

1   **Methane Production and Emissions in Trees and Forests**

2

3   Kristofer R. Covey\*<sup>1,2</sup>

4   <sup>1</sup>Environmental Studies and Sciences Program

5   Skidmore College

6   815 N Broadway

7   Saratoga Springs NY, 128660

8   [kcovey@skidmore.edu](mailto:kcovey@skidmore.edu)

9   ORCID ID#: 0000-0002-3282-9564

10

11   <sup>2</sup>Yale University, School of Forestry and Environmental Studies

12   195 Prospect St.

13   New Haven, CT 06511 USA

14   [kristofer.covey@yale.edu](mailto:kristofer.covey@yale.edu)

15   Phone: 518.321.3706

16

17   J. Patrick Megonigal\*<sup>3</sup>

18   <sup>3</sup>Smithsonian Environmental Research Center

19   647 Contees Wharf Road

20   Edgewater, MD 21037

21   [megonigalp@si.edu](mailto:megonigalp@si.edu)

22   V: 443-482-2346

23   ORCID ID#: 0000-0002-2018-7883

24   \*Authors for correspondence

25

26

27

28

29

30

31

32

33

34

35

36 **Abstract**

37

38 Forest ecosystem CH<sub>4</sub> research has focused on soils, but trees are also important sources  
39 and sinks in forest CH<sub>4</sub> budgets. Living and dead trees transport and emit CH<sub>4</sub> produced  
40 in soils; living trees and deadwood emit CH<sub>4</sub> produced inside trees by microorganisms;  
41 and trees produce CH<sub>4</sub> through an abiotic photochemical process. We review the state of  
42 the science on the production, consumption, transport, and emission of CH<sub>4</sub> by living and  
43 dead trees, and the spatial and temporal dynamics of these processes across hydrologic  
44 gradients inclusive of wetland and upland ecosystems. Emerging research demonstrates  
45 that tree CH<sub>4</sub> emissions can significantly increase the source strength of wetland forests,  
46 and modestly decrease the sink strength of upland forests. Scaling from stem or leaf  
47 measurements to trees or forests is limited by knowledge of the mechanisms by which  
48 trees transport soil-produced CH<sub>4</sub>, microbial processes produce and oxidize CH<sub>4</sub> inside  
49 trees, a lack of mechanistic models, the diffuse nature of forest CH<sub>4</sub> fluxes, complex  
50 overlap between sources and sinks, and extreme variation across individuals.  
51 Understanding the complex processes that regulate CH<sub>4</sub> source-sink dynamics in trees  
52 and forests requires cross-disciplinary research and new conceptual models that transcend  
53 the traditional binary classification of wetland versus upland forest.

54

55 **Keywords**

56 tree, forest, methane, tree microorganism, anaerobic metabolism, methane oxidation,  
57 climate, greenhouse gases

58

## I. Introduction

Forests are a dominant feature of the global carbon cycle and play an important role in regulating climate and climate change (Bonan, 2008; Pan *et al.*, 2011). Research on forests in the context of the global carbon cycle is focused primarily on carbon dioxide (CO<sub>2</sub>) dynamics because the fluxes are large, and carbon sequestration in wood and soil organic matter influence century-scale projections of radiative forcing (Canadell & Raupach, 2008). Less attention is directed to forests as sources and sinks of other carbon trace gases such as methane (CH<sub>4</sub>). Soils are fairly well characterized in forest CH<sub>4</sub> budgets, but trees were only recently recognized as sources or sinks of this important greenhouse gas (Carmichael *et al.*, 2014; Saunois *et al.*, 2016). We review evidence that CH<sub>4</sub> dynamics in forests are far more complex than previously believed due to a combination of plant, microbial, and abiotic processes mediated by living and dead trees.

Methane causes 32-45 times more radiative forcing in a century than CO<sub>2</sub> on a mass basis (Neubauer & Megonigal, 2015) and contributes ~20% of radiative forcing (Denman, 2007; Myhre *et al.*, 2013; Neubauer & Megonigal, 2015). Because CH<sub>4</sub> is more responsive than CO<sub>2</sub> to changes in sources or sinks (Hansen *et al.*, 2000), forest CH<sub>4</sub> budgets are a meaningful aspect of management directed at slowing the pace of global climate change (UNFCCC, 2016). A more nuanced understanding of forests is needed across fundamental forest-climate interactions to improve Earth system models and manage forests for climate mitigation (Canadell & Raupach, 2008). It is increasingly clear that forest CH<sub>4</sub> cycling is one such interaction.

