberia, Sierra de Bolaños.

Range.—Known only from western Jalisco (type localities and Cerro Tequila.)

MEGALEPIDURUS GROUP

Species.—Two.

Range.—Western central Veracruz and eastern Puebla.

KEY TO SPECIES OF THE MEGALEPIDURUS GROUP OF SCELOPORUS

1. Dorsal scales 44 to 56; scales between femoral pore series 2 to 5; basal sub-caudals keeled in females; males with distinct blue areas on sides of belly.

   pictus (p. 121)

   Dorsal scales 54 to 62; scales between series of femoral pores 4 to 8; basal sub-caudals smooth in females; males immaculate below.

   mecalepidurus (p. 121)

SCLOPORUS MEGALEPIDURUS Smith


Type.—EHT-HMS No. 7543 (EHT field No. 2908); E. H. Taylor and H. M. Smith collectors.

Type locality.—Near Totalco, Veracruz.

Range.—Known only from northern Puebla, western Veracruz, and Tlaxcala (Apizaco).

SCLOPORUS PICTUS Smith


Type.—Amer. Mus. Nat. Hist. No. 18744; Paul D. R. Rüthling, collector.

Type locality.—Santa Catarina, Puebla.

Range.—Central Puebla and central western Veracruz.

TORQUATUS GROUP

Species.—Twenty-one forms, representing 10 species, are recognized.

Range.—Arizona and Texas south to Guatemala.

This group was in 1938 termed the poinsettii group, inasmuch as Sceloporus poinsettii Baird and Girard (1854) was the oldest name in the group after exclusion of S. torquatus Wiegmann, 1828. The latter name was considered unavailable because it was once (Wagler, 1828) placed in the genus Tropidurus, in which the combination Tropidurus torquatus (Wied, 1820) had already been made (Wied, 1824). Secondary homonyms, such as this, have not, however, been regarded in recent years as permanently suppressed; they are suppressed only as long as they actually remain homonyms. Accordingly, we revert to the name torquatus for this group.
KEY TO SPECIES OF THE TORQUATUS GROUP OF SCELOPORUS

1. Supracaudals in a single series, with no scale divided.  
   Supracaudals in 2 series; or, if in one series, with one or more scales divided.  

2. Femoral pores 8–14; dorsal scales 25–35.  
   Femoral pores more than 14; or dorsal scales more than 35.  

3. Dorsal nuchal collar broadly interrupted medially by a space about 5 scales wide; throat mottled.  
   Dorsal nuchal collar broad, complete; throat not mottled.  

4. Femoral pore counts on both sides usually total more than 21; dorsal scales usually 31 or more.  
   Femoral pore counts on both sides usually total less than 22; dorsal scales generally less than 31.  

5. Dorsal scales 31 or less.  
   Dorsal scales 31 or less.  

6. Nuchal collar divided on each side of neck, the area between lighter in color;  
   dorsal color light, with dark and light spots irregularly placed; size large (maximum snout-vent measurement 129 mm.); dorsal scales more strongly keeled and mucerinita.  
   Nuchal collar broad, complete; dorsal color dark, without light spots; maximum snout-vent measurement 98 mm.; dorsal scales more weakly keeled and mucerinita.  

7. Lateral scales about one-half as large as median dorsals.  
   Lateral scales as large as or larger than dorsals.  

8. Dorsal scales 40 or more; nuchal collar narrow, with light borders broken; a  
   light line on side of head and another on side of neck; each dorsal scale usually with a light median spot.  
   Dorsal scales usually less than 40; nuchal collar broad, with unbroken light  
   borders; no light lines on sides of head and neck; no spots on dorsal scales.  

9. Lateral scales with the terminal point arising well within the free posterior  
   margin; dorsal scales 41 or more.  
   Lateral scales with the terminal point arising at or very near the free posterior  
   margin; head scales not microscopically rugose; no oblique dark blue lines  
   on throat.  

10. Dorsal scales 47 to 54; throat with very distinct, oblique, dark blue lines;  
   head scales not microscopically rugose; oblique dark bands on sides of body  
   distinct; maximum snout-vent measurement 79 mm.  

11. Dorsal scales 55 or more.  
   Dorsal scales less than 55.  

12. Dorsal scales 47 to 53.  
   Dorsal scales less than 47.  

13. Tail with very distinct, broad, alternating dark and light bands, most distinct  
   toward tip, where they are complete; supracaudals in 2 complete rows;  
   head scales very irregular; light borders of nuchal collar broad; a broad light  
   band across neck behind occiput; preocular usually entire; inner row of  
   labiomental scales generally terminating at a point posterior to suture  
   between second and third infralabials.  

---

12a For one other species of this group, see footnote 72a.
Tail without distinct alternating dark and light bands of nearly equal width; bands about tail not complete toward tip; dorsal scales 40 or less. 14

14. Dorsal scales 31 or less. 15
Dorsal scales 32 or more. 16

15. A middorsal series of very large, dark blutches, sometimes fused bandlike; all except males with a pair of parallel, closely placed dark lines down middle of throat; collar 4 scales long or less.

mucronatus mucronatus (p. 124)

No middorsal series of dark blutches; parallel throat lines absent; collar involving 4½ scale lengths or more... mucronatus omiltemanus (p. 124)

16. Each dorsal scale row, in adult males, with a longitudinal light line; dorsals 30 to 38. mucronatus aureolus (p. 124)
Scale rows not with continuous lines. 17

17. Nuchal black collar 4 or 5 scales wide, bordered anteriorly and posteriorly by a light band 1 or 1½ scales wide; each border interrupted medially by a light scale; snout-vent length often more than 100 mm., reaching 143 mm. cyanogenys (p. 125)
Not so; maximum snout-vent length 100 mm. 18

18. Lateral body scales distinctly decreasing in size laterally, at a point halfway between axilla and groin, distinctly smaller than dorsal scales. 19
Lateral body scales not decreasing in size laterally at least up to a point halfway between axilla and groin, where they are still subequal to, or even a little larger than, middorsal scales. 20

19. Dorsal scales on lower foreleg about half size of those on upper foreleg; dark transverse streaks in lateral belly patches; nuchal collar covering six scales medially, or more. jarrovii sugillatus (p. 128)
Dorsal scales on lower foreleg but little if any smaller than those on upper foreleg; no dark streaks in lateral belly patches; nuchal collar less than four scales long middorsally. jarrovii immucronatus (p. 128)

20. Supraoculars essentially in 1 row; if an outer row is evident, it is composed of scales much smaller than those of inner row, and usually number no more than two. jarrovii jarrovii (p. 128)
Supraoculars in 2 rows, those of outer row a little smaller than those of inner, usually numbering 3 or more. 21

21. Adult males black above and below, with orange areas and spots on sides of head, belly, and tail; only throat, underside of tail, and posterior surface of hind leg not black; females somewhat similar, very dark above, the collar poorly defined, young with poorly defined, narrow, light borders on neck collar; dorsal scales average 37.5. jarrovii oberon (p. 129)
Adult males light brown above, with very broad, very well defined nuchal collar; a median area on belly white except in very largest males; sides of abdomen blue, black-edge; females and young with more distinct light borders on nuchal collar; dorsal scales average 40.6. jarrovii minor (p. 128)

SCLELOPORUS SERRIFER SERRIFER Cope


S61316—50—9
Type.—U.S.N.M. No. 34868; Arthur Schott collector.

Type locality.—Yucatán, here restricted to Mérida.

Range.—Northern part of the Yucatán Peninsula. Recorded only
from the state of Yucatán; intergrades with S. s. plioporus are re-
corded from Balchacaj, Campeche.

SCLEPORUS SERRIFER PLIOPOUS Smith


Type locality.—Four miles east of Encero, Veracruz.

Range.—Gulf coast of Mexico from southern Tamaulipas southward
through the basal part of the Yucatán Peninsula to Petén, Guatemala. Recorded from the states of Tamaulipas, San Luis Potosí, Veracruz, and Tabasco (Tenosique); intergrades with S. s. serrifer are recorded
from Balchacaj, Campeche.

SCLEPORUS MUCRONATUS MUCRONATUS Cope


Type.—U.S.N.M. Nos. 25074–9; No. 25077 designated as lectotype
(Smith, op. cit., 1938, p. 583).

Type locality.—Mirador, Veracruz.

Range.—Parts of Hidalgo, Veracruz, Puebla, and México, at high
elevations.

SCLEPORUS MUCRONATUS AUREOLUS Smith


Type.—U.S.N.M. No. 112232; H. M. Smith collector.

Type locality.—Two miles west of Acultzingo, Veracruz.

Range.—Western central Veracruz and southern Puebla.

SCLEPORUS MUCRONATUS OMILTEMANUS Günther

Sceloporus omiltemanus Günther, Biologia Centrali-Americana, Reptilia and
Batrachia, 1890, p. 66, pl. 32, fig. A.


Type locality.—Omilteme, Guerrero, 8,000 feet elevation.

Range.—Central highlands of Oaxaca and the Sierra Madre del Sur in Guerrero.77

SCELOPORUS POINSETTII Baird and Girard


Type.—U.S.N.M. No. 2948 cotypes, 2 specimens, and No. 2952, 2 specimens, “Sonora”; J. H. Clark collector.

Type locality.—“Rio San Pedro of the Rio Grande del Norte, and the province of Sonora,” here restricted to the former locality.

Range.—Southern New Mexico east to central Texas, south through western Nuevo León and southern Coahuila to central Durango. Recorded in Mexico from Chihuahua, Coahuila, Durango, and Nuevo León.

SCELOPORUS CYANOGENYS Cope


Sceloporus torquatus poinsettii, Burt, Copeia, 1932, No. 3, p. 158 (part).


Type locality.—Monterrey, Nuevo León.

Range.—Southern Texas from Devils River and Starr County to central Tamaulipas and Nuevo León. Recorded in Mexico only from the states of Tamaulipas and Nuevo León.

SCELOPORUS BULLERI Boulenger


Type locality.—La Cumbre de los Arrastrados, Jalisco.

Range.—Western and southern parts of Jalisco.

SCELOPORUS TORQUATUS TORQUATUS Wiegmann 78


77 A record for Chiapas (Smith, Publ. Field Mus. Nat. Hist., zool. ser., vol. 26, 1939, p. 221) is in error; the specimen referred to is an S. przewalskii.

78 See footnote 76 for use of this name.


Type locality.—Mexico, here restricted to México, D. F.

Range.—Central Mexico, including Hidalgo, west-central Veracruz, México, Distrito Federal, northern Puebla, eastern Morelos, southern Guanajuato, and northern Michoacán. Recorded only from the states cited.

SCELOPORUS TORQUATUS BINOCULARIS Dunn


Type locality.—Trail from Pablillo to Alamar, Nuevo León.

Range.—Known only from the general region of the type locality.

SCELOPORUS TORQUATUS MELANOGASTER Cope


Type.—U.S.N.M. No. 9877; Alfredo Dugès collector.

Type locality.—“Probably from Guanajuato” = Noria (Michoacán), or Tupátaro, near Cuerámaro, Guanajuato fide Dugès (La Naturaleza, ser. 2, vol. 1, 1887, pp. 114–115), here restricted to Tupátaro.

Range.—Northern Jalisco, west through most of Guanajuato, north through central and southern San Luis Potosí and Zacatecas. Recorded from Aguascalientes, Guanajuato, Jalisco, Michoacán, San Luis Potosí, and Zacatecas.

SCELOPORUS LINEOLATERALIS Smith

**Type.—** EHT–HMS, No. 4323; E. H. Taylor and H. M. Smith collectors.

*Type locality.*—Six miles northeast of Pedriceña, Durango.

*Range.*—Durango.

**SCELOPORUS ORNATUS ORNATUS** Baird


*Type.—** U.S.N.M. No. 2845; Darius Nash Couch collector.

*Type locality.*—Patos, Coahuila.

*Range.*—Southeastern Coahuila.

**SCELOPORUS ORNATUS CAERULEUS** Smith


*Type.—** Univ. Kansas Mus. Nat. Hist. (DHD–HMS field No. 350); D. H. Dunkle and H. M. Smith, collectors (lost?).

*Type locality.*—Five miles south of San Pedro, Coahuila.

*Range.*—Known only from the type locality.

**SCELOPORUS DUGESII DUGESII** Bocourt


*Type locality.*—Colima.

*Range.*—Along the coastal ranges of western Mexico from southern Nayarit to Colima. Recorded from the states of Jalisco, Colima, and Nayarit.

**SCELOPORUS DUGESII INTERMEDIUS** (Dugès)

*Tropidolepis intermedius Dugès*, La Naturaleza, vol. 1, 1869, p.143 (nomen nudum).

*Sce1oporus intermedius Dugès*, La Naturaleza, vol. 4, 1877, pp. 29–34, pl. 1, figs. 21–32.


*Sce1oporus Westphalii Dugès*, La Naturaleza, vol. 4, 1877, p. 30 (substitute name for S. intermedius, which Dugès retained for his description of the species in 1877, although he says “Quería haberle dado el nombre de Sc. Westphalii”).

*Type.—** Mus. Alf. Dugès; two cotypes; collector unknown.
Type locality.—La Noria, near Zamora, hacienda of D. Epifanio Jiménez (in Michoacán).

Range.—Known from Guanajuato and northern and central Michoacán.

**SCELOPORUS JARROVIJ JARROVIJ Cope**


Type.—U.S.N.M. Nos. 8494, two specimens, and 8495, cotypes.

Type locality.—Southern Arizona.

Range.—Central Arizona east to western New Mexico, and south through Chihuahua and western Sonora to extreme western Zacatecas and extreme northern Nayarit. Recorded in Mexico from Chihuahua, Durango, and Sonora.

**SCELOPORUS JARROVIJ SUGILLATUS Smith**


Type.—U.S.N.M. No. 112100; H. M. Smith collector.

Type locality.—Lake No. 4, Zempoala, México. 10,000 feet elevation.

Range.—High elevations in Ajusco range. Known only from the type locality.

**SCELOPORUS JARROVIJ IMMUCRONATUS Smith**


Type.—EHT-HMS No. 9358A (EHT-HMS field No. 500); E. H. Taylor and H. M. Smith collectors.

Type locality.—Ten miles north of El Pinalito, Hidalgo.

Range.—Western Querétaro through Hidalgo, eastern San Luis Potosí, to south-central Veracruz. Recorded from each state cited.

**SCELOPORUS JARROVIJ MINOR Cope**


Type.—U.S.N.M. Nos. 26166–26167.

Type locality.—Zacatecas, here restricted to Valparaiso Mountains.

Range.—Northern Querétaro and Guanajuato through México, western Zacatecas, and most of San Luis Potosí, to western Nuevo León. Recorded from each state cited.
SCELOPORUS JARROVII OBERON Smith and Brown


Type.—U.S.N.M. No. 105823; H. M. Smith and Rozella Smith, collectors.

Type locality.—Arteaga, Coahuila.

Range.—Southeastern Coahuila. Recorded only from the type locality, from Diamond Pass, Mount Zapalinamé, near Saltillo, and from Sierra Guadalupe.

VARIABILIS GROUP

Species.—Nine forms, representing five species, are recognized in this group. One form, S. variabilis olloporus Smith, is extralimital, occurring from Guatemala to Costa Rica.

Range.—Southern central Texas south to Costa Rica, entirely on Atlantic slopes west of the Isthmus of Tehuantepec.

KEY TO SPECIES OF THE VARIABILIS GROUP OF SCELOPORUS

1. Series of femoral pores separated medially by no more than 6 scales........ 2
   Series of femoral pores separated medially by 10 or more scales........ 3

2. Dorsal scales 69 to 76; scales around body 70 to 81; dorsal scale rows at nape 18 to 21.......................... parvus scutulatus (p. 131)
   Dorsal scales 58 to 69; scales around body 61 to 69; dorsal scale rows at nape 15 to 18.......................... parvus parvus (p. 131)

3. Ventral interfemoral scales separated from ventral thigh scales by a group of small scales one-third or one-fourth size of adjacent scales; a rudimentary gular fold; lateral scales much less than half size of ventral scales; dorsal scales 69 to 83........................................ couchii (p. 132)
   Ventral interfemoral scales more or less continuous with ventral thigh scales; no rudimentary gular fold; lateral scales more than one-half size of ventral scales; dorsals usually less than 69........................... 4

4. Dorsal scales 36 to 47; subnasal usually absent; shank and posterior surface of lower foreleg distinctly banded; spots on back distinct in both sexes; preocular usually divided; frontoparietals usually in contact medially.. tepensis (p. 130)
   Dorsal scales 47 or more........................................ 5

5. Males and females immaculate below; femoral pores usually 9 or less on each side (occasionally more in females); dorsal scales 48 to 55; frontoparietals usually separated by an azygous scale; preocular usually entire; subnasal rarely present.............................. 6
   Males with red, blue-bordered areas on sides of abdomen; subnasal usually present; frontoparietals usually in contact medially; preocular usually divided.................................................. 7

6. Postrostrals usually 2, never 4; scales around body usually 53 to 58.
   cozumelae (mainland) (p. 130)
   Postrostrals usually 4, rarely 2 or 3; scales around body usually 59 to 64.
   cozumelae (typical) (p. 130)

7. Dorsal scales 59 or more........................................ 8
   Dorsal scales usually less than 59; dorsolateral light lines 1 and 2 half scale rows wide posteriorly; maximum snout-vent measurement about 74 mm. 9
8. Dorsolateral light stripes very distinct, 2 and 2 half scale rows wide posteriorly; females with sides of belly marked as in males, but less distinctly; maximum snout-vent measurement 71 mm. __variabilis smithi__ (p. 131)

Dorsolateral light stripes not so distinct, 1 and 2 half scale rows wide posteriorly; spots between dorsolateral light stripes very distinct; females with sides of belly immaculate; maximum snout-vent measurement 53 mm. __variabilis marmoratus__ (p. 131)

9. Femoral pores 12 or more __variabilis variabilis__ (p. 130)

Femoral pores 11 or less __variabilis oUoporus__ (p. 129)

**SCOLEPORUS COZUMELAE Jones**


**Type.**—U.S.N.M. No. 13904; “U. S. Fish Commission” collector.

**Type locality.**—Cozumel Island, Quintana Roo.

**Range.**—Northern half of Yucatan Peninsula and adjacent Islands. Recorded from Cozumel and Mujeres Islands and the states of Yucatán and Quintana Roo.

**SCOLEPORUS TEAPENSIS Günther**


**Type locality.**—Teapa, Tabasco.

**Range.**—Restricted to Atlantic slopes from southern Veracruz and northeastern Oaxaca, eastward through Chiapas, Tabasco, and Campeche, and through the Petén region of Guatemala to British Honduras, south to Cobán, Alta Verapaz. Recorded from each state cited in Mexico.

**SCOLEPORUS VARIABILIS VARIABILIS Wiegmann**

_Sceloporus variabilis variabilis_ Wiegmann, Herpetologia Mexicana, 1834, p. 51.


**Type.**—? Zool. Mus. Berlin; F. Deppe collector.

**Type locality.**—Mexico, here restricted to Veracruz, Veracruz.

**Range.**—Southern Tamaulipas along the Atlantic coast to south-central Veracruz (Río Blanco), inland to eastern Querétaro and Puebla; through eastern Oaxaca and western Guatemala, reaching the coast only in Chiapas. Recorded from each state mentioned, and from San Luis Potosí, Guanajuato, and Hidalgo.
SCELOPORUS VARIABILIS SMITHI Hartweg and Oliver


_Type locality._—Guengola Mountain, 6 kilometers northwest of the city of Tehuantepec, Oaxaca.

_Range._—Eastern Oaxaca.

SCELOPORUS VARIABILIS MARMORATUS Hallowell


_Type._—Presumably lost.

_Type locality._—San Antonio, Tex.

_Range._—Atlantic lowlands from San Antonio, Tex., south to southern Tamaulipas, west to Coahuila. Recorded in Mexico only from the states of Coahuila, Nuevo León, and Tamaulipas.

SCELOPORUS PARVUS PARVUS Smith


_Type._—EHT-HMS No. 7120 (EHT field No. 279); E. H. Taylor and H. M. Smith collectors.

_Type locality._—Hills 5 miles west of Sabinas Hidalgo, Nuevo León.

_Range._—Northern Nuevo León to southern San Luis Potosí, west of the Sierra Oriental. Recorded only from the two states cited, and from Coahuila (Arteaga).

SCELOPORUS PARVUS SCUTULATUS Smith


_Type._—EHT-HMS No. 7129; E. H. Taylor and H. M. Smith collectors.

_Type locality._—Thirty miles north of Matehuala, San Luis Potosí.

_Range._—Northwestern San Luis Potosí south along the Sierra Oriental to southern Hidalgo. Recorded only from the two states cited.
Sceloporus couchii 


Type.—U.S.N.M. No. 2739 (9 specimens), cotypes; Lt. Darius Nash Couch collector.

Type locality.—Santa Catarina, Nuevo León (specimens collected by Lt. Couch were also at hand from “Pesqueria Grande,” Mexico).

Range.—Central and northern Nuevo León and eastern Coahuila. In the United States, reported from southern Texas.79

MERRIAMI GROUP

Species.—One, with two subspecies.

Range.—Southwestern Texas and adjacent Coahuila.

KEY TO SPECIES OF THE MERRIAMI GROUP OF SCELOPORUS

1. Anterior section of frontal usually divided; frontoparietals usually divided; outer row of labiomental scales rarely terminating with the first scale wedged between first infralabial and first postmental; head scales rugose; subcaudal surface distinctly banded; gular bars extensive, confluent medially.
   
   merriami annulatus (p. 133)

   Anterior section of frontal rarely divided; frontoparietals rarely divided; outer row of labiomental scales terminating with the first scale wedged between first infralabial and first postmental; head scales smooth; subcaudal surface nearly or quite immaculate, not barred; gular bars short, usually separate medially.-----------------------------merriami merriami (p. 132)

SCELOPORUS MERRIAMI MERRIAMI Stejneger


Type.—U.S.N.M. No. 33039; W. Lloyd collector.

Type locality.—East Painted Cave, near mouth of the Pecos River, Tex.

Range.—The Rio Grande and its immediate tributaries from western Brewster County to southeastern Val Verde County, Tex. The species occurs on the bluffs along the Mexican side of the river, but no specific records for either Coahuila or Chihuahua (in both of which states it should occur) are known.

79 Except possibly for the type specimen of Lysoptychus lateralis, no specimen of S. couchii has ever, to our knowledge, been taken in Texas. We consider it doubtful that the specimen came from there.
SCELOPORUS MERRIAMI ANNULATUS Smith


*Type.*—EHT-HMS No. A787; E. H. Taylor and J. S. Wright collectors.

*Type locality.*—East slope of the Chisos Mountains, Brewster County, Tex.

*Range.*—Southern and central Brewster County, Tex., and eastern Coahuila (Cuatro Ciénegas, Jaral).

MACULOSUS GROUP

*Species.*—One.

*Range.*—Eastern Durango.

SCELOPORUS MACULOSUS Smith


*Type.*—EHT-HMS No. 7638; E. H. Taylor and H. M. Smith collectors.

*Type locality.*—Fourteen miles northeast of Pedriceña, Durango.

*Range.*—Known only from east-central Durango.

CHRYSOSTICTUS GROUP

*Species.*—One.

*Range.*—Yucatán Peninsula.

SCELOPORUS CHRYSOSTICTUS Cope


*Type.*—U.S.N.M. No. 24865, six cotypes.

*Type locality.*—Yucatán, here restricted to Chichen Itzá.

*Range.*—The Yucatán Peninsula, and the area at its eastern base. Recorded in Mexico from Yucatán, Campeche, and Quintana Roo.

SINIFERUS GROUP

*Species.*—Four, one of which includes two subspecies.

*Range.*—Pacific slopes from Guerrero to Nicaragua.

KEY TO SPECIES OF THE SINIFERUS GROUP OF SCELOPORUS

1. Nasals and anterior internasals broadly in contact with rostral; no postrostrals; postanals enlarged in males; ventral scales notched.—ochoterenai (p. 135)

Nasals and anterior internasals separated from rostral by two or more postrostrals; ventral scales not notched. 2
2. One canthal ................................................................. 3
   Two canthals .......................................................... 4
3. Femoral pores 11–12; dorsal scales 38–44 - carinatus (p. 135)
   Femoral pores 3–6; dorsal scales 28–37 - squamosus (p. 135)
4. Femoral pores 3–11; postanals not or slightly enlarged in males.
   siniferus siniferus (p. 134)
   Femoral pores 12–14; postanals distinctly enlarged in males.
   siniferus cupreus (p. 134)

**SCELOPORUS SINIFERUS SINIFERUS** Cope


*Type.*—U.S.N.M. Nos. 30453–30471, cotypes.

*Type locality.*—Pacific side of the Isthmus of Tehuantepec, here restricted to Tehuantepec (city), Oaxaca.

*Range.*—Pacific slopes from western Guerrero to extreme western Guatemala, and inland to Morelos. Recorded in Mexico from Guerrero, Morelos, Oaxaca, and Chiapas.

**SCELOPORUS SINIFERUS CUPREUS** Bocourt 10


---

10 It is with some doubt that the name *cupreus* is applied to the highland form of *siniferus*. This was discussed in some detail previously (Publ. Field Mus. Nat. Hist., zool. ser., vol. 29, 1939, p. 317). Specimens then available showed a higher femoral pore count for the types of *cupreus* and of *cochranae* (the latter from Mount Zempoaltepec, Oaxaca), for which the name *cupreus* was used, than for the few specimens referred to *siniferus* then available from the vicinity of Oaxaca City. However, new material from the latter region (including EHT—HMS 19195-19202, 19479-80) reveals a variation from 35 to 50 (av. 41.2) in 19 counts of dorsal scales, and from 6 to 15 (av. 9.4) in 40 counts of femoral pores.

From these data it is apparent that the highland population of *siniferus* is recognizably different from the typical subspecies and deserves a name. Handicapped by lack of adequate material from Mount Zempoaltepec and its environs, we find it difficult to determine whether the population from that area is the same as that from the Oaxaca region, since the single specimen available has enlarged postanals and broad dorsolateral light stripes. Typically *siniferus* (including Oaxaca specimens) lacks enlarged postanals; none has been observed with them well developed. Likewise, distinct and broad dorsolateral stripes do not occur in male *siniferus* although they are present in females. None of the other characters previously thought to be peculiar to the Mount Zempoaltepec specimen are actually so, as shown by data on the Oaxaca series. Moreover, there is some slight variation in the size of the basal caudal scales and in the width and distinctness of the dorsolateral light stripes in males. In view of this variability in the only two characters defining the Mount Zempoaltepec specimen (*cupreus*, sensu stricto), we believe it best under the circumstances to extend the concept of this name to include the Oaxaca specimens; it should then, of course, be treated as a subspecies, if the present interpretation of the name is correct.
Type locality.—Oaxaca, here restricted to Mount Zempoaltepec.
Range.—Highlands of central Oaxaca.

SCLEOPORUS SQUAMOSUS Bocourt


Type locality.—"Environs de Guatemala et de l'Antigua" [=Volcán Antigua], 1,500 meters.

Range.—Pacific slopes from eastern Chiapas to Costa Rica. Recorded only from Chiapas in Mexico.

SCLEOPORUS CARINATUS Smith


Type.—EHT-HMS No. 15205 (EHT-HMS field No. 4866); E. H. Taylor and H. M. Smith collectors.

Type locality.—Near Tuxtla Gutiérrez, Chiapas.

Range.—Plateau of Chiapas.

SCLEOPORUS OCHOTERENAI Smith


Type.—EHT-HMS No. 7158 (EHT field No. 1075); E. H. Taylor and H. M. Smith collectors.

Type locality.—Two miles north of Mazatlán, Guerrero (12 miles south of Chilpancingo, Guerrero).

Range.—Morelos and Guerrero.

UTIFORMIS GROUP

Species.—One.

Range.—Western Mexico, Sinaloa to Guerrero.

SCLEOPORUS UTIFORMIS Cope


Type.—U.S.N.M. No. 42089; John Xantus collector.

Type locality.—Near Colima, Colima.
Range.—Pacific slopes from southern Sinaloa to western Guerrero, inland about 125 miles in the southern part of its range. Recorded only from Sinaloa, Nayarit, Jalisco, Colima, Michoacán, and Guerrero.

**SCALARIS GROUP**

Species.—Seven forms are recognized at present, representing four species.

Range.—High elevations from southeastern Arizona and central Nuevo León southward to the edge of the plateau in Oaxaca.

**KEY TO SPECIES OF THE SCALARIS GROUP OF SCELPORUS**

1. Nasals and internasals in contact with rostral; no postrostrals. _jalapae_ (p. 136) Nasals and internasals separated from rostral by two postrostrals.  
2. Dorsal scales 50 or more; lateral scales in slightly, although distinctly, oblique rows.  
3. One canthal  
4. Two canthals, the first occasionally forced above canthal ridge by contact of second canthal and subnasal  
5. Tibia/head proportion usually less than 0.90; scales of second pair of postmentals separated medially; dorsal scales usually more than 40; maximum snout-vent measurement 61 mm. _slevini_ (p. 138) 
6. Males with much black in ventral coloration; black shoulder spot with the light blue spot, if present, on its anterior edge; tail with a continuous dark median dorsal stripe; tibia/head proportion usually less than 0.90; maximum snout-vent measurement 65 mm. _unicanthalis_ (p. 138) 

**SCELPORUS JALAPAE** Günther


**Type locality.**—Jalapa, Veracruz.
Range.—Central Veracruz south through eastern Puebla to central Oaxaca. Recorded only from the states cited.

**SCELOPORUS AENEUS AENEUS Wiegmann**


_Type._—Zool. Mus. Berlin; F. Deppe collector.
_Type locality._—Mexico, by inference, here restricted to Tres Cumbres.

Range.—Western Puebla, to central western Michoacán, and Jalisco; north on the plateau to northern Guanajuato. Recorded from the states cited and from Distrito Federal, México, Morelos, and questionably, Jalisco.

**SCELOPORUS AENEUS BICANTHALIS Smith**


_Type._—EHT-HMS No. 7939; E. H. Taylor, collector.
_Type locality._—Cofre de Perote, near Cruz Blanca, Veracruz.

Range.—From northern Hidalgo along the eastern escarpment of the plateau to central Oaxaca, including parts of Puebla, México, and Veracruz. Recorded only from the states cited.

**SCELOPORUS GOLDMANI Smith**


_Type._—Univ. Michigan Mus. Zool. No. 80896; C. L. Lundell collector.
_Type locality._—Charcas, San Luis Potosí.

Range.—Southern Coahuila southward in the central part of the plateau of Mexico to central San Luis Potosí. Recorded only from the states cited.

**SCELOPORUS SCALARIS SCALARIS Wiegmann**


_Type._—Zool. Mus. Berlin; F. Deppe collector.
_Type locality._—Mexico, here restricted to México, D. F.

Range.—Central Durango, southward over the central and southern parts of the Mexican Plateau except (apparently) Guerrero. Re-
corded from Distrito Federal and the states of Durango, Guanajuato, Hidalgo, Jalisco, México, Michoacán, Puebla, and Zacatecas.

**SCLEPORUS SCALARIS UNICANTHALIS Smith**


*Type.*—EHT-HMS No. 7699; H. M. Smith collector.

*Type locality.*—Magdalena, Jalisco.

*Range.*—Known only from the southern border of the Mexican plateau in central Jalisco, from the eastern border of Lake Chapala westward to Nayarit. Recorded only from the state of Jalisco.

**SCLEPORUS SCALARIS SLEVINI Smith**


*Type.*—Calif. Acad. Sci. No. 48013; J. R. Slevin collector.

*Type locality.*—Miller Peak, Huachuca Mountains, Cochise County, Ariz.

*Range.*—Southern Arizona south to northern Durango, and east to to Nuevo León in Mexico. Recorded from the states cited and from Sonora, Chihuahua, and Coahuila.

**PYROCEPHALUS GROUP**

*Species.*—Three.

*Range.*—Pacific slopes from southwestern Chihuahua to northwestern Oaxaca.

**KEY TO SPECIES OF THE PYROCEPHALUS GROUP OF SCLEPORUS**

1. Scales on posterior surface of thigh granular; postfemoral dermal pocket present .......................................................... *gadoviae* (p. 138)
   Scales on posterior surface of thigh larger, imbricate; no postfemoral dermal pocket .......................................................... 2

   2. Dorsal scales larger, 36-41; females not red-headed; males with unbroken lateral belly patches; dark spot on interparietal not enclosing or touching light pineal spot .......................................................... *nelsoni* (p. 139)
   Dorsal scales smaller, 41-50; females red-headed, conspicuously barred below on throat; males (and usually females) with a series of broad dark bars on each side of belly; a dark spot surrounding light pineal spot. .......................................................... *pyrocephalus* (p. 139)

**SCLEPORUS GADOVIAE Boulenger**


*Type locality.*—Mezquiritlan, north of Chilpancingo, Guerrero.
Range.—Southern Michoacán through Guerrero, Morelos, southern Puebla to northern and western Oaxaca. Recorded only from the states cited.

**SCELOPORUS PYROCEPHALUS Cope**


*Type.—U.S.N.M. No. 31495; John Xantus collector.*

*Type locality.—Near Colima, Colima.*

*Range.—Central Jalisco, south and east through Colima and Michoacán to Guerrero. Recorded only from states cited.*

**SCELOPORUS NELSONI Cochran**


*Type.—U.S.N.M. No. 47676; E. W. Nelson and E. A. Goldman collectors.*

*Type locality.—Plomosas, Sinaloa.*

*Range.—Pacific slopes from southwestern Chihuahua to northwestern Jalisco. Recorded from Chihuahua, Sonora, Sinaloa, Nayarit, and Jalisco.*

**Genus SATOR Dickerson**


*Genotype.—Sactor grandeaeus* Dickerson.

*Range.—Santa Cruz and Cerralvo Islands in the Gulf of California.*

*Species.—Two.*

**KEY TO SPECIES OF SATOR**

1. A poorly defined anterior gular fold, without granules; a posterior (true) gular fold immediately in front of arm insertions, but not continuous across throat; median lateral scales very small, often granular, abruptly differentiated from dorsals and ventrals

---

*grandeaeus* (p. 140)

A well defined anterior gular fold, with granules; no evidence whatever, even at sides in front of arm insertions, of a posterior gular fold; median lateral scales larger, gradually merging with dorsals and ventrals

---

*angustus* (p. 139)

**SATOR ANGUSTUS Dickerson**


*Type locality.—Santa Cruz Island, Gulf of California, Baja California.*

*Range.—Restricted to the type locality.*

---

861316—50—10
SATOR GRANDAEVUS Dickerson


**Type locality.**—Cerralvo Island, Gulf of California, Baja California.

**Range.**—Restricted to the type locality.

**Genus UROSAURUS** Hallowell


**Genotype.**—Urosaurus gracius Hallowell.

**Range.**—Southern Texas west to California, entering Utah and Colorado, southward through Baja California and many adjacent islands, and through mainland Mexico from the western Texas border south to the plateau and along the western slopes to Chiapas.

**Species.**—Ten species and 21 forms are recognized; all 10 species and 19 forms are known to occur or are to be expected in Mexico.

**KEY TO MEXICAN ***SPECIES OF UROSAURUS***

1. Enlarged anterodorsal femoral scales smooth. .......... auriculatus (p. 146)
   Enlarged anterodorsal femoral scales strongly keeled. ........... 2

2. Enlarged dorsals in a single broad band, uninterrupted by an intervening series of smaller scales. ................................................................. 3
   Enlarged dorsals separated into two or more parallel series by the presence of a vertebral series of smaller scales. ....................................................... 7

3. Tail two or more times length of head and body combined. .......... gracius (p. 144)
   Tail less than twice length of head and body combined. ............. 4

4. Dermal folds, when present, not heavily crested with tubercles; blue abdominal patches only in males; enlarged dorsals comparatively small. ...... 5
   Dermal folds present, always crested with tubercles of fairly large size; abdominal blue patches sometimes in females as well as males. .......... 6

5. Enlarged dorsals larger, from 17 to 24 in length of head from tip of snout to posterior edge of interparietal; gular region in males deep yellow or orange. .......... nigricaudus (p. 145)
   Enlarged dorsals smaller, 32 to 36 in length of head from tip of snout to posterior edge of interparietal; gular region in males usually blue. ........ microscutatus (p. 145)

6. Four to seven rows of enlarged dorsal scales; abdomen of both sexes with a blue wash and/or blue patches; dorsolateral folds not converging in sacral region. ................................................................. gadovi (p. 145)
   About three rows of enlarged dorsal scales; only males with a blue abdomen; dorsolateral folds converging in the sacral region to form prominent ridges. .......... irregularis (p. 146)

7. Three or fewer rows of lateral tubercles. .............. 12
   Four or more rows of lateral tubercles. .......... 8

---

81 Adapted from Mittleman, op. cit., pp. 127-133.
8. Enlarged dorsals commencing caudal of a line joining the anterior points of insertion of the forelimbs; dorsals weakly keeled, rounded posteriorly, prominently pavemented; general habitus not at all rugose unnicus (p. 147)

Enlarged dorsals commencing craniad of a line joining the anterior points of insertion of the forelimbs, or else equal with such a line; dorsals prominently keeled, usually mucronate or spinose, imbricate; ventrals imbricate; general appearance rugose...

9. Form rugose; enlarged dorsals strongly carinate and prominently mucronate; tubercles of lateral and dorsolateral folds well developed; ventrals mucronate; gular surface generally stippled, with a light median area; blue abdominal patches of males quite extensive... bicarinatus bicarinatus (p. 146)

General appearance somewhat less rugose; enlarged dorsals not so often mucronate; tubercles smaller, sometimes absent; ventrals less mucronate, occasionally rounded; gular surfaces evenly stippled; abdominal blue of males sometimes restricted to small sternal patches.

bicarinatus anonymorphus (p. 146)

Ventral submucronate to mucronate; dorsolateral and lateral tubercles well developed; enlarged dorsals commencing on the shoulders just craniad of a line joining the anterior points of insertion of the forelimbs....

11. Ventrals mucronate, prominently carinated laterally; gular scales with a tendency toward pavementation, especially anteriorly; gular surfaces evenly stippled; abdominal blue of males restricted to small sternal patches.

bicarinatus uniculus (p. 147)

Ventral submucronate (occasionally rounded), only faintly keeled on the lateral portions of the belly, or else not at all; gular scales imbricate; gular region with an even blue wash, and only barely flecked if at all; abdominal blue of males evenly distributed... bicarinatus tuberculatus (p. 147)

12. Enlarged dorsals commencing on nape

13. Enlarged dorsals commencing on shoulders or caudal of them...

14. Enlarged dorsals strongly keeled; scales of primary and secondary series almost equal in size; postfemoral dermal pocket absent or rudimentary; no prominent lateral pattern of dark whorls...

clarionensis (p. 142)

Enlarged dorsals not so rugose, scales of primary series prominently larger than those of the secondary series; postfemoral dermal pocket regularly present; a distinct lateral pattern of dark whorls... ornatus schottii (p. 144)

15. Tail two or more times length of head and body combined... graciosus (p. 144)

Tail less than twice length of head and body combined...

16. Enlarged dorsals extending onto basal portion of tail for a distance equal to length of femur, or more; entire gular region, including sublabials, a uniform bright blue...

ornatus caeruleus (p. 143)

Enlarged dorsals extending onto basal portion of tail for a distance equal to less than length of femur; entire gular region including sublabials never completely blue...

17. Enlarged dorsals often irregularly arranged; lateral tubercles not affecting diagonal arrangements; average size less than 45 mm. from snout to vent...

18. Enlarged dorsals regularly arranged in parallel series on either side of the vertebrals; tubercles in parallel diagonal series; average size greater than 45 mm. from snout to vent...
17. Scales of primary series not twice as large as those of the secondary series; largest of the dorsals inferior in size to enlarged femorals and tibials; ventral interhumeral and interfemoral areas immaculate, or but slightly stippled. *ornatus schmidtii* (p. 143)

Scales of primary series almost twice as large as those of secondary series; largest of dorsals equal to, or larger than, enlarged femoral and tibial scales; ventral interhumeral and interfemoral areas heavily maculated.

*ornatus ornatus* (p. 142)

18. Largest of dorsals equal to, or larger than enlarged femorals; vertebrae extending onto basal portion of tail for a distance equal to half, or slightly more, of length of femur; entire gular region in males, except sublabials, an intense blue; head length/head width ratio averaging 81 percent.

*ornatus chirieahuae* (p. 143)

Largest of dorsals inferior in size to enlarged femorals; vertebrae extending only onto rump, or but slightly farther; no uniform intense blue color present in male gular region; head length/head width ratio averaging 75 percent or less. 

19. Enlarged dorsals separated into 2 parallel series by width of vertebral series, which is greater in width than broadest of enlarged dorsals; prefrontals and frontonasals usually 3 each; general coloration pallid, light tan above, whitish below, males with bright-blue abdominal patches; average head length/width ratio 75.4 percent; average length, snout to vent, 55.1 mm.

*ornatus symmetricus* (p. 144)

Enlarged dorsals separated by a vertebral series whose width is less than that of largest of dorsal scales; prefrontals 2, rarely 3 (by inclusion of an azygous); frontonasals 5; general color variable, but usually dark brown or gray with dark cross bands, and heavily stippled, spotted, or blotched ventrally; abdominal patches in males dark blue to indigo; average head length/head width ratio 70.6 percent; average length, snout to vent, 46.4 mm.

*ornatus linearis* (p. 143)

**ORNATUS GROUP**

*Species.*—Three species and 11 forms are included; all but two subspecies of *ornatus* occur or may be expected to occur in Mexico.

*Range.*—Utah, central Texas, and southern Nevada southward to extreme northern Mexico, reaching farther southward only along the coast to southern Sinaloa; Clarion Island.

**UROSAURUS CLARIONENSIS** *(Townsend)*


*Type.*—U.S.N.M. No. 15904; C. H. Townsend collector.

*Type locality.*—Clarion Island, Revillagigedo Islands, Mexico.

*Range.*—Known only from the type locality (administered by Colima).

**UROSAURUS ORNATUS ORNATUS** *(Baird and Girard)*


Type.—U.S.N.M. No. 2750, two cotypes, male and female.
Type locality.—Restricted to Río San Pedro [=Devils River], Val Verde County, Tex.
Range.—Central and southern Texas, south to northern Coahuila. Has been taken along the Rio Grande River on the American side, and very probably occurs in Coahuila.

UROSAURUS ORNATUS CAERULEUS (Smith)

Type.—Univ. Kansas Mus. Nat. Hist., No. 19237 (Smith and Dunkle field No. 132); Hobart M. Smith and David H. Dunkle collectors.
Type locality.—Thirty miles north of Chihuahua City, Chihuahua.
Range.—Central Chihuahua. Reported only from the type locality and from 20 miles south of Chihuahua.

UROSAURUS ORNATUS CHIRICAHUA (Mittleman)

Type locality.—Pinery Canyon, Chiricahua Mountains, 6,000 feet, Cochise Co., Ariz.
Range.—So far as known, restricted to the Chiricahua and Dos Cabezas Mountains, Ariz.; of probable occurrence in Mexico.

UROSAURUS ORNATUS LINEARIS (Baird)

Type.—U.S.N.M. No. 2759 (now lost); Caleb Kennerly collector. Neotype: U.S.N.M. No. 62077, Los Nogales, Sonora, Mexico; F. J. Dyer, collector.
Type locality.—Los Nogales, Sonora, Mexico.
Range.—Southern Arizona and southern New Mexico, southward to northern Sonora and Chihuahua.

UROSAURUS ORNATUS SCHMIDTI (Mittleman)
UROSARUS ORNATUS SCHOTTII \(^{11}\) (Baird)


**Type.**—U.S.N.M. No. 2761 (now lost); A. Schott collector.


**Range.**—Central Sonora and southward to southern Sinaloa, and the Tres Marias Islands; Tiburón Island.

UROSARUS ORNATUS SYMMETRICUS (Baird)


**Type.**—U.S.N.M. No. 2760 (lost). Neotype: U.S.N.M. No. 2744; Fort Yuma, Imperial County, Calif.; M. Thomas collector.

**Type locality.**—Fort Yuma, Imperial County, Calif.

**Range.**—In United States: southern California and adjacent parts of Arizona south to western Sonora and northern Baja California. Reported from Sonora: 2 miles south of Nogales, Pinetos Camp, 32 miles south of Nogales, Duros Millos, Gran Desierto; Baja California: “Colorado River valley and desert to Delta.”

UROSARUS GRACIOSUS Hallowell


\(^{11}\) The status of this form is questioned by Oliver, loc. cit., who feels that the name should be *Uta ornata lateralis* Boulenenger.


Type locality.— “Lower California” [=southern California], here restricted to Winterhaven (=Fort Yuma), Calif.

Range.— Southern Nevada, western Arizona, southern California, and northern Baja California. Reported from Baja California: 85 miles south of Mexicali, San Felipe. It may occur also in Sonora but is not recorded from there.

NIGRICAUDUS GROUP

Species.— Four.

Range.— Jalisco, Michoacán, and Baja California.

UROSOURUS NIGRICAUDUS (Cope)


Type.— U.S.N.M. No. 5307, 12 cotypes; John Xantus collector.

Type locality.— Cape San Lucas, Baja California.

Range.— Southern tip of Baja California, with adjacent coastal islands, Espíritu Santo, Ballena, San José, and Magdalena.

UROSOURUS MICROSCUTATUS (Van Denburgh)


Type.— Stanford Univ. Mus. No. 1221; J. M. Stowell collector.

Type locality.— San Pedro Mártir Mountains, Baja California.

Range.— Extreme southern California through all of Baja California except the southern fifth. Reported from Baja California: Puerto Escondido, San Xavier, San Quintín, San Pedro Mártir, Mount San Matías, and the Islands of San Francisco, San José, Danzante, Coronado, Carmen, San Marcos, Santa Magdalena.

UROSOURUS GADOVI (Schmidt)


Type locality.— Cofradía, Jalisco, Mexico.

UROSAURUS IRREGULARIS (Fischer)


Type.—Municipal Nat. Hist. Coll. Bremen, Germany, No. 437; collector unknown.

Type locality.—“Aus dem Hochlande von Mexico.”

Range.—Mexico highlands. No specific locality known.

BICARINATUS GROUP

Species.—Three, with a total of six forms.

Range.—Pacific slopes from southwestern Chihuahua south to Chiapas; Socorro Island.

UROSAURUS AURICULATUS (Cope)


Type.—U.S.N.M. No. 7027; Grayson collector.

Type locality.—Socorro Island, Revillagigedo Islands.

Range.—Known only from the type locality.

UROSAURUS BICARINATUS BICARINATUS (Duméril)


Type locality.—Mexico, here restricted to Cuernavaca, Morelos.

Range.—Pacific slopes from Michoacán to central Guerrero, and up the basin of the Río Balsas to southern Puebla. Reported from Puebla: “Puebla,” Izúcar de Matamoros, Tlapalalá, Chiautla; Morelos: Cuernavaca; Guerrero: Agua del Obispo, Cocoyul, Chilpancingo, Mesquititlán, Iguala, Río Balsas, etc.; Michoacán: San Blas, Jorullo.

UROSAURUS BICARINATUS ANONYMORPHUS (Mittleman)

Uta anonymorpha MITTLEMAN, Herpetologica, vol. 2, 1940, pp. 34-38, pl. 3, fig. 2.

**CHECKLIST OF REPTILES OF MEXICO**


*Type.*—U.S.N.M. No. 46988; E. W. Nelson and E. A. Goldman collectors.

*Type locality.*—Tehuantepec, Oaxaca.


**UROSAURUS BICARINATUS NELSONI** (Schmidt)


*Type.*—U.S.N.M. No. 46836; E. W. Nelson and E. A. Goldman collectors.

*Type locality.*—Cuicatlán, Oaxaca.

*Range.*—Known only from the type locality.

**UROSAURUS BICARINATUS TUBERCULATUS** (Schmidt)


*Type locality.*—Colima, Colima, Mexico.


**UROSAURUS UNICUS** (Mittleman)


*Type.*—U.S.N.M. No. 14248; Edward Wilkinson collector.

*Type locality.*—Chihuahua (Batopilas?)

*Range.*—Known only from the type locality.

**Genus UTA** Baird and Girard

Genotype.—*Uta stansburiana* Baird and Girard.

Range.—Western Texas to California, north to Washington, south through Baja California and adjacent islands, and into the northern tier of Mexican states from Sonora to Coahuila.

Species.—Nine are here listed, with a total of eleven forms; only one form (*U. s. stansburiana*) is extralimital, making a total for the genus of 12 forms. The entire genus is badly in need of careful revision, however; considerable shuffling of the nominal species and subspecies is to be expected eventually.

**KEY TO MEXICAN SPECIES OF UTA**

1. Dorsal scales shorter, not imbricate, not mucronate, often with intervening granules; a dark blotch behind axilla.------------------------ 2
   Dorsal scales larger, imbricate at least centrally, usually without intervening granules; caudal scales imbricate, keeled, and strongly mucronate.---- 4

2. Scales on base of tail not imbricate; not unicolor above.--------- 3
   Scales on base of tail imbricate, strongly keeled and mucronate; no dark or light markings above, except rarely a few pale blue dots.

   **nolascensis** (p. 151)

3. Basal caudals weakly keeled, not or but shortly mucronate; about 110–125 dorsals from interparietal to back of thighs; 26–30 of largest dorsals equal length of head to back of interparietal plate; gular region bluish

   **stellata** (p. 150)

   Basal caudals keeled and strongly mucronate; about 106–116 dorsals from interparietal to backs of thighs; 21–25 of largest dorsals equal length of head to back of interparietal plate; gular region blackish.---- **palmeri** (p. 151)

4. Dorsal scales from interparietal to rear of thighs 70–81. **squamata** (p. 149)
   Dorsal scales 82 or more.---------------------------------------- 5

5. No stripes whatever in dorsal pattern of either young or adults; dorsals 92–117, generally more than 100. **taylori** (p. 150)
   Stripes present in young and some adults; dorsals larger or smaller.---- 6

6. No distinct dark blue blotch behind axilla. **mannophorus** (p. 149)
   A distinct dark blue spot or blotch behind axilla.---------------- 7

7. Two rows of postrostrals; both internasals separated by 2 scales from rostral.----------------------------------------------- 8
   One row of postrostrals; one or both internasals separated from rostral by only one scale.------------------------------------- 10

8. Dorsal scales generally 17–23 in a head length (from snout to rear edge of interparietal).--------------------------- 9
   Dorsal scales generally 23–28 in a head length. **stansburiana hesperis** (p. 149)

9. Size larger (64 mm. snout to vent). **martinensis** (p. 149)
   Size smaller (48 mm.).------------------------------------------ **concina** (p. 150)

10. Hind leg longer, 74–85 percent of body length. **stansburiana elegans** (p. 148)
    Hind leg shorter, 65–79 percent of body length.

   **stansburiana stejnegeri** (p. 150)

**UTA STANSBURIANA ELEGANS** Yarrow


Type.—U.S.N.M. No. 12666.

Type locality.—La Paz, Baja California.

Range.—Southern two-thirds of Baja California and on Smiths, Mejía, Ángel de la Guarda, Isla Partida, Isla Raza, Sal Si Puedes, North San Lorenzo, South San Lorenzo, Tortuga, San Marcos, Ildefonso, East and West Las Galeras, Monserrate, San José, San Francisco, Espíritu Santo, and Ballena Islands.

**UTA MANNOPHORUS** Dickerson


Type.—U.S.N.M. No. 64260; C. H. Townsend collector.

Type locality.—Carmen Island, Gulf of California.

Range.—Known only from type locality and Danzante and Coronado Islands, Baja California.

**UTA SQUAMATA** Dickerson


Type.—U.S.N.M. No. 64259; C. H. Townsend collector.

Type locality.—Santa Catalina Island, Gulf of California.

Range.—Known only from type locality.

**UTA STANSBURIANA HESPERIS** Richardson


Type locality.—Arroyo Seco Cañon, near Pasadena, Los Angeles County, Calif.

Range.—San Joaquin Valley of central California southward on the Pacific slope through northwestern Baja California to San Bartolomé Bay; Los Coronados Islands.

**UTA MARTINENSIS** Van Denburgh


Type.—California Acad. Sci. No. 4698; R. H. Beck collector.
Type locality.—San Martín Island, Baja California.
Range.—Known only from the type locality.

UTA CONCINNA Dickerson


_Type._—U.S.N.M. No. 64257; C. H. Townsend collector.
_Type locality._—San Martin Island, Baja California.
_Range._—Known only from the type locality.

UTA STELLATA Van Denburgh


_Type._—California Acad. Sci. No. 4704, R. H. Beck collector.
_Type locality._—San Benito Islands, Baja California.
_Range._—Known only from the type locality.

UTA STANSBURIANA STEJNEGERI Schmidt


_Type locality._—Mouth of Dry Cañon, Alamogordo, Otero County, N. Mex.
_Range._—Western Texas through southern Nevada to eastern California, southward into northeastern Baja California and the northern tier of Mexican states from Sonora to western Coahuila. Reported from _Baja California_: Cocopah Mountains, Volcano Lake, etc.; _Sonora_: Costa Rica Ranch, Sonoyta, Pozo de Luis, Tepoca Bay, San Pedro Bay, Ortiz, Batamotal, Guaymas, Empalme; Islands of Tiburón, Patos, Pelican, and San Esteban; _Coahuila_: 10 miles east of Torreón, San Pedro, 5 miles south of San Pedro, 30 miles west of La Rosa, Jaral, Torréon; _Durango_: Avilco, 5 miles north of Conejos, 25 miles north of Bermejillo, Lerdo; _Chihuahua_: 15 miles south of Juárez.

UTA TAYLORI Smith


_Type._—EHT-HMS No. 10692 (originally field No. 320a), male; E. H. Taylor collector.
_Type locality._—Ten miles northwest of Guaymas, Sonora.

The possibility of intergradation between _Uta taylori_ and _Uta stansburiana_, as recently suggested, is too remote to consider. _Uta stansburiana_ extends throughout the known range of _taylori_.

\[^1\] The possibility of intergradation between _Uta taylori_ and _Uta stansburiana_, as recently suggested, is too remote to consider. _Uta stansburiana_ extends throughout the known range of _taylori_.
Range.—Known only from the region near the type locality. Reported from Sonora: La Posa, 54 miles southwest of Hermosillo.

**UTA PALMERI Stejneger**


*Type.*—U.S.N.M. No. 16002; Edward Palmer collector.

*Type locality.*—San Pedro Mártir Island, Sonora.

*Range.*—Known only from the type locality.

**UTA NOLASCENSIS Van Denburgh and Slevin**


*Type.*—California Acad. Sci. No. 50508; Joseph R. Slevin collector.

*Type locality.*—San Pedro Nolasco Island, Sonora.

*Range.*—Known only from the type locality.

**Family XANTUSIIDAE Baird**


*Genera.*—Four genera known; the only extralimital genus is *Cricosaura* of Cuba.

*Range.*—Southwestern United States to Panama; Cuba.

**KEY TO MEXICAN GENERA OF THE XANTUSIIDAE**

1. Supraoculars present, as small scales above eyes (medial to superciliaries); dorsal scales uniformly granular.  
   Xantusia (p. 154)

   Supraoculars absent, no small scales above eyes (except superciliaries); dorsal scales heterogeneous in size.  
   2

2. Distinct, vertical rows of well-differentiated, enlarged, keeled scales on sides of body, separated from each other by granular area.  
   Lepidophyina (p. 151)

No distinct, vertical rows of enlarged scales on sides of body.  
   Gaigeia (p. 153)

**Genus LEPIDOPHYMA A. Duméril**


*Genotype.*—*Lepidophyма flavimaculatus* A. Duméril.

*Range.*—From Hidalgo, Mexico, south to Costa Rica.

*Species.*—Two, with five forms, one of which (*flavimaculata obscura*) is extralimital.
KEY TO MEXICAN SPECIES OF LEPIDOPHYMA

1. All (except basal) whorls of enlarged scales on tail separated from each other dorsally by 4 rows of scales; femoral pores 15 to 22; median prefrontal normally present, sometimes absent._flavimaculata flavimaculata_ (p. 152)
   All whorls of enlarged scales on tail separated from each other dorsally by 3 rows of scales (rarely feeble evidence of a fourth row); femoral pores less than 14; median prefrontal present or absent.  
   2

2. A median prefrontal smithii smithii (p. 152)
   No median prefrontal.  
   3

3. Scales in posterior temporal region (anterior to ear) minute and very uniform in size, except for a series of relatively large, projecting auricular lobules and a row of larger scales beside the upper temporals; whorls on tail relatively little differentiated smithii occulor (p. 153)
   Scales in posterior temporal region (anterior to ear) larger, irregular in size; auricular lobules poorly defined or absent; no scales bordering upper temporals; tail whorls strongly differentiated smithii tehuanae (p. 152)

LEPIDOPHYMA FLAVIMACULATA FLAVIMACULATA A. Duméril

Lepidophyia flavimaculatus A. Duméril, in Duméril and Duméril, Catalogue méthodique de la collection des reptiles (Paris Museum), 1851, pp. 138–139.


Type.—Mus. Hist. Nat. Paris; P. M. A. Morelet collector.

Type locality.—Province de Petén, Guatemala, here restricted to Río de la Pasión.

Range.—Atlantic slopes from southern Veracruz to British Honduras, excluding the Yucatán Peninsula. Recorded in Mexico from Veracruz: Río de las Playas; Tabasco: Río de las Playas; Chiapas: across the Usumacinta River from Piedras Negras, Petén, Guatemala.

LEPIDOPHYMA SMITHII SMITHII Bocourt


Type locality.—“Tehuantepec, and west coast of Guatemala,” restricted to “Guatemala,” here restricted to Mazatenango.

Range.—Pacific slopes of Chiapas and western Guatemala. Recorded in Mexico only from Chiapas: La Esperanza near Escuintla.

LEPIDOPHYMA SMITHII TEHUANAE Smith


On the basis of the figures given of the types, Smith believes they represent a population characteristic of Chiapas and Guatemala, and not of the region of Tehuantepec city.
Type.—U.S.N.M. No. 111488; H. M. Smith collector.

Type locality.—Cerro Arenal, 30 kilometers west of Tehuantepec, Oaxaca.

Range.—Pacific slopes of Oaxaca and Chiapas (?) in the vicinity of the Isthmus of Tehuantepec. Reported from Tres Cruces, El Limón, Cerro Arenal, La Concepción, Santa Efigenia, mountains near Santo Domingo, near Tehuantepec, Cafetal Concordia; ? Chiapas: Tonalá.

LEPIDOPHYMA SMITHII OCCULOR Smith


Type.—U.S.N.M. No. 47133; E. W. Nelson and E. A. Goldman collectors.

Type locality.—Jalpan, Querétaro.

Range.—Atlantic slopes presumably from northern Veraeruiz into southern Tamaulipas. Recorded only from San Luis Potosí: Tamazunchale; Querétaro: Jalpan.

Genus GAIGEIA Smith


Genotype.—Lepidophyma gaigeae Mosauer.

Range.—Hidalgo and Oaxaca.

Species.—Four.

KEY TO MEXICAN SPECIES OF GAIGEIA

1. All whorls on tail complete, none restricted to dorsal surface. gaigeae (p. 154)

   Some of scale whorls on tail restricted to dorsal surface.----------------------------- 2

2. Only one row of scales on ventral surface between whorls of enlarged scales

   (i. e., every third whorl restricted to dorsal surface)----------------------------- 3

   Proximally 2 and distally 3 rows of scales on ventral surface between whorls of

   enlarged scales--------------------------------------------- sylvatica (p. 154)

3. Numerous, very closely approximated, enlarged, keeled scales, separated by

   small granules, present on sides of body; 2 or 3 rows of granules in vertebral

   region--------------------------------------------- radula (p. 153)

   Dorsal scales practically uniform in size; 4 rows of granules in vertebral region.

   dontomasi (p. 153)

GAIGEIA DONTOMASI Smith


Type.—U.S.N.M. No. 111473; Thomas MacDougall collector.

Type locality.—Lachiguiri, Oaxaca, 7,100 feet elevation.

Range.—Known only from the type locality.

GAIGEIA RADULA Smith


Type.—U.S.N.M. No. 111472; H. M. Smith collector.
Type locality.—San José Manteca, 5 kilometers from San Carlos Yautepec, Oaxaca.

Range.—Known only from the type locality.

**Gaigeia Sylvatica** (Taylor)

*Lepidophyма sylvatica* Taylor, Copeia, 1939, pp. 131–133, figs. 1, 2.


Type.—EHT-HMS No. 16259; E. H. Taylor collector.

Type locality.—Seven miles north of Zuicaltipan, Hidalgo.

Range.—Known only from the type locality.

**Gaigeia Gaigeae** (Mosauer)


Type.—Mus. Comp. Zool. No. 42145; Walter Mosauer collector.

Type locality.—Durango, Hidalgo.

Range.—The immediate vicinity of the type locality.

**Genus Xantusia** Baird


Genotype.—*Xantusia vigilis* Baird.

Range.—Southern California, Arizona, southern Utah, Baja California, and adjacent islands.

Species.—Five, one of which includes two subspecies; three species occur in Mexico.

**KEY TO MEXICAN SPECIES OF XANTUSIA**

1. Ventral plates in 12 series
2. Ventral plates in 14 series henshawi (p. 155)
2. A single frontal, eye large vigilis (p. 154)
2. A pair of frontals, eye small gilberti (p. 155)

**Xantusia Vigilis** Baird


Type.—U.S.N.M. No. 3063 (three cotypes); John Xantus collector.

Type locality.—“Fort Tejon,” California.

Range.—Southwestern Utah and eastern California southward into northern Baja California as far as San Felipe Bay and San Matías Pass.
XANTUSIA HENSHAWI Stejneger


Type.—U.S.N.M. No. 20339; H. W. Henshaw collector.

Type locality.—Witch Creek, San Diego County, Calif., elevation 2,700 feet.

Range.—Extreme southwestern California and northern Baja California, in the San Pedro Mártir Mountains.

XANTUSIA GILBERTI Van Denburgh


Type.—California Acad. Sci. No. 401; Gustav Eisen collector.

Type locality.—San Francisquito, Sierra Laguna, Baja California.

Range.—Cape region of southern Baja California.

Family SCINCIDAE Gray


Genera.—About 60, of which three occur in Mexico. Of these three, none are confined to the Western Hemisphere. Only two other genera occur in the Americas: Cryptoblepharus (South America), a wanderer from the Pacific Islands, and Neoseps (Florida).

Range.—Southern Canada south through Central America and South America to Argentina; West Indies; Africa and Madagascar; south Asia; China; Japan; Philippine Islands; Dutch Indies; Pacific Islands and Australia.

KEY TO MEXICAN GENERA OF SCINCIDAE

1. Two supranasals between rostral and frontonasal; frontoparietal always divided; lower eyelid with a translucent disk or not; palatine bones in contact or not. ................................. 2

Supranasals absent, the single frontonasal in contact with rostral; frontoparietal divided or not; an undivided translucent disk on lower eyelid; palatine bones in contact on median line. .................................. Scincella (p. 156)

2. An undivided translucent disk on lower eyelid; palatine bones in contact on median line. .......................................................... Mabuya (p. 155)

Lower eyelid without a translucent disk, or if so it is divided into several parts; palatine bones separated on median line. ................................. Eumeces (p. 160)

Genus MABUYA Fitzinger

Mabuya Fitzinger, Neue Classification der Reptilien, 1826, pp. 23, 52.

Genotype.—Lacertus mabouya Lacépède (by absolute tautonymy, fide M. A. Smith, Fauna British India, etc., 1935, Reptilia and Am-
phibia, vol. 2, p. 257). This is contrary to opinion 92 of the International Commission on Zoological Nomenclature, which states that *Scincus sloani*ii of Daudin is the type. Fitzinger did not list that form.85

Range.—Southern Mexico on both coasts; Central America, South America; Africa, Madagascar; southern Asia, Philippines, East Indies.

Species.—About 85 species, only one of which occurs in Mexico.

**MABUYA MABOUYA MABOUYA** (Lacépède)


Genotype.—*Scincus lateralis* Say.

Range.—Greater part of eastern United States east of the Rocky Mountains; Mexico from Hidalgo southward through Central America to Panama. Ceylon, southern Asia, China, Philippines, East Indies, Tasmania, Africa, New Zealand.

Species.—About 50 species. Seven species and 10 forms are known in the Americas, all except *S. incertum* of Guatemala occurring in Mexico (even this exception may actually occur).

**KEY TO MEXICAN SPECIES OF SCINCELLA**

1. Frontoparietal divided; many median subcaudals in contact on each side with 2 scales. 2

Frontoparietal single; median subcaudals in contact with only 1 scale on each side 6

---


84 This is the only name based upon continental (as opposed to island) specimens north of South America. It may be revived, for undoubtedly geographically segregated variants of taxonomic magnitude do exist within the enormous range now included within this supposedly single race.

85 Adoption here of this name is the decision of solely the senior author.
2. Usually 3 or more pairs of nuchals; upper tertiary temporal (occasionally split) in contact with parietal; 26-28 scale rows around middle of body; limbs short, separated by 10-12 scales when adpressed. \textit{laterale} (p. 159)

Two pairs of nuchals, or fewer; upper tertiary temporal separated from parietal by contact of upper secondary temporal and nuchal. \textit{cherriei} (p. 158)

3. Limbs, when adpressed, touching or overlapping in adults, forelimb reaching rear corner of eye; scale rows at middle of body often 30 or 32. \textit{caudaequinas} (p. 158)

Limbs, when adpressed, separated from each other by 1 or more scale lengths; scale rows at middle of body rarely 30 and apparently never 32. \textit{silvicolum} (p. 158)

4. “Usually 1 pair of nuchals or less; lamellae under fourth toe 19 or fewer; dorsolateral light stripe continuous”. (Smith) \textit{gemmingeri} (p. 159)

Usually 2 pairs of nuchals; lamellae under fourth toe 20 (constant?); dorsolateral light stripe dark-spotted, interrupted. \textit{silvicolum} (p. 158)

5. Usually nuchals on the two sides 1-2 or more (89 percent); lateral light line along head and neck poorly defined; axilla-groin/snout-vent percentage 59; 65 mm. maximum snout-vent measurement. \textit{cherriei} (p. 158)

Usually nuchals on the two sides 1-1 or less (88 percent); lateral light line along head and neck fairly well defined; axilla-groin/snout-vent percentage 50; 54 mm. maximum snout-vent measurement. \textit{cherriei} (p. 158)

6. Scale rows 30 or more

Scale rows less than 30

7. Tail blue or blue-gray, with traces of a cross-banded pattern evidenced by feeble dark bars visible laterally or by transverse series of light spots; legs longer, usually overlapping in adults

Tail pink or reddish, with no traces of a cross-banded pattern, but instead with a longitudinal dark line on each side at least at base; legs shorter, never overlapping in adults; dorsal scales 65 to 79, usually 69 or more. \textit{cherriei} (p. 158)

8. Dorsal scales 65 to 72, average 69.1. \textit{cherriei} (p. 158)

Dorsal scales 59 to 67, average 63.2. \textit{cherriei} (p. 157)

9. Tail blue or blue-gray, with traces of a cross-banded pattern evidenced by feeble dark bars visible laterally or by transverse series of light spots; legs longer, usually overlapping in adults; dorsal scales 54 to 60, average 57. \textit{cherriei} (p. 158)

Tail pink or reddish, with no traces of a cross-banded pattern, but instead with a longitudinal dark line on each side at least at base; legs shorter, never overlapping in adults; dorsal scales 58 to 72, average over 66. \textit{cherriei} (p. 158)

\textbf{SCINCELLA \textit{cherriei} (Cope)}


\textit{Type locality}.—Palmar, Costa Rica.
Range.—Tabasco and northern Chiapas on Atlantic slopes, and Costa Rica on Pacific slopes, eastward to Panama. Recorded in Mexico only from Tabasco: Teapa; Chiapas: Palenque.

SCINCELLA CHERRIEI IXBAAC (Stuart)

Scincella cherriei ixbaac, MIDDLEMAN, Herpetologica, vol. 6, 1950, p. 20.

Type.—Univ. Michigan Mus. Zool. No. 80820; Milton Trautman.
Type locality.—Chichen Itzá, Yucatán.

Range.—The peninsula of Yucatán, southward as far as Campeche and northern Petén, Guatemala. Recorded in Mexico only from Yucatán: Chichen Itzá; Campeche: Tres Brazos.

SCINCELLA CHERRIEI STUARTI (Smith)

Scincella cherriei stuarti, MIDDLEMAN, Herpetologica, vol. 6, 1950, p. 20.

Type.—U.S.N.M. No. 115174; H. M. Smith collector.
Type locality.—Potrero Viejo, Veracruz.
Range.—Central Veracruz, in foothills, southward to the Isthmus of Tehuantepec; recorded only from the states of Veracruz and Oaxaca (between Río Grande and La Gloria).

SCINCELLA CAUDAEOQUINAE (Smith)


Type.—Univ. Illinois Mus. Nat. Hist. No. 10131; J. P. Craig.
Type locality.—Salto Cola de Caballo, 25 miles south of Monterrey, Nuevo León.
Range.—Eastern foothills from central Nuevo León southward to southern San Luis Potosí (known only from the type locality and 10 miles west of Naranjo, San Luis Potosí).

SCINCELLA SILVICOLA (Taylor)

Scincella silvicola, MIDDLEMAN, Herpetologica, vol. 6, 1950, p. 20.

Type.—EHT-HMS No. 10033, adult female; E. H. Taylor collector.
Type locality.—Forested hill about 10 miles southeast of Córdoba, near San Lorenzo, Veracruz.
Range.—Atlantic slopes in central Veracruz and in the headwaters of the Río Papaloapam in Oaxaca. Recorded from Veracruz: San José de Gracia; Oaxaca: Cuicatlán.

**SCINCELLA GEMMINGERI GEMMINGERI** *(Cope)*


*Lygosoma (Mocoa) Gemmingeri*, Bocourt, Mission scientifique au Mexique. . .

Études sur les reptiles, livr. 7, 1881, pp. 449–450.


**Type.**—U.S.N.M. No. 6331, four cotypes; François Sumichrast collector.

**Type locality.**—Orizaba, Veracruz.

**Range.**—Eastern Hidalgo, central and southern Veracruz, and Oaxaca south to Tehuantepec, on the slopes of the plateau and in lowlands. Recorded only from Veracruz: Tequeyutepec, La Perla, Río Verde, Jalapa, Orizaba; *Hidalgo*; Zacualtipan (possibly referable to *L. g. forbesorum*); Oaxaca: Ixcuintepec, Tres Cruces, San José Manteca, 17 miles north of Niltepec, Cafetal Concordia.

**SCINCELLA GEMMINGERI FORBESORUM** *(Taylor)*


**Type.**—EHT-HMS No. 10043, E. H. Taylor collector.

**Type locality.**—La Placita, Hidalgo, 8 miles south of Jacala, elevation 7,000 feet.

**Range.**—Known only from higher mountains of Hidalgo, in the vicinity of the type locality.

**SCINCELLA LATERALE** *(Say)*


**Type.**—U.S.N.M. No. 3152.

**Type locality.**—Mississippi River, below Cape Girardeau, Mo.

**Range.**—New Jersey and southward to the Gulf and to central Coahuila, excluding the lower Río Grande Valley. Reported from Coahuila: Nogales (Sabinas River near Múzquiz).
SCINCELLA ASSATA ASSATA (Cope)


Type locality.—Volcan Isalco, El Salvador.

Range.—Pacific slopes, southeastern Chiapas southward to Honduras. Recorded in Mexico from Chiapas: Huchuetan, Escuintla.

SCINCELLA ASSATA TAYLORI (Oliver)


Type locality.—Santiago, Colima.

Range.—Colima to southwestern Chiapas, in lower areas. Recorded from Colima: Santiago, Paso del Río; Guerrero: Between Rincón and Cajones, Mazatlán, Chilpancingo, Agua del Obispo, Tierra Colorada; Jalisco: Tenacatita; Chiapas: Tonalá; Oaxaca: Matías Romero, Santa Esfigenia, Tapanatepec.

Genus EUMECES Wiegmann

Eumeces WIEGMA N, Herpetologia Mexicana, 1834, pt. 1, p. 36.
(type, Lamprosauros guttulatus Hallowell = Plestiodon obsoletum Baird and Girard).


Range.—Southern Canada southward through the United States, Mexico, and Central America to Nicaragua; Bermuda; North Africa; southwestern Asia; China; French Indo-China; Japan.

Species.—About 58 species and 76 forms are recognized; 23 forms of 22 different species occur in Mexico.

KEY TO MEXICAN SPECIES OF EUMECES

1. Median dorsal scales at least three times the width of adjoining scales—2
   Dorsal scales not greatly widened, rarely twice as wide as adjoining scales—3
2. Scales in 21 rows about middle of body; 3 broad black stripes begin on snout, pass back and break up on middle of back........ schwartzei (p. 162)
Scales in 19 rows about middle of body; no stripes, but scales with black dots.................................................................altamirani (p. 163)

3. Three supraoculars; no postnasal; 1 postmental.............................................. 4
Four supraoculars; postnasals and postmentals variable.................................... 5

4. Parietals enclose interparietal; scale rows about middle of body, 24; seventh labial in contact with upper secondary temporal..........................dugesii (p. 169)
Parietals not enclosing interparietal; scale rows usually 24; seventh labial not in contact with upper secondary temporal...lynxe furcirostris (p. 163)

5. Postmentals normally 2................................................................. 6
Postmental normally 1................................................................. 12

6. Body without white stripes at any time; lateral scale rows generally oblique; young black with a blue tail; head with cream spots; adults olive with darker areas on scales; limbs large, overlapping when adpressed; scales 26–28 rows; 125 mm. snout to vent.........................obsoletus (p. 165)
Body with white stripes in young, retained in adult, or approaching uniform olive coloration in adults, especially old males; lateral scale rows parallel; young never black but usually with blue or pink (red) or orange tail... 7

7. Dorsolateral light stripes distinct and confined to third scale row. multivirgatus (p. 165)
Dorsolateral light stripes not involving third scale row at all, or else including also second scale row.................................................. 8

8. Median light line beginning between arms running forward bifurcating on head; or median line lost and the bifurcating lines only, remain on head............................. 9
No trace of a median light line or bifurcating lines on head; usually 4 light lines, 2 lateral and 2 dorsolateral.................................................. 10

9. Parietals enclose interparietal; scale rows usually 28 at middle of body; postnasal usually present; median white line to shoulder; limbs short, not touching when adpressed............................................callicephalus (p. 164)
Parietals not enclosing interparietal; bifurcating lines on head, but median line obsolete; postnasal absent; limbs touch in young when adpressed, in adults separated by 3 or 4 scale lengths; postmental single or divided; 26–28 scale rows; maximum size about 70 mm........tetragrammus (p. 165)

10. Parietals enclose interparietal; 24 scale rows; seventh labial broadly in contact with upper secondary temporal; tail orange in young; 4 light lines retained in adults; limbs fail to touch when adpressed.........................lagunensis (p. 167)
Parietals do not enclose interparietal.................................................. 11

11. Snout-vent length greater than 75 mm., or interparietal nearly parallel-sided; specimens under 50 mm. snout-vent length with pink tail (no blue); 8 supralabials on each side (98 percent)...gilbertirubricaudatus (p. 167)
Snout-vent length not more than 75 mm.; interparietal bluntly wedge-shaped; juveniles, usually adults, with blue tail; 7 supralabials on one or both sides (94 percent)............................................................skiltonianus (p. 167)

12. A median light line bifurcating on frontal.............................................. 13
Median light line present or absent, if present bifurcating behind frontal... 14

13. Median line from middle of body bordered with darker, and bifurcating on anterior half of frontal; limbs separated when adpressed; usually 24 scales about body.................................................................lynxe lynxe (p. 163)
Median light line bifurcating on the posterior part of frontal, or all lines lost in adult olive coloration; limbs overlapping when adpressed; subcaudals widened; maximum snout to vent length, 100 mm...sumichrasti (p. 164)

14. Parietals enclosing interparietal.................................................. 15
Parietals not enclosing interparietal.................................................. 17
15. Limbs long, overlapping when adpressed; strong, wide dorsolateral light lines; seventh labial broadly in contact with secondary upper labial; no primary temporal; frontoparietals narrowly separated; scale rows, 28; snout to vent, 65 mm. \( \text{colimensis} \) (p. 169)

Limbs shorter, separated when adpressed. \( \text{indubitus} \) (p. 168)

16. Seventh labial not touching upper secondary temporal; primary temporal as large as upper secondary temporal; 24 scale rows; maximum length snout to vent, 51 mm. \( \text{parvulus} \) (p. 166)

Seventh labial broadly in contact with upper secondary temporal; primary temporal present, smaller than upper secondary temporal; 22–24 scale rows around middle of body; maximum length snout to vent, 66 mm. \( \text{dicei} \) (p. 169)

17. Primary temporal absent; limbs short, not touching when adpressed; seventh labial in contact with upper secondary temporal; 22 scale rows; dorsolateral lines growing dim posteriorly. \( \text{indubitus} \) (p. 168)

Primary temporal present. \( \text{parviauriculatus} \) (p. 166)

18. Four narrow dark dorsal lines and narrow dorsolateral light lines; 22–24 scale rows; no postnasal; limbs widely separated when adpressed; maximum snout to vent length, 76 mm. \( \text{copei} \) (p. 166)

Dorsal pattern not of 4 narrow black lines. \( \text{brevilineatus} \) (p. 164)

19. Bifurcating lines on head; lateral and dorsolateral light lines rarely reaching farther back than arm; scale rows, 26–28; maximum snout to vent length, 66 mm. \( \text{brevirostris} \) (p. 168)

No bifurcating lines on head. \( \text{parviauriculatus} \) (p. 166)

20. A postnasal; usually bronze olive, with lateral brown stripe, with sometimes trace of a dorsolateral light line; 26 scale rows; the limbs adpressed, the toes may or may not touch. \( \text{humilis} \) (p. 166)

No postnasal.