Despite efforts to constrain and refine the strength of the many sources and few sinks of atmospheric CH<sub>4</sub>, the global CH<sub>4</sub> budget remains highly uncertain (Saunois *et al.*, 2016). The total size of the global CH<sub>4</sub> pool is well-constrained in the range of 539-609 Tg CH<sub>4</sub> yr<sup>-1</sup>, but mismatches between bottom-up models and top-down estimates leave considerable uncertainty about individual components (Dlugokencky *et al.*, 2011; Allen, 2016; Saunois *et al.*, 2017).

Wetland ecosystems are the largest natural source of CH<sub>4</sub> globally and forested wetlands are ~60% of total global wetland area (Matthews & Fung, 1987; Prigent *et al.*, 2007), suggesting that forested wetlands are a significant global source of CH<sub>4</sub>. Reports of a discrepancy between emissions-based estimates and satellite-based estimates of CH<sub>4</sub> sources in tropical forests (Frankenberg *et al.*, 2008) sparked new interest in tree surfaces as an overlooked source (Terazawa *et al.*, 2007; Gauci *et al.*, 2010). Most of the research effort on wetland CH<sub>4</sub> cycling has been in herbaceous wetland systems, but emerging literature on soil- and plant-mediated CH<sub>4</sub> emissions in wetland forests indicates that this source alone may account for 5-10% of global CH<sub>4</sub> emissions (Pangala *et al.*, 2017).

Upland ecosystems on freely drained soils are recognized as CH<sub>4</sub> sinks in global budgets, and have been the focus of studies on CH<sub>4</sub> consumption by soils (Le Mer & Roger, 2001; Saunois *et al.*, 2016). Transient periods of CH<sub>4</sub> emission have been reported in nominally upland forests, but such emissions are cryptic and easily overlooked (Megonigal & Guenther, 2008). It is now clear that all biological surfaces in upland and wetland forests have the potential to emit or consume CH<sub>4</sub> (Carmichael *et al.*, 2014).

The emphasis on wetland forests as *net* atmospheric CH<sub>4</sub> sources and upland forests as *net* sinks masks the complex interplay of aerobic and anaerobic processes that occur to varying degrees in all forest ecosystems (Fig. 1). The outcome of this dynamic

can change the radiative balance of forests over temporal scales of minutes to decades and spatial scales of microsites to biomes. It is perhaps because of the focus on forests as either net sources or net sinks that research on the interrelated processes of CH<sub>4</sub> production and oxidation has centered exclusively on just one process or the other. This perspective fundamentally limits our ability to fully represent the dynamic nature of forests in budgets and Earth system models. The goal of this review is to emphasize the common processes that exist across all forested ecosystems in order to advance a holistic understanding of carbon cycling and the radiative balance of forest ecosystems.

## II. Tree CH<sub>4</sub> Fluxes

Global budgets, Earth system models, and carbon accounting policies generally assume that the contribution of CH<sub>4</sub> in upland forests can be measured as the rate of exchange at the soil surface (Saunois *et al.*, 2016). The focus on soil fluxes reflects the difficulty of enclosing whole trees in gas flux chambers, the most common method for quantifying trace gas fluxes. Improved instrumentation and growing interest in the role of forests in global CH<sub>4</sub> dynamics is providing new insights on variation in tree CH<sub>4</sub> fluxes across tree species, tissue types within living trees, and stages of dead tree decay. This review draws from 84 studies on CH<sub>4</sub> dynamics in living trees and deadwood (Table S1).

### 1. Fluxes modeled from stem CH<sub>4</sub> concentration in upland forests

Gas concentrations inside tree stems are useful for judging the potential of trees to act as net sources or sinks of a gas, and to efficiently assess sources of variation before investing in flux measurements. Such measurements are common in upland forests on freely drained soils where CH<sub>4</sub> fluxes are low and variation in time, space, species and environmental gradients is large. There are no published reports of *in situ* wood CH<sub>4</sub> concentrations from wetland forests to our knowledge. CH<sub>4</sub> can accumulate to very high concentrations in upland trees (Mukhin & Voronin, 2007; Covey *et al.*, 2012; Hietala *et al.*, 2015), in some cases reaching >65% of total stem gas (Bushong, 1907). These high concentrations explain dramatic images of flaming trees (Fig. 2), and suggest that CH<sub>4</sub> emission rates from tree surfaces are restricted by slow diffusion through trunk wood (Sorz & Hietz, 2006; Wang *et al.*, 2017). Super-ambient CH<sub>4</sub> concentrations in trees were first reported 120 years ago in cottonwood (Bushong, 1907). Subsequent studies confirmed this observation (Zeikus & Ward, 1974; Wang *et al.*, 2016), and extended it to many other tree species (Covey *et al.*, 2012; Wang *et al.*, 2017).