21. Scale rows 20 around body; ear small, median dorsal scales widened. \( \text{parviauriculatus} \) (p. 166)

Scale rows more than 20. \( \text{brevirostris} \) (p. 168)

22. Scale rows 22 (rarely 24); body slender, dorsolateral lines distinct to tail; seventh labial usually touching upper secondary temporal; snout to vent 56 mm. \( \text{ochoterena} \) (p. 169)

Scale rows usually 24, rarely 22; dorsolateral lines to tail or not; parietal usually not enclosed but occasionally may be enclosed, variable. \( \text{ochoterena} \) (p. 166)

**Eumeces schwartzei** Fischer


Type.—Naturh. Mus., Hamburg, Germany.**

Type locality.—“Einer kleinen Insel in der Laguna de Términos (Campeche Bai).”

Range.—Campeche, Tabasco, Yucatán; in Central America it is known from Guatemala and British Honduras. Reported from Campeche: Tres Brazos, Encarnación, Isla de Carmen; Tabasco: Tenosique; Yucatán: Chichen Itzá.

** This specimen was actually taken aboard ship in a cargo of dyewood and collected on board ship, alive, collector unknown. It was sent to the Zoologischen Garten in Hamburg. At its death it was sent to the Naturhistorischen Museum of Hamburg.
EUMECES ALTMIRANI Dugès


Type.—In Museo "Alfredo Dugès" in Colegio del Estado de Guanajuato in Guanajuato (without number); Federico Altamirano collector.

Type locality.—"Regiones Cálidas del Estado de Michoacán" (later designated by Dugès as Apatzingán de la Constitución, Michoacán).

Range.—Known only from Michoacán, on the southern slopes of the plateau. Reported from El Sabino, Apatzingán.

EUMECES LYNXE LYNXE (Wiegmann)

Scincus quinquelineatus var., Wiegmann, Isis von Oken, 1828, p. 373 (non Linnaeus).


Eupreps lynxe Wiegmann, Herpetologia Mexicana, 1834, pp. 36-37.


Plestiodon Bellii Gray, Catalogue of the specimens of lizards in the collection of the British Museum, 1845, p. 92 (type locality not designated, here restricted to El Chico, Hidalgo).

Type.—Zool. Mus., Berlin; F. Deppe collector.

Type locality.—"Specimen nostra prope Chico inventit Deppe." Very probably (and here restricted to) El Chico (or Mineral El Chico) near Pachuca, Hidalgo, where the species has been found to be abundant.

Range.—Known definitely from Hidalgo, Veracruz, San Luis Potosí and Puebla; recorded, probably erroneously, from Guerrero, Guanajuato and Michoacán. Reported from Hidalgo: Zacualtipan, San Miguel, Guerrero, Durango, El Chico; Veracruz: Jalapa, Mount Orizaba; San Luis Potosí: Alvarez; Puebla: Zacatlán.

EUMECES LYNXE FURCIROSTRIS Cope


Type.—Acad. Nat. Sci. Philadelphia No. 11327; Dr. Flohr collector.

Type locality.—Jalapa, Veracruz.

Range.—Puebla, central northern Veracruz, and possibly eastern Hidalgo. Reported from Puebla: Tezuitlán; Veracruz: Toxtlacuaya, Jalapa, Las Vigas, Cofre de Perote; Hidalgo: Zacualtipan.
EUMECES SUMICHRASTI (Cope)


Type.—U.S.N.M. No. 6601; François Sumichrast collector.

Type locality.—“Orizava” ex errore, = Potrero, Veracruz. The specimen bears Sumichrast’s original tag, “Potrero, No. 4, F. Sumichrast.” Later Sumichrast published notes that he had found the species “en los encinales de Potrero, cerca de Cordoba a una altura de 500 metros.”

Range.—Veracruz to Chiapas in lowlands; Central America; Honduras, British Honduras, and Guatemala. Reported from Veracruz: Potrero, Jalapa; Chiapas: Palenque, La Esperanza, Mineral de Santa Fé.

EUMECES BREVILINEATUS Cope


Type.—Lectotype, designated by Taylor, loc. cit., U.S.N.M. No. 10159, one of four cotypes; G. W. Marnock collector.

Type locality.—Helotes, Bexar County, Tex.

Range.—Tamaulipas and Nuevo León; in the United States: southern Texas. Reported from Tamaulipas: 26 kilometers north of El Limón; Nuevo León: 4 miles west of Sabinas Hidalgo, 31 miles south of Sabinas Hidalgo.

EUMECES CALUCEPHALUS Bocourt


Type locality.—Guanajuato [Guanajuato].

Range.—Sonora (Álamos), Chihuahua, Durango, Zacatecas, Jalisco, Guanajuato, Michoacán, Querétaro (Huaxteca Potosina), Nayarit; in United States: Arizona (southeastern).
EUMECES TETRAGRAMMUS (Baird)


_Type._—U. S. N. M. No. 3124; Darius Nash Couch collector.

_Type locality._—“Lower Rio Grande.” (Later indicated by Baird, as “Matamoras, Mex.” A second specimen was at hand collected by Dr. Kennerly at “Salado River,” U. S. N. M. No. 3139.)

_Range._—Tamaulipas, San Luis Potosí, and Veracruz, in lowlands; in the United States: southern Texas. Reported from _Tamaulipas:_ 3½ miles west of El Forlón, San José, Matamoros; _San Luis Potosí:_ Antiguo Morelos, Ébano; _Veracruz:_ south of Tampico.

EUMECES OBSELOTUS (Baird and Girard)


_Type._—U. S. N. M., No. 3133; John H. Clark collector.

_Type locality._—“Valley of the Río San Pedro of the Río Grande del Norte,” now Devils River, Tex.

_Range._—Tamaulipas, Nuevo León, Coahuila, and Chihuahua. In United States: Arizona, New Mexico, and Texas, north to Utah, Colorado, Nebraska, and Kansas. Reported from _Tamaulipas:_ Matamoros; _Nuevo León:_ Santa Catarina; _Chihuahua:_ Chihuahua (city); _Coahuila:_ Cuatro Ciénegas.

EUMECES MULTIVIRGATUS (Hallowell)


_Type._—Acad. Nat. Sci. Philadelphia No. 9371; Dr. Hammond collector.

_Type locality._—“Posa Creek, 460 miles west of Fort Riley, Kansas” (= Cow Creek,88a Larimer County, Colo.).

_Range._—The central high plains and proplateau areas south from southwestern Nebraska to extreme western Texas, northern Arizona,

88a The original tag attached to the specimen definitely states Cow Creek. The “Posa” is a bad interpretation of poor chirography.
and presumably western Chihuahua. Recorded in Mexico only from Chihuahua: no specific locality.\(^5^9\)

**Eumeces Humilis** Boulenger


*Type locality.*—Presidio [Sinaloa]. There is a strong presumption that the locality is in Sinaloa although not known positively to be so.

*Range.*—Sinaloa. Known only from the type locality. (Specimens previously referred to this species from Texas and New Mexico have been referred to another form, *Eumeces taylori*, by Smith.)

**Eumeces Parvulus** Taylor


*Type.*—U.S.N.M. No. 56903; collector unknown. Originally in the Julius Hurter collection.

*Type locality.*—Tepic, Nayarit.

*Range.*—Colima, Nayarit and Sinaloa. Reported from Colima: Paso del Río; Nayarit: Tepic, Compostela, Minimán; Sinaloa: Plomo-sas.

**Eumeces Parviauriculatus** Taylor


*Type.*—U.S.N.M. No. 47536; E. A. Goldman collector.

*Type locality.*—Near Álamos, Sonora.

*Range.*—Sonora and western Chihuahua. Reported from Sonora: Near Álamos; Chihuahua: Mojárachic.

**Eumeces Copei** Taylor


*Type.*—EHT-HMS No. 29717 (EHT field No. 3859); H. M. Smith and E. H. Taylor collectors.

*Type locality.*—Ten miles southeast of Asunción, in the western part of the state of México.

---

\(^5^9\) This specimen, U. S. N. M. No. 30833, very likely represents a distinct species (see Taylor, loc. cit., p. 333), but we hesitate to attempt a characterization of it because of certain obvious abnormalities the specimen possesses, and because of its intimate and entirely uncertain relationship with populations farther to the north in Arizona and New Mexico.
Range.—Puebla, Distrito Federal, Morelos, México, and Michoacán. Reported from Puebla: Near Río Frío (México); México: Río Frío, near Salazar, 10 miles southeast of Asunción, 8 miles west of Villa Victoria, 15 kilometers west of Toluca, 3 miles west of Zinacatepec; Morelos: Tres Marias, Lagunas de Zempoala; Distrito Federal: Santa Lucía; Michoacán: Cerro de Tecolote, Mount Tancítaro.

**EUMECES SKILTONIANUS** (Baird and Girard)


_Type._—U.S.N.M. No. 3172, two cototypes, smaller one designated as lectotype; Rev. George Gary (or Geary) collector.

_Type locality._—Oregon, here restricted to The Dalles.

Range.—Southern British Columbia to northwestern Baja California and adjacent islands, and eastward to western Montana and Utah. Reported in Baja California from Los Coronados Islands, Todos Santos Islands; Ensenada, San José, Rancho San José, Alcatraz, San Pedro Mártir Mountains, Arroyo Encantada, San Quintín, etc.

**EUMECES GILBERTI RUBRICAUDATUS** Taylor

_Eumeces quadrilineatus_ Hallowell (nec Blyth), Reports of explorations and surveys, to ascertain the most practicable and economical route for a railroad from the Mississippi River to the Pacific Ocean, 1853–6, vol. 10, pt. 4, Zool., Rept., 1859, p. 10, pl. 9, figs. 3a, b, c, d (type locality, “Upper California, near Mohave river and in San Bernardino Valley,” here restricted to Mojave River; U. S. Nat. Mus.).


_Type._—California Acad. Sci. No. 39002.

_Type locality._—Tehachapi Mountains, Calif.

Range.—Southern California and extreme northwestern Baja California. Reported in Mexico only from Baja California: San Antonio del Mar, North Coronado Island.

**EUMECES LAGUNENSIS** Van Denburgh


Type.—Originally California Acad. Sci. Nos. 400 and 402, cotypes (destroyed in the earthquake and fire in 1906); Gustav Eisen collector. Neotype.—U. S. N. M. No. 67398; W. M. Mann collector; February 1924, “on the Trail between Loreto and Comondú.”

Type locality.—San Francisquito, Sierra de la Laguna, Baja California.

Range.—Confined to the southern third of Baja California in the mountains. Reported from Baja California: Comondú; San Francisquito, Sierra de la Laguna; between Loreto and Comondú.

EUMECES BREVIROSTRIS ( Günther)


Type locality.—Oaxaca, here restricted to the city of Oaxaca.

Range.—Durango to Oaxaca and east to Veracruz, in highlands. Reported from Guerrero: Omilteme; Oaxaca: Cerro San Felipe, La Parada, San José Lachiguiri, Tehuantepec; Veracruz: Laguna, Perote, San Bernardino, Totalco, Orizaba; Puebla: Near Río Frío (México), Texmelucan, El Seco (Km. 205); Durango: Ciudad, El Salto, Coyote; Jalisco: La Cumbre of the Arrastrados, Talpa, Mascota, Sierra de Juanocatlan, La Laguna; Michoacán: No specific record.

EUMECES INDubitus Taylor


Type.—EHT–HMS No. 29715 (EHT field No. 1731); E. H. Taylor and H. M. Smith collectors.

Type locality.—Kilometer 63 (Mexico-Cuernavaca highway), near Cuernavaca, Morelos.

Range.—Known from the states of Morelos, México, and Michoacán. Reported from Morelos: near Cuernavaca, Tepoztlán, Lagunas de Zempoala, Huajintlán; México: Asunción; Michoacán: 15 miles southeast of Zitácuaro, Puerto Hondo.
EUMECES DUGESII Thominot


_Type locality._—"Province Guanajuato," here restricted to the city of Guanajuato.

_Range._—Guanajuato, Jalisco (?), and Michoacán. Reported from Guanajuato: Guanajuato; Michoacán: Carapa, Rancho San José, 9 miles west of Zacapú, Tangancicuaro, Patamban, El Soledad (Tancitaro), Apatzingán; Jalisco: Nevado de Colima.90

EUMECES COIMENSIS Taylor


_Type._—Chicago Nat. Hist. Mus. No. 1649; collector unknown.
_Type locality._—Colima, Colima.
_Range._—Known only from the type locality.

EUMECES DICEI Ruthven and Gaige


_Type locality._—Marmolejo, Tamaulipas.
_Range._—Tamaulipas and Nuevo León. Reported from Nuevo León: Pablillo, Cieneguillas south of Galeana; Tamaulipas: Marmolejo.

EUMECES OCHOTERAEE Taylor


_Type._—EHT-HMS No. 29716 (EHT Field No. 1015); E. H. Taylor and H. M. Smith collectors.
_Type locality._—Mazatlán; "4 mi. north of Chilpancingo, Guerrero" (actually about 15 km. south of Chilpancingo).
_Range._—Known only from Guerrero; probably confined to the Sierra Madre del Sur. Reported from Agua del Obispo, 7 miles east of Chilpancingo, Mazatlán, vicinity of Chilpancingo, between Rincón and Cajones, Chilapa.

90 Gadow (Proc. Zool. Soc. London, 1905, pp. 195, 218–219) records a "fuscirostris" (=furcirostris) from this locality. Inasmuch as the most distinctive feature of _d. furcirostris_ is the possession of three supra-oculars, it seems highly probable that Gadow observed this character in his specimen from the Nevado. In such case the specimen would be _dugesii_, the only species in that area that possesses this character.
Family ANELYTROPSIDAE Cope


Genera.—A single genus, Anelytropsis Cope, is known.
Range.—Foothills of the plateau in central eastern Mexico.

Genus ANELYTROPSIS Cope


Genotype.—Anelytropsis papillosus Cope.
Range.—Central eastern part of Mexico.
Species.—One.

ANELYTROPSIS PAPILLOSUS Cope


Type.—Unknown; originally two cotype specimens.
Type locality.—Near Jalapa, Veracruz.
Range.—Eastern San Luis Potosí and eastern central Veracruz. Reported from San Luis Potosí: 20 miles south of Valles; Veracruz: Jalapa, Motzorongo.

Family TEIIDAE Gray


Genera.—About 40 genera, 3 of which, Cnemidophorus, Ameiva, and Gymnophthalmus, occur in Mexico.

KEY TO MEXICAN GENERA OF TEIIDAE

1. Anterior nasal plates not separated by a frontonasal; frontoparietal present; inner finger well developed; scales granular; eyelids present............ 2
Nasal plates widely separated by a frontonasal; inner finger absent; no eyelids; scales cycloid, quincuncial; prefrontals present but no frontoparietals.

Gymnophthalmus (p. 192)

2. Central gular scales not or scarcely enlarged as compared with adjacent lateral scales; tongue somewhat widened posteriorly, and notched behind; no basal sheath between base of tongue and larynx........ Cnemidophorus (p. 174)
Central gular scales enlarged, often abruptly so; tongue not, or less widened posteriorly, a basal sheath evident between the base of tongue and larynx.

Ameiva (p. 170)

Genus AMEIVA Meyer

Ameiva Meyer, Synopsis reptilium . . . sistema generum methodum . . , 1795, p. 27.

Genotype.—Lacerta americana Seba [=Ameiva ameiva (Linnaeus)].
Range.—Tamaulipas and Jalisco to Brazil; West Indies.
Species.—About 13, with some 28 forms. Two species with 11 subspecies occur in Mexico.
CHECKLIST OF REPTILES OF MEXICO

KEY TO MEXICAN SPECIES OF AMEIVA

1. Outer row of ventrals considerably smaller than the others.
   festiva edwardsii (p. 174)

   Outer row of ventrals as large as the others... 2

2. Preanal scales in 2 rows. 3
   Preanal scales in one row or with no more than 1 posterior scale divided... 9

3. Two rows of granules between third supraocular and superciliaries; third
   supraoculars generally completely separated from frontoparietals by
   granules. undulata parva (p. 173)
   A single row of granules between third supraoculars and superciliaries; third
   supraoculars in contact with frontoparietals anteriorly... 4

4. Median gulars abruptly enlarged. 6
   Median gulars little enlarged, gradually merging with lateral gulars... 5

5. Upper lateral vertical light lines from axilla to groin, 12 or more light lines.
   undulata gaigeae (p. 172)
   Upper lateral vertical lines from axilla to groin, 11 or less light lines.
   undulata hartwegi (p. 171)

6. Median gulars irregular or no more than 2 regular (87 percent); lamellae
   under fourth toe 28 or more; no upper lateral light stripe, although large
   spots take its place in adult males. undulata podarga (p. 172)
   At least 3 median gulars regular (100 percent in all except u. amphigramma,
   with 87 percent); lamellae variable; upper lateral light stripe present or
   absent... 7

7. Dorsolateral dark stripes present except in some large adults, in which the
   upper lateral light spots if present are much narrower than the spaces between
   them; lamellae on the fourth toe usually (88 percent) 27 or less.
   undulata stuarti (p. 173)
   No dorsolateral dark stripes; upper lateral light spots as wide as or wider
   than spaces between, or represented by a continuous upper lateral light
   stripe; lamellae variable... 8

8. Upper lateral light spots or lines not contacting dorsolateral light area or
   line in adult males, separated by a narrow dark area; lateral gulars not or
   scarcely enlarged. undulata amphigramma (p. 172)
   Upper lateral light spots merged with dorsolateral light line in adult males;
   lateral gulars markedly enlarged. undulata thomasi (p. 173)

9. Last preanal scale generally (86 percent) divided; lateral markings showing
   little tendency to be arranged vertically; middorsal markings greatly re-
   duced. undulata dextra (p. 173)
   Last preanal scale generally entire; lateral markings tending to be arranged
   vertically; middorsal markings well developed... 10

10. Upper lateral light spots in adult males large, wider than intervening dark
    spaces; generally (95 percent) 5 or less rows of preanals.
    undulata sinistra (p. 174)
    Upper lateral light spots in adult males small, narrower than intervening
    dark spaces in all males; frequently (65 percent) 6 or more rows of preanals
    undulata undulata (p. 174)

AMEIVA UNDULATA HARTWEGI Smith

pl. 2, fig. b.

861316—50—12
Type.—U.S.N.M. No. 108600; H. M. Smith collector.

Type locality.—“Across the Río Usumacinta from Piedras Negras, Guatemala, in Chiapas, Mexico.”

Range.—Atlantic slopes of Mexico and Guatemala from the vicinity of the southeastern end of Laguna de Términos south and eastward across the base of the Yucatán Peninsula to northwestern Honduras. Recorded in Mexico only from Chiapas (the type locality); it probably occurs also in Campeche and Quintana Roo.

AMEIVA UNDULATA GAIGEEAE Smith and Laufe

Type—EHT-HMS No. 11927; Progreso, Yucatán; Hobart M. Smith collector.

Type locality.—Progreso, Yucatán.

Range.—Northern half of the Yucatán Peninsula and southward to the island Carmen along the extreme eastern coast. Reported from Yucatán: Chichen Itzá, La Vega, Tunkas; Quintana Roo: Cobá, Mujeres Island; Campeche: Champotón.

AMEIVA UNDULATA PODARGA Smith and Laufe

Type.—EHT-HMS No. 14471; Hobart M. Smith and David H. Dunkle collectors.

Type locality.—Seven miles west of Ciudad Victoria, Tamaulipas.

Range.—Known from southern Tamaulipas and eastern San Luis Potosí and probably extends into Veracruz and Hidalgo lowlands. Reported from Tamaulipas: Alta Mira, Victoria, Hacienda La Clementina near Forlón, Antiguo Morelos; San Luis Potosí: near Ciudad del Maíz, Río Guayala near Magiscatzin, near Valles, Huichihuayán, Tamazunchale.

AMEIVA UNDULATA AMPHIGRAMMA Smith and Laufe

Type.—EHT-HMS No. 11983; H. M. Smith collector.

Type locality.—San Andrés Tuxtla, Veracruz.

Range.—Northern Veracruz southward at low elevations to the Isthmus of Tehuantepec, westward into valleys extending into extreme eastern Oaxaca and probably northeastern Puebla. Reported from Veracruz: Atoyac, Boca del Río, Cuatotolapam, Lake Catemaco, Potrero Viejo, Rodríguez Clara, San Andrés Tuxtla, etc.; Oaxaca: Cosolapa, Matías, Agua Fría, Tuxtepec; Tabasco: La Vent; Puebla: 1 mile northeast of Huauchinango.
AMEIVA UNDULATA THOMASI Smith and Laufe
Type.—EHT-HMS No. 15327, H. D. Thomas, collector.
Type locality.—La Libertad, Chiapas, near Río Cuilco where it crosses the Guatemalan border.
Range.—Upper tributaries of Río Grijalva in the interior of Chiapas and adjacent Guatemala. Recorded in Mexico only from the type locality.

AMEIVA UNDULATA STUARTI Smith
Type.—U.S.N.M. No. 108601, H. M. Smith collector.
Type locality.—Palenque, Chiapas.
Range.—Atlantic slopes of Mexico from the middle of the Isthmus of Tehuantepec eastward in the lowlands to the southern borders of Laguna de Términos and to Tenosique, Tabasco; southward up the valley of the Río Grijalva at least as far as Tuxtla Gutiérrez, Chiapas. Reported from Chiapas: Palenque, San Ricardo, Tuxtla Gutiérrez; Campeche: Balchacaj, Tres Brazos; Tabasco: Tenosique, Frontera, Teapa; Oaxaca: Isthmus of Tehuantepec (intergrades?).

AMEIVA UNDULATA PARVA Barbour and Noble
Type.—Mus. Comp. Zool. No. 5831; Van Patten collector.
Type locality.—Guatemala (restricted by Smith and Laufe to Mazatenango).
Range.—Pacific slopes from the Isthmus of Tehuantepec in Oaxaca to Costa Rica. Reported in Mexico from Oaxaca: near Niltepec; Chiapas: Tapachula, Tonalá, La Esperanza, Huchuetán.

AMEIVA UNDULATA DEXTRA Smith and Laufe
Type.—EHT-HMS No. 11966; E. H. Taylor and H. M. Smith collectors.
Type locality.—Near Rincón, Guerrero.
Range.—Southern slope of the Sierra Madre del Sur. Known from Oaxaca and Guerrero. Reported from Oaxaca: Juquila; Guerrero: Chilpancingo, Cocoyul, Los Cajones, Rincón, Órganos, El Treinta, Acapulco, 8 miles east of Coyuca.
AMEIVA UNDULATA SINISTRA Smith and Laufe


*Type.*—EHT-HMS No. 11908; H. M. Smith collector.

*Type locality.*—Manzanillo, Colima.

*Range.*—Pacific slopes from Jalisco to the arid Balsas basin in Michoacán, thence inland along the northern drainage of the Río Balsas to Puebla. Reported from *Colima*: Colima, Quesería, Hda. Paso del Río, Salvador, Pascuales, Periquillo, Manzanillo; *Jalisco*: Tenacatita, Ixtapa, Tenacatita Bay; *Michoacán*: Uruapan; *Puebla*: Chiautla; *Morelos*: Puente de Ixtla.

AMEIVA UNDULATA UNDULATA (Wiegmann)

*Cnemidophorus undulatus* Wiegmann, Herpetologia Mexicana, 1834, p. 27.


*Type.*—Zool. Mus. Berlin; F. Deppe collector.


*Range.*—Pacific slopes of the Isthmus of Tehuantepec as far west as Puerto Ángel, and eastward to Niltpec. Known only in the state of *Oaxaca*: El Limón, Palmar, Tres Cruces, Cafetal Concordia, Juchitán, Puerto Ángel, Totontepec, Tehuantepec, Ranchería La Manga.

AMEIVA FESTIVA EDWARDSII Bocourt


*Type locality.*—Izabal and Santa María de Panzós, near Río Polochic, Guatemala; here restricted to Panzós.

*Range.*—Atlantic foothills from the Isthmus of Tehuantepec through Honduras, in heavy forests. Recorded in Mexico only from *Chiapas*: Ixtacomitán, Palenque, across the Río Usumacinta from Piedras Negras (Petén, Guatemala).

Genus CNEMIDOPHORUS Wagler


**Genotype.**—*Seps murinus* Laurenti.

**Range.**—North America south from Oregon, Wisconsin, and Maryland through Central America to southern Brazil and Bolivia.

**Species.**—About 18, and a total of about 47 species and subspecies; 15 species and 41 forms, as here recognized, occur in Mexico. These 18 species represent 5 distinct groups, all but one of which (the *lemniscatus* group, restricted to Central and South America, of 3 species and 5 forms as recognized by Burt, *op. cit.*), occur in Mexico.

KEY TO MEXICAN SPECIES OF CNEIMIDOPHORUS

1. Only one frontoparietal scale  
   Two frontoparietals

2. Dorsal surface unicolor, not striped or reticulated. *hyperythrus pictus* (p. 186)
   Dorsal surface striped or reticulated, not unicolor

3. Lateral stripes more or less broken; at least a few spots or cross bars present; body often tessellated; adults large. *ceralbensis* (p. 188)
   Lateral stripes distinct and in even, continuous lines; spots and cross bars absent; size always small

4. Usually 2 or 3 longitudinal light lines in vertebral area
   Usually 1 median dorsal light line, forked anteriorly or not

5. Usually 3 light lines along middle of back; second supraocular usually without granules between it and frontal. *hyperythrus hyperythrus* (p. 187)
   Usually 2 light lines; second supraocular usually at least in part separated from frontal by granules. *hyperythrus beldingi* (p. 186)

6. Dorsal stripe not of same intensity and width as lateral stripes but fainter and usually narrower
   Dorsal stripe of same intensity and width as lateral stripes

7. More than half of second supraocular usually in contact with frontal.
   *hyperythrus schmidtii* (p. 187)
   Half or less than half of second supraocular usually in contact with frontal.
   *hyperythrus beldingi* (p. 186)

8. Ground color of sides (between the 2 distinct lateral stripes) reddish gray; dorsal ground color gray or black, abruptly contrasted with lateral ground color. *hyperythrus danheimae* (p. 186)
   Ground color of sides not usually reddish gray, but often reddish or brownish; dorsal ground color of same general shade as that of the sides and not in sharp contrast to it. *hyperythrus caeruleus* (p. 186)

9. Supraocularears normally 3; striped at least in the young
   Supraocularears normally 4, or 3, no stripes at any stage, but a pattern of small, scattered light spots on a dark background

10. Scales between midventral scutes and anus 4 to 8; snout-vent measurement never exceeding 92 mm
    Scales between midventral scutes and anus 9 to 15; snout-vent measurement reaching 138 mm

11. Adults and subadults spotted, the spots often dim. *guttatus guttatus* (p. 179)
    Adults and subadults (as well as young) striped, the stripes sometimes broken up into spots. *guttatus immutabilis* (p. 180)

---

81 The species possessing this character were formerly regarded as members of the genus *Verticaria* Cope. Burt (U. S. Nat. Mus. Bull. 154, 1931, p. 14) has concluded, however, that the group is not a natural one, since one species (*ceralbensis*) is related to the *tessellatus* group, while the others are related to the *sejJS* group. The character is, furthermore, subject to a slight variation.
12. An accessory scute between frontoparietal and parietal.  

*deppii cozumelus* (p. 179)

No accessory scute. .......................................................... 13

13. Femoral pores usually 17 or fewer (81–82 percent) ........................................ 14

Femoral pores usually 18 or more (94 percent) ........... *deppii deppii* (p. 178)

14. Lower precocular generally (79 percent) in contact with loreal; a broad mid-

dorsal light band. .................................................. *deppii lineatissimus* (p. 179)

Lower precocular generally (73 percent) not in contact with loreal; narrow light

stripes on back as on sides. ............................... *deppii oligoporus* (p. 179)

15. Enlarged scales immediately preceding gular fold (mesoptychials) small (not

more than 3 times diameter of smallest scales in median part of preguar fold); or, if larger, no evidence whatever of longitudinal light lines, or else at least a few dark spots on throat below level of ear openings. 16

Mesoptychials larger; a lined pattern always present in the young, generally

in adults; no dark spots on throat below level of ear openings; or, if less than

3 times diameter of smallest scales in median part of preguar fold, 7 light

lines and intervening dark spaces absolutely straight, continuous and
disconnected. .......................................................... 28

16. Mesoptychials moderately or considerably enlarged, more than 3 times

diameter of smallest scales in median part of preguar fold. 17

Mesoptychials smaller. ........................................... 19

17. No evidence of a longitudinal arrangement of lines on back. ............................. 18

Dorsal surface distinctly lined .................................. *tesselatus* (p. 188)

18. No spotting in area above level of tympanum and anterior to insertion of

forearm, spots relatively well defined, very small, not yellowish.

*bacatus* (p. 187)

Spots present in area above tympanum and anterior to insertion of forearm;
spots often obscure and diffuse, usually orange or yellowish.

*catalinensis* (p. 188)

19. Ventral surfaces of tail and limbs more or less suffused with red or pink. 20

Not so ........................................................................... 21

20. Black markings on temporal regions absent or few or ill-defined; dorsal and

lateral markings on body showing both transverse and longitudinal arrange-

ment in at least large specimens; each dark unit or spot square or rectangul-

ar in outline, not noticeably rounded. ...................... *tigris rubidus* (p. 191)

Black markings on temporal region well defined; dorsal and lateral markings

on body showing a predominantly longitudinal arrangement; back covered

by alternate longitudinal chains of black and white, individual spots in
black chain rarely square or rectangular. .................... *tigris celeripes* (p. 190)

21. Dorsal markings consisting of either fine reticulations or light spots, and these

markings without longitudinal arrangement. .............. 22

Dorsal markings with at least some indication of longitudinal arrange-

ment. ........................................................................... 23

22. Labials deep or moderate brown or black .................. *tigris Martyris* (p. 191)

Labials light gray, slate or light brown .................... *tigris canus* (p. 190)

23. Dorsal light lines only 3 on each side (exclusive only of a pair of lateral lines),

twice as wide as interspaces. .................................. *maximus* (p. 188)

Dorsal light lines more numerous or narrower, or both. 24

24. Dorsal pattern of 6–8 persistent longitudinal light stripes ............................... 25

Dorsal pattern of fewer than 6 unbroken light stripes; at least lower lateral

stripe on each side broken by dark vertical bars. .............. 27
25. Dark fields between dorsal light stripes unspotted and unbroken throughout life; entire ventrum black in adults. \textit{tigris aethiops} (p. 189)
At least some of dark fields broken into rows of spots; ventrum not entirely black in adults. .................................................... 26

26. Middorsal dark field or row of spots distinctly wider than paravertebral light stripes; throat white, spotted or barred with black. \textit{tigris multiscutatus} (p. 191)
Middorsal dark field or row of spots as wide as or slightly narrower than paravertebral light stripes; throat and chest black; belly spotted. \textit{tigris gracilis} (p. 188)

27. Four longitudinal light stripes persisting unbroken in middorsum of adults, the dark fields between represented by rows of black spots; sides with bold vertical bars. \textit{tigris tigris} (p. 189)
Six longitudinal light stripes persisting unbroken in adults, or, if any broken, dorsal pattern generally light and without any bold markings. \textit{tigris marmoratus} (p. 190)

28. Anterior nasal in contact with second upper labial; femoral pores never more than 14. ....................................................... 42
Anterior nasal usually separated from second upper labial; if in contact, femoral pores usually more than 14. ........................................ 29

29. Enlarged scales preceding gular fold relatively small, frequently grading into granular scales of fold. ........................................ 30
Enlarged scales preceding gular fold relatively large, abruptly differentiated from granular scales of throat. ........................................ 31

30. Postantebrachials enlarged; ventral surfaces light bluish in adults; juvenile striped pattern retained throughout life; maximum snout-vent length 70 mm.; dorsal scales relatively large, 50 to 70 in a transverse row between enlarged belly plates. \textit{inornatus} (p. 184)
Postantebrachials not enlarged; belly, chest, or gular region dark (black) in adults; snout-vent length reaching 95 mm.; dorsal scales 85 or more in a transverse row. \textit{tigris aethiops} \(^2\) (p. 189)

31. In addition to a dorsolateral light stripe on tail, a lateral light stripe present, bordered below by a black line (near base of tail) continuing onto otherwise unicolor posterior surface of thigh; no light spots in dark field between stripes in adults; postantebrachials not enlarged in either sex; neither sex dark blue or orange below. \textit{sexlineatus} (p. 184)
No lateral light stripe on tail, at least not distinguishable from ventral tail color; thighs mottled or unicolor, no posterior light stripe except sometimes feebly indicated at insertion of leg; light spots often present in dark fields between stripes; postantebrachials distinctly enlarged generally, sometimes less so in females; males frequently dark blue on belly or chest, or else cream, not at all bluish. ....................................................... 32

32. Only 5 light lines, including a vertebral, 2 dorsolateral and 2 lateral lines through the ear; ventral surfaces cream even in males, never blue; light lines very broad, more than half width of intervening dark spaces in adults; no marking between light lines, except series of small spots in dark spaces in juveniles (disappearing in adults). \textit{burti} (p. 185)
Not so. ............................................................................. 33

\(^2\)Young only key out here; adults key out through couplet 16 to couplet 25.
33. Space between paravertebral light stripes (i.e., those originating even with parietal scales, not with median interparietal) narrower or at least no wider than space between dorsolateral and upper lateral light stripes; if stripes not visible, they are replaced by dorsal pattern of dark cross bars. 34
Space between paravertebral light stripes wider; if stripes not visible, they are replaced by numerous scattered light spots. 37

34. Adults without a cross-barred pattern. 35
Adults cross-barred. 36

35. Dark interspaces or rows of dark spots continuing to area above hind legs. sackii stictogrammus (p. 183)
Dark interspaces or rows of spots disappearing anterior to area above hind legs. sackii semifasciatus (p. 184)

36. Postantebaschials generally scutelike. sackii scalaris (p. 182)
Postantebaschials generally polygones, little enlarged. sackii australis (p. 181)

37. Hind legs mottled in adults, but not with small light spots; dorsal pattern cross-barred in adults. sackii sackii (p. 180)
Hind legs with small light spots in adults; dorsal pattern not cross-barred. 38

38. Stripes absent in adult, replaced by numerous, scattered light dots especially toward rear half of body. sackii bocourtii (p. 181)
Stripes present in adults. 39

39. Collar black in adults; stripes obsolete except for lateral stripes; keels on caudal scutes nearly parallel. sackii mariarium (p. 183)
Collar light; stripes not obsolete; keels on caudal scales strongly oblique. 40

40. Ventral surfaces of tail and limbs pink. sackii angusticeps (p. 183)
Ventral surfaces of tail and limbs cream or white. 41

41. Stripes becoming rows of light spots in adults. sackii communis (p. 182)
Stripes persistent. sackii gularis (p. 183)

42. Femoral pores 11–15 in known specimens. Iabilis (p. 185)
Femoral pores 19–20 in known specimens. gadovi (p. 185)

DEPPII GROUP

Species.—Six forms, belonging to two species, now recognized.

Range.—Central Veracruz on the Atlantic coast and Nayarit on the Pacific southward to Costa Rica.

CNEMIDOPHORUS DEPPII DEPPII Wiegmann

Cnemidophorus deppei Wiegmann, Herpetologia Mexicana, 1834, p. 29.—Bocourt, Mission scientifique au Mexique . . . , Études sur les reptiles, livr. 3, 1874, p. 281, pl. 20c, fig. 5, pl. 20d, fig. 1.


Type.—Zool. Mus. Berlin; F. Deppe collector.

Range locality.—Mexico, restricted to Tehuantepec, Oaxaca.

Range.—Southern Guerrero on Pacific slopes and the Salamá Basin in Guatemala on Atlantic slopes, south to Costa Rica. Recorded in Mexico only from the states of Guerrero, Oaxaca, and Chiapas.

**Cnemidophorus Deppii Cozumelus Gadow**


**Type locality**.—Cozumel Island, east coast of Quintana Roo.

**Range**.—The eastern portion of the Yucatán Peninsula. Recorded in Mexico only from Quintana Roo (Mujeres and Cozumel Island).

**Cnemidophorus Deppii Lineatissimus Cope**


**Type**.—U.S.N.M. Nos. 24937-40, 32299-32314, 20 cotypes; Hoge collector.

**Type locality**.—Colima and Guadalajara (Jalisco), here restricted to Colima, Colima.

**Range**.—Pacific slopes from Nayarit to central and perhaps southern Guerrero. The range interdigitates with that of *C. d. deppii* in central Guerrero. Recorded only from Nayarit, Jalisco, Colima, Michoacán, Guerrero, and Puebla (Chiautla).

**Cnemidophorus Deppii Oligoporus Smith**


**Type**.—Chicago Nat. Hist. Mus. No. 29145; Julius Friesser.

**Type locality**.—Perez, Veracruz.

**Range**.—Atlantic slopes from northern Veracruz to Campeche. Recorded only from the states of Veracruz and Campeche (Ciudad del Carmen).

**Cnemidophorus Guttatus Guttatus Wiegmann**


**Type.**—Zool. Mus. Berlin; F. Deppe collector.

**Type locality.**—Mexico, here restricted to Veracruz, Veracruz.

**Range.**—Atlantic slopes, from central Veracruz to the Isthmus of Tehuantepec. Recorded only from the states of Veracruz and Oaxaca.

**Cnemidophorus guttatus immaculata Cope**


*Cnemidophorus microlepidopus* Cope, loc. cit. (U.S.N.M. No. 30187; “West Tehuantepec,” here restricted to the city of Tehuantepec, Oaxaca; F. Sumichrast collector).


**Type.**—U.S.N.M. No. 30141; F. Sumichrast collector.

**Type locality.**—“West Tehuantepec,” Oaxaca, here restricted to the city of Tehuantepec.

**Range.**—Pacific slopes from Colima to Chiapas. Recorded only from the states of Colima (Manzanillo), Michoacán, Guerrero, Morelos, Oaxaca, and Chiapas (Tonalá).

**Sexlineatus** Group

**Species.**—Six, with a total of 16 forms as at present recognized.

**Range.**—Southern California, southeastern Wyoming, Wisconsin, and Maryland southward to the Gulf of Mexico, to Guatemala and into northwestern Baja California.

**Remarks.**—The arrangement presented herewith for this group is far from final. We have, however, made an attempt to correlate zoogeography with variations pointed out by others, and the tentative conclusions we trust will at least be a step forward. The present picture does make some geographic sense, as previous arrangements have not.

**Cnemidophorus sackii sackii** Wiegmann


Type.—Zool. Mus. Berlin No. 884; F. Deppe collector.
Type locality.—"Mexico," restricted to Cuernavaca, Morelos.
Range.—The Rio Balsas Valley. Recorded from numerous localities in Morelos, Guerrero, Puebla, and Michoacán.

Cnemidophorus Sackii Bocourt Boulenger


Cnemidophorus sexlineatus sackii, Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 4, 1874, pp. 276–277, pl. 20c, fig. 6.


Type.—Brit. Mus. Nat. Hist., three cotypes; No. 1857.10.28.81 here designated lectotype.

Type locality.—"Mexico" and "California," here restricted to Oaxaca, Oaxaca.

Range.—Uplands from central Oaxaca through central Chiapas into central Guatemala; range perhaps discontinuous. Recorded in Mexico only from Oaxaca: Oaxaca, Santo Domingo; and Chiapas: Piedra Parada.

Cnemidophorus Sackii Australis Gadow


Gadow (op. cit., p. 358) states that Peters's cotypes are immature and therefore not readily placed. The original description is of little assistance. We do not follow Gadow's allocation of the name, with a distinct southern race, for two reasons: (1) The name can simply be disposed of, if the types are unidentifiable, by placing it in synonymy; and (2) we believe it quite probable that the types actually came from southwestern Puebla in the Balsas Basin, where a number of Peters's species are known to be restricted. This latter area is occupied by C. s. sackii, of which Peters's name can reasonably be held a synonym.

Type locality.—Not specified, Lagunas or Cuicatlán, Oaxaca; here restricted to Cuicatlán, Oaxaca.

Range.—Foothills about the central Oaxacan highlands, excluding the Balsas Basin, in the upper headwaters of the Río Papaloapam and Río Tehuantepec, and near the Isthmus of Tehuantepec.

Cnemidophorus Sackii Communis Cope


Type.—Numerous cotypes, all lost.

Type locality.—Not definitely stated: Colima, Guadalajara, Córdoba, Cobán (Guatemala) or San Antonio (Texas); here restricted to Colima, Colima.

Range.—Most of the central plateau of Mexico, from Chihuahua and presumably Nuevo León southward to central Puebla; the Pacific coast from the Balsas basin northward to southern Sinaloa and perhaps Sonora. Recorded from ? Sonora, Chihuahua, Durango, Coahuila, Tamaulipas, San Luis Potosí, Zacatecas, Hidalgo, Querétaro, Guanajuato, Aguascalientes, Jalisco, Sinaloa, Nayarit (including Isabel Island), Colima, Michoacán, México, Distrito Federal, Puebla, and Veracruz.

Cnemidophorus Sackii Scalaris Cope


Cnemidophorus gularis scalaris verus Cope, op. cit., p. 45 (no types designated; Chihuahua, Chihuahua).

Cnemidophorus gularis gularis obsoletus Cope, loc. cit. (as above).

Type.—U.S.N.M. Nos. 8319, 14302, seven cotypes (No. 14302a, lectotype); John Potts collector.

Type locality.—Not specifically designated, either "Mexican plateau south of Chihuahua," or "City of Chihuahua;" here restricted to Chihuahua, Chihuahua.
Range.—Central and eastern Chihuahua, western Coahuila and northern Durango. Recorded from each state mentioned.

CNEMIDOPHORUS SACKII STICTOGRA/MMUS Burger


*CNemidophorus gularis octolineatus*, Smith (nec Baird), Handbook of lizards, 1946, pp. 409–412, figs. 120–127, pl. 114 (part).

*Type.—*U.S.N.M. (Hensley-Burger No. 768).

*Type locality.—*Yank Springs, 6 miles southeast of Ruby, Santa Cruz County, Ariz.

*Range.—*Central Arizona and Mexico southward through Sonora and probably northern Sinaloa, and the northern parts of Chihuahua and Coahuila. Recorded in Mexico from Sonora and Chihuahua.

CNEMIDOPHORUS SACKII MARIARUM Günther


*Type locality.—*Tres Marías Islands.

*Range.—*Restricted to the Tres Marías Islands, on each one of which it has been taken.

CNEMIDOPHORUS SACKII ANGUSTICEPS Cope


*Type.—*U.S.N.M. Nos. 24876–24878, three cotypes.

*Type locality.—*“Yucatán,” here restricted to Chichen Itzá.

*Range.—*The northern and western portions of the Yucatán Peninsula. Recorded from Yucatán: Chichen Itzá, Progreso, Tunkas; Quintana Roo: Cobá, Cozumel Island; Campeche: Champotón.

CNEMIDOPHORUS SACKII GULARIS Baird and Girard


*Cnemidophorus gularis sericeus* Cope, Trans. Amer. Philos. Soc., vol. 17, 1892, p. 48 (U.S.N.M. No. 15650; Wm. Taylor collector; San Diego, Tex.).

*Type.*—U.S.N.M. Nos. 3022, 2989, 14 cotypes (lectotype No. 3022a); Colonel Graham collector.

*Type locality.*—“Indianola and the Valley of the Rio Grande del Norte,” restricted to mouth of Devils River, Tex.

*Range.*—Oklahoma and most of Texas southward through Tamaulipas and eastern Nuevo León to northern Veracruz. Recorded in Mexico from Tamaulipas, San Luis Potosí, Veracruz, and Nuevo León.

**Cnemidophorus gularis semifasciatus** Cope


*Type.*—U.S.N.M. No. 9248; Lieutenant Couch collector.

*Type locality.*—Agua Nueva, Coahuila.

*Range.*—Big Bend area of Texas and adjacent Coahuila. Recorded in Mexico only from the state of Coahuila.

**Cnemidophorus inornatus** Baird


*Cnemidophorus gularis velox* Springer, Copeia, No. 169, 1928, p. 102 (Butler Univ. No. 848; Pueblo Bonito, San Juan County, N. Mex., by present restriction).

*Type.*—U.S.N.M. No. 3032; Lieutenant Couch collector.

*Type locality.*—Pesquería Grande (= García), Nuevo León.

*Range.*—Western Texas to extreme southeastern Arizona, southward into Chihuahua, Coahuila, and Nuevo León. Recorded in Mexico only from the states cited.
CNEMIDOPHORUS BURTI Taylor


Type.—EHT-HMS No. 13117; E. H. Taylor collector.
Type locality.—La Posa, 10 miles northwest of Guaymas, Sonora.
Range.—Southwestern Sonora. Known only from the vicinity of Guaymas.

CNEMIDOPHORUS SEXLINEATUS (Linnaeus)


Type.—Not known.
Type locality.—“Carolina,” here restricted to Charleston, S. C.
Range.—Eastern United States from Wisconsin, southeastern Wyoming, and Maryland southward to the Gulf as far west as, perhaps, the mouth of the Rio Grande. No definitely reliable records are known for Mexico, but we are informed by Bryce Brown that the species occurs near Brownsville, Tex., and may be expected across the river in Tamaulipas. Records available from Matamoros, Tamaulipas, may or may not belong here.

CNEMIDOPHORUS GADOVI Burger


Type.—U.S.N.M. No. 40042.
Type locality.—Hermosillo, Sonora.
Range.—Known only from the type locality.

CNEMIDOPHORUS LABIALIS Stejneger


Type.—U.S.N.M. No. 15596; L. Belding collector.
Type locality.—“Cerros Island,” Pacific coast, Baja California.
Range.—Northwestern Baja California between San Quintín and Punta Eugenia, Cedros Island.
Species.—One, represented by six forms as at present recognized.

Range.—Extreme southwest California south throughout the peninsula of Baja California and on adjacent islands.

**CNEMIDOPHORUS HYPERYTHRUS BELDINGI** (Steeneger)


*Type.*—U.S.N.M. No. 11980; L. Belding collector.

*Type locality.*—"Cerros" Island, Baja California.

*Range.*—Extreme southwestern California southward on Pacific slopes (not Gulf) to Cedros Island.

**CNEMIDOPHORUS HYPERYTHRUS CAERULEUS** (Dickerson)


*Type locality.*—Carmen Island, Baja California.

*Range.*—Known only from the type locality.

**CNEMIDOPHORUS HYPERYTHRUS DANHEIMAЕ** Burt


*Type.*—California Acad. Sci. No. 435; Walter E. Bryant collector.

*Type locality.*—San José Island, Gulf of California, Baja California.

*Range.*—Known only from the type locality.

**CNEMIDOPHORUS HYPERYTHRUS PICTUS** (Van Denburgh and Slevin)


*Type.*—California Acad. Sci. No. 49155; Joseph R. Slevin collector.

*Type locality.*—Monserrate Island, Baja California.

*Range.*—Known only from the type locality.
CNEMIDOPHORUS HYPERYTHRUS SCHMIDTI (Van Denburgh and Slevin)


*Type.*—California Acad. Sci. No. 50512; Joseph R. Slevin collector.

*Type locality.*—San Marcos Island, Gulf of California, Baja California.

*Range.*—The central fifth of the peninsula of Baja California from about lat. 28° south to 25° 30' N. (the Vizcaino Desert south to Comondú), and adjacent islands.

CNEMIDOPHORUS HYPERYTHRUS HYPERYTHRUS Cope


Verticaria franciscensis Van Denburgh and Slevin, loc. cit. (Calif. Acad. Sci. No. 50513; San Francisco Island, Baja California; J. R. Slevin collector).—Van Denburgh, op. cit., pp. 568-570.

*Type.*—U.S.N.M. No. 5299; John Xantus collector.

*Type locality.*—Cape San Lucas, Baja California.

*Range.*—The southern third of the peninsula of Baja California and its adjacent islands on both the Gulf and Pacific sides, south from about lat. 25°30'.

TESSELATUS GROUP

*Species.*—Six, one of which is represented by eleven subspecies, as now recognized.

*Range.*—Western United States from Idaho and Oregon south through Baja California and western Texas to southern Coahuitla, Chihuahua, and Sonora.

CNEMIDOPHORUS BACATUS Van Denburgh and Slevin


*Type.*—California Acad. Sci. No. 49152; Joseph R. Slevin collector.

*Type locality.*—San Pedro Nolasco Island, Sonora.

*Range.*—Known only from the type locality.

861316—50—13
CNEMIDOPHORUS CATALINENSIS Van Denburgh and Slevin


Type.—California Acad. Sci. No. 50507; Joseph R. Slevin collector. Type locality.—Santa Catalina Island, Baja California.

Range.—Known only from the type locality.

CNEMIDOPHORUS CERALBENSIS (Van Denburgh and Slevin)


Type.—California Acad. Sci. No. 50510; Joseph R. Slevin collector. Type locality.—Ceralbo Island, Baja California.

Range.—Known only from the type locality.

CNEMIDOPHORUS TESSELLATUS (Say)


Type.—Lost.

Type locality.—Arkansas River, near Castle Rock Creek, Colo. (= Beaver Creek, Fremont County).

Range.—Western Texas, eastern New Mexico, and probably southern Colorado, southward into adjacent northern Chihuahua and Coahuila. No reliable Mexican records are known to us; Yarrow (U. S. Nat. Mus. Bull. 24, 1883, p. 43) records it from “between Panos and St. Luis, Mexico,” and Cope (U. S. Nat. Mus. Bull. 32, 1887, p. 45) cites it from “between Mexico City and Chihuahua,” but neither record definitely refers to the species as now understood, and even if they did, the locality of collection is very uncertain.

CNEMIDOPHORUS MAXIMUS Cope


Type.—U.S.N.M. No. 5297; John Xantus collector.
Type locality.—Cape San Lucas, Baja California.
Range.—Southern Baja California, southward from La Paz Bay, or perhaps Magdalena Bay.

CNEMIDOPHORUS TIGRIS TIGRIS Baird and Girard

Type.—U.S.N.M. No. 4103; Capt. Howard Stansbury collector.
Type locality.—“Valley of the Great Salt Lake,” Utah, restricted to Salt Lake City, Utah.
Range.—Southern Idaho and southeastern Oregon south through western Utah and southeastern California to extreme northeastern Baja California, and extreme northwestern Sonora. Recorded in Mexico only from Baja California and Sonora (between Sonoyta and Puerto Peñasco).

CNEMIDOPHORUS TIGRIS GRACILIS Baird and Girard

Type.—U.S.N.M. No. 3034; J. L. LeConte collector.
Type locality.—“Desert of Colorado,” restricted to Yuma, Ariz.
Range.—Extreme eastern California in Colorado River Valley, southern Arizona south of the plateau, extreme southwestern New Mexico, northern Sonora. Recorded in Mexico only from the state of Sonora.

CNEMIDOPHORUS TIGRIS AETHIOPS Cope

Cnemidophorus estebanensis Dickerson, op. cit., p. 474 (U.S.N.M. No. 64446; C. H. Townsend collector; San Esteban Island, Sonora).
Cnemidophorus punctilineatus Dickerson, op. cit., p. 475 (U.S.N.M. No. 64447; C. H. Townsend collector; Tiburón Island, Sonora).

Type.—U.S.N.M. Nos. 64240-5, cotypes; Jenkins and Evermann collectors.

Type locality.—Hermosillo, Sonora.

Range.—Southern Sonora and adjacent islands (Tiburón, San Esteban).

Cnemidophorus tigris marmoratus Baird and Girard


Type.—U.S.N.M. No. 3024; two cotypes; J. H. Clark collector.

Type locality.—“Between San Antonio and El Paso,” restricted to El Paso, Tex.

Range.—Southwestern New Mexico to southwestern Coahuila. Recorded in Mexico only from Chihuahua, Durango (5 km. west of Torreón), and Coahuila.

Cnemidophorus tigris canus Van Denburgh and Slevin


Type.—California Acad. Sci. No. 49153; Joseph R. Slevin collector.

Type locality.—Sal Si Puedes Island, Gulf of California, Baja California.

Range.—The type locality, and North and South San Lorenzo Islands (on the latter, intergrades with C. t. martyris).

Cnemidophorus tigris celeripes Dickerson


Type locality.—San José Island, Baja California.

Range.—Known only from the type locality.
CHECKLIST OF REPTILES OF MEXICO

**Cnemidophorus tigris Martyris Stejneger**


_Type._—U.S.N.M. No. 15620; E. Palmer collector.

_Type locality._—San Pedro Mártir Island, Gulf of California, Sonora.

_Range._—The type locality (intergrades with *C. t. canus* on South San Lorenzo Island).

**Cnemidophorus tigris Rubidus Cope**


_Type._—U.S.N.M. Nos. 15149–15155, cotypes; U. S. Fish Commission collector.

_Type locality._—Santa Margarita Island, Baja California.

_Range._—The southern third of Baja California, south of San Marcos Island, including adjacent islands in the Gulf and off the western edge of the peninsula (notably Carmen, Santa Margarita, and Magdalena Islands, Baja California).

**Cnemidophorus tigris Multiscutatus Cope**


_Type._—U.S.N.M. Nos. 15160–3; U. S. Fish Commission donor.

_Type locality._—Cedros Island, Baja California.
Range.—Southern California west of the Sierra Nevada, south to central Baja California, exclusive of the extreme northeastern corner; islands in the Gulf of California near the center of the Peninsula (Isla Partida, Ángel de la Guarda).

Genus GYMNOPHTHALMUS Merrem


Genotype.—_Lacerta quadrilineata_ Linnaeus [= _Gymnophthalmus lineata_ (Linnaeus)].

Range.—Central Argentina north through South and Central America. In Mexico, Oaxaca and possibly Chiapas.

Species.—Seven. Only one occurs in Mexico.

GYMNOPHTHALMUS SUMICHRASTI (Cope)


Type.—U.S.N.M. No. 30245–46; François E. Sumichrast collector.

Type locality.—“Western part of the State of Tehuantepec” (“Near Ventose Bay” fide Dunn in Stuart, loc. cit.).

Range.—Southern Oaxaca near the Isthmus of Tehuantepec eastward on Pacific slopes possibly to Honduras. Recorded in Mexico only from Oaxaca: Tehuantepec, Bahía Ventosa, Mount Guengola, Cacoprieto. A specimen, presumably of this species, was seen near San Ricardo, Chiapas, by the junior author.

Family HELODERMIDAE Gray


Genera.—A single genus, _Heloderma_ Wiegmann, is known.

Range.—Southern Utah to the Isthmus of Tehuantepec, on Pacific slopes.

Genus HELODERMA Wiegmann


Genotype.—*Trachyderma horridum* Wiegmann.

*Range.*—Southern Utah to the Isthmus of Tehuantepec.

*Species.*—Two.

KEY TO SPECIES OF *HELODERMA*

1. Tail equal to or longer than body; head and neck about half of body length; predominating color black; pterygopalatine teeth present... *horridum* (p. 193)

Tail about two-thirds of body length; head and neck about one-third of body length; colors pink, yellowish, and black; none or rarely 1 or 2 pterygopalatine teeth... *suspectum* (p. 193)

**HELODERMA SUSPECTUM** Cope


*Type.*—U.S.N.M. No. 2971, three cotypes; Major Emory collector.

*Type locality.*—Sierra de la Unión, "Sonora" (=Arizona).

*Range.*—Sonora as far south as Guaymas; in the United States, Arizona, southern Nevada, and southwestern Utah. Recorded from Sonora: Guadalupe Cañon, San Pedro Bay, La Posa about 10 miles northwest of Guaymas, Monument 88, San Bernardino, Niggerhead Mountain, 10 miles south of Noria, etc.

**HELODERMA HORRIDUM** (Wiegmann)


*Heloderma Hernandesii* Wiegmann, Herpetologia Mexicana, pt. 1, 1834, p. 25 (for a theoretical form).

*Type.*—Zool. Mus. Berlin; Ferdinand Deppe collector.

*Type locality.*—"Vivit in ferventibus terrae Mexicanae regionibus," here restricted to Huajintlan, Guerrero.

*Range.*—Coastal areas from Oaxaca to Sonora, extending up the Río Balsas Valley to Morelos. Recorded from Sonora: Álamos, Guirocoba; *Sinaloa*: Presidio near Mazatlán, San Blas; *Jalisco*: Autlán, Barranca de San Cristóbal; *Morelos*: Cañon del Lobo near Cuernavaca; *Oaxaca*: Tapanatepec, Salina Cruz, Jimiltepec, Juchitán, Tehuantepec, Quiengola and Mixtequilla Mountains; *Michoacán*: Apatzingán, Parécuaro, Oropeo, Etúcuaro, Puruarán; *Colima*: Paso del Río, Colima; *Guerrero*: Huajintlán; *Chiapas*: Rancho San Bartolo.

---

A sight record for *Zongolica*, Veracruz (Martín del Campo, Anal. Inst. Biología, vol. 6, 1935, p. 297) is unacceptable and may be referable to *Xenoserus*. Likewise unacceptable are records for Guanajuato and Yucatán. We are unable to find authoritative records for Nayarit, although Martín del Campo (*loc. cit.*) cites the state.
Family ANGUIDAE Cope


Genera.—Eleven, of which nine are restricted to the Americas, one (Anguis) is restricted to the Old World, and one (Ophisaurus) occurs in both hemispheres. Five occur in Mexico.97

Range.—Southern Canada to Argentina, West Indies, Europe, northern Africa, southern Asia.

KEY TO MEXICAN GENERA OF ANGUIDAE

1. No lateral fold.----------------------------------------Celestus (p. 194)  
A lateral fold present.-------------------2

2. Lateral fold weakly developed, containing few or no granules; sides of neck with coarse granules; head much widened and depressed; pterygoid teeth absent; minimum scales in a transverse row across neck 4–6; arboreal species.-----------------------------------------------Abronia (p. 196)

Lateral fold moderately to well developed, containing a moderate or large granular area; sides of neck with fine granules; head thicker, not depressed; pterygoid teeth various; minimum scales in a transverse row across neck no less, often more numerous; terrestrial species.--------------------------3

3. No anterior internasals, but instead a pair of enlarged supranasals, which much resemble internasals; no unpaired median internasals; nasal in contact with rostral; pterygoid teeth well developed.---------------------------Elgaria (p. 205)

Anterior internasals, posterior internasals and supranasals all present; nasal seldom in contact with rostral; pterygoid teeth various.---------------4

4. One or more median, unpaired postrostrals; frontals in contact with maxilla; pterygoid teeth well developed; tail extremely elongate.------------------------Gerrhonotus (p. 202)

No postrostrals; frontals separated from maxilla; pterygoid teeth absent or vestigial; tail normal.---------------------------Barisia (p. 198)

Genus CELESTUS Gray


Genotype.—Celestus striatus Gray.

Range.—West Indies, Central America, and southern Mexico.

Species.—About 18, two (possibly three) of which occur in Mexico.

97 Ophisaurus ventralis, cited from Jalapa, Veracruz, as early as 1854 (Yarrow, U. S. Nat. Mus. Bull. 24, 1884, p. 46), and retained in accounts of the species even up to the present time, has never been authoritatively recorded from Mexico and probably does not occur there. It almost certainly does not occur at Jalapa. It is possible that the record arose from a simple misunderstanding of Peale and Green's Scincus ventralis (=Gerrhonotus lioccephalus) which occurs in the same general area. Regardless of the nature of the error, we emphatically believe the record is erroneous.

99 Supranasals absent in Gerrhonotus lioccephalus australis, which has a postrostral.

Dunn (Notulae Nat., No. 4, 1939, p. 3) expresses the opinion that forms here referred to Celestus, characterized by absence of sheaths on the claws, are congeneric with Diploglossus (Wiegmann, Herpetologia Mexicana, 1834, p. 36; type "Sc. fasciatus Gray"—Diploglossus fasciatus), which possesses sheaths.
KEY TO MEXICAN SPECIES OF CELESTUS

1. Median prefrontal in contact with 2 supraoculars, wider than long; no lateral prefrontals (fused with median loreal); 2 dorsolateral light stripes in adults separated by 4 and 2 half scale rows. **atiitianensis** (p. 195)

Median prefrontal in contact only with anterior supraocular, as long as wide; small lateral prefrontals usually present; adults not with 2 dorsolateral light stripes. **enneagrammus** (p. 195)

2. Snout normal, not especially elongate or flattened; lamellae under fourth toe, 15–18; scales in 33–35 rows; first labial reaching to middle of naris; distance between anterior and posterior loreals less than the length of either; no vertical light bars on sides in young or adults. **rozellae** (p. 195)

Snout flattened, elongate; lamellae under fourth toe, 23–26; scales in 31–33 rows; first labial reaching to anterior border of naris; distance between anterior and posterior loreals as great or greater than length of either; vertical light bars present on sides in young and adults.

**CELESTUS ROZELLAЕ** Smith

*Diploglossus steindachneri*, Günther (nec Cope), Biologia Centrali-Americana, Reptilia and Batrachia, 1885, p. 34, pl. 22, fig. A.


Type.—U.S.N.M. No. 113526; Rozella Smith collector.

Type locality.—Palenque, Chiapas.

Range.—Atlantic slopes, presumably from the Isthmus of Tehuantepec to British Honduras. Recorded in Mexico only from the type locality.

**CELESTUS ATIITLANENSIS** Smith, new species

*Diploglossus* (Celestus) steindachneri, Bocourt (nec Cope), Mission scientifique au Mexique . . . , Études sur les reptiles, livr. 6, 1879, pp. 383–384, pl. 22, fig. 3.

Type.—Mus. Hist. Nat. Paris, specimen figured as above.

Type locality.—Atitlán, Guatemala.

Range.—Known only from the type specimen from Atitlán, Guatemala, but probably occurring along Pacific slopes from Chiapas to Nicaragua.

**CELESTUS ENNEAGRAMMUS** (Cope)


---

1 Diagnosis.—Allied to *C. enneagrammum* and *C. rozellae*, but with median prefrontal in contact with two (instead of one) supraoculars, no lateral prefrontals (presumably fused to median loreals), a very broad median prefrontal (broader than long), and a pattern in the single known adult (112 mm. snout to vent) consisting of two dorsolateral light stripes separating three broad dark stripes from each other. Similar to *C. bistattatus* Boulenger but with three loreals instead of two, seven supralabials to a point below middle of eye instead of six.—H. M. S.
**Genus ABRONIA Gray**


**Genotype.** _Gerrhonotus deppii_ Wiegmann [= _Abronia deppii_ (Wiegmann).]

**Range.** Hidalgo, south to Guerrero and Chiapas. In Central America, Guatemala.

**Species.** Nine species, 10 forms; 6 species and 7 forms in Mexico.

**KEY TO MEXICAN FORMS OF ABRONIA**

1. Postmental unpaired. ................................. 2
   Postmental paired. ................................. 3

2. Scale rows 30-33; parietal separated from supraoculars or very narrowly in contact; dorsal markings usually indistinct, those on neck, if present, combining to form a large single mark. _ochoterenai_ (p. 197)
   Scale rows 39; parietals broadly in contact with the supraocular; markings on back distinct, those on neck discrete, not forming a large single mark. _matudai_ (p. 196)

3. Suboculars missing or reduced to minute scales; one anterior temporal bordering orbit. _deppii_ (p. 197)
   Suboculars present, well developed; 2 anterior temporals bordering orbit. 4

4. Area of granular scales on sides of neck very narrow; no granular zone in lateral fold. _oaxacae_ (p. 197)
   Area of granular scales covering entire sides of neck; a granular zone in lateral fold. 5

5. Prominent light markings along the sides; dorsal osteoderms absent over posterior part of body; dorsal scales about 31-34 (average about 33); minimum number of scales in a single nuchal transverse row usually 6.
   _taeniata taeniata_ (p. 197)
   No light markings on sides of body; dorsal osteoderms present over the whole body. 6

6. Dorsal scale rows 25 to 29; minimum number of scales in a transverse nuchal row, 4 or 5 (75 percent); chin and lower labials white with occasionally a few scattered dark blotches. _taeniata graminea_ (p. 198)
   Dorsal scale rows 30-31; minimum number of scales in a transverse nuchal row, 6; infralabials darker than chin and as dark as granular area of neck, with indistinct lighter bands. _fuscolabialis_ (p. 198)

**ABRONIA MATUDAI** (Hartweg and Tihen)


Type.—Univ. Michigan Mus. Zool. No. 88331; Eizi Matuda collector.

Type locality.—Volcán de Tacaná, Chiapas, 2,000 meters.

Range.—Known only from the type locality.

**ABRONIA OCHOTERENAI** (Martín del Campo)


Type.—Instituto de Biología, México, cotypes, male and female; Mario del Toro collector.

Type locality.—Santa Rosa, Comitán, Chiapas.

Range.—Known only from the type locality.

**ABRONIA DEPPII** (Wiegmann)


Type.—Zool. Mus. Berlin; F. Deppe collector.

Type locality.—Mexico. Here restricted to Omilteme, Guerrero.

Range.—Sierra Madre del Sur. Known only from the vicinity of Omilteme, Guerrero.

**ABRONIA OAXACAE** ( Günther)

Gerrhonotus oaxacae Günther, Biologia Centrali-Americana, Reptilia and Batrachia, 1885, p. 36, pl. 24, figs. A, A', A''.


Type.—Brit. Mus. Nat. Hist., 2 adult males, 1 young; A. Boucard collector.

Type locality.—“Oaxaca,” Mexico.

Range.—Mountains in Oaxaca. Reported definitely only from “Luvina, Tehuantepec, Oaxaca.”

**ABRONIA TAENIATA** (Wiegmann)

Gerrhonotus taeniatus Wiegmann, Isis von Oken, vol. 21, 1828, p. 379; Herpetologia Mexicana, 1834, p. 32, pl. 9, fig. 1, 2.


Gerrhonotus (Abronia) taeniatus, Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 5, 1878, pp. 327–329, pl. 21A, figs. 4, 4a, and 5.

---

1 This arrangement has been suggested verbally by Dr. Tihen.
Genus Barisia Gray


Genotype.—Gerrhonotus imbricatus Wiegmann [=Barisia imbricata imbricata (Wiegmann)].

Range.—Chihuahua and Coahuila south to Chiapas; Central America to Panamá.

Species.—Nine, with 16 forms; 8 species, 12 forms, occur in Mexico.

KEY TO MEXICAN SPECIES OF BARISIA

1. Postmental unpaired; prefrontal bones in contact........................................... 2
   Postmental paired; prefrontal bones separated by frontonasal.......................... 4

2. Eighteen to 20 longitudinal dorsal scale rows; frontonasal normally present. 3
   Fourteen longitudinal dorsal scale rows; frontonasal normally absent. viridiflava (p. 200)

3. Upper postnasal separated from lower by a loreonasal contact. moreleti rafaeli (p. 199)

1 It is known that Deppe collected in this region.
Upper and lower postnasals in contact .......... *moreleti temporalis* (p. 199)
4. Superciliary series complete; pterygoid teeth absent .................. 5
   Superciliary series incomplete; pterygoid teeth vestigial .................. 8
5. Supranasals unexpanded; an anterior canthal present .................. 6
   Supranasals expanded; no anterior canthal .................................. 7
6. Scales of neck keeled; anterior loreal in contact with anterior canthal. gadovii gadovii (p. 200)
   Scales of neck smooth; anterior loreal separated from anterior canthal by a contact of posterior loreal with upper postnasal .......................... *gadovii levigata* (p. 200)
7. Frontonasal and postrostral present ...................................... *modesta* (p. 201)
   No frontonasal or postrostral present ......................................... *antauges* (p. 200)
8. One loreal; more than 33 dorsal scale rows ................................ 9
   Two loreals; fewer than 33 dorsal scale rows ................................ *rudicollis* (p. 202)
9. More than 1 superciliary (usually 3); transverse dorsal scale rows usually fewer than 47 ............................................................... 10
   A single (middle) superciliary element; transverse dorsal scale rows usually more than 47 ............................................................... *levicollis* (p. 202)
10. Fewer than 16 longitudinal dorsal scale rows; contact of anterior superciliary with loreal usually smaller than that of first medial supraocular with loreal ............................................................. 11
   Sixteen longitudinal dorsal scale rows; contact of anterior superciliary with loreal as great or greater than that of first medial supraocular with loreal. *imbricata imbricata* (p. 201)
11. Lowest primary temporal in contact with penultimate as well as with antepenultimate supralabial; 39-45 transverse dorsal scale rows. *imbricata ciliaris* (p. 202)
   Lowest primary temporal not in contact with penultimate supralabial; 35-39 transverse dorsal scale rows .................................................. *imbricata planifrons* (p. 201)

**BARISIA MORELETI RAFAELI** (Hartweg and Tihen)


*Type locality.*—16 km. south of Siltepec, Chiapas, 2,300 meters elevation.

*Range.*—High mountains of southern Chiapas. Reported from Cerro Paxtal, 1,500 meters; Chiquihuite, Volcán de Tacaná, 2,500 meters; Cerro Malé.

**BARISIA MORELETI TEMPORALIS** (Hartweg and Tihen)


*Type locality.*—Eleven km. southeast of Ciudad de las Casas, Chiapas, 2,300 meters elevation.

*Range.*—Known only from the type locality.
BARISIA GADOVII GADOVII (Boulenger)


_Type locality._—Omilteme, Guerrero.
_Range._—Sierra Madre del Sur, in Guerrero. Known only from mountains west of Chilpancingo and the type locality.

BARISIA GADOVII LEVIGATA Tihen


_Type._—U.S.N.M. No. 47212; E. W. Nelson and E. A. Goldman collectors.
_Type locality._—“Valley of Oaxaca,” Oaxaca.
_Range._—Mountains of central Oaxaca. Reported from “mountains west of Oaxaca city.”

BARISIA VIRIDIFLAVA (Bocourt)


_Gerrhonotus antauges_ (part), Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 5, 1878, pp. 346–349, pl. 21B, figs. 7, 7a.


_Type._—Mus. Hist. Nat. Paris; “un seul exemplaire qui a été cédé par M. Boucard, comme provenant des collections de M. F. Sumichrast.”
_Type locality._—Mexico. Restricted by Tihen to the highlands of central Oaxaca, near the city of Oaxaca.
_Range._—Mountains north of Oaxaca (city). Reported from “summit of Cerro San Felipe north of Oaxaca” (city).

BARISIA ANTAUGES Cope


_Type._—U. S. N. M. No. 30221; François E. Sumichrast collector.
_Type locality._—Orizaba, Veracruz [=Volcán Citlaltépetl].
_Range._—Volcán Citlaltépetl (Mount Orizaba), Veracruz; known only from the type locality.
BARISIA MODESTA (Cope)


Type.—U. S. Nat. Mus., three specimens; collector unknown.
Type locality.—“Probably Guatemala,” here restricted to Mount Orizaba.

Range.—Region of Mount Orizaba (Volcán Citlaltépetl), Veracruz (not known from Guatemala).

BARISIA IMBRICATA IMBRICATA (Wiegmann)

Gerrhonotus (Barisia) imbricatus, Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 5, 1878, pl. 21B, figs. 1, 1a, 2, 2a, and livr. 6, 1879, pp. 363–365.
Gerrhonotus lichenigerus Wagler, Descriptiones et icones amphibia-rum, 1833, pl. 34, fig. 2 (type locality here restricted to Mexico, D. F.).—WIEGMANN, Herpetologia Mexicana, 1834, pl. 10, figs. 2–5.
Gerrhonotus adspersus Wiegmann, Herpetologia Mexicana, 1834, pl. 10 (type locality, Mexico by inference, here restricted to San Martín, México; Zool. Mus. Berl.; F. Deppe collector).

Type.—Zool. Mus. Berlin, two cotypes; F. Deppe collector.
Type locality.—Mexico, here restricted to México, D. F.
Range.—The periphery and southern part of the central Mexican Plateau, from Veracruz to Jalisco. Reported from numerous localities in the states of Jalisco, Michoacán, México, Morelos, Puebla, Oaxaca, Veracruz, Hidalgo, and Guanajuato, and from Distrito Federal.

BARISIA IMBRICATA PLANIFRONS (Bocourt)

Gerrhonotus (Barisia) planifrons Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 5, 1878, pl. 21C, figs. 1, 1a; livr. 6, 1879, pp. 361–363 (pl. 21C was published with the name, in livr. 5, 1878; this constitutes the “type description”).

Type.—Mus. Hist. Nat. Paris; Adolphe Boucard collector.
Type locality.—“Oaxaca.”
Range.—Oaxaca, mountainous areas; not known from a definite locality.
BARISIA IMBRICATA CILIARIS (Smith)


Type.—U.S.N.M. No. 47496; E. W. Nelson and E. A. Goldman collectors.

Type locality.—Sierra Guadelupe, Coahuila.

Range.—The northern portion of the plateau from extreme southern Coahuila to Guanajuato, east to San Luis Potosí and west to Sinaloa. Reported from Coahuila: Sierra Guadelupe; Zacatecas: Sierra Madre; Durango: Coyotes, Inde; San Luis Potosí: mountains near San Luis Potosí (city), near Xilitla, near Jesús María; Guanajuato: San Felipe, Sierra de Santa Rosa; Hidalgo: Durango; Nuevo León: Pablillo, Galeana; Sinaloa: Escuinapa.

BARISIA LEVICOLLIS Stejneger


Type.—U.S.N.M. No. 9362; "evidently obtained by one of the surveying parties of the United States and Mexican Boundary Survey," fide Stejneger.

Type locality.—“Mexican boundary.” Probably Chihuahua, fide Thiен, loc. cit.

Range.—Chihuahua and probably adjacent areas in Sonora. Reported from Chihuahua: north Chihuahua, Colonia García, Meadow Valley, 65 miles east of Batopilas, Samachique.

BARISIA RUDICOLLIS (Wiegmann)


Gerrhonotus (Barissia) rudicollis, Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 5, 1878, pl. 21B, figs. 3, 3a; and livr. 6, 1879, pp. 367-369.


Type.—Zool. Mus. Berlin; F. Deppe collector.

Type locality.—Mexico. Here restricted to Hacienda de la Gavía, México (state).

Range.—Western México and perhaps adjoining parts of Michoacán. Reported from México: Hacienda de la Gavía.
Genus GERRHONOTUS Wiegmann


Genotype.—Gerrhonotus tessellatus Wiegmann [=Gerrhonotus liocephalus Wiegmann].

Range.—Central Texas southward through the plateaux of Mexico to southwesteren Chiapas.

Species.—One. Five forms recognized, all occurring in Mexico.

KEY TO SPECIES OF GERRHONOTUS

1. Three loreals (lorecanths) on each side. 2
   Four or more lorecanths on each side. 3

2. Supranasals absent; azygous prefrontal much longer than broad; frontal widely separated from interparietal; all except lowermost anterior temporal touch fifth medial supraocular. liocephalus australinus (p. 204)
   Supranasals present; azygous prefrontal as broad as or broader than long; frontal in contact with or very narrowly separated from interparietal; only 2 uppermost anterior temporals in contact with fifth supraocular. liocephalus liocephalus (p. 203)

3. Dorsal bands obsolete; venter nearly without marking; 52-60 dorsal scales (average 55.6); tail to body ratio, 2.5-2.6; caudals 157-163; second primary temporal touches the fifth medial supraocular. liocephalus liocephalus (p. 204)
   Dorsal bands distinct; venter mottled and flecked. liocephalus loweryi (p. 204)

4. Dorsal scales 45-54 (average 49); tail-body ratio, 1.75-2.1; caudals, 116-137. liocephalus infernalis (p. 204)
   Dorsal scales 49-52 (average 51); tail-body ratio 2.3; caudals (approx.) 140. liocephalus ophiusus (p. 204)

GERRHONOTUS LIOCEPHALUS LIOCEPHALUS Wiegmann


Gerrhonotus liocephalus, Bocourt, Mission scientifique au Mexique ..., Etudes sur les reptiles, livr. 5, 1878, pp. 342-346, pl. 21A, figs. 1, 2, 2a.


Gerrhonotus tessellatus Wiegmann, Herpetologia Mexicana, pt. 1, 1834, pp. 32-33, pl. 10, fig. 3 (substitute name for preceding).


Type.—Zool. Mus. Berlin; F. Deppe collector.

Type locality.—Mexico (Oaxaca, fide Bocourt loc. cit., p. 344, here restricted to Tlapancingo).

Range.—Central plateau region, from Guanajuato to Guerrero and Oaxaca. Reported from Distrito Federal: Magdalena, Mixiuheca; Guerrero: Omitlteme; México: Temascaltepec; Oaxaca: Llano Ocotal, 861316—50—14
La Concepción, Tlapacingo, Tres Cruces; Puebla: Cacaloapam; Morelos: Cuernavaca; Guanajuato: Silao.

**GERRHONOTUS LIOCEPHALUS LOWERYI** Tihen


*Type.*—La. State Univ. Mus. No. 480; Marcella Newman collector.

*Type locality.*—"Xilitla region" (neighborhood of Xilitla, San Luis Potosí).

*Range.*—Known only from the type locality and possibly also from Ciudad del Maíz, San Luis Potosí.

**GERRHONOTUS LIOCEPHALUS OPHIURUS** Cope


*Type.*—U.S.N.M. No. 30206; François E. Sumichrast collector.

*Type locality.*—Orizaba, Veracruz, Mexico.

*Range.*—Eastern foothills of the plateau. Known only from Veracruz: Orizaba, Jicaltepec, Córdoba, Cerro Gordo; Puebla: Hueytamalco (Teziutlán).

**GERRHONOTUS LIOCEPHALUS INFERNALIS** Baird


*Type.*—? U.S.N.M. No. 3090; C. B. R. Kennerly collector.

*Type locality.*—Devils River, Tex.

*Range.*—San Luis Potosí, Coahuila and probably adjoining states of Chihuahua and Nuevo León, northward to central Texas across the Rio Grande valley from Rio Grande City to mouth of Devil’s River. Reported from Coahuila: Carmen Mountains, Cerro Encarnación, Sierra Guadalupe south of La Cuchilla, Monclova; San Luis Potosí: Alvarcez.

**GERRHONOTUS LIOCEPHALUS AUSTRINUS** Hartweg and Tihen


*Type.*—Univ. Michigan Mus. Zool. No. 94921; Eizi Matuda collector.

*Type locality.*—3,200 meters elevation on Cerro Malé, Porvenir, Chiapas.

*Range.*—Known only from the type locality.
Genus Elgaria Gray


Genotype.—Cordylus (Gerrhonotus) multicarinatus Blainville = Elgaria multicarinata multicarinata.

Range.—Chihuahua and Baja California, northward through California to British Columbia, Utah, and Montana.

Species.—Six species and 13 forms. Six forms occur in Mexico.

Key to Mexican Species of Elgaria

1. Alternate black and white marks on labials (white spots somewhat ocellate).                 kingii group 4
2. Temporal scales all or partly keeled.                                                                      multicarinata group 2
3. Temporal scales smooth; granular area of lateral fold white with gray or black reticulations; scales on arm smooth; 2 rows of scales keeled on thigh; lateral body scales smooth.                        cedrosensis (p. 206)
4. Granular area of lateral fold white crossed by black bands usually continuous with dark body bands; eight dorsal scale rows keeled moderately, lateral rows faintly keeled; 4 internasals of about same size; arm scales smooth; thigh scales with 2 rows keeled.                        paucicarinatus (p. 206)
5. Dorsal longitudinal scale rows usually 16; scales from occiput to posterior part of thigh in 51 to 56 transverse rows (average, 52.9); transverse row of four anterior temporals, uppermost always in contact with uppermost secondary temporal; granular area in fold usually black. kingii kingii (p. 205)
6. Dorsal longitudinal scale rows usually 14; scales from occiput to posterior part of thigh 55 to 60 (average 57.3); four anterior temporals, uppermost separated from uppermost secondary temporal (60 percent); granular area in fold usually gray.                           nobilis (p. 206)


Type.—Brit. Mus. Nat. Hist., adult (in bad state); from the T. Bell collection, collector unknown.

Type locality.—Mexico [so stated by Boulenger, Catalogue of Lizards . . . , vol. 2, 1885, p. 275]. Tihen suggests "presumably Chihuahua." Here restricted to Mojarachic.

Range.—Eastern Sonora and western Chihuahua, probably southward into Durango and Sinaloa. Reported from Chihuahua: Madera, Mojarachic, Chiricahui, Colonia García (possibly intergrades with with nobilis); Sonora: (no specific locality).


Type locality.—Fort Webster Copper Mines of the Gila [Santa Rita del Cobre], N. Mex.

Range.—Southwestern New Mexico and southeastern Arizona. While no records are available from Mexico, specimens have been taken within a few hundred yards of the border, and the form must occur in Mexico (Sonora).


Type locality.—Fort Webster Copper Mines of the Gila [Santa Rita del Cobre], N. Mex.

Range.—Southwestern New Mexico and southeastern Arizona. While no records are available from Mexico, specimens have been taken within a few hundred yards of the border, and the form must occur in Mexico (Sonora).


Type locality.—Fort Webster Copper Mines of the Gila [Santa Rita del Cobre], N. Mex.

Range.—Southwestern New Mexico and southeastern Arizona. While no records are available from Mexico, specimens have been taken within a few hundred yards of the border, and the form must occur in Mexico (Sonora).


Type locality.—Fort Webster Copper Mines of the Gila [Santa Rita del Cobre], N. Mex.

Range.—Southwestern New Mexico and southeastern Arizona. While no records are available from Mexico, specimens have been taken within a few hundred yards of the border, and the form must occur in Mexico (Sonora).
CHECKLIST OF REPTILES OF MEXICO

ELGARIA MULTICARINATA NANA (Fitch)

Gerrhonotus scincicauda nanus Fitch, Copeia, 1934, p. 7.


Type locality.—South Island, Los Coronados Islands, Baja California.

Range.—Los Coronados Islands, Baja California.

ELGARIA MULTICARINATA WEBBII (Baird)


Type.—USNM. No. 3078; T. H. Webb collector.

Type locality.—“San Diego to El Paso, “probably from near vicinity of San Diego, San Diego Co. California [?]” (Fitch, loc. cit.). Here restricted to San Diego, Calif.

Range.—Southern California and extreme northern Baja California. Reported in Baja California from San Pedro Mártil Mountains, Alcatraz.

Family XENOSAURIDAE Cope


Genera.—A single genus, Xenosaurus Peters, is known.

Range.—San Luis Potosí to Guatemala.

Genus XENOSAURUS Peters


Genotype.—Xenosaurus fasciatus Peters = Xenosaurus grandis (Gray).

Range.—San Luis Potosí to Guatemala.

Species.—Three.

KEY TO SPECIES OF XENOSAURUS

1. Arm with widely spaced tubercles on its dorsal surface; a row of supraoculars each 1½ to 2 times as long as wide; cream, black spotted on venter... 2

2. Arm regularly covered with tubercles on dorsal surface; supraoculars not forming a series of more or less regular enlarged scales; venter uniformly gray or gray-white without spots or flecks......... newmanorum (p. 208)
2. Scales bordering posterior gular fold small, rather widely separated from each other, considerably smaller than median chest scales..._grandis_ (p. 208)
Scales bordering posterior gular fold larger, in contact with each other or narrowly separated, about as large as median chest scales..._rackhami_ (p. 208)

**XENOSAURUS NEWMANORUM** Taylor


*Type.*—Louisiana State Univ. No. 85; Marcella Newman collector.

*Type locality.*—“Xilitla Region” (near Xilitla, San Luis Potosí).

*Range.*—Known only from the type locality.

**XENOSAURUS GRANDIS** (Gray)


*Type locality.*—Córdoba, Veracruz (probably from nearby mountains).

*Range.*—Foothills of central Veracruz southward to the Isthmus. Reported from *Veracruz*: Huanusco [=Huatusco], Córdoba, Orizaba; *Oaxaca*: “Oaxaca,” Tehuantepec.4

**XENOSAURUS RACKHAMI** **Stuart**


*Type locality.*—Finca Volcán, 49 km. east of Cobán, Alta Verapaz, Guatemala, 4,000 feet.

*Range.*—The highlands of central eastern Chiapas and adjacent Guatemala. Recorded in Mexico only from *Chiapas*: Santa Rosa (near Comitán).

Family **ANNIELLIDAE** **Boulenger**


*Range.*—Southwestern California and northwestern Baja California.

---

Genus ANNIELLA Gray


Genotype.—Anniella pulchra Gray.

Range.—Southern California and northern Baja California.

Species.—Two, one with two subspecies.

KEY TO MEXICAN SPECIES OF ANNIELLA

1. Rostral sharply pointed in profile; fourth supralabial largest.
   
   Rostral rounded in profile; second supralabial largest.

   geronimensis (p. 209)

   pulchra pulchra (p. 209)

ANNIELLA GERONIMENSIS Shaw


Type.—L. M. Klauber No. 7543.

Type locality.—Isla San Gerónimo, Baja California.

Range.—Known only from the type locality.

ANNIELLA PULCHRA PULCHRA Gray


Type locality.—California, here restricted to San Diego.

Range.—Pacific slopes of central and southern California, and extreme northern Baja California, west of the desert and the Sierra San Pedro Mártir. Recorded in Baja California: San Salado Cañon, San José, San Quintín, Los Coronados Islands.

Subclass ARCHOSAURIA

Archosauria Romer, Vertebrate paleontology, 1945, p. 597.

Orders.—A single living order, the Loricata, exists.

Order LORICATA

Loricata Merrem, Tentamen systematis amphibiorum, 1820, p. 34.

Families.—Three families exist at the present time, two of which occur in Mexico.
KEY TO LIVING FAMILIES OF LORICATA

1. Dentary teeth 25–26, none received by sockets in upper jaw; maxillary bones broadly in contact with each other on dorsal surface of skull; mandibular symphysis extraordinarily long, only 2 or 3 teeth at rear of mandible not opposite the symphysis. Gavialidae

Dentary teeth 20 or fewer, all or nearly all received by sockets in upper jaw; maxillary bones never in contact with each other on dorsal surface of skull; mandibular symphysis shorter, 4 or more teeth at rear of mandible not opposite the symphysis. 2

2. Fourth mandibular tooth fitting into a pit in upper jaw, and 17 to 22 dentary teeth. Alligatoridae (p. 211)

Fourth mandibular tooth fitting into a notch in upper jaw or, if into a pit, dentary teeth 16 or fewer. Crocodylidae (p. 210)

Family CROCODYLIDAE Gray


Genera.—Four genera and 17 species and subspecies are recognized at the present time; one genus occurs in the Americas.

Range.—World wide in tropical lands.

Genus CROCODYLUS Laurenti


Genotype.—Crocodylus niloticus Laurentii.

Species.—Twelve species, two with two subspecies, are recognized; only four species, one with two subspecies, occur in the New World, and two in Mexico.

Range.—Northern Australia, New Guinea, East Indies, Siam, Ceylon, tropical Africa and India, Madagascar; southern Florida, Greater Antilles, northern Mexico south to Ecuador and Venezuela.

KEY TO MEXICAN SPECIES OF CROCODYLUS

1. Premaxillo-maxillary suture transverse. moreletii (p. 211)

Premaxillo-maxillary suture extending posteriorly in an arc to level of posterior border of seventh tooth. acutus (p. 210)

CROCODYLUS ACUTUS ACUTUS Cuvier


* Not in Mexico; restricted to India.

* Allocation of the Mexican and, for that matter, Central American specimens is at present very uncertain. South American (Colombia, Ecuador) specimens are referable to the race C. a. lewpanus, and Santo Domingo specimens to C. a. acutus, but no study has been made of material from elsewhere.
† *Crocodilus americanus* Laurenti, Specimen medicum exhibens synopsin reptilium, 1768, p. 54 (type, pl. 106 in Seba’s “Locupletissimi Rerum Naturalium Thesauri Occurata Descriptio,” vol. 1, 1734; type locality, America, here restricted to Veracruz, Veracruz; unidentifiable).—Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 1, 1870, pp. 30–31, pl. 8, fig. 1.  


*Crocodilus pacificus* Duméril and Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 1, 1870, pp. 31–33, pl. 9, fig. 5 (type presumably in Mus. Hist. Nat. Paris; type locality, Río Nagualate, Guatemala).  


*Type locality.—Santo Domingo (Republic).*  

*Range.—Greater Antilles, Central America north to Tamaulipas and Sinaloa. Reported from various localities in the states of Quintana Roo (Isla de Mujeres), Campeche, Colima, Guerrero, Michoacán, Oaxaca, Tabasco, Tamaulipas, Veracruz, and María Magdalena Island, Nayarit.*  

**CROCODYLUS MORELETII** Duméril and Duméril  

*Crocodilus moreletii* Duméril and Duméril, Catalogue méthodique de la collection des reptiles, 1851, p. 28.—Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 1, 1870, pp. 37–38, pl. 9, fig. 2.—Schmidt, Publ. Field Mus. Nat. Hist., zool. ser., vol. 12, 1924, pp. 79–84, pl. 6, figs. 2, 7, 8.  


*Type locality.—Lake Petén, Guatemala.*  

*Range.—Atlantic slopes from Guatemala to Tamaulipas. Reported in Mexico from the states of Campeche, Chiapas (Palenque), Colima, Tabasco, Tamaulipas, Veracruz; records from Colima (by Dugès) and the Pacific side of Chiapas (Tapachula, by Sumichrast) are to be considered erroneous until corroborated.*  

**Family ALLIGATORIDAE** Gray  


*Genera.—Four, of which only one occurs in Mexico.*  

1 *Alligator mississippiensis* is recorded from “Mexico” by Garman (Bull. Essex Inst., vol. 16, 1884, p. 11) and Boettger (Kat. Rept. Senckenb. Mus., pt. 1, 1893, p. 19), but we regard the records as erroneous until verified;
Genus CAIMAN Spix

Caiman Spix, Animalia nova sine specie lacertarum . . . Brasiliam . . ., 1825, p. 3.

Genotype.—Caiman fissipes Spix (=Crocodilus latirostris Daudin).
Species.—Two species, one with three subspecies, are recognized; one occurs in Mexico.
Range.—The Isthmus of Tehuantepec to Argentina.

CAIMAN CROCODILUS FUSCUS (Cope)


Type.—Unknown.
Type locality.—Río Magdalena, Colombia.
Range.—Oaxaca and perhaps Michoacán to Colombia. Reported from the states of Oaxaca: Tapanatepec, Agua Fría; and Chiapas: Tonalá, Belén, Colonia Soconusco. A record from the Balsas Valley in Michoacán (by Gadow) may be correct, but one from Yucatán (Tozzer) is, we believe, certainly incorrect.

SPECIES INQUIRENDAE

A few species whose natural ranges remain unknown have been included in their appropriate systematic place in the preceding pages. A few others, described from "Mexico," have since been found to occur elsewhere. Among these are the following:

HERPETOCHALCIS HETEROPUS Boettger


The type locality was stated to be perhaps California, or Mexico, or Central America. The name is apparently a synonym of Chalcides heteropus Lichtenstein, Nomencl. Mus. Zool. Berol., 1856, p. 17 (=Bachia heteropa) described from La Guaira. Venezuela, and restricted in range to that country.

LEIOSAURUS BELLI Duméril and Bibron


The type locality was stated to be "Mexico," but the species (and genus) has since been discovered to be restricted to Argentina.
ANEUPORUS OCCIPITALIS Bocourt

Aneuporus occipitalis Bocourt, Mission scientifique au Mexique . . ., Études sur les reptiles, livr. 4, 1874, pp. 215–217, pl. 18, fig. 1.

This has since been renamed Tropidurus bocourti by Boulenger, since Bocourt's name is suppressed as a homonym of Tropidurus occipitalis (Peters). The species occurs in Peru.

SPHAERODACTYLUS ANTHRACINUS Cope


Three other species recorded from "Mexico," but with a type locality elsewhere, are known to be restricted to extralimital areas:

BACHIA DORBIGNYI (Duméril and Bibron)

Bachia dorbignyi (Duméril and Bibron), Müller, Reisen in den Vereinigten Staaten, Canada und Mexico, vol. 3, pt. 3, 1865, p. 604.

This species, still recognized by this name, occurs in Bolivia and Chile.

ECPHYMOTES OBTUSIROSTRIS (Wiegmann)

Ecphymotes obtusirostris (Wiegmann), Müller, op. cit., p. 600.

This name is a synonym of Anisolepis undulatus (Wiegmann), which species occurs in Brasil.

TROPIDURUS PTYCHOPLEURUS Lichtenstein

Tropidurus ptychopleurus Lichtenstein, Müller, op. cit., p. 602.

This name is a synonym of Liolaemus tenuis (Duméril and Bibron), which species occurs in Chile.

One other name, supposedly based upon a Mexican specimen, has been impossible to place:
DASIA MICROCEPHALUS (Hallowell)


_Type locality._—Mexico.

_Remarks._—The first words in Hallowell’s original description are “Syn. Scincus ventralis, Peale and Green.” Then follows the description. We believe the citation of Peale and Green’s name as a synonym (an earlier one at that) of Hallowell’s own name could be construed to make his name unavailable, that is, actually a synonym of *Scincus ventralis* and thus a part of the synonymy of *Gerrhonotus l. microcephalus*. The fact remains that the animal described by Hallowell, and presumably the type of his name, is far different from *Gerrhonotus*. We recommend association of the name with the species represented by the animal described and not assignment to the synonymy of *Scincus ventralis*.

The type is not in good condition (portions of the head mutilated), but appears to belong to a non-American genus, probably _Dasia_.

It may possibly be one of the original series of three specimens of Peale and Green’s *Scincus ventralis*, since only two of them are now present in the series labeled as cotypes, and thus would be explained Hallowell’s citation of _Scincus ventralis_ as a synonym of his species. It is also possible that a confusion of specimens occurred, the original *Gerrhonotus* being exchanged for the present type of *microcephalus*, which Hallowell erroneously thought was one of Peale and Green’s cotypes. One of these alternatives must be true: Either a peculiar skink, unknown except by the type of *microcephalus*, occurs in Mexico, or else some shift of specimens occurred in the Philadelphia Academy collections between 1830 and 1856. We favor the latter alternative.

Still another species is represented by a specimen questionably secured in Mexico, while the range of the species is suspected of being entirely extralimital.

**Diploglossus Monotropis** (Kuhl)

_Diploglossus monotropis_ (KuhL), Beiträge zur Zoologie und vergleichende Anatomie, 1820, p. 128.

The species is known to occur from Costa Rica to Ecuador. The U. S. National Museum has one specimen questionably from “Colima.” Occurrence in Mexico is highly questionable.

---

8 We are indebted to Dr. E. R. Dunn for this information.
STATE LISTS

The accompanying lists have been constructed in the same way as those for the snakes (U. S. Nat. Mus. Bull. 187, 1945, pp. 202-203) and amphibians (U. S. Nat. Mus. Bull. 194, 1948, pp. 100-101). Table 2 shows for each state and each of the indicated groups, first, the number of forms recorded; second, the number of genera represented (these two numbers separated by a diagonal line); and third, the relative position of the state (so far as number of forms is concerned) as compared with other states (this number in parentheses). The lists of snakes and amphibians have been brought more or less up to date by inclusion of the additional forms defined and recorded by various authors such as Blair, Bogert, Burger, Conant, Klauber, Smith, Tanner, and Taylor (see Univ. Kansas Sci. Bull., vol. 33, 1950, pp. 313-380, for references).
<table>
<thead>
<tr>
<th>State</th>
<th>Serpentes</th>
<th>Amphibia</th>
<th>Testudines</th>
<th>Sauroid, Amphibioidea</th>
<th>Loricata</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aguascalientes</td>
<td>1/1 (23)</td>
<td>4/3 (23)</td>
<td>1/1 (12)</td>
<td>3/2 (22)</td>
<td>0/0 (3)</td>
<td>9/7 (27)</td>
</tr>
<tr>
<td>Baja California</td>
<td>65/20 (5)</td>
<td>15/8 (17)</td>
<td>7/7 (6)</td>
<td>87/21 (1)</td>
<td>0/0 (3)</td>
<td>174/56 (3)</td>
</tr>
<tr>
<td>Campeche</td>
<td>25/21 (17)</td>
<td>16/11 (16)</td>
<td>9/7 (4)</td>
<td>27/15 (15)</td>
<td>2/1 (1)</td>
<td>79/55 (19)</td>
</tr>
<tr>
<td>Chiapas</td>
<td>71/41 (3)</td>
<td>47/21 (3)</td>
<td>6/4 (7)</td>
<td>48/22 (6)</td>
<td>2/2 (1)</td>
<td>174/90 (3)</td>
</tr>
<tr>
<td>Chihuahua</td>
<td>43/18 (10)</td>
<td>17/7 (15)</td>
<td>5/4 (8)</td>
<td>37/14 (8)</td>
<td>0/0 (3)</td>
<td>102/43 (10)</td>
</tr>
<tr>
<td>Coahuila</td>
<td>39/17 (11)</td>
<td>13/7 (19)</td>
<td>7/5 (6)</td>
<td>37/13 (8)</td>
<td>0/0 (3)</td>
<td>96/42 (12)</td>
</tr>
<tr>
<td>Colima</td>
<td>36/30 (12)</td>
<td>17/13 (15)</td>
<td>5/3 (8)</td>
<td>29/16 (11)</td>
<td>2/1 (1)</td>
<td>89/63 (13)</td>
</tr>
<tr>
<td>Distrito Federal</td>
<td>21/15 (18)</td>
<td>14/11 (18)</td>
<td>2/1 (11)</td>
<td>10/6 (20)</td>
<td>0/0 (3)</td>
<td>47/33 (22)</td>
</tr>
<tr>
<td>Durango</td>
<td>28/15 (16)</td>
<td>13/6 (19)</td>
<td>3/2 (10)</td>
<td>31/12 (10)</td>
<td>0/0 (3)</td>
<td>75/35 (20)</td>
</tr>
<tr>
<td>Guanajuato</td>
<td>25/17 (17)</td>
<td>12/8 (20)</td>
<td>2/1 (11)</td>
<td>20/8 (16)</td>
<td>0/0 (3)</td>
<td>59/34 (21)</td>
</tr>
<tr>
<td>Guerrero</td>
<td>71/41 (3)</td>
<td>42/18 (4)</td>
<td>5/4 (8)</td>
<td>49/20 (5)</td>
<td>1/1 (2)</td>
<td>168/84 (4)</td>
</tr>
<tr>
<td>Hidalgo</td>
<td>31/20 (14)</td>
<td>29/10 (7)</td>
<td>1/1 (12)</td>
<td>23/9 (16)</td>
<td>0/0 (3)</td>
<td>84/40 (15)</td>
</tr>
<tr>
<td>Jalisco</td>
<td>54/34 (7)</td>
<td>22/12 (11)</td>
<td>2/2 (11)</td>
<td>35/15 (9)</td>
<td>0/0 (3)</td>
<td>113/63 (9)</td>
</tr>
<tr>
<td>Mexico</td>
<td>14/9 (20)</td>
<td>25/12 (8)</td>
<td>1/1 (12)</td>
<td>16/6 (18)</td>
<td>0/0 (3)</td>
<td>59/28 (21)</td>
</tr>
<tr>
<td>Michoacán</td>
<td>67/34 (4)</td>
<td>24/12 (10)</td>
<td>3/2 (10)</td>
<td>38/16 (7)</td>
<td>1/1 (2)</td>
<td>133/65 (7)</td>
</tr>
<tr>
<td>Morelos</td>
<td>36/26 (12)</td>
<td>25/12 (9)</td>
<td>1/1 (12)</td>
<td>20/12 (16)</td>
<td>0/0 (3)</td>
<td>82/51 (16)</td>
</tr>
<tr>
<td>Nayarit</td>
<td>39/26 (11)</td>
<td>19/12 (13)</td>
<td>5/4 (9)</td>
<td>18/9 (17)</td>
<td>1/1 (2)</td>
<td>82/52 (17)</td>
</tr>
<tr>
<td>Nuevo León</td>
<td>39/23 (11)</td>
<td>16/8 (16)</td>
<td>4/3 (9)</td>
<td>26/9 (16)</td>
<td>0/0 (3)</td>
<td>85/43 (15)</td>
</tr>
<tr>
<td>Oaxaca</td>
<td>99/47 (1)</td>
<td>52/23 (2)</td>
<td>11/8 (2)</td>
<td>73/26 (2)</td>
<td>2/2 (1)</td>
<td>237/106 (2)</td>
</tr>
<tr>
<td>Puebla</td>
<td>52/24 (8)</td>
<td>33/15 (5)</td>
<td>2/1 (11)</td>
<td>37/13 (8)</td>
<td>0/0 (3)</td>
<td>124/53 (8)</td>
</tr>
<tr>
<td>Querétaro</td>
<td>5/4 (21)</td>
<td>3/2 (24)</td>
<td>0/0 (13)</td>
<td>7/5 (21)</td>
<td>0/0 (3)</td>
<td>15/11 (25)</td>
</tr>
<tr>
<td>Quintana Roo</td>
<td>16/13 (19)</td>
<td>3/3 (24)</td>
<td>8/7 (5)</td>
<td>16/12 (18)</td>
<td>1/1 (2)</td>
<td>44/36 (23)</td>
</tr>
<tr>
<td>San Luis Potosí</td>
<td>75/37 (2)</td>
<td>30/15 (6)</td>
<td>3/2 (10)</td>
<td>35/17 (9)</td>
<td>0/0 (3)</td>
<td>143/71 (6)</td>
</tr>
<tr>
<td>Sinaloa</td>
<td>29/25 (15)</td>
<td>21/13 (12)</td>
<td>5/4 (8)</td>
<td>26/17 (13)</td>
<td>0/0 (3)</td>
<td>81/59 (18)</td>
</tr>
<tr>
<td>Sonora</td>
<td>62/31 (6)</td>
<td>21/11 (12)</td>
<td>12/9 (1)</td>
<td>54/19 (4)</td>
<td>0/0 (3)</td>
<td>149/70 (5)</td>
</tr>
<tr>
<td>Tabasco</td>
<td>34/27 (13)</td>
<td>18/12 (14)</td>
<td>8/6 (5)</td>
<td>22/14 (15)</td>
<td>2/1 (1)</td>
<td>84/60 (16)</td>
</tr>
<tr>
<td>Tamaulipas</td>
<td>39/26 (11)</td>
<td>19/13 (13)</td>
<td>5/4 (8)</td>
<td>23/13 (14)</td>
<td>2/1 (1)</td>
<td>88/57 (14)</td>
</tr>
<tr>
<td>Tlaxcala</td>
<td>4/4 (22)</td>
<td>5/4 (22)</td>
<td>0/0 (13)</td>
<td>3/2 (25)</td>
<td>0/0 (3)</td>
<td>12/10 (26)</td>
</tr>
<tr>
<td>Veracruz</td>
<td>99/46 (1)</td>
<td>71/25 (1)</td>
<td>10/8 (3)</td>
<td>58/22 (3)</td>
<td>2/1 (1)</td>
<td>240/102 (1)</td>
</tr>
<tr>
<td>Yucatán</td>
<td>47/32 (9)</td>
<td>16/13 (16)</td>
<td>8/7 (5)</td>
<td>27/16 (12)</td>
<td>0/0 (3)</td>
<td>98/68 (11)</td>
</tr>
<tr>
<td>Zacatecas</td>
<td>16/11 (19)</td>
<td>10/5 (21)</td>
<td>0/0 (13)</td>
<td>13/6 (19)</td>
<td>0/0 (3)</td>
<td>39/22 (24)</td>
</tr>
<tr>
<td>AGUASCALIENTES</td>
<td>CHECKLIST OF REPTILES OF MEXICO</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>----------------</td>
<td>--------------------------------</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Kinosternon integrum</td>
<td>Sceloporus spinosus spinosus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus sackii communis</td>
<td>Sceloporus torquatus melanogaster</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>BAJA CALIFORNIA</strong></td>
<td><strong>BAJA CALIFORNIA</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amyda emoryi</td>
<td>Eumeces skiltonianus skiltonianus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Caretta caretta gigas</td>
<td>Gambelia wislizenii wislizenii</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Clemmys marmorata pallida</td>
<td>Petrosaurus repens</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dermochelys coriacea</td>
<td>Petrosaurus thalassinus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eretmochelys imbricata</td>
<td>Phrynopsoma cerroense</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gopherus agassizii</td>
<td>Phrynopsoma coronatum coronatum</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pseudemys scripta nebulosa</td>
<td>Phrynopsoma coronatum blainvillii</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bipes biporus</td>
<td>Phrynopsoma coronatum frontale</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anniella geronimensis</td>
<td>Phrynopsoma corneum jamesi</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anniella pulchra pulchra</td>
<td>Phrynopsoma m'callii</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Callisaurus draconoides draconoides</td>
<td>Phrynopsoma platyrhinos platyrhinos</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Callisaurus draconoides carmenensis</td>
<td>Phrynopsoma solare</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Callisaurus draconoides erinitus</td>
<td>?Phyllodactylus tuberculatus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Callisaurus draconoides gabbii</td>
<td>Phyllodactylus uniceps</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus catalinensis</td>
<td>Sator angustus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus ceralbensis</td>
<td>Sator grandaevus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus hypertyrus beldingi</td>
<td>Sauromalus ater</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus hypertyrus caeruleus</td>
<td>Sauromalus australis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus hypertyrus danheimae</td>
<td>Sauromalus hispidus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus hypertyrus hypertyrus</td>
<td>Sauromalus klauberi</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus hypertyrus pictus</td>
<td>Sauromalus slevini</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus hypertyrus schmidtii</td>
<td>Sceloporus gracius vandenburgianus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus labialis</td>
<td>Sceloporus magister lineatulus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus maximus</td>
<td>Sceloporus magister magister</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus tigris canus</td>
<td>Sceloporus magister monserratensis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus tigris celeripes</td>
<td>Sceloporus magister rufidorsum</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus tigris multiscreatus</td>
<td>Sceloporus magister zosteromus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cnemidophorus tigris rubidus</td>
<td>Sceloporus occidentalis biseriatus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Coleonyx variegatus variegatus</td>
<td>Sceloporus oreutti lieki</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Coleonyx variegatus abbotti</td>
<td>Sceloporus oreutti oreutti</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Coleonyx variegatus peninsularis</td>
<td>Streptosaurus mearnsi</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Coleonyx variegatus slevini</td>
<td>Streptosaurus slevini</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Crotaphytus collaris baileyi</td>
<td>Uma notata notata</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Crotaphytus insularis</td>
<td>Urosaurus graciusos</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ctenosaura hemilopha</td>
<td>Urosaurus microscutatus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dipsosaurus carmenensis</td>
<td>Urosaurus nigricaudus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dipsosaurus catalinensis</td>
<td>Urosaurus ornatus symmetricus</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dipsosaurus dorsalis dorsalis</td>
<td>Uta concinna</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dipsosaurus dorsalis lucasensis</td>
<td>Uta mannophorius</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Elgaria cedrosensis</td>
<td>Uta martinensis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Elgaria multicarinata nana</td>
<td>Uta squamata</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Elgaria multicarinata webbii</td>
<td>Uta stansburiana elegans</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Elgaria paucicarinata</td>
<td>Uta stansburiana hesperis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eumeces gilberti rubriceudatus</td>
<td>Uta stansburiana stejnegeri</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eumeces lagunensis</td>
<td>Uta stellata</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Xantusia gilberti</td>
<td>Xantusia henshawi</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Xantusia vigilis</td>
<td>Xantusia vigilis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Chelydra serpentina
Claudius angustatus
Dermatemys mawii
Eretmochelys imbricata
Geoemyda areolata
Kinosternon acutum
Kinosternon cruentatum cruentatum
Kinosternon leucostomum
Pseudemys scripta ornata
Ameiva undulata gaigeae
Ameiva undulata stuarti
Anolis kidderi
Anolis lemurinus bourgeaci
Anolis limifrons rodriguezi
Anolis sagrei mayensis
Anolis tropidonotus
Anolis ustus
Basiliscus vittatus
Cnemidophorus deppii oligoporus
Geoemyda pulcherrima incisa
Kinosternon abaxillare
Kinosternon cruentatum cruentatum
Kinosternon leucostomum
Lepidochelys olivacea
Pseudemys scripta ornata
Abronia matudai
Abronia ochoterenai
Ameiva festiva edwardsi
Ameiva undulata hartwegi
Ameiva undulata parva
Ameiva undulata stuarti
Ameiva undulata thomasi
Anolis biporcatus
Anolis humilis uniformis
Anolis limifrons rodriguezii
Anolis pentapion
Anolis serceus
Barisia moreleti rafaeli
Barisia moreleti temporalis
Basiliscus vittatus
Celestus rozellae
Cnemidophorus deppii deppii
Cnemidophorus guttatus immutabilis
Cnemidophorus sackii bocourtii
Coleonyx elegans elegans
Corythophanes cristatus
Corythophanes hernandezii
Corythophanes percarinatus
Cnemidophorus sackii angusticeps
Coleonyx elegans elegans
Corythophanes cristatus
Ctenosaura similis similis
Enyaliosaurus erythromelas
Eumeces schwartzei
Hemidaeulytus turcicus turcicus
Iguana iguana rhinolopha
Laemancus serratus
Scincella cherriei ixbaac
Mabuya mabouya mabouya
Sceloporus chrysostictus
Sceloporus lundelli lundelli
Sceloporus serrifer plioporus
Sceloporus teapensis
Sphaerodactylus glaucus glaucus
Crocodylus acutus acutus
Crocodylus moreletii

CHIAPAS

Ctenosaura similis similis
Eumeces sumichrasti
Gerrhonotus liocephalus austrinus
Gonatodes fuscus
Heloderma horridum
Iguana iguana rhinolopha
Scincella assata assata
Scincella assata taylori
Scincella cherriei cherriei
Lepidophyma flavimaculata flavimaculata
Lepidophyma smithii smithii
Lepidophyyma smithii tehuanae
Mabuya mabouya mabouya
Phrynosoma asio
Phyllodactylus magnus
Sceloporus carinatus
Sceloporus malachiticus aphaninitus
Sceloporus malachiticus taeniocnemis
Sceloporus melanorhinus stuarti
Sceloporus prezygus
Sceloporus siniferus siniferus
Sceloporus squamosus
Sceloporus teapensis
Sceloporus variabilis variabilis
Urosaurus bizarritus anaynomorphus
Xenosaurus rackhami
Caiman crocodilus fuscus
Crocodylus moreletii
### CHIHUAHUA

- *Chrysemys picta bellii*
- ?*Gopherus berlandieri*
- *Kinosternon hirtipes*
- *Kinosternon sonoriense*
- *Terrapene ornata*
- *Anolis nebuloides*
- *Barisia levicollis*
- *Cnemidophorus inornatus*
- *Cnemidophorus sackii communis*
- *Cnemidophorus sackii stictogrammus*
- *Cnemidophorus sackii scalaris*
- *Cnemidophorus tigris marmoratus*
- *Crotaphytus collaris baileyi*
- *Ctenosaura hemilopha*
- *Elgaria kingii kingii*
- *Eumeces callicephalus*
- *Eumeces multivirgatus*
- *Eumeces obsoletus*
- *Eumeces parviauriculatus*
- *Gambelia wislizenii wislizenii*
- *Holbrookia maculata approximans*
- *Holbrookia texana*
- *Phrynosoma cornutum*
- *Phrynosoma douglassii hernandesii*
- *Phrynosoma modestum*
- *Phrynosoma orbiculare orbiculare*
- *Phyllodactylus homolepidurus*
- *Sceloporus clarkii clarkii*
- *Sceloporus grammicus disparilis*
- *Sceloporus jarrovi jarrovi*
- *Sceloporus magister magister*
- *Sceloporus nelsoni*
- *Uta stansburiana stejnegeri*

### COAHUILA

- *Amyda enoryi*
- *Gopherus berlandieri*
- *Kinosternon flavescens flavescens*
- *Kinosternon flavescens stejnegeri*
- *Pseudemys scripta elegans*
- *Pseudemys scripta gaigeae*
- *Terrapene coahuila*
- *Barisia imbricata ciliaris*
- *Cnemidophorus inornatus*
- *Cnemidophorus sackii communis*
- *Cnemidophorus sackii semifasciatus*
- *Cnemidophorus sackii scalaris*
- *Cnemidophorus tigris marmoratus*
- *Coleonyx brevis*
- *Crotaphytus collaris baileyi*
- *Crotaphytus reticulatus*
- *Eumeces obsoletus*
- *Gambelia wislizenii wislizenii*
- *Gerrhonotus liocephalus infernalis*
- *Holbrookia maculata approximans*
- *Holbrookia maculata dickersonae*
- *Holbrookia texana*
- *Scincella laterale*
- *Phrynosoma cornutum*
- *Phrynosoma modestum*
- *Sceloporus caudus*
- *Sceloporus couchii*
- *Sceloporus goldmani*
- *Sceloporus grammicus disparilis*
- *Sceloporus jarrovi jarrovi*
- *Sceloporus merriani annulatus*
- *Sceloporus olivaceus*
- *Sceloporus ornatus caeruleus*
- *Sceloporus ornatus ornatus*
- *Sceloporus parvus parvus*
- *Sceloporus poinsettii*
- *Sceloporus scalaris slevini*
- *Sceloporus undulatus consobrinus*
- *Uma exsul*
- *Uta stansburiana stejnegeri*
<table>
<thead>
<tr>
<th>COLIMA</th>
<th>DISTRITO FEDERAL</th>
<th>DURANGO</th>
</tr>
</thead>
<tbody>
<tr>
<td>Geoemyda rubida</td>
<td>Kinosternon hirtipes</td>
<td>Kinosternon flavescens stejnegeri</td>
</tr>
<tr>
<td>Geoemyda pulcherrima pulcherrima</td>
<td>Kinosternon integrum</td>
<td>Kinosternon sonoriense</td>
</tr>
<tr>
<td>Kinosternon hirtipes</td>
<td>Lepidochelys olivacea</td>
<td>Pseudemys scripta gaigeae</td>
</tr>
<tr>
<td>Kinosternon integrum</td>
<td>Ameiva undulata sinistra</td>
<td>Anolis nebulosus</td>
</tr>
<tr>
<td>Lepidochelys olivacea</td>
<td>Anolis nebuloides</td>
<td>Barisia imbricata imbricata</td>
</tr>
<tr>
<td>Ameiva undulata sinistra</td>
<td>Anolis nebulosus</td>
<td>Chemidophorus sackii communis</td>
</tr>
<tr>
<td>Anolis nebuloides</td>
<td>Anolis schmidti</td>
<td>Chemidophorus sackii communis</td>
</tr>
<tr>
<td>Anolis nebulosus</td>
<td>Basiliscus vittatus</td>
<td>Chemidophorus guttatus immutabilis</td>
</tr>
<tr>
<td>Anolis schmidti</td>
<td>Chemidophorus deppii lineatissimus</td>
<td>Chemidophorus sackii communis</td>
</tr>
<tr>
<td>Basiliscus vittatus</td>
<td>Coleonyx elegans nemoralis</td>
<td>Coleonyx elegans nemoralis</td>
</tr>
<tr>
<td>Chemidophorus deppii lineatissimus</td>
<td>Ctenosaura pectinata</td>
<td>Ctenosaura pectinata</td>
</tr>
<tr>
<td>Chemidophorus guttatus immutabilis</td>
<td>Eumees colimensis</td>
<td>Eumees colimensis</td>
</tr>
<tr>
<td>Chemidophorus sackii communis</td>
<td>Eumees parvulus</td>
<td>Eumees parvulus</td>
</tr>
<tr>
<td>Coleonyx elegans nemoralis</td>
<td>Phrynosoma orbicolare orbicolare</td>
<td>Heloderma horridum</td>
</tr>
<tr>
<td>Ctenosaura pectinata</td>
<td>Iguana iguana rhinolopha</td>
<td>Kinosternon flavescens stejnegeri</td>
</tr>
<tr>
<td>Eumees colimensis</td>
<td>?Laemanctus longipes</td>
<td>Kinosternon sonoriense</td>
</tr>
<tr>
<td>Eumees parvulus</td>
<td>Scincella assata taylori</td>
<td>Pseudemys scripta gaigeae</td>
</tr>
<tr>
<td>Heloderma horridum</td>
<td>Mabuya mabouya mabouya</td>
<td>Anolis nebulosus</td>
</tr>
<tr>
<td></td>
<td>Phrynosoma asio</td>
<td>Barisia imbricata imbricata</td>
</tr>
<tr>
<td></td>
<td>Phrynosoma orbicolare dugesii</td>
<td>Chemidophorus sackii communis</td>
</tr>
<tr>
<td></td>
<td>Phyllocaelyclus lanei</td>
<td>Chemidophorus guttatus immutabilis</td>
</tr>
<tr>
<td></td>
<td>Sceloporus dugesii dugesii</td>
<td>Chemidophorus sackii communis</td>
</tr>
<tr>
<td></td>
<td>Sceloporus grannicus microlepidotus</td>
<td>Chemidophorus sackii communis</td>
</tr>
<tr>
<td></td>
<td>Sceloporus horridus oligoporus</td>
<td>Chemidophorus sackii scalaris</td>
</tr>
<tr>
<td></td>
<td>Sceloporus melanorhinus calligaster</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td></td>
<td>Sceloporus pyrocephalus</td>
<td>Sceloporus spinosus spinosus</td>
</tr>
<tr>
<td></td>
<td>Sceloporus utiformis</td>
<td>Sceloporus torquatus torquatus</td>
</tr>
<tr>
<td></td>
<td>Urosaurus auriculatus</td>
<td>Sceloporus aeneus aeneus</td>
</tr>
<tr>
<td></td>
<td>Urosaurus bicornatus tuberculatus</td>
<td>Sceloporus grannicus microlepidotus</td>
</tr>
<tr>
<td></td>
<td>Urosaurus clarionensis</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td></td>
<td>Crocodylus acutus acutus</td>
<td>Sceloporus spinosus spinosus</td>
</tr>
<tr>
<td></td>
<td>Crocodylus moreletii</td>
<td>Sceloporus torquatus torquatus</td>
</tr>
<tr>
<td></td>
<td>DISTRITO FEDERAL</td>
<td>DURANGO</td>
</tr>
<tr>
<td></td>
<td>Sceloporus aeneus aeneus</td>
<td>Phrynosoma cornutum</td>
</tr>
<tr>
<td></td>
<td>Sceloporus grannicus microlepidotus</td>
<td>Phrynosoma douglassii brachycereum</td>
</tr>
<tr>
<td></td>
<td>Sceloporus scalaris scalaris</td>
<td>Phrynosoma modestum</td>
</tr>
<tr>
<td></td>
<td>Sceloporus spinosus spinosus</td>
<td>Phrynosoma orbicolare orbicolare</td>
</tr>
<tr>
<td></td>
<td>Sceloporus torquatus torquatus</td>
<td>?Phyllocaelyclus lanei</td>
</tr>
<tr>
<td></td>
<td>Sceloporus aeneus aeneus</td>
<td>Sceloporus grannicus microlepidotus</td>
</tr>
<tr>
<td></td>
<td>Sceloporus scalaris scalaris</td>
<td>Sceloporus horridus oligoporus</td>
</tr>
<tr>
<td></td>
<td>Sceloporus spinosus spinosus</td>
<td>Sceloporus jarrovii jarrovii</td>
</tr>
<tr>
<td></td>
<td>Sceloporus torquatus torquatus</td>
<td>Sceloporus lineolateralis</td>
</tr>
<tr>
<td></td>
<td>Sceloporus maculosus</td>
<td>Sceloporus magister magister</td>
</tr>
<tr>
<td></td>
<td>Sceloporus poinsettii</td>
<td>Sceloporus poindsettii</td>
</tr>
<tr>
<td></td>
<td>Sceloporus scalaris scalaris</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td></td>
<td>Sceloporus slevini</td>
<td>Sceloporus scalaris slevini</td>
</tr>
<tr>
<td></td>
<td>Sceloporus spinosus spinosus</td>
<td>Sceloporus undulatus consobrinus</td>
</tr>
<tr>
<td></td>
<td>Sceloporus undulatus consobrinus</td>
<td>Uta stansburiana stejnegeri</td>
</tr>
</tbody>
</table>
# GUANAJUATO

<table>
<thead>
<tr>
<th>Kinosternon hirtipes</th>
<th>Phrynosoma orbiculare orbiculare</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kinosternon integrum</td>
<td>Sceloporus aeneus aeneus</td>
</tr>
<tr>
<td>Barisia imbricata ciliaris</td>
<td>Sceloporus dugesii intermedius</td>
</tr>
<tr>
<td>Barisia imbricata imbricata</td>
<td>Sceloporus grammicus disparilis</td>
</tr>
<tr>
<td>Cnemidophorus sackii communis</td>
<td>Sceloporus grammicus microlepidotus</td>
</tr>
<tr>
<td>Eumeces callicephalus</td>
<td>Sceloporus jarrovii minor</td>
</tr>
<tr>
<td>Eumeces dugesii</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td>Gerrhonotus liocephalus liocephalus</td>
<td>Sceloporus spinosus spinosus</td>
</tr>
<tr>
<td>Holbrookia maculata dickersonae</td>
<td>Sceloporus torquatus melanogaster</td>
</tr>
<tr>
<td>Laemanctus serratus</td>
<td>Sceloporus torquatus torquatus</td>
</tr>
<tr>
<td>Phrynosoma boucardi</td>
<td>Sceloporus variabilis variabilis</td>
</tr>
</tbody>
</table>

# GUERRERO

<table>
<thead>
<tr>
<th>Chelonia mydas</th>
<th>Hemidactylus frenatus</th>
</tr>
</thead>
<tbody>
<tr>
<td>Geoemyda pulcherrima pulcherrima</td>
<td>Iguana iguana rhinolopha</td>
</tr>
<tr>
<td>Geoemyda rubida</td>
<td>Scinella assata taylori</td>
</tr>
<tr>
<td>Kinosternon integrum</td>
<td>Mabuya mabouya mabouya</td>
</tr>
<tr>
<td>Lepidochelys olivacea</td>
<td>Phrynosoma asio</td>
</tr>
<tr>
<td>Abronia deppii</td>
<td>Phrynosoma taurus</td>
</tr>
<tr>
<td>Ameiva undulata dextra</td>
<td>Phyllocaecylus borai</td>
</tr>
<tr>
<td>Anolis dunni</td>
<td>Phyllocaecylus delcampi</td>
</tr>
<tr>
<td>Anolis gadovii</td>
<td>Phyllogaecylus lanei</td>
</tr>
<tr>
<td>Anolis liogaster</td>
<td>Phyllocaecylus magnatuberculatus</td>
</tr>
<tr>
<td>Anolis megapholidotus</td>
<td>Phyllocaecylus magnus</td>
</tr>
<tr>
<td>Anolis nebuloides</td>
<td>?Sceloporus asper</td>
</tr>
<tr>
<td>Anolis nebulosus</td>
<td>Sceloporus formosus scitulus</td>
</tr>
<tr>
<td>Anolis taylori</td>
<td>Sceloporus gadoviae</td>
</tr>
<tr>
<td>Barisia gadovii gadovii</td>
<td>Sceloporus grammicus grammicus</td>
</tr>
<tr>
<td>Basiliscus vittatus</td>
<td>Sceloporus horridus horridus</td>
</tr>
<tr>
<td>Bipes canaliculatus</td>
<td>Sceloporus horridus oligoporus</td>
</tr>
<tr>
<td>Bipes traclydylus</td>
<td>Sceloporus melanorhinus calligaster</td>
</tr>
<tr>
<td>Cnemidophorus deppii deppii</td>
<td>Sceloporus mucronatus omiltemanus</td>
</tr>
<tr>
<td>Cnemidophorus deppii lineatissimus</td>
<td>Sceloporus ochoterenai</td>
</tr>
<tr>
<td>Cnemidophorus guttatus immutablis</td>
<td>Sceloporus pyrocephalus</td>
</tr>
<tr>
<td>Cnemidophorus sackii sackii</td>
<td>Sceloporus siniferus siniferus</td>
</tr>
<tr>
<td>Coleonyx elegans nemoralis</td>
<td>Sceloporus stejnegeri</td>
</tr>
<tr>
<td>Ctenosaura pectinata</td>
<td>Sceloporus utiformis</td>
</tr>
<tr>
<td>Eumeces brevirostris</td>
<td>Urosaurus bicarinatus bicarinatus</td>
</tr>
<tr>
<td>Eumeces ochoterenai</td>
<td>Urosaurus bicarinatus anonymorphus</td>
</tr>
<tr>
<td>Gerrhonotus liocephalus liocephalus</td>
<td>Crocodylus acutus acutus</td>
</tr>
<tr>
<td>Heloderma horridum</td>
<td></td>
</tr>
<tr>
<td>HIDALGO</td>
<td></td>
</tr>
<tr>
<td>--------------------------------</td>
<td>---------------------------</td>
</tr>
<tr>
<td>Kinosternon hirtipes</td>
<td>Phrynosoma boucardi</td>
</tr>
<tr>
<td>Abronia taeniata taeniata</td>
<td>Phrynosoma orbiculare orbiculare</td>
</tr>
<tr>
<td>Barisia imbricata imbricata</td>
<td>Sceloporus aeneus bicanthalis</td>
</tr>
<tr>
<td>Barisia imbricata ciliaris</td>
<td>Sceloporus grammicus disparilis</td>
</tr>
<tr>
<td>Chelidonophorus sackii communis</td>
<td>Sceloporus grammicus microlepidotus</td>
</tr>
<tr>
<td>Eumeces lynxe furcirostris</td>
<td>Sceloporus jarrovi immucronatus</td>
</tr>
<tr>
<td>Eumeces lynxe lynxe</td>
<td>Sceloporus muenonatus muenonatus</td>
</tr>
<tr>
<td>Gaigeia gaigeae</td>
<td>Sceloporus parvus scutulatus</td>
</tr>
<tr>
<td>Gaigeia sylvatica</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td>Laemanctus serratus</td>
<td>Sceloporus spinosus spinosus</td>
</tr>
<tr>
<td>Scincella gemmingeri gemmingeri</td>
<td>Sceloporus torquatus torquatus</td>
</tr>
<tr>
<td>Scincella gemmingeri forbesorum</td>
<td>Sceloporus variabilis variabilis</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>JALISCO</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Kinosternon integrum</td>
<td>Sceloporus asper</td>
</tr>
<tr>
<td>Pseudemys scripta ornata</td>
<td>Sceloporus bulleri</td>
</tr>
<tr>
<td>Ameiva undulata sinistra</td>
<td>Sceloporus clarkii boulenger</td>
</tr>
<tr>
<td>Anclis nebulosus</td>
<td>Sceloporus dugesii dugesii</td>
</tr>
<tr>
<td>Barisia imbricata imbricata</td>
<td>Sceloporus grammicus microlepidotus</td>
</tr>
<tr>
<td>Basiliscus vittatus</td>
<td>Sceloporus heterolepis</td>
</tr>
<tr>
<td>Chelidonophorus deppii lineatissimus</td>
<td>Sceloporus horridus albiventeris</td>
</tr>
<tr>
<td>Chelidonophorus sackii communis</td>
<td>Sceloporus horridus oligoporus</td>
</tr>
<tr>
<td>Ctenosaura pectinata</td>
<td>Sceloporus melanorhinus calligaster</td>
</tr>
<tr>
<td>Eumeces brevirostris</td>
<td>Sceloporus nelsoni</td>
</tr>
<tr>
<td>Eumeces callicephalus</td>
<td>Sceloporus pyrocephalus</td>
</tr>
<tr>
<td>Eumeces dugesii</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td>Heloderma horridum</td>
<td>Sceloporus scalaris unicanthalias</td>
</tr>
<tr>
<td>Holbrookia maculata approximans</td>
<td>Sceloporus spinosus spinosus</td>
</tr>
<tr>
<td>Iguana iguana rhinolopa</td>
<td>Sceloporus torquatus melanogaster</td>
</tr>
<tr>
<td>Scincella assata taylori</td>
<td>Sceloporus utiformis</td>
</tr>
<tr>
<td>Phrynosoma orbiculare orbiculare</td>
<td>Urosaurus bicarinatus tubereulatus</td>
</tr>
<tr>
<td>?Phyllocladus lanei</td>
<td>Urosaurus gadovi</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>MÉXICO</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Kinosternon hirtipes</td>
<td>Sceloporus aeneus bicanthalis</td>
</tr>
<tr>
<td>Barisia imbricata imbricata</td>
<td>Sceloporus grammicus microlepidotus</td>
</tr>
<tr>
<td>Barisia rudicollis</td>
<td>Sceloporus jarroviii sugillatus</td>
</tr>
<tr>
<td>Chelidonophorus sackii communis</td>
<td>Sceloporus jarroviii minor</td>
</tr>
<tr>
<td>Eumeces copei</td>
<td>Sceloporus muenonatus muenonatus</td>
</tr>
<tr>
<td>Eumeces indubitus</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td>Gerrhonotus liocephalus liocephalus</td>
<td>Sceloporus spinosus spinosus</td>
</tr>
<tr>
<td>Phrynosoma orbiculare orbiculare</td>
<td>Sceloporus torquatus torquatus</td>
</tr>
</tbody>
</table>
### MICHOACÁN

<table>
<thead>
<tr>
<th>Geoemyda rubida</th>
<th>Iguana iguana rhinolopha</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kinosternon hirtipes</td>
<td>Mabuya mabouya mabouya</td>
</tr>
<tr>
<td>Kinosternon integrum</td>
<td>Phrynosoma asio</td>
</tr>
<tr>
<td>Ameiva undulata sinistra</td>
<td>Phrynosoma orbiculare orbiculare</td>
</tr>
<tr>
<td>Anolis nebulosus</td>
<td>Phylodactylus lanei</td>
</tr>
<tr>
<td>Barisia imbricata imbricata</td>
<td>Sceloporus aeneus aeneus</td>
</tr>
<tr>
<td>Basiliscus vittatus</td>
<td>Sceloporus asper</td>
</tr>
<tr>
<td>Cnemidophorus deppii lineatissimus</td>
<td>Sceloporus dugesii intermedius</td>
</tr>
<tr>
<td>Cnemidophorus guttatus immutabilis</td>
<td>Sceloporus gadoviae</td>
</tr>
<tr>
<td>Cnemidophorus sackii communis</td>
<td>Sceloporus grammicus microlepidotus</td>
</tr>
<tr>
<td>Cnemidophorus sackii sackii</td>
<td>Sceloporus horridus oligoporus</td>
</tr>
<tr>
<td>?Coleonyx elegans nemoralis</td>
<td>Sceloporus melanorhinus calligaster</td>
</tr>
<tr>
<td>Ctenosaura pectinata</td>
<td>Sceloporus pyrocephalus</td>
</tr>
<tr>
<td>Enyaliosaurus clarki</td>
<td>Sceloporus scalaris scalaris</td>
</tr>
<tr>
<td>Eumeces altamirani</td>
<td>Sceloporus spinosus spinosus</td>
</tr>
<tr>
<td>Eumeces brevirostris</td>
<td>Sceloporus torquatus torquatus</td>
</tr>
<tr>
<td>Eumeces calicephalus</td>
<td>Sceloporus torquatus melanogaster</td>
</tr>
<tr>
<td>Eumeces copei</td>
<td>Sceloporus utiformis</td>
</tr>
<tr>
<td>Eumeces dugesii</td>
<td>Urosaurus bicornatus bicornatus</td>
</tr>
<tr>
<td>Eumeces indubitus</td>
<td>Urosaurus gadovi</td>
</tr>
<tr>
<td>Heloderma horridum</td>
<td>Crocodylus acutus acutus</td>
</tr>
</tbody>
</table>

### MORELOS

<table>
<thead>
<tr>
<th>Kinosternon integrum</th>
<th>Heloderma horridum</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ameiva undulata sinistra</td>
<td>Phrynosoma orbiculare orbiculare</td>
</tr>
<tr>
<td>Anolis nebulosus</td>
<td>Sceloporus aeneus aeneus</td>
</tr>
<tr>
<td>Barisia imbricata imbricata</td>
<td>Sceloporus gadoviae</td>
</tr>
<tr>
<td>Cnemidophorus guttatus immutabilis</td>
<td>Sceloporus grammicus microlepidotus</td>
</tr>
<tr>
<td>Cnemidophorus sackii sackii</td>
<td>Sceloporus horridus horridus</td>
</tr>
<tr>
<td>Ctenosaura pectinata</td>
<td>Sceloporus ochoterenai</td>
</tr>
<tr>
<td>?Enyaliosaurus clarki</td>
<td>Sceloporus siniferus siniferus</td>
</tr>
<tr>
<td>Eumeces copei</td>
<td>Sceloporus torquatus torquatus</td>
</tr>
<tr>
<td>Eumeces indubitus</td>
<td>Urosaurus bicornatus bicornatus</td>
</tr>
<tr>
<td>Gerrhonotus liocephalus liocephalus</td>
<td></td>
</tr>
</tbody>
</table>

### NAYARIT

<table>
<thead>
<tr>
<th>Chelonia mydas</th>
<th>Iguana iguana rhinolopha</th>
</tr>
</thead>
<tbody>
<tr>
<td>Geoemyda pulcherrima pulcherrima</td>
<td>Peropus mutilatus</td>
</tr>
<tr>
<td>Kinosternon hirtipes</td>
<td>Phylodactylus lanei</td>
</tr>
<tr>
<td>Kinosternon integrum</td>
<td>Sceloporus asper</td>
</tr>
<tr>
<td>Terrapene nelsoni</td>
<td>Sceloporus clarkii boulengeri</td>
</tr>
<tr>
<td>Anolis nebulosus</td>
<td>Sceloporus dugesii dugesii</td>
</tr>
<tr>
<td>Cnemidophorus deppii lineatissimus</td>
<td>Sceloporus horridus albiventris</td>
</tr>
<tr>
<td>Cnemidophorus sackii communis</td>
<td>Sceloporus melanorhinus calligaster</td>
</tr>
<tr>
<td>Cnemidophorus sackii mariarium</td>
<td>Sceloporus nelsoni</td>
</tr>
<tr>
<td>Ctenosaura pectinata</td>
<td>Sceloporus utiformis</td>
</tr>
<tr>
<td>Eumeces calicephalus</td>
<td>Urosaurus ornatus schottii</td>
</tr>
<tr>
<td>Eumeces parvulus</td>
<td>Crocodylus acutus acutus</td>
</tr>
</tbody>
</table>
NUEVO LEÓN

Amyda emoryi  Phrynosoma cornutum
Gopherus berlandieri  Phrynosoma modestum
Pseudemys floridana texana  Phrynosoma orbiculare orbiculare
Pseudemys scripta elegans  Sceloporus couchii
Barisia imbricata ciliaris  Sceloporus cyanogenys
Cnemidophorus inornatus  Sceloporus grammicus disparilis
Cnemidophorus sackii gularis  Sceloporus jarrovii minor
Coleonyx brevis  Sceloporus olivaceus
Crotaphytus collaris baileyi  Sceloporus parvus parvus
Crotaphytus reticulatus  Sceloporus poinsettii
Eumees brevilineatus  Sceloporus scalaris slevini
Eumees dicei  Sceloporus torquatus binocularis
Eumees obsoletus  Sceloporus undulatus consobrinus
Holbrookia texana  Sceloporus variabilis marmoratus
Scincella caudaequinae

OAXACA

Chelonia mydas  Cnemidophorus sackii australis
Dermatemys mawii  Cnemidophorus sackii bocourtii
Eretmochelys imbricata  Coleonyx elegans elegans
Geoemyda pulcherrima incisa  Coleonyx elegans nemoralis
Geoemyda rubida  Corythophanes hernandezii
Kinosternon cruentatum cruentatum  Ctenosaura acanthura
Kinosternon integrum  Ctenosaura pectinata
Lepidochelys olivacea  Ctenosaura similis similis
Pseudemys scripta ornata  Enyaliiosaurus quinquecarinatus
Pseudemys umbra  Eumees brevirostris
Staurotypus salvini  Gaigeia dontomasi
Abronia fuscolabialis  Gaigeia radula
Abronia oaxacae  Gerrhonotus lioccephalus lioccephalus
Ameiva undulata amphigrama  Gymnophthalmus sumichrasti
Ameiva undulata dextra  Heloderma horridum
Ameiva undulata parva  Iguana iguana rhinolopha
Ameiva undulata stuarti  Laemanctus deborrei
Ameiva undulata undulata  Laemanctus longipes
Anolis heliactin  Laemanctus serratus
Anolis limifrons rodriguezii  Scincella assata taylori
Anolis milleri  Scincella cherriei stuarti
Anolis nebuloides  Scincella gemmingeri gemmingeri
Anolis nebulosus  Scincella silvicola
Anolis sericeus  Lepidophyma smithii tehuanae
Anolis tropidonotus  Mabuya mabouya mabouya
Barisia gadovii levigata  Phrynosoma asio
Barisia imbricata imbricata  Phrynosoma braconnieri
Barisia imbricata planifrons  Phyllocaecylus magnus
Barisia viridiflava  Phyllocaecylus muralis
Basiliscus vittatus  Sceloporus aeneus bicanthalis
Celestus enneagrammus  Sceloporus edwardtaylori
Cnemidophorus deppii deppii  Sceloporus formosus formosus
Cnemidophorus guttatus guttatus  Sceloporus gadoviae
Cnemidophorus guttatus immutabilis  Sceloporus grammicus grammicus
| Sceloporus grammicus microlepidotus | Sceloporus variabilis smithi |
| Sceloporus horridus horridus | Sceloporus variabilis variabilis |
| Sceloporus jalapae | Sphaerodactylus glaucus glaucus |
| Sceloporus malachiticus salvini | Sphaerodactylus glaucus torquatus |
| Sceloporus melanorhinus melanorhinus | Urosaurus bicarinatus anonymorphus |
| Sceloporus mucronatus olimtemanus | Urosaurus bicarinatus nelsoni |
| Sceloporus siniferus cupreus | Xenosaurus grandis |
| Sceloporus siniferus siniferus | Caiman crocodilus fuscus |
| Sceloporus spinosus caeruleopunctatus | Crocodylus acutus acutus |

**PUEBLA**

- *Sceloporus* grammicus microlepidotus
- *Sceloporus* horridus horridus
- *Sceloporus* jalapae
- *Sceloporus* malachiticus salvini
- *Sceloporus* melanorhinus melanorhinus
- *Sceloporus* mucronatus olimtemanus
- *Sceloporus* siniferus cupreus
- *Sceloporus* siniferus siniferus
- *Sceloporus* spinosus caeruleopunctatus
- *Sceloporus* teapensis

**QUERÉTARO**

| Sceloporus variabilis variabilis |

**QUINTANA ROO**

| Chelonia mydas | Terrapene mexicana yucatana |
| Eretmochelys imbricata | Ameiva undulata gaigeae |
| Geoemyda areolata | Anolis cozumelae |
| Kinosternon creaseri | Anolis limifrons rodriguezii |
| Kinosternon cruentatum consors | Anolis tropidonotus |
| Lepidochelys kempii | Aristelliger georgeensis |
| Pseudemys scripta ornata | Basiliscus vittatus |

* Morfín (Informe rendido por la comisión geográfica-exploradora de Quintana Roo al C. Secretario de Fomento, 1918, pp. 1-57, pls. 1-10) records, in a previously overlooked paper, a number of snakes from Quintana Roo. They are *Boa imperator* (= *Constrictor constrictor imperator*), *Bothrops atrix* (sic, = *B. atra asper*), *Crotalus basiliscus* (= *C. d. durissus*), and *Elaps fulvius* (= *Micrurus affinis mangensis*), all from "between Payo Obispo and Champoton."
Cnemidophorus deppii cozumelus
Cnemidophorus sackii angusticeps
Coleonyx elegans elegans
Corythophanes herlandezii
Ctenosaura similis similis

Iguana iguana rhinolopha
Mabuya mabouya mabouya
Sceloporus chrysostictus
Sceloporus cozzumelae
Croecydylus aequus aequus

SAN LUIS POTOSÍ

Kinosternon hirtipes
Kinosternon integrum
Terrapene mexicana mexicana
Amoiva undulata podarga
Anolis petri
Anolis scirceus
Anelytropsis papillosus
Barisia imbricata ciliaris
Corythophanes herlandezii
Cnemidophorus sackii communis
Cnemidophorus sackii gularis
Crotaphytus collaris baileyi
Ctenosaura acanthura
Eumeces lynxe lynxe
Eumeces tetragrammus
Gerrhonotus liocephalus infernalis
Gerrhonotus liocephalus loweryi
Holbrookia maucleata approximans
Holbrookia maucleata dickersonae
Holbrookia texana

Laemanetus serratus
Scincella caudaequiniae
Lepidophyma smithii ocellor
Phrynosoma cornutum
Phrynosoma modestum
Phrynosoma orbiculare orbiculare
Sceloporus cautus
Sceloporus goldmani
Sceloporus grannicus disparilis
Sceloporus jarrovii minor
Sceloporus jarrovii immueronatus
Sceloporus olivaceus
Sceloporus parvus parvus
Sceloporus parvus scutatulatus
Sceloporus serrifer ploiporus
Sceloporus spinosus spinosus
Sceloporus torquatus melanogaster
Sceloporus variabilis variabilis
Xenosaurus newmanorum

SINALOA

Caretta caretta gigas
Geoemyda pulcherrima pulcherrima
Kinosternon hirtipes
Kinosternon integrum
Pseudemys scripta ornata
Anolis nobolusus
Anolis utowanae
Barisia imbricata ciliaris
Callisaurus draconoides bogerti
Callisaurus draconoides brevipes
Cnemidophorus sackii communis
Coleonx fasciatus
Ctenosaura hemilopa
Ctenosaura pectinata
Dipsosaurus dorsalis sonoriensis
Eumeces humilis

Eumeces parvulus
Heloderma horridum
Holbrookia maucleata elegans
Holbrookia maucleata thermophila
Iguana iguana rhinolopha
Peropus mutilatus
Phrynosoma solare
Phylodactylus lavei
Sceloporus clarkii boulenieri
Sceloporus horridus albiventris
Sceloporus nelsoni
Sceloporus utfiformis
?Sphaerodactylus glauces torquatus
Urosaurus bicornatus tuberculatus
Urosaurus ornatus schottii

SONORA

Chelonia mydas
Dermochelys coriacea
Eretmochelys imbricata
Geoemyda pulcherrima pulcherrima
Gopherus agassizii

Kinosternon flavescens stejnegeri
Kinosternon integrum
Kinosternon sonoriense
Lepidochelys olivacea
Pseudemys scripta hiltoni
Terrapene klauberi
Terrapene ornata
Anolis nebuloides
Callisaurus draconoides brevipes
Callisaurus draconoides gabbii
Callisaurus draconoides insatus
Callisaurus draconoides ventralis
Cnemidophorus bacatus
Cnemidophorus burti
Cnemidophorus gadowi
Cnemidophorus sackii communis
Cnemidophorus sackii stictogrammus
Cnemidophorus tigris aethiops
Cnemidophorus tigris gracilis
Cnemidophorus tigris martyris
Cnemidophorus tigris sonoriensis
Crotaphytus dickersonae
Crotaphytus collaris baileyi
Ctenosaura hemilopha
Dipsosaurus dorsalis dorsalis
Dipsosaurus dorsalis sonoriensis
Elgaria kingii kingii
Eumeces calliephalus
Eumeces parviauriculatus
Gambelia wislizenii wislizenii
Heloderma horridum
Heloderma suspectum
Holbrookia maculata approximans
Holbrookia maculata thermophila
Holbrookia texana
Phrynosoma cornutum
Phrynosoma ditmarsi
Phrynosoma douglasii hernandesi
Phrynosoma m'callii
Phrynosoma modestum
Phrynosoma platyrhinos goodiei
Phrynosoma solare
Phyllocaudia homolepidurus
Sauromalus obesus townsendi
Sauromalus varius
Sceloporus clarkii clarkii
Sceloporus clarkii boulengeri
Sceloporus jarrovi jarrovi
Sceloporus magister magister
Sceloporus nelsoni
Sceloporus scalaris slevini
Sceloporus undulatus virgatus
Uma notata cowlesi
Urosaurus bicearinatus tuberculatus
Urosaurus ornatus linearis
Urosaurus ornatus schottii
Urosaurus ornatus symmetricus
Uta nolascensis
Uta palmeri
Uta stansburiana stejnegeri
Uta taylori

TABASCO

Claudius angustatus
Dermatemys mawii
Geochelidon areolata
Kinosternon acutum
Kinosternon cruentatum cruentatum
Kinosternon leucostomum
Pseudemys scripta ornata
Staurotypus triporeatus
Ameiva undulata amphigrama
Ameiva undulata stuarti
Anolis capito
Anolis humilis uniformis
Anolis laeviventris
Anolis lemurinus bourgaei
Anolis sagrei mayensis
Anolis sericeus
Anolis tropidonotus
Basiliscus vittatus
Coleonx elegans elegans
Ctenosaura similis similis
Eumeces schwartzei
Hemidactylus turecius turecius
Iguana iguana rhinolopha
Laemanctus deborrei
Scinax cherriei cherriei
Lepidophyma flavimauculata flavimauculata
Mabuya mabouya mabouya
Sceloporus serrifer ploporus
Sceloporus teapensis
Sphaeroeca glaucus glaucus
Crocodylus acutus acutus
Crocodylus moreletii
TAMAULIPAS

Amyda emoryi
Gopherus berlandieri
Pseudemys scripta cataspila
Pseudemys scripta elegans
Terrapene mexicana mexicana
Ameiva undulata podarga
Anolis sericeus
Basiliscus vittatus
Cnemidophorus sackii communis
Cnemidophorus sackii gularis
Coleonyx brevis
Crotophytus reticulatus
Ctenosaura acanthura
Eumeces brevilineatus
Eumeces dicel

Eumeces obsoletus
Eumeces tetragrammmus
Hemidaactylus turcicus turcicus
Holbrookia texana
Laemaneatius serratus
Phrynosoma cornutum
Sceloporus eyanogenys
Sceloporus grammicus disparilis
Sceloporus olivaceus
Sceloporus serrifer plioporus
Sceloporus spinosus spinosus
Sceloporus variabilis marmoratus
Sceloporus variabilis variabilis
Croodylus acutus acutus
Croodylus moreletii

TLAXCALA

Phrynosoma orbiculare orbiculare
Sceloporus grammicus microlepidotus

VERACRUZ

Chelonia mydas
Chelydra serpentina
Claudius angustatus
Dermatemys mawii
Geomys areolata
Kinosternon acutum
Kinosternon herrerai
Kinosternon integrum
Kinosternon leucostomum
Pseudemys scripta cataspila
Staurotypus triporeatus
Terrapene mexicana mexicana
Abronia taeniata graminea
Abronia taeniata taeniata
Ameiva undulata amphigamma
Anelytropsis papillosus
Anolis barkeri
Anolis cymbops
Anolis leaviventris
Anolis lemurinus bourgeaei
Anolis petersii
Anolis sericeus
Anolis tropidonotus
Barisia antaages
Barisia imbricata imbricata
Barisia modesta
Basiliscus vittatus
Celestus enneagrammuss
Cnemidophorus depii oligoporus

Cnemidophorus guttatus guttatus
Cnemidophorus sackii communis
Cnemidophorus sackii gularis
Coleonyx elegans elegans
Corythophanes cristatus
Corythophanes hondizensii
Ctenosaura acanthura
Ctenosaura similis similis
Eumeces brevirostris
Eumeces lynx furcirostris
Eumeces lynx lynx
Eumeces sumichrasti
Eumeces tetragrammus
Gerrhonotus locephalus ophiurus
Hemidaactylus mabouia
Hemidaactylus turcicus turcicus
Holbrookia propinqua piperata
Iguana iguana rhinolopha
Laemaneatus longipes
Laemaneatus serratus
Seinelea cherici stuarti
Seinelea gemmingeri gemmingeri
Seinelea silvicola
Lepidophyma flavimaculata flavimaculata
Mabuya mabouya mabouya
Phrynosoma orbiculare cortezi
Sceloporus aeneus bicanthalis
Sceloporus formosus formosus
Sceloporus grammicus microlepidotus
Sceloporus jalapae
Sceloporus jarrovii immucronatus
Sceloporus malachiticus salvini
Sceloporus megalepidurus
Sceloporus muenronatus muenronatus
Sceloporus muenronatus aureolus
Sceloporus pictus
Sceloporus serrifer plioporus

Caretta caretta caretta
Chelonia mydas
Dermatemys mawii
Geoemyda areolata
Kinosternon creaseri
Kinosternon cruentatum consors
Pseudemys scripta ornata
Terrapene mexicana yucatana
Ameiva undulata gaigeae
Anolis beckeri
Anolis kidnieri
Anolis lemurinus bourgeai
Anolis limifrons rodriguezii
Anolis sagrei mayensis
Anolis tropidonotus
Anolis ustus
Basiliscus vittatus
Cnemidophorus sackii angusticeps

YUCATÁN

Caretta caretta caretta
Chelonia mydas
Dermatemys mawii
Geoemyda areolata
Kinosternon creaseri
Kinosternon cruentatum consors
Pseudemys scripta ornata
Terrapene mexicana yucatana
Ameiva undulata gaigeae
Anolis beckeri
Anolis kidnieri
Anolis lemurinus bourgeai
Anolis lemurinus bourgeai
Anolis limifrons rodriguezii
Anolis sagrei mayensis
Anolis tropidonotus
Anolis ustus
Basiliscus vittatus
Cnemidophorus sackii angusticeps

ZACATECAS

Barisia imbricata ciliaris
Cnemidophorus sackii communis
Eumeces callicephalus
Holbrookia maculata dickersonae
Phrynosoma modestum
Phrynosoma orbiculare orbiculare
Sceloporus cautus

Sceloporus grammicus disparilis
Sceloporus jarrovii minor
Sceloporus scalaris scalaris
Sceloporus spinosus spinosus
Sceloporus torquatus melanogaster
Sceloporus undulatus consobrinus
INDEX

abaxillare, Kinosternon, 22, 218.
abottii, Coleonyx variegatus, 42, 44, 217.
abnormis, Dermatemys, 19.
Abonia, 194, 196.
deppii, 196, 197, 221.
fuscolabialis, 196, 198, 224.
matusnai, 196, 218.
oxaceae, 196, 197, 224.
ochoteronai, 196, 197, 218.
taeniata graminea, 196, 198, 225, 228.
taeniata taeniata, 196, 197, 222, 225, 228.
taeniatus, 197.

acanthinus, Sceloporus, 108.

acuta, Kinosternon scorpioides, 23.
acutirostris, Anolis, 64.

acutum, Kinosternon, 22, 23, 227, 228.
Crocodylus acutus, 210, 218, 220, 221, 223, 225, 226, 227, 228, 229.
adspersus, Gerrhonotus, 201.
aeneus, Sceloporus, 137.
Sceloporus aeneus, 136, 137, 220, 221, 222, 223, 225.
aethiops, Cnemidophorus tessellatus, 189.
Cnemidophorus tigris, 177, 189, 227.

Agama collaris, 91.
cornuta, 99.
(Phrynosoma) coronata, 102.
cristata, 68, 69.
orbicularis, 97.
torquata, 126.
undulata, 105.

agassizii, Chelonia, 17.
Gopherus, 28, 217, 226.
Testudo, 28.
Xerobates, 28.

agilis, Scincus, 156.
Agkistrodon, 3.
Akleistops, 151.
guatemalensis, 151, 152.
albiventer, Sceloporus horridus, 110, 116, 222, 226.
albogularis, Gonatodes, 45.
Gymnodaemus, 43.

alfaronis, Cnemidophorus, 178.
allacea, Mabuia, 156.
Alligator chiapensis, 212.
lacordairei, 211.
mississippiensis, 211.
Alligatoridae, 210, 211.
alpha, Sceloporus grammicus, 120.
altamirani, Eumeces, 160, 161, 163, 223.
alticoronatus, Laemaneus, 70, 229.
ambygrammatus, Eumeces skiltonianus, 167.
Amblystomidae, 2.
Ameiva, 170.
amica, Ameiva, 170.
edwardsii, 174.
festiva edwardsii, 171, 174, 218.
tesselata, 188.
undulata amphigrama, 171, 172, 224, 225, 227, 228.
undulata dextra, 171, 173, 221, 224.
undulata gaigeae, 171, 172, 218, 225, 228.
undulata hartwegi, 171, 218.
undulata parva, 171, 173, 218, 224.
undulata podarga, 171, 172, 226, 228.
undulata stuarti, 171, 173, 218, 224, 227.
undulata thomasi, 171, 173, 218.
undulata undulata, 171, 174, 224.
amica, Ameiva, 170.
americana, Lacerta, 170.
americanus, Basiliscus, 71.
Crocodylus, 210, 211.
Amoebopsis, 154.
amphigrama, Ameiva undulata, 171, 172, 224, 225, 227, 228.
Amphisbaenia, 2, 12, 37.
Amyda, 18.
cartilaginea, 18.
emoriy, 18, 217, 219, 224, 228.
javanica, 18.
spinifera, 18.

Anapsida, 12.
Ancistrodon, 3.
Anelytropsidae, 39, 170.
Anelytropsis, 170.
papillosus, 170, 226, 228.
Aneuporus occipitalis, 213.
Anguidae, 39, 40, 194.
Anguis, 194.
angustatus, Claudius, 26, 218, 227, 228.
angusticeps, Cnemidophorus, 183.
Anolis sagrei, 55.

sagrei mayensis, 56, 59, 218, 227, 229.
sallaei, 67.
schiedii, 56, 57, 58, 63, 64.
schmidtii, 56, 58, 60, 220.

sagrei dus, 68, 67, 218, 224, 226, 227, 229,

stuarti, 63.
sulcifrons, 61.
tayloiri, 57, 61, 221.
tropidonotus, 56, 59, 60, 218, 224, 225, 227, 228, 229.

uniformis, 59, 60.
ustus, 58, 66, 218, 229.
ustus veraepacis, 67.

utowanae, 57, 62, 226.
viridis, 55.
wiegmanni, 63.

anonymera, Uta, 146.

Uta bicarinata, 146.
anonymerus, Urosaurus bicarinatus, 141, 146, 218, 221, 225.

Anota, 55.
calidarium, 101.
gooden, 102.
m'callii, 95, 99.
modes, 101.

platyrhina, 101.

antagues, Barisia, 199, 200, 228.

gerrhonotus, 200.
antarinus, Sphaerodactylus, 52, 213.

approximans, Holbrookia, 83.


Archosaura, 12, 209.

areolata, Emys, 30.

Geoemyda, 29, 30, 218, 225, 227, 228, 229.

Aristelliger, 40, 41, 51.
georgeensis, 51, 225.

irregularis, 51.
lar, 51.

praesignis, 51.

arizonae, Cnemidophorus, 184.

armata, Iguana (Ctenosaura), 74.

artieulata, Cyclura, 74.

asio, Batrachosoma, 102.

Phrynosoma, 96, 102, 218, 220, 221, 223, 224.

asper, Sceloporus, 107, 109, 221, 222, 223.

Aspidonectes emoryi, 18.

Aspidoseelis, 174.


assatum, Leiolopisma assatum, 160.

Lygosoma assatum, 160.

assatus, Lampropholis, 160.

ater, Sauromalus, 79, 80, 217.

Athea, 12, 13

atillanensis, Celestus, 195.

atrir, Bothrops, 225.

auriculatus, Anolis, 64.

Sceloporus mucronatus, 123, 124, 225, 229.

auriculata, Uta, 146.
auriculatus, Urosaurus, 140, 146, 220.
australis, Cnemidophorus communis, 181.
Cnemidophorus sackii, 178, 181, 224.
Sauromalus, 79, 80, 217.
austrianus, Gerrhonotus locephalus, 203, 204, 218.
bacatus, Cnemidophorus, 176, 187, 227.
baccatus, Anolis, 57, 62.
Bachia, 293, 295, 297.
Barisia, 194, 198.
Barisia, 194, 198.
Bailey, Crotaphyatus, 92.
Crotaphythus collaris, 92, 217, 219, 220, 224, 226, 227.
bakeri, Ctenosaura, 73.
balsas, Cnemidophorus mexicanus, 181.
Basiliscus, 54, 71.
americanus, 71.
basiliscus, 71.
(Cristasauroidea) nuchalis, 72.
vittatus, 71, 218, 220, 221, 222, 223, 224, 225, 227, 228.
basilius, Basiliscus, 71.
Crotalus, 225.
Batrachosoma, 94.
asio, 102.
bekkari, Anolis, 56, 58, 228.
bartolomae, Cnemidophorus, 191.
Basiliscus, 54, 71.
austrinicus, 204, 222.
austinicus, 204, 222.
bacatus, Anolis, 57, 62.
Bachia, 293, 295, 297.
Barisia, 194, 198.
Basiliscus, 54, 71.
americanus, 71.
basiliscus, 71.
(Cristasauroidea) nuchalis, 72.
vittatus, 71, 218, 220, 221, 222, 223, 224, 225, 227, 228.
basilius, Basiliscus, 71.
Crotalus, 225.
Batrachosoma, 94.
asio, 102.
bekkari, Anolis, 56, 58, 228.
bartolomae, Cnemidophorus, 191.
Basiliscus, 54, 71.
austrinicus, 204, 222.
americanus, 71.
basiliscus, 71.
(Cristasauroidea) nuchalis, 72.
vittatus, 71, 218, 220, 221, 222, 223, 224, 225, 227, 228.

bi-carinatus, Phymatolepis, 146.
Bimanus, 38.
bianocularis, Sceloporus, 126.
Sceloporus ferrarioperezi, 126.
Sceloporus torquatus, 122, 126, 224.
Bipedidae, 37.
bipes, 37, 38.
biporatus, 38, 217.
canaliculatus, 38, 221.
tridaeylatus, 38, 221.
biporata, Dactyloa, 55, 65.
biporatus, Anolis, 58, 65, 67, 218.
Staurotyphus, 27.
biporus, Bipes, 38, 217.
Euchirotes, 38.
bi-seriatus, Sceloporus, 118.
biseriatus, Sceloporus occidentalis, 117, 118, 217.
bischoffi, Holbrookia, 84.
biscaucus, Crocodilus, 211.
bissa, Caretta, 16.
bivittata, Daconura, 71, 72.
bivittatus, Anolis petersi, 65.
blainvillii, Phrynosoma, 103.
Phrynosoma coronatum, 97, 103, 217.
Blepharactisis, 192.
speciosa, 192.
Boa imperator, 225.
bocourtii, Cnemidophorus, 181.
Cnemidophorus communis, 181.
Cnemidophorus sackii, 178, 181, 218, 224.
Gerrhonotus, 200.
Tropidurus, 213.
bocouriil, Eumeces, 166.
bogerti, Callisaurus draconoides, 86, 89, 226.
Coleonyx variegatus, 42, 44.
bordai, Phyllodactylus, 46, 47, 221.
Bothrops atratus, 225.
boucardii, Phrynosoma, 96, 98, 222.
boucardi, Phrynosoma, 98.
Tapaya, 98.
bouleneger, Sceloporus, 113.
Sceloporus clarkii, 111, 113, 222, 223, 226, 227.
boulenegerianus, Anolis, 65.
bourgaei, Anolis, 66, 67.
Anolis lemurinus, 58, 66, 218, 227, 228, 229.
brachycercum, Phrynosoma douglasi, 96, 100, 220.
Brachydaeylatus, 41.
nitratius, 41.
brachylopha, Ctenosaura teres, 75.
braeconnieri, Goniadactylus, 45.
Phrynosoma, 95, 100, 224, 225.
brevigulare, Cnemidosternum, 25.
brevilineatus, Eumeces, 162, 164, 224, 228.
brevipedis, Callisaurus draconoides, 86, 88, 226, 227.
brevirostris, Ctenosaura, 75.
Eumeces, 162, 168, 220, 221, 222, 223, 224, 225, 228.
Mabouia, 168.
brevis, Colonyx, 42, 43, 219, 220, 224, 228.
bufonius, Phrynospoma, 99.
bullaris, Anolis, 56.
Dactyloa, 55.
bulleri, Sceloporus, 122, 125, 222.
bunker, Holbrookia, 82, 84, 219.
burti, Cnemidophorus, 177, 185, 227.

Cachryx, 76, 77.
defensor, 77.
erhythmomalas, 77.
caerulea, Uta, 143.
Verticaria, 186.
caeruleopunctatus, Sceloporus spinosus, 110, 116, 225.
caeruleus, Cnemidophorus hypyrthrus, 175, 186, 217.
Sceloporus ornatus, 122, 127, 219.
Urosaurus ornatus, 141, 143, 219.
Caiman, 212.
crocodilus fuscus, 212, 218.
fissipes, 212.
calidiarum, Anota, 101.
Phrynospoma, 101.
calligaster, Sceloporus melanochninus, 110, 113, 220, 221, 222, 223.
callirostris, Emys, 32.

Callisaurus, 55, 55.
carmenensis, 87.
crinitus, 87.
dracoides, 85, 86.
dracoides bogerti, 86, 89, 226.
dracoides brevipes, 86, 88, 226, 227.
dracoides carmenensis, 86, 87, 217.
dracoides crinitus, 86, 87, 217.
dracoides dracoindes, 86, 217.
dracoides gabbei, 86, 87, 217, 227.
dracoides inusitatus, 86, 88, 227.
dracoides splendidus, 86, 87, 217.
dracoides ventralis, 86, 88, 227.
inusitatus, 88.

Canailluculatus, Bipes, 33, 221.
Chirotes, 39.
canus, Cnemidophorus, 190.
Cnemidophorus tessellatus, 190.
Cnemidophorus tigris, 176, 190, 217.

Caouana, 14, 15.
caouana, Testudo, 15, 16.
capito, Anolis, 58, 65, 227.

Caretta, 14, 15.
bissa, 16.
caretta, 16.
caretta caretta, 15, 16, 229.
caretta gigas, 15, 16, 217, 226.
gigas, 16.

Caretta nasuta, 15.
reivwaga, 15.
caretta, Caretta, 16.
Caretta caretta, 15, 16, 229.
Testudo, 16.
Carettochelyidae, 18.
Carettoidea, 12, 14.

carinatus, Sceloporus, 134, 135, 218.
carmenensis, Callisaurus, 87.

Callisaurus draconoides, 86, 87, 217.
Dipsosaurus, 78, 217.
carneus, Anolis, 65.
carolina, Cistudo, 34.
Terrapene, 34.
carolinensis, Anolis, 55, 56.
cartilaginea, Amyda, 18.
caspica, Clemmys caspica, 33.
catalinensis, Cnemidophorus, 176, 188, 217.

Dipsosaurus, 78, 217.
castasplia, Emys, 32.
Pseudemys scripta, 31, 32, 228.
caudaequinae, Leiolepispsma, 158.
Scinellia, 157, 158, 224, 226.
Caudata, 2.
cautus, Sceloporus, 117, 219, 226, 229.
cedrosensis, Elgaria, 205, 206, 217.
Gerrhonotus, 206.
celeripes, Cnemidophorus, 190.
Cnemidophorus tessellatus, 190.
Cnemidophorus tigris, 176, 190, 217.

Celestus, 194.
atillanensis, 195.
enneagrammunus, 195, 224, 228.
rozella, 195, 218.
striatus, 194.

Chelone, 17.
Chelona, 17.

Chelonia, 14, 17.
agassizii, 17.
dussumieri, 15.
lachrymata, 16.
macula, 17.
mydas, 17, 221, 223, 224, 225, 226, 228, 229.
olivacea, 14, 15.
pseudo-caretta, 16.
pseudo-mydas, 16.

virgata, 17.
Cheloniss, 17.
Cheloniidae, 14.
Chelonura, 20.
Chelopus rubida, 30.
Chelydra, 20.
rossignoni, 20.
serpentina, 20, 21, 218, 228.
Chelydridae 19, 20.
cheriei, Leiolopisma, 158.
Leiolopisma cheriei, 157.
Lygosoma assatum, 157.
Lygosoma cheriei, 157.
Mooc, 157.
Scinella cheriei, 157, 218, 227.
chiapasis, Alligator, 212.
chiricahuae, Urosaurus ornatus, 142, 143.
Uta ornata, 143.
Chirotes canaliculatus, 39.
Chloremys, 19.
Chrysemys, 29, 33.
nebulosa, 32.
picta bellii, 34, 219.
scripta elegans, 32.
chryosictus, Sceloporus, 105, 133, 218, 226, 229.
ciliaris, Barisia imbricata, 199, 202, 219, 220, 221, 222, 224, 226, 229.
Gerrhonotus leviscollis, 202.
cinereus, Sphaerodactylus, 52.
Cinosternidae, 21.
Cinosternon, 3.
cruentatum, 23.
effeldtii, 23.
hirtipes, 25.
leucostomum, 25.
punetatum, 26.
shawianum, 23.
Cinosternum berendtianum, 23.
brevigulare, 25.
cobanum, 25.
henrici, 25.
hirtipes, 25.
rustellum, 25.
scorpioides integrum, 25.
sonoriense, 26.
triliratum, 23.
Cistudo carolina, 34.
mexicana, 34.
(Onychotria) mexicana, 34.
oranta, 36.
yucatana, 35.
clarionensis, Urosaurus, 141, 142, 220.
Uta, 142.
clarki, Ctenosaura, 76.
Enyaliusaurus, 76, 223.
clarkii, Sceloporus, 113.
Sceloporus clarkii, 111, 113, 219, 227.
Claudius, 21, 26.
angustatus, 26, 218, 227, 228.
megalochephalus, 26.
pictus, 27.
severus, 27.
clausa, Testudo, 34.

Clemmys, 29, 37.
caspica caspica, 33.
guttata, 37.
marmorata pallida, 37, 217.
Clemaspis, 45.
Cnemidophorus, 1, 4, 170, 174.
alfaronis, 178.
angusticeps, 183.
arizonae, 184.
bacatus, 176, 187, 227.
bartolomae, 191.
bocourtii, 181.
burti, 177, 185, 227.
canus, 190.
catalinensis, 176, 188, 217.
celeripes, 190.
cer albensis, 175, 188, 217.
communis, 182.
communis australis, 181.
communis bocourtii, 181.
communis copei, 181, 182.
communis occidentalis, 182.
costatus, 181.
decemlineatus, 178.
deppei cozumel, 179.
deppii, 178.
deppii cozumelus, 176, 179, 226.
deppii deppii, 176, 178, 218, 221, 224.
deppii lineatissimus 176, 179, 220, 221, 222, 223, 225.
deppii oligoporos, 176, 179, 218, 228.
dickersonae, 191.
disparilis, 189.
estebanensis, 189.
gadovi, 178, 185, 227.
gracilis, 189.
ghamili, 188.
gularis, 183.
gularis gularis, 183.
gularis gularis obsoletus, 182.
gularis gularis verus, 182.
gularis mecki, 184.
gularis octolineatus, 183.
gularis scalaris, 182.
gularis sealous, 183.
gularis semifasciatus, 184.
gularis sericeus, 183.
gularis velox, 184.
guttatus, 183.
guttatus guttatus, 175, 179, 224, 228.
guttatus immutabilis, 175, 180, 218, 220, 221, 223, 224.
guttatus striatus, 180.
hyphythrus, 174.
hyphythrus beldingi, 175, 186, 217.
hyphythrus caeruleus, 175, 186, 217.
hyphythrus danheimiae, 175, 186, 217.
hyphythrus hyphythrus, 175, 187, 217.
hyphythrus pictus, 175, 186, 217.
Cnemidophorus hyperythrus schmidtii, 175, 187, 217.
immutabilis, 180.
inornatus, 177, 184, 219, 224.
labialis, 178, 185, 217.
lavittis, 178.
lineatissimus, 179.
mariarum, 183.
marmoratus, 190.
martyris, 191.
maximus, 176, 188, 217.
melanostethus, 189, 190.
mexicanus, 181.
mexicanus balas, 181.
mexicanus typica, 181.
microlepidopus, 180.
mitaegus, 183.
octolineatus, 184.
perplexus, 184, 185.
punctelineatus, 189.
rubidus, 191.
sackii, 181.
sackii angusticeps, 178, 183, 218, 226, 229.
sackii anstralis, 178, 181, 224.
sackii bocouri, 178, 181, 218, 224.
sackii gularis, 178, 183, 224, 226, 228.
sackii mariarum, 178, 183, 223.
sackii sackii, 178, 180, 221, 223, 225.
sackii scalaris, 183, 182, 219, 220.
sackii semicircularis, 178, 184, 219.
sackii stictogrammus, 178, 183, 219, 227.
septemvittatus, 184.
sexlineatus, 177, 185.
sexlineatus sackii, 180, 181.
sexlineatus tigris, 182.
stejegeri, 191.
tessellatus, 3, 176, 188.
tessellatus, 3.
tessellatus aethiops, 189.
tessellatus canus, 190.
tessellatus celeripes, 190.
tessellatus martyris, 191.
tessellatus multiscutatus, 191.
tessellatus rubidus, 191.
tessellatus stejegeri, 191.
tessellatus tessellatus, 189.
tigris aethiops, 177, 189, 227.
tigris canus, 176, 190, 217.
tigris celeripes, 176, 190, 217.
tigris gracilis, 177, 189, 227.
tigris marmoratus, 177, 190, 219, 220.
tigris martyris, 176, 191, 227.
tigris multiscutatus, 177, 191, 217.
tigris rubidus, 176, 191.
tigris tigris, 177, 189, 217, 227.
undulatus, 174.
unicolor, 150.
Cnemidophorus vandenburghi, 191.
variolosus, 190.
coahuila, Terrapene, 34, 36, 219.
cobanensis, Anolis, 63.
cobanum, Cinosternum, 25.
Cocconeus, 55, 61.
cochranae, Sceloporus, 134.
Coleonyx, 41, 42.
brevis, 42, 43, 219, 220, 224, 228.
elegans, 41, 42, 43.
elegans elegans, 42, 218, 224, 226, 227, 228, 229.
elegans nemoralis, 42, 43, 220, 221, 223, 224.
fasciatus, 42, 43, 220, 226.
variatus, 44.
variatus abbottii, 42, 44, 217.
variatus bogerti, 42, 44.
variatus peninsularis, 42, 44, 217.
variatus slevini, 42, 45, 217.
variatus sonoriensis, 42, 45, 227.
variatus variegatus, 42, 44, 217, 227.
coleonyx, Gymnodactylus, 42.
colinensis, Eumeces, 162, 169, 220.
collaris, Agama, 91.
Crotaphytus, 92.
Crotaphytus collaris, 92.
Colpochelys, 15.
communis, Cnemidophorus, 182.
completa, Ctenosaura, 73.
concentrica, Testudo, 36.
concina, Testudo, 31.
Uta, 148, 150, 217.
consobrinus, Sceloporus, 118.
Sceloporus undulatilis, 117, 118, 219, 220, 224, 229.
consors, Kinosternon cruentatum, 22, 24, 225, 229.
conspicuosa, Ctenosaura, 73.
Constrictor constrictor imperator, 225.
durissus durissus, 225.
copei, Anolis, 65.
Cnemidophorus communis, 181, 182.
Eumeces, 162, 166, 220, 222, 223, 225.
copoi, Crotaphytus, 94.
Cophosaurus, 81.
texana, 85.
texana, 85.
Cordylus (Gerrhonotus) multicarinatus, 205.
corriacea, Dermochelys, 13, 217, 226.
Testudo, 13.
cornuta, Agama, 99.
Tapaya, 99.
coronata, Agama (Phrynosoma), 102.
cornatum, Phrynosoma, 94, 96.
Phrynosoma coronatum, 97, 102, 217.
cortezii, Phrynosoma orbiculare, 96, 98, 225, 228.
Tapaya, 98.
Corythaeolus, 71.
Corythophanes, 3, 54, 68.
chamaeleopis, 68.
cristatus, 68, 69, 218, 228, 229.
hernandezii, 68, 218, 224, 226, 228, 229.
mexicanus, 68.
percarinatus, 68, 69, 218.
Corythophanes, 3, 68, 69.
costatus, Cnemidophorus, 181.
couchii, Sceloporus, 105, 129, 132, 219, 224.
cowlesi, Uma notata, 89, 90, 227.
cozumelus, Cnemidophorus, 179.
cozumela, Cnemidophorus, 176, 179, 226.
crinitus, Callisaurus, 87.
Cristasaura, 151.
Cristasaura, 71, 72.
mittrella, 71.
cristata, Agama, 68, 69.
cristatus, Corythophanes, 68, 69, 218, 228, 229.
Crocodilus bicuscutatus, 211.
latirostris, 212.
mexicanus, 211.
pacificus, 211.
acutus acutus, 210, 218, 220, 221, 223, 225, 226, 227, 228, 229.
americanus, 210, 211.
moreletii, 210, 211, 218, 220, 227, 228, 229.
niloticus, 210.
Crotalus basiliscus, 225.
polystictus, 6.
Crotaphytus, 55, 91.
baileyi, 92.
collaris, 92.
collaris baylei, 92, 217, 219, 220, 224, 226, 227.
collaris collaris, 92.
collaris dickersonae, 93.
copeli, 94.
dickersonae, 92, 93, 227.
dorsalis, 77, 78.
fasicatus, 94.
fasciatus, 94.
gambeli, 94.
inclusus, 92, 93, 217.
reticulatus, 92, 93, 219, 224, 228.
wislizenii, 94.
Dactyloa, 55.
biporata, 55, 65.
bullaris, 55.
(Anolis) laeviventris, 62.
nebulous, 55, 65.
schiedii, 63.
Dactyloperus, 51.
damulus, Anolis, 57, 61.
danheimae, Cnemidophorus hyperythrus, 175, 186, 217.
Dasia microcephalus, 214.
deborrei, Laemanctus, 70, 71, 224, 227.
deeumlineatus, Cnemidophorus, 178.
defensor, Cachryx, 77.
Cotesauras, 77.
Enyallosaurus, 76, 77, 229.
delcampi, Phyllocladactylus, 46, 47, 221.
delicatissima, Iguana, 72.
delicatissimus, Sceloporus, 131.
INDEX

Enyaliosaurus, 54, 75.
clariki, 76, 223.
defensor, 76, 77, 229.
erythromelas, 76, 77, 218.
quinquecarinatus, 76, 224.
Epaphelus, 192.
sumichrasti, 192.
Eremia, 15.
Eremochelys, 14, 16.
imbricata, 16, 217, 218, 224, 225, 226.
squama, 17.
erythromelas, Cachryx, 77.
Ctenosaura, 76, 77.
Enyaliosaurus, 76, 77, 218.
espiritensis, Verticaria, 187.
estebanensis, Cnemidophorus, 189.
Eublepharis fasciatus, 43.
variegatus, 44.
Euchelonia, 17.
Euchirotes, 38.
biporus, 38.
Eumeeces, 155, 160.
altamirani, 160, 161, 163, 223.
bocourtii, 166.
brevilineatus, 162, 164, 224, 228.
brevirostris, 162, 163, 220, 221, 222, 223, 224, 225, 228.
collimensis, 162, 169, 220.
copei, 162, 166, 220, 222, 223, 225, 228.
dicei, 162, 169, 224, 228.
dugesii, 161, 169, 221, 222, 223.
gilberti rubricaudatus, 161, 167, 217.
humilis, 162, 166, 226.
indubitus, 162, 168, 222, 223.
lagunensis, 161, 167, 217.
lynxe fureirostris, 161, 163, 222, 223, 228.
lynxe lynxe, 161, 163, 222, 225, 226, 228.
obsoluto, 161, 165, 219, 224, 228.
ochotereni, 162, 169, 221.
parviiuriculatus, 162, 166, 219, 227, 229.
pavulonius, 162, 166, 220, 223, 226.
pavimentatunus, 160.
quadricineatus, 167.
riviroseae, 164.
schmidtii, 164.
schwartzei, 160, 162, 218, 227, 229.
skiltonianus ambygrammus, 167.
skiltonianus lagunensis, 168.
skiltonianus skiltonianus, 161, 167, 217.
sumichrasti, 161, 164, 218, 228.
tetragrammus, 161, 165, 226, 228.

Euphryne, 79.
obesus, 79, 81.

Euprepes lynxe, 163.
Euprepis microcephalus, 214.
exsul, Hemidactylus, 51.
Uma, 89, 219.
fantasticus, Sphaerodactylus, 52.
fasciatus, Coleonyx, 42, 43, 220, 226.
Crotaphytus, 94.
Eublepharis, 43.
Xenosaurus, 207, 208.
fasciolatus, Crotaphytus, 94.
ferrariperesi, Sceloporus, 126.
imbratustrum, Gerrhonotus, 197.
fissipes, Caiman, 212.
flavescens, Kinosternon flavescens, 22, 214, 219.
Platythyra, 21, 24.
flavilenta, Holbrookia maculata, 83.
flavimaculata, Lepidophyryn flavimaculata, 152, 218, 227, 228.
flavimaculatus, Lepidophyryn, 151, 152.
flavomaculatum, Lepidophyryn flavomaculatum, 152.
floridanus, Sceloporus spinosus, 117.
forbesorum, Leioloipisma, 159.
Leioloipisma gemmingeri, 159.
Seicelluma gemmingeri, 157, 159, 222.
formosus, Sceloporus, 107, 108.
Sceloporus formosus, 107, 108, 224, 225, 228.
Tropidolepis, 108.
franciscensis, Verticaria, 187.
frenatus, Hemidactylus, 40, 50, 221.
frontale, Phrynosema blainvillii, 103.
Phynosema coronatum, 97, 103, 217.
frontalis, Phrynosema, 103.
fulvius, Elaps, 225.
fulvus, Sceloporus, 135.
fureirostris, Eumeeces lynxe, 161, 163, 222, 225, 228.
fuscouraturs, Anolis, 55.
fuscolabialis, Abronia, 196, 189, 224.
Gerrhonotus, 198.
fuscus, Caiman crocodilus, 212, 218.
Gonatodes, 45, 218.
Gonatodes albogularis, 45.
Gymnochelys, 45.
Pterosuchus, 212.
Stenodactylus, 45.
gabbii, Callisaurus draconoides, 86, 87, 217, 227.
Callisaurus ventralis, 87.
gadovi, Cnemidophorus, 178, 185, 227.
Urosaurus, 212, 145, 222, 223.
Uta, 145.
gadoviae, Sceloporus, 138, 221, 223, 224, 225.
gadovii, Anolis, 56, 61, 221.
Barisia gadovii, 199, 200, 221.
Gerrhonotus, 200.
gaiigae, Ameiva undulata, 171, 172, 218, 225, 228.
Gaigeia, 153, 154, 222.
Lepidophyryn, 153, 154.
Pseudemys scripta, 31, 33.
Sceloporus undulatus, 111, 112, 229.
Gerrhonotus moreleti temporalis, 199.
multi-carinatus nanus, 207.
multifasciatus, 206.
nobilis, 206.
oaeanus, 197.
obscuros, 200.
olivaceus, 201.
ophiurus, 204.
paucecarinatus, 206.
(Barissa) planifrons, 201.
rudicollis, 202.
scincicauda ignavus, 207.
scincicauda nanus, 207.
sceincicuda webbi, 207.
taeniatus, 197.
tesselatus, 203.
vaseconelosii ochoterenai, 197.
viridiflavus, 200.
webbi, 207.

Gigas, Caretta, 16.
Gigas, Caretta, 15, 16, 217, 226.
gilberti, Xantusia, 154, 155, 217.
glaucus, Sphaerodactylus, 53.
Sphaerodactylus glaucus, 52, 53.
218, 225, 227, 229.
goldmani, Sceloporus, 136, 137, 219, 226.
Terrapene, 35.

Gonatodes, 40, 41, 45.
albogularis, 45.
albogularis fusculus, 45.
fuscus, 45, 218.

Goniacyclus brachionerii, 45.
godei, Anota, 102.

Phrynosoma, 102.
Phrynosoma platyrhinos, 96, 101, 217.

Gopherus, 28.
agassizii, 28, 217, 226.
berlandierii, 28, 219, 224.

gracilis, Cnemidophorus, 189.
Cnemidophorus tigris, 177, 189, 227.
graciosa, Uta, 144.
gracieosa, Sceloporus, 106.
Urosaurus, 140, 141, 144, 217.
Urosaurus ornatus, 145.

gramhami, Cnemidophorus, 188.
graminea, Abonia taeniata, 196, 198, 225, 228.

gramineae, Gerrhonotus, 198.
grammicus, Sceloporus, 105, 119.
Sceloporus gramminicus, 119, 221, 224.
grandaevius, Sator, 139, 140, 217.
grandis, Cubina, 208.

Xenosaurus, 207, 208, 225, 229.
grayi, Emys, 33.
grayii, Poriodogaster, 151.
guatemalensis, Akleistops, 151, 152.
guentheri, Sceloporus, 108.
guenteri, Anolis, 64.
gularis, Cnemidophorus, 183.
Cnemidophorus gularis, 183.
Cnemidophorus saekli, 178, 183, 224, 226, 228.

Uta, 144.
guenteri, Anolis, 57, 63.
guttata, Clemmys, 37.
guttatus, Cnemidophorus, 183.
Cnemidophorus guttatus, 175, 179, 224, 228.
guttulatus, Lamprosaurus, 160, 165.
Gymnodactylus albugularis, 45.
coleonyx, 42.
fuseus, 45.
seapularis, 42.
Gymnophiona, 2.
Gymnophthalmus, 39, 170, 192.
lineata, 192.
sumichrasti, 192, 224.

Harlanii, Phrynosoma, 99.
hartwegi, Ameiva undulata, 171, 218.
heleactin, Anolis, 58, 67, 224.
Heloderma, 192.
hernandesii, 193.
horridum, 193, 208, 218, 220, 221, 222, 223, 224, 226, 227.
suspectum, 193, 227.
Heloderminae, 40, 192.
Hemichirotes, 38.
tridaetalyus, 38.
Hemidaetlyus, 40, 41, 49.
exul, 51.
frenatus, 40, 50, 221.
javanicus, 49.
mabouia, 40, 50, 223.
mabuya, 49.
multilatus, 51, 52.
navarri, 51, 52.
tureicus, 40, 50.
tureicus tureicus, 50, 218, 227, 228, 229.

Hemilophia, Ctenosaura, 73, 75, 217, 219, 226, 227.
hemilophia, Ctenosaura, 73, 75, 217, 219, 226, 227.
hemilophia, Ctenosaura, 73, 75, 217, 219, 226, 227.

Holbrookia, 54, 81.
approximans, 83.
bischoffii, 84.
bunkerii, 82, 84, 219.
dickersonae, 83.
elegans, 84.
elegans elegans, 84.
elegans thermophila, 84.
lacerata, 83.
maculata, 82.
maculata dickersonae, 82, 83, 219, 220, 221, 226, 229.
maculata elegans, 82, 84, 226.
maculata flavilenta, 83.
maculata lacerata, 82, 83, 219.
maculata pulchra, 82, 84.
maculata thermophila, 82, 84, 226, 227.
propinqua, 85.
propinqua piperata, 82, 85, 228.
propinqua propinqua, 82, 83, 85.
pulchra, 84.
texana, 82, 85, 219, 220, 224, 226, 227, 228.
thermophila, 84.

Homolepidurus, Phyllodactylus, 46, 48, 219, 227.
Phyllodactylus tuberculosus, 48.

Homolosaurus, 86.
ventralis, 86, 88.

Horridus, Sceloporus horridus, 110, 116, 221, 223, 225.
humeralis, Sceloporus, 134.
humilis, Eumeces, 162, 166, 226.

Hyperythra, Verticaria hyperythra, 187.
hyperythrus, Cnemidophorus, 174.
Cnemidophorus hyperythrus, 175, 187, 217.

Idiodaetlyus, 51.
georgeensis, 51.

Iguana, 72.

Iguana, 54, 72.

(Idiotaclaeis) armata, 74.
(Idiotaclaeis) bellii, 74.
delicatissima, 72.
hernandesii, 72.

Iguana iguana, 72.
iguana rhinolophia, 72, 218, 220, 221, 222, 223, 224, 225, 226, 227, 228.
iguana rhinolophia, 72, 218, 220, 221, 222, 223, 224, 225, 226, 227, 228.
(Ignysillophus) rhinolophus, 72.
(Ignysillophus) similis, 73.
tuberculata, 72.

Iguana, 72.
Iguana iguana, 72.
Lacerta, 72.

Iguanidae, 40, 53.
imbricata, Barisia imbricata, 198, 199, 201, 220, 221, 222, 223, 224, 225, 228.
Eretmochelys, 16, 217, 218, 224, 225, 226.
Testudo, 16.
imbricatus, Gerrhonotus, 198, 201.
immueronatus, Sceloporus jarrovi, 123, 128, 222, 225, 226, 229.
immutabilis, Cnemidophorus, 180.
Cnemidophorus guttatus, 175, 180, 218, 220, 221, 223, 224.
imperator, Boa, 225.
Constrictor constrictor, 225.
impetigosus, Anolis, 56, 60.
inertia, Scinella, 156.
incisa, Emys, 30.
Geoemyda pulcherrima, 29, 30, 218, 224.
indubitatus, Eumeces, 162, 168, 222, 223.
infernalis, Gerrhonotus, 204.
Gerrhonotus ioecephalus, 203, 204, 219, 226.
inornatus, Cnemidophorus, 177, 184, 219, 224.
Sphaerodactylus, 53.
insulana, Ctenosaura, 75.
insularis, Crotaphytus, 92, 93, 217.
integrum, Cninsternum, 25.
Cinosternum scorpion, 25.
Kinosternon, 22, 25, 217, 220, 221, 222, 223, 224, 225, 226, 228.
Kinosternum, 21, 23.
intermedius, Anolis, 63.
Sceloporus, 127.
Sceloporus dugesii, 122, 127, 221, 223.
Tropidolepis, 127.
interrupta, Ctenosaura, 75.
inuatus, Callisaurus, 88.
Callisaurus draconoides, 86, 88, 227.
Callisaurus ventralis, 88.
irregularis, Aristelliger, 51.
Phymatomolp (Uta), 146.
Urosaurus, 140, 146.
Uta, 146.
ixbae, Leiolopisma cherriei, 158.
Lygosoma assatum, 158.
Scinella cherriei, 157, 158, 218, 229.
jaeobi, Anolis, 68.
jalapae, Sceloporus, 136, 225, 229.
jamesi, Phrynosaurs, 102.
Phrynosaurs coronatum, 97, 102, 217.
jarrovi, Sceloporus, 127.
javanea, Amyda, 18.
Javaenic, Hemidactylus, 49.
kempii, Lepidochelys, 14, 15, 225.
Thalassochelys (Colpochelys), 15.
kidderi, Anolis, 58, 66, 218, 229.
klingi, Elgaria, 205.
Elgaria klingi, 205, 219.
Gerrhonotus, 205, 206.
Kinosternidae, 19, 21.
Kinosternon, 3, 21.
abaxillare, 22, 218.
aeutum, 22, 23, 218, 227, 228.
creaseri, 22, 23, 225, 229.
cruentatum consors, 22, 24, 225, 229.
cruentatum cruentatum, 22, 23, 218, 224, 227.
flavesens flavesens, 22, 24, 219.
flavesens stejnegeri, 22, 24, 219, 220, 226.
herrera, 22, 24, 228.
integrum, 22, 25, 217, 220, 221, 222, 223, 224, 225, 226, 228.
lucostomum, 22, 25, 218, 225, 227, 228.
longicaudatum, 21.
scorpioides, 21.
scorpioides acuta, 23.
soroniense, 22, 26, 219, 220, 226.
Kinosternum integrum, 21.
mexicanum, 23.
soroniense, 26.
lauberi, Terrapene, 34, 35, 227.
Sauromalus, 79, 80, 217.
labilis, Cnemidophorus, 178, 185, 217.
lacerta, Holbrookia, 83.
Holbrookia maculata, 82, 83, 219.
Lacerta acanthura, 74.
americana, 170.
iguana, 72.
lumbricoidea, 39.
mexicana, 39.
orbicularis, 94, 95, 97.
quadrilineata, 192.
sexlineata, 174, 185.
tureca, 50.
Lacerta mabouya, 155, 156.
orbicularis, 97.
lachrymata, Chelonia, 16.
lacordairei, Alligator, 211.
Laemanius, 54, 69.
aiccoronatus, 70, 229.
deborrei, 70, 71, 224, 227.
longipe, 69, 70, 71, 220, 224, 228.
serratus, 70, 218, 221, 222, 224, 226, 228.
laeviventris, 63, 227, 228.
Daetlya (Anolis), 62.
lagunensis, Eumeces, 161, 167, 217.
Eumeces skiltonianus, 168.
Plestiodon, 168.
Plestiodon skiltonianus, 168.
Lampropholis assatus, 160.
Lamprosaurus, 160.
guttulatus, 160, 165.
lancedolia, Iguana (Ctenosaura), 74.
lanei, Phylodactylus, 47, 48, 220, 221, 222, 223, 226.
lar, Aristelliger, 51.
laterale, Leiolopisma, 159.
Lygosoma, 159.
Seincell, 157, 159, 219.
lateralis, Lysoptychus, 105, 132.
Sceloporus, 132.
Seincus, 159.
Uta, 144.
Uta ornata, 144.
Uta (Phymatoilepis), 144.
latirostris, Crocodilus, 212.
lativittis, Cnemidophorus, 178.
Leiolopisma, 156.
assatum assatum, 160.
assatum taylori, 160.
caudaequinæ, 158.
cheriei, 158.
cherrièr cherrièr, 157.
cherrièr ibxaæ, 158.
cherrièr stuarti, 158.
forbesorum, 139.
gemmingeri, 159.
gemmingeri forbesorum, 159.
gemmingeri gemmingeri, 159.
laterale, 159.
silvicolum, 158.
safarlii, 156.
Leiosaurus belli, 212.
leminscatus, Gerrhonotus, 204.
Lepidochelys, 14.
kempii, 14, 15, 225.
olivacea, 14, 15, 218, 220, 221, 224, 226.
Lepidophyra, 151.
flavimaculata flavimaculata, 152,
218, 227, 228.
flavimaculata obscura, 151.
flavimaculatus, 151, 152.
flavomaculatum flavomaculatum, 
152.
gaiæae, 153, 154.
smithii, 152.
smithii ocellorum, 152, 153, 225, 226.
smithii smithii, 152, 218.
smithii tenuae, 152, 218, 224.
sylvatica, 154.
Lepidosaurus, 12, 37.
leucostoma, Swanka, 25.
leucostomum, Cinos startling, 25.
Kinos startling, 22, 25, 218, 225, 227, 228.
levicollis, Barisia, 199, 202, 219.
Gerrhonotus levicollis, 202.
levigata, Barisia gadovii, 199, 200, 224.
lichenigerus, Gerrhonotus, 201.
lucki, Sceloporus, 114.
Sceloporus oreutii, 111, 114, 217.
Limnochelone, 19.
micruran, 19.
linearis, Urosaurus ornatus, 142, 143, 
219, 227.
Uta ornamenta, 143.
lineata, Gymnophthalmus, 192.
lineatissimus, Cnemidophorus, 179.
Cnemidophorus deppli, 176, 179, 
220, 221, 222, 223, 225.
lineatus, Sceloporus, 115.
Sceloporus magister, 110, 115, 217.
lineolateralis, Sceloporus, 122, 126, 220.
lineolatus, Sphaerodactylus, 52.
liocephalus, Gerrhonotus, 203.
Gerrhonotus liocephalus, 203, 214, 
220, 221, 222, 223, 224, 225.
lhogaster, Anolis, 57, 62, 221.
Liolaemus tenuis, 213.
littoralis, Malaclemmys, 36.
Malaclemmys terrapin, 36.
Malaclemys terrapin, 36.
longicaudatum, Kinosternon, 21.
longicaudatus, Tapajà orbiolares, 97.
longipes, Laemaniæs, 69, 70, 71, 220, 
224, 228.
Loricata, 2, 12, 209.
loweryi, Gerrhonotus liocephalus, 203, 
204, 226.
lucasensis, Diposaurus dorsalis, 78, 
217.
lumbricoides, Lacerta, 39.
lunea, Sceloporus, 107.
lundelli, Sceloporus lundelli, 111, 112, 
218.
Lygosoma assatum assatum, 160.
assatum cherrièr, 157.
assatum ibxaæ, 158.
cherrièr cherrièr, 157.
cherrièr stuarti, 158.
(Mocaa) gemmingeri, 159.
laterale, 159.
silvicolum, 158.
safarlii, 156.
Lonomia belli, 212.
lemnisactus, Gerrhonotus, 204.
nasuta, Caretta, 15.
navarri, Hemidactylus, 51, 52.
nebuloides, Anolis, 58, 66, 219, 220, 221, 224, 225, 227.
nebulosa, Chrysemys, 32, 217.
Daectylus, 55, 65.
Pseudemys scripta, 31, 32, 217.
nebulosus, Anolis, 58, 65, 220, 221, 222, 223, 224, 225, 226.
nelsoni, Phrynosoma, 103.
Terrapene, 34, 35, 223.
Urosaurus bicarinatus, 141, 147, 225.
Uta, 147.
Uta bicarinata, 147.
memoralis, Coleonyx elegans, 42, 43, 220, 221, 223, 224.
Neoseps, 155.
newmanorum, Xenosaurus, 207, 208, 226.
nigricauda, Uta, 145.
nigracaudus, Urosaurus, 140, 145, 217.
nobilis, Elgaria, 206.
Elgaria kingii, 205, 206.
Gerrhonotus, 206.
nolascensis, Uta, 148, 151, 227.
Norops tropidonotus, 60.
vucatianicus, 60.
nota, Uma, 59.
Una notata, 89, 217.
nuchalis, Basiliscus (Cristasaura), 72.
oaxae, Abronia, 196, 197, 224.
Gerrhonotus, 197.
oberon, Sceloporus jarrovii, 123, 129, 219.
obesus, Euphyne, 79, 81.
Sauromalus obesus, 79, 81.
obscura, Lepidophyma flavimaculata, 151.
obseurus, Gerrhonotus, 200.
Sceloporus, 109.
obsolatum, Plestiodon, 160, 165.
obsolus, Cnemidophorus gularis gularis, 182.
Eumeces, 161, 165, 219, 224, 228.
obtusirostris, Echymnotus, 213.
occidentalis, Cnemidophorus communis, 182.
occipitalis, Aneuporus, 213.
Tropidurus, 213.
oculor, Lepidophyma smithii, 152, 153, 225, 226.
ochoterenai, Abronia, 196, 197, 218.
Eumeces, 162, 169, 221.
Gerrhonotus vasconceliosii, 197.
Phrynosoma, 103.
Sceloporus, 133, 135, 221, 223.
octolineatus, Cnemidophorus, 181.
Cnemidophorus gularis, 183.
Oedicoryphus, 71.
oligoporus, Cnemidophorus deppii, 176, 179, 218, 228.
Sceloporus, 116.
Sceloporus horridus, 110, 116, 220, 221, 222, 223.
Oligosoma gemmingeri, 159.
olivacea, Chelonias, 14, 15.
Lepidochelys, 14, 15, 218, 220, 221, 224, 226.
olivaceus, Gerrhonotus, 201.
Sceloporus, 111, 117, 219, 224, 226, 228.
ollopaurus, Sceloporus variabilis, 129, 130.
olliltemanus, Sceloporus, 124.
Sceloporus mueronatus, 123, 124, 221, 225.
Sceloporus torquatus, 124.
Onychotria, 34.
mexicana, 34.
Ophthalmurкус, 194.
ophius, Gerrhonotus, 204.
Gerrhonotus iiocephalus, 203, 204, 225, 228.
Porogasterus, 204.
orbiculare, Agama, 97.
orbiculare, Phrynosoma, 96, 97, 98.
Phrynosoma orbiculare, 96, 97, 219, 220, 222, 223, 224, 225, 226, 228, 229.
orbiculare, Lacerta, 94, 95, 97.
Lacertus, 97.
Tapaya, 94, 98.
oreutii, Sceloporus oreutii, 111, 114, 217.
ornata, Cistudo, 36.
Emys, 31.
Pseudemys scripta, 31, 33, 218, 222, 224, 225, 226, 227, 229.
Terrapene, 34, 36, 219, 227.
Uta, 142.
Uta ornata, 142.
ornatus, Urosaurus ornatus, 142.
Sceloporus, 127.
Scelopurus ornatus, 122, 127, 219.
pacidicus, Crocodilus, 211.
paleari, Cyclura, 76.
pallida, Clemmys marmorata, 37, 217.
palmeri, Uta, 148, 151, 227.
pananensis, Anolis, 61.
papillosus, Anelytropsis, 170, 226, 228.
Paraloma, 71.
parkeri, Ctenosaura, 75.
parva, Ameiva undulata, 171, 173, 218, 224.
Uta, 149.
parviamiculatus, Eumeces, 162, 166, 219, 227.
parviseutata, Uta, 145.
parvulus, Eumeces, 162, 166, 220, 223, 226.
parvus, Sceloporus, 131.
Sceloporus parvus, 129, 131, 219, 224, 226.
paucicarinatus, Elgaria, 205, 206, 217.
Gerrhonotus, 206.
Phrynosoma platyrhinos, 95, 101.
platyrhinos goodei, 96, 102, 227.
platyrhinos platyrhinos, 96, 101, 217.
regale, 104.
schmidtii, 103.
solare, 96, 104, 217, 226, 227.
solaris, 104.
spinimentum, 102.
taurus, 95, 104, 221, 225.
wiegmanni, 97.
Phyllocladus, 41, 46.
bordai, 46, 47, 221.
delcampi, 46, 47, 221.
homolepidurus, 46, 48, 219, 227.
lanei, 47, 48, 220, 221, 222, 223, 226.
magnatuberculatus, 47, 48, 221.
magnus, 47, 48, 218, 221, 224.
mentalis, 46.
muralis, 46, 47, 224.
pulecher, 46.
tuberculatus, 47, 49, 217.
tuberculatus homolepidurus, 48.
unetus, 46, 47, 217.
(Diplopachytylus) unetus, 47.
xanti, 49.
Phymatelepis, 140, 144.
bicarinatus, 140.
bi-carinatus, 146.
(Uta) irregularis, 146.
picta, Testudo, 33.
Verticaria, 186.
Xantusia, 155.
pictus, Claudius, 27.
Cnemidophorus hypothyrsus, 175, 186, 217.
Sceloporus, 121, 225, 229.
pilsbryi, Sceloporus, 120.
viarara, Holbrookia propinqua, 82, 85, 228.
planiceps, Phrynosoma, 99.
Phrynosoma cornutum, 99.
planifrons, Barisia imbricata, 199, 201, 224.
Gerrhonotus (Barissia), 201.
Platypholis, 160.
platyrhina, Anota, 101.
platyrhinos, Dolosaurus, 101.
Phrynosoma, 95, 101.
Phrynosoma platyrhinos, 96, 101, 217.
Platysternidae, 18.
Platyhyra, 21.
flavescens, 21, 24.
Plestiodon bellii, 163.
lagenensis, 168.
multivirgatum, 165.
obsoletum, 160, 165.
quinqueleptum, 163.
skiltonianum, 167.
skiltonianus lagenensis, 168.
tetragrammus, 165.
Platysternidae, 2.
Pleurodira, 12.
pleurostictus, Sceloporus, 119.
INDEX

plioporus, Sceloporus serrifer, 122, 124,
218, 226, 227, 229.
Plistodon sunichrasti, 164.
Proepus, 49.
podarga, Ameiva undulata, 171, 172,
226, 228.
poinsettii, Sceloporus, 121, 122, 125,
219, 220, 224.
Sceloporus torquatus, 125.
polyphemus, Testudo, 28.
polystictus, Crotalus, 6.
Poroiodogaster, 151.
grayii, 151.
praesignis, Aristelliger, 51.
prezygus, Sceloporus, 107, 109, 218.
Pristiecerus, 55.
propinqua, Holbrookia, 85.
Holbrookia propinqua, 82, 83, 85.
propus, Chamaelea, 59.
Pseudemys, 29, 31.
elegans, 32.
florida, texana, 31, 33, 224.
scripta, cataspila, 31, 32, 228.
scripta, elegans, 31, 32, 219, 228.
scripta, gaigeae, 31, 33.
scripta, hiltoti, 31, 32, 226.
scripta, nebulosa, 31, 32, 217.
scripta, ornata, 31, 33, 218, 222,
224, 225, 226, 227, 229.
texana, 33.
troostii, elegans, 32.
ubra, 31, 33, 224.
pseudo-caretta, Chelonia, 16.
pseudo-mydas, Chelonia, 16.
Pterogastenes, 203.
Pterogasterus modestus, 201.
ophrus, 204.
ptychopleurus, Tropidurus, 213.
pulcher, Phyllodactylus, 46.
pulcherrima, Emys, 30.
Geemyda pulcherrima, 29, 30, 220,
221, 223, 226.
pulchra, Anniella, 209.
Anniella pulchra, 209, 217.
Holbrookia, 84.
Holbrookia maculata, 82, 84.
punctata, Testudo, 37.
punctatissimus, Sphaerodactylus, 52.
punctatum, Cinosternum, 26.
punctilineatus, Chrysemys, 189.
pyrocephalus, Sceloporus, 105, 138, 139,
220, 221, 222, 223.
pyrhocephalus, Sceloporus, 139.
quadrilineata, Lacerta, 192.
quadrilineatus, Eumeces, 167.
quinquecarinata, Cyclura, 76.
quinquecarinatus, Enyalius, 76, 224.
quinquelineatum, Plestiodon, 163.
quinquelineatus, Scelus, 163.
rackhami, Xenosaurus, 208, 218.
radula, Gaigea, 153, 224.
rafaeli, Barisia moreleti, 198, 199, 218.
Gerrhonotus moreleti, 199.
Rapara, 20.
rapiacuda, Gekko, 49.
Thecadactylus, 49.
rapiacuda, Thecadactylus, 49, 229.
regale, Phrynops, 104.
remivaga, Caretta, 15.
reps, Petrosaurus, 90, 91, 217.
Uta, 90.
Reptilia, 12.
reticulatus, Crotaphytus, 92, 93, 219,
224, 228.
Rhinelemys, 29.
Rhinelemmys mexicana, 30.
rhinolophus, Iguana iguana, 72, 218, 220,
221, 222, 223, 224, 225, 227, 228.
rhinocephalus, Iguana [Hypsilocephalus], 72.
rhodostictus, Callisaurus, 87.
Rhynchocephalia, 37.
rodriguezi, Anolis, 64.
Anolis limifrons, 57, 64, 218, 224,
225, 229.
rossignoni, Chelydra, 20.
Emysauras, 20.
rostellum, Cinosternum, 25.
rovirosae, Eumeces, 164.
rozellae, Celestus, 205, 218.
rubida, Chelopus, 30.
Geoemyda, 29, 30, 220, 221, 223, 224.
rubidus, Chrysemys, 191.
Chrysemys tessellatus, 191.
Chrysemys tigris, 176, 191, 217.
rubigenous, Anolis, 64.
rubracaudatus, Eumeces gigleri, 161,
167, 217.
rubriventris, Sceloporus, 119.
rudicollis, Barisia, 109, 202, 222.
Gerrhonotus, 202.
rufidorsum, Sceloporus, 115.
Scleropus magnus, 112, 115, 217.
rufopunctata, Uma, 89.
ruthveni, Anolis, 60.
saeki, Chrysemys, 181.
Chrysemys saeki, 178, 180,
221, 223, 225.
Chrysemys sexlineatus, 180, 181.
sagraei, Anolis, 55.
Salientia, 2.
sallaei, Anolis, 67.
salvini, Sceloporus, 108.
Sceloporus malachiticus, 107, 108,
225, 229.
salvini, Staurotypus, 27, 224.
Staurotypus (Stauromys), 27.
Sator, 55, 139.
angustus, 139, 217.
grandeau, 139, 140, 217.
Sauria, 2, 12, 37, 39.
Sauromalus, 20.
Sauromalus, 54, 79.
at, 79, 80, 217.
australis, 79, 80, 217.
hispidus, 79, 80, 217.
kauber, 79, 80, 217.
obesus obesus, 79, 81.
obesus townsendi, 79, 81, 227.
Sauromalus obesus tumidus, 79, 81.

clevini, 79, 80, 217.
townsendi, 81.
vara1us, 79, 81, 227.

scalaris, Cnemidophorus gularis, 182.
Cnemidophorus sackii, 178, 182, 219, 220.

Sce1oporus, 105, 137.
Sce1oporus scalaris, 136, 137, 220, 221, 222, 223, 229.

scapularis, Gymnodactylus, 42.

Sce1oporus, 55, 105.
acanthinus, 108.
aeneus, 137.
aeneus aeneus, 136, 137, 220, 221, 223, 225.
aeneus bicanthalis, 136, 137, 222, 224, 225, 228.
asper, 107, 109, 221, 222, 223.
biocularis, 126.
bullengeri, 113.
bulli1er, 122, 125, 222.
carinatus, 154, 155, 218.
cautus, 117, 219, 226, 229.
chrysostictus, 105, 133, 218, 226, 229.
clarkii, 113.
clarkii bullengeri, 111, 113, 222, 223, 226, 227.
clarkii clarkii, 111, 113, 219, 227.
cochranae, 134.
consobrinus, 118.
couchi, 105, 129, 132, 219, 224.
cozumelae, 129, 130, 226, 229.
cupreus, 134.
cytogenes, 123, 125, 224, 228.
delicatissimus, 131.
digueti, 114.
dispar, 120.
disparilis, 120.
dugesii, 127.
dugesii dugesii, 122, 127, 220, 222, 223.
dugesii intermedius, 122, 127, 221, 223.
edwardtaylori, 110, 112, 224.
ferraripeperi, 126.
ferraripeperi binocularis, 126.
ferraripeperi melanogaster, 126.
formosus, 107, 108.
formosus formosus, 107, 108, 224, 225, 228.
formosus scitulus, 107, 109, 221.
fulvus, 135.
gadoviae, 138, 221, 223, 224, 225.
goldmani, 136, 137, 219, 226.
graciosus, 106.
graciosus vandenburgianus, 119, 217.
grannicius, 105, 119.
grannicius alpha, 120.
grannicius disparilis, 119, 120, 219, 220, 221, 222, 224, 226, 228, 229.

Sce1oporus grammicus grammicus, 119, 221, 224.
grannicius microlepidotus, 119, 120, 220, 221, 222, 223, 225, 228, 229.
guenteri, 108.
heterolepis, 119, 121, 222.
heterurus, 120.
horridus albiventris, 110, 116, 222, 226.
horridus horridus, 110, 116, 221, 223, 225.
horridus oligoporus, 110, 116, 220, 221, 222, 223.
humaralis, 134.
intermedius, 127.
jalapae, 136, 225, 229.
jarrovi, 128.
jarrovi immu1eronatus, 123, 128, 222, 224, 225, 226, 229.
jarrovi jarrovii, 122, 123, 128, 219, 220, 227.
jarrovi minor, 123, 128, 221, 222, 224, 225, 226, 229.
jarrovi oberon, 123, 129, 219.
jarrovi sugillatus, 123, 128, 222.
lateralis, 132.
llicki, 114.
lineatulus, 115.
lineolateralis, 122, 126, 220.
lunae, 107.
lundelli gaigeae, 111, 112, 229.
lundelli lundelli, 111, 112, 218.
maclurus, 105, 133, 220.
magister lineatulus, 110, 115, 217.
magister monserratensis, 112, 115, 217.
magister rufidorsum, 112, 115, 217.
magister zosteromus, 112, 115, 217.
malachiticus malachiticus, 107.
malachiticus salvini, 107, 108, 225, 229.
malachiticus smaragdinus, 107.
malachiticus taeniocnemis, 107, 218.
mormoratus, 131.
megalepidurus, 106, 121, 225, 228, 229.
melanogaster, 126.
melanorhinus, 112, 113.
melanorhinus calligaster, 110, 113, 220, 221, 222, 223.
melanorhinus melanorhinus, 110, 112, 225.
melanorhinus stuarti, 110, 113, 218.
merriami, 105, 132.
merriani annulatus, 132, 133, 219.
merriani merriami, 132.
microlepidopterus, 120.
microlepidotus, 120.
microlepidotus disparilis, 120.
microlepidotus microlepidotus, 120.
microlepis, 120.
monserratensis, 115.
INDEX

249

Sceloporus muconatus aureolus, 123, 124, 225, 229.
muconatus muconatus, 123, 124, 222, 225, 229.
muconatus omiltemanus, 123, 124, 221, 225.
obscurs, 109.
occidentalis biseriatus, 117, 118, 217.
ochoterenai, 135, 135, 221, 223.
oligoporus, 116.
olivaceus, 111, 117, 219, 224, 226, 228.
omiltemanus, 124.
ornatus, 111, 114, 217.
ornatus orcutti, 114, 114, 217.
orcutti, 131.
parvus, 131.
parvus parvus, 129, 131, 219, 224, 226.
parvus scutulatus, 129, 131, 222, 226.
pietus, 121, 225, 229.
pilbryi, 120.
pleurostictus, 119.
poinsetti, 121, 122, 125, 219, 220, 224.
prezvgus, 105, 107, 219, 220.
pyrocephalus, 105, 138, 139, 220, 221, 222, 223.
pyrocephalus, 139.
rubriventris, 119.
rufidorsum, 115.
salvini, 108, 137.
salvini, 108, 135, 137.
salvini scalaris, 161, 137, 220, 221, 222, 223, 229.
salvini unianthalis, 136, 138, 222.
serrifer plioporus, 122, 124, 218, 226, 227, 229.
serrifer serrifer, 122, 123, 218, 228, 229.
siniferus, 105, 134.
siniferus eupreus, 134, 225.
siniferus siniferus, 134, 218, 221, 222, 223, 225.
spinulosus, 106, 107, 116.
spinulosus caeruleopunctatus, 110, 116, 225.
spinulosus floridanus, 117.
squamosus, 134, 135, 218.
stejngeri, 107, 109, 221.
taeniocomimus, 107.
teapensis, 129, 130, 218, 225, 227, 229.
torquatus, 105, 106, 121.
torquatus binocularis, 122, 126, 224.
torquatus cyanogenys, 125.

Sceloporus torquatus melanogaster, 122, 126, 217, 221, 222, 223, 226, 229.
torquatus minor, 128.
torquatus muconatus, 124, 125.
torquatus omiltemanus, 124.
torquatus poineettii, 125.
torquatus torquatus, 122, 125, 220, 221, 222, 223, 225, 229.
undulatus, 106, 107.
undulatus consobrinus, 117, 118, 219, 220, 224, 229.
undulatus virgatus, 117, 118, 219, 227.
utfornix, 105, 135, 220, 221, 222, 223, 226.
vandenburgianus, 119.
variabilis, 105, 130.
variabilis marmoratus, 130, 131, 219, 224, 228.
variabilis olloporus, 129, 130.
variabilis smithi, 130, 131, 225, 226.
variabilis variabilis, 130, 218, 221, 222, 225, 226, 228, 229.
viviparus, 108.
westphalii, 127.
zosteromus, 115.
schiedii, Anolis, 57, 58, 63, 64.
Daectyla, 63.
schlegeli, Dermoehelys, 15.
Sphargis coriacea, 13.
schmidtii, Anolis, 56, 58, 60, 220.
Chemidophorus hyperythrus, 175, 187, 217.
Eumeces, 164.
Phrynosoma, 103.
Urosaurus ornat, 142, 143, 219.
Uta ornat, 143.
Varidushyperythra, 187.
schotti, Urosaurus ornatus, 141, 144, 223, 226, 227.
Uta, 144.
Uta ornat, 144.
Scincella, 155, 156.
assata tayloiri, 157, 160, 218, 220, 221, 222, 224.
caudaquinac, 157, 158, 224, 226.
cherriei cherriei, 157, 218, 227.
cherriei ixbaac, 157, 158, 218, 229.
cherriei stuarti, 157, 158, 224, 228.
gemmingeri forbesorum, 157, 159, 222.
gemmingeri gemmingeri, 157, 159, 222, 224, 228.
incerta, 156.
laterale, 157, 159, 219.
sivico, 157, 158, 224, 228.
seinecauda, Tropidolepis, 205.
Seicidae, 40, 155.
Seineus agilis, 156.
lateralis, 159.
pavimentatus, 160.
quinquelinctus, 163.
sloanii, 156.
telfairii, 156.
Scincus ventralis, 203, 214.
seitulus Sceloporus formosus, 107, 109, 221.
semifasciatus, 21.
seoploides, Kinosternon, 21.
securillis, Sceloporus parvus, 129, 131, 222, 229.
sealurus, Cnemidophorus gularis, 183.
semicristata, Cyclura, 74.
semifasciatus, Cnemidophorus gularis, 184.
Scincidophorus sackii, 178, 184, 219.
Seps murinus, 175.
septemvittatus, Cinemidophorus, 184.
serricra, Verticaria, 186.
serricraus, Anolis, 58, 67, 218, 224, 226, 227, 228.
Scincidophorus gularis, 183.
Serpentes, 2, 12, 37.
serrata, Chelydra, 20, 21, 218, 228.
Testudo, 20, 21.
serratus, Laemcactus, 70, 218, 221, 222, 224, 226, 228, 229.
serrifer, Sceloporus serrifer, 122, 123, 218, 229.
severus, Claudius, 27.
sexlineata, Lacerta, 174, 185.
sexlineatus, Cinemidophorus, 177, 185.
shawianum, Cinemosternon, 23.
shawii, Cyclura, 74.
Siderolamprus, 194.
enneagrammus, 194.
silvicola, Scincella, 157, 158, 224, 228.
silvicolum, Leiolopisma, 158.
similis, Ctenosaura, 73, 218, 224, 226, 227, 228, 229.
Cyclura (Ctenosaura), 73.
Iguana (Ctenosaura), 73.
siniferus, Sceloporus, 105, 134.
Sceloporus siniferus, 131, 218, 221, 223, 225.
skiltonianum, Plestiodon, 167.
skiltonianus, Eumees, 161, 167, 217.
slevini, Coleonyx variegatus, 42, 45, 217.
Sauromalus, 79, 80, 217.
Streptosaurus, 91, 217.
Uta, 91.
sloanii, Scincus, 156.
smaragdinus, Sceloporus malachiticus, 107.
smithi, Sceloporus variabilis, 130, 131, 225.
smithii, Lepidophyma, 152.
Lepidophyma smithii, 152, 218.
solare, Phrynosoma, 96, 104, 217, 226, 227.
solaris, Phrynosoma, 104.
sonoriense, Cinemosternon, 26.
Kinosternon, 22, 26, 219, 220, 226.
Kinosternum, 26.
Thyrostenon, 26.
sonoriensis, Coleonyx variegatus, 42, 45, 227.
Dipsosaurus dorsalis, 77, 79, 226, 227.
Spasmocnemis, 51.
speciosa, Blepharactisis, 192.
spengleri, Testudo, 29.
Sphaerodactylus, 41, 52.
anthracinus, 52, 213.
cinerous, 52.
fantasticus, 52.
glaucus, 53.
glaucus glaucus, 52, 53, 218, 225, 227, 229.
glaucus torquatus, 52, 53, 225, 226.
inornatus, 53.
lineolatus, 52.
punetatisimus, 52.
sputator, 52.
torquatus, 53.
Sphargis coriacea schlegelii, 13.
spinihera, Amyda, 18.
spinitionum, Phrynosoma, 102.
spinosus, Sceloporus, 106, 107, 116.
Sceloporus spinosus, 110, 111, 116, 126, 217, 220, 221, 222, 223, 225, 228, 229.
splendidus, Callisaurus, 87.
Callisaurus draconoides, 86, 87, 217.
sputator, Sphaerodactylus, 52.
Squamata, 12, 37.
squamata, Eretmochoelys, 17.
Uta, 148, 149, 217.
squamosus, Sceloporus, 134, 135, 218.
stansburiana, Uta, 148.
Uta stansburiana, 148.
Stauremys, 27.
Staurotypus, 21, 27.
biporeatus, 27.
marmoratus, 27.
levinii, 27, 224.
(Sstauremys) salvinii, 27.
triporeatus, 27, 227, 228.
estindechneri, Diplogloasus, 195.
stejnegeri, Cinemidophorus, 191.
Cinemidophorus tessellatus, 191.
Kinosternon flavescens, 22, 24, 219, 220, 226.
Sceloporus, 107, 109, 221.
stellata, Uta, 148, 150, 217.
Stenodactylus fuscus, 45.
variegatus, 44.
stictogrammus, Cnemidophorus sackii, 178, 183, 219, 227.
Streptosaurus, 55, 91.
mearnsi, 91, 217.
slevini, 91, 217.
striatus, Celestus, 194.
Cinemidophorus guttatus, 180.
<table>
<thead>
<tr>
<th>Species</th>
<th>Category</th>
</tr>
</thead>
<tbody>
<tr>
<td>troruca, Agama, 126.</td>
<td>torquata, Agama, 126.</td>
</tr>
<tr>
<td>troruatus, Sceloporus, 105, 106, 121.</td>
<td>Sceloporus torruatus, 122, 125, 220, 221, 222, 223, 225, 229.</td>
</tr>
<tr>
<td>Tropidurus, 121.</td>
<td>Tropidurus bicarinatus, 140, 146, 220.</td>
</tr>
</tbody>
</table>
| townsendi, Sauromalus, 81. | bica
rinatus an

omorphus, 141, 146, 218, 221, 225. |
| Sauromalus obesus, 79, 81, 227. | bicarinatus bicarinatus, 141, 146, 221, 223, 225. |
| Trachyderma, 192. | bicarinatus nelsoni, 141, 147, 225. |
| horridum, 192, 193. | bicarinatus tuberculatus, 141, 147, 220, 222, 226, 227. |
| Trachypilus, 55. | clarionensis, 142, 142, 220. |
| tridactylus, Bipes, 38, 221. | gadovi, 140, 145, 222, 223. |
| Hemichirotes, 38. | graciosus, 140, 141, 144, 217. |
| triliraturn, Cinosternum, 23. | irregularis, 140, 146. |
| Trionichidae, 18. | microscutatus, 140, 145, 217. |
| Trionychidae, 18. | nigricaudus, 140, 145, 217. |
| Triporcatus, Stylonychia, 27. | ornatus chiri
cahuae, 142, 143. |
| Triporcatus, Staurotypus, 27, 227, 228. | ornatus graciosus, 145. |
| Tropidogaster, 94. | ornatus linearis, 142, 143, 219, 227. |
| Tropidolepis, 105, 205. | ornatus ornatus, 142. |
| formosus, 108. | ornatus schmidtii, 142, 143, 219. |
| intermedius, 127. | ornatus schottii, 111, 144, 223, 226, 227. |
| scinincta, 205. | ornatus symmetricus, 142, 144, 217, 227. |
| tropidonotus, Anolis, 56, 59, 60, 218, 224, 225, 227, 228, 229. | unicus, 141, 147, 219. |
| Norops, 60. | ustus, Anolis, 58, 66, 218, 229. |
| Tropidopilus, 55. | Uta, 1, 55, 146, 147. |
| Tropidurus boe
courtii, 213. | anony

ompha, 146. |
| occipitalis, 213. | auriculata, 146. |
| ptychopleurus, 213. | bica

rinata, 146. |
| torquatus, 121. | bica

rinata an

omorpha, 146. |
| tuberculata, Iguana, 72. | bi-

carinata, 146. |
| Uta, 147. | bi-

carinata bi-
carinata, 146. |
| Uta bi-carinata, 147. | bi-

carinata nelsoni, 147. |
| tuberculata, Urosaurus bicarinatus, 141, 147, 220, 222, 226, 227. | bi-

carinata tuberculata, 147. |
| tuberculatus, Gekko, 50. | caerulae, 143. |
| Phylodactylus, 47, 49, 217. | clarionensis, 142. |
| tmidius, Sauromalus obesus, 79, 81. | concinna, 148, 150, 217. |
| turecia, Lacerta, 50. | elegans, 148, 149. |
| tureicus, Hemidactylus, 40, 50. | gadovi, 145. |
| Hyla
dactylus tureicus, 50, 218, 227, 228, 229. | gracilis, 144. |
| typica, Chemidophorus mexicanus, 181. | gularis, 144. |
| Uma, 54, 89. | irregularis, 146. |
| exsul, 89, 219. | lateralis, 144. |
| notata, 89. | (Phymatolepis) lateralis, 144. |
| notata cowlesi, 89, 90, 227. | mannophorus, 148, 149, 217. |
| notata notata, 50, 217. | martinensis, 148, 149, 217. |
| Rufopunctata, 89. | mearnsi, 91. |
| umbra, Emys, 33. | microscutata, 145. |
| Pseudemys, 89, 31, 33, 224. | nelsoni, 147. |
| unctus, Diplogractus, 47. | nigricauda, 145. |
| Phylodactylus, 46, 47, 217. | nolascensis, 148, 151, 227. |
| Phylodactylus (Diplogractus), 47. | ornata, 142. |
| undulata, Agama, 105. | ornata chiri
cahuae, 143. |
| Ameiva undulata, 71, 174, 224. | ornata lateralis, 144. |
| undulatus, Cnemidophorus, 174. | ornata linearis, 143. |
| Sceloporus, 106, 107. | ornata ornata, 142. |
| unica, Uta, 147. | ornata schmidtii, 143. |
| unicanthalis, Sceloporus scalaris, 136, 138, 222. | ornata schottii, 144. |
| unicolor, Cnemidophorus, 180. | ornata symmetrica, 144. |
| unicus, Urosaurus, 141, 147, 219. | palmeri, 148, 151, 227. |
| uniformis, Anolis, 59, 60. | parva, 149. |
| Anolis humilis, 56, 60, 218, 227. | Urosaurus, 55, 140. |
INDEX

Uta parviscutata, 145.
repens, 90.
schottii, 144.
slevini, 91.
squamata, 148, 149, 217.
stansburiana, 148.
stansburiana elegans, 148, 150, 217.
stansburiana hesperis, 148, 149, 217.
stansburiana stansburiana, 148.
stellata, 148, 150, 217.
syinmetrica, 144.
taylori, 148, 150, 227.
thalassiiia, 90.
tuberculata, 147.
unica, 147.
Utiformis, Sceloporus, 105, 135, 220, 221, 222, 223, 226.
Uutowanac, Anolis, 57, 62, 226.
vandenburghi, Cnemidophorus, 191.
vandenburgianus, Sceloporus, 119, 217.
Sceloporus graciosus, 119, 217.
variabilis, Sceloporus, 105, 130.
Sceloporus variabilis, 130, 218, 221, 222, 225, 226, 228, 229.
variegatus, Coleonyx, 44.
Coleonyx variegatus, 42, 44, 217, 227.
Eublepharis, 44.
Stenodactylus, 44.
variolosus, Cnemidophorus, 190.
varius, Sauromalus, 79, 81, 227.
velox, Cnemidophorus gularis, 184.
ventralis, Callisaurus, 88.
Callisaurus draconoides, 86, 88, 227.
Callisaurus ventralis, 88.
Homalosaurus, 89, 88.
Scincus, 203, 214.
venusta, Emys, 31.
veraepacis, Anolis ustus, 67.
Verticaria, 174.
beldingi, 186.
caerulea, 186.
ceralbensis, 188.
estephrinis, 187.
franciscensis, 187.
hyperythra beldingi, 186.
hyperythra hyperythra, 187.
hyperythra schmidti, 187.
picta, 186.
serieca, 186.
Verus, Cnemidophorus gularis gularis, 182.
vigilis, Xantusia, 154, 217.
virgata, Chelonia, 17.
virgatus, Sceloporus undulatus, 117, 118, 219, 227.
viridiflava, Barisia, 198, 200, 224.
viridiflavus, Gerrhonotus, 200.
viridis, Anolis, 55.
Testudo, 17.
vittatus, Basiliscus, 71, 218, 220, 221, 222, 223, 224, 225, 227, 228, 229.
webbii, Elgaria multicaudata, 205, 207, 217.
Gerrhonotus, 207.
Gerrhonotus scincicauda, 207.
westphalii, Sceloporus, 127.
wiegmanni, Anolis, 63.
Phrynosoma, 97.
wislizenii, Crotaphytus, 94.
Gambelia, 93, 94.
Gambella wislizenii, 94, 217, 219, 227.
xanti, Phylodactylus, 49.
Xantusia, 151, 154.
gilberti, 154, 155, 217.
henshawi, 154, 155, 217.
picta, 155.
vigilis, 154, 217.
Xantusiidae, 39, 151.
Xenosauridae, 40, 207.
Xenosaurus, 207.
Fasciatus, 207, 208.
grandis, 207, 208, 225, 229.
newmanorum, 207, 208, 226.
rackhami, 208, 218.
Xerobates agassizii, 28.
berlandieri, 28.
yucatana, Cistudo, 35.
Terrapene, 35.
Terrapene mexicana, 34, 35, 225.
yucatanticus, Norops, 60.
Zablepsis, 154.
zosteromus, Sceloporus, 115.
Sceloporus magister, 112, 115, 217.