Methane emissions from upland trees have been modeled from concentration data using a modified version of Fick's Law (Covey *et al.*, 2012), but concentration data alone cannot be interpreted as conclusive evidence that CH<sub>4</sub> is emitted from trees at meaningful rates. The only study to compare measured and modelled stem CH<sub>4</sub> fluxes reported that they were poorly correlated at diurnal scales, but better correlated at seasonal scales when sample sizes are large (Wang *et al.*, 2017). The processes that regulate gas dynamics in tree stems are poorly understood compared to soils, but were reviewed in detail for CO<sub>2</sub> by Teskey *et al.* (2008). Stem CO<sub>2</sub> efflux rates differ from modeled rates due to factors such as the temperature dependence of stem respiration, translocation of dissolved CO<sub>2</sub> by the transpiration stream, and CO<sub>2</sub> consumption by cortical photosynthesis (Teskey & McGuire, 2007). CH<sub>4</sub> shares each of these characteristics with CO<sub>2</sub>, including the

149 existence of both sources and sinks, and transport in the transpiration stream.

150 Advances in modeling tree CH<sub>4</sub> fluxes from concentration data, and in scaling  
151 flux data to whole, mature trees requires detailed process studies that link sites of  
152 production and oxidation to pathways of transport in lateral and longitudinal dimensions.  
153 Direct measurement of CH<sub>4</sub> fluxes are required to develop and validate models, and high  
154 frequency measurements may be a particularly insightful analytical tool in such efforts.  
155 For example, flux measurements on a *Liriodendron tulipifera* stem at 45 minute intervals  
156 over three days showed a diurnal cycle that peaked in late afternoon, at about the time of  
157 minimum tree diameter and 4 hours after peak sap flux (Fig. 3; Pitz and Megonigal  
158 2017). The timing of the CH<sub>4</sub> emissions peak suggests it is related to physical factors  
159 such as stem water content that control gas diffusion rates (Wang *et al.*, 2017). A second  
160 species (*Fagus grandifolia*) behaved differently, and it is certain that more extensive data  
161 sets of this type will show even more complex patterns. High frequency records over  
162 seasons and weather events, combined with knowledge of gas and heat transfer kinetics  
163 in trees will enable inferences about the processes controlling CH<sub>4</sub> production, transport,  
164 and emissions. A strategy that combines near-continuous measurements to elucidate fine-  
165 scale processes, easily deployed manual stem chambers (Siegenthaler *et al.*, 2016) for  
166 high replication, and improved stem flux models is likely to be the best scaling approach  
167 for forests.

## 168 169 2. Methane fluxes from direct measurement

170 Direct measurements show that all trees – living or dead – have the potential to be CH<sub>4</sub>  
171 sources, CH<sub>4</sub> sinks or both. Most *in situ* tree flux measurements are made on trunks, and  
172 show either net positive or null emissions, with net consumption a less common result  
173 (Table 1). Variation in CH<sub>4</sub> fluxes from tree surfaces arises from species, ages, tissue  
174 types, site characteristics, and environmental conditions. When averaged over many  
175 individuals or time points at a given site, variability ranges from emissions of nearly  
176 17,000  $\mu\text{mol m}^{-2} \text{ h}^{-1}$  to consumption of 0.7  $\mu\text{mol m}^{-2} \text{ h}^{-1}$  (Table 1). Methane emissions  
177 are generally higher from wetland than upland trees, presumably reflecting a far larger  
178 contribution from soil-derived CH<sub>4</sub> in wetter forests. Within upland or wetland forests,  
179 emissions from living trees tend to be higher than dead trees, and emissions from fresh  
180 deadwood are higher than from decayed debris (Table 1). This pattern suggests that the  
181 endogenic CH<sub>4</sub> emitted by trees is produced from a non-structural photosynthate source  
182 that declines after tree death.

183 The lowest rates of site-wide tree emissions are from a three-month study of the  
184 conifer *P. sylvestris* in an upland forest, with median trunk CH<sub>4</sub> emission of 0.01 to 0.001  
185  $\mu\text{mol m}^{-2} \text{ stem h}^{-1}$  (Machacova *et al.*, 2016). Emissions were lower in a relatively dry plot  
186 than a wet plot. Low emission rates are consistent with reports of low CH<sub>4</sub> concentrations  
187 inside the stems of gymnosperms, but this is the only published study of a gymnosperm  
188 and the only boreal site studied. Average rates are 1-2 orders of magnitude higher in other  
189 upland forests, all of which are dominated by angiosperm species (Table 1). The highest  
190 upland rates reported were made in a *Populus davidiana* forest, and were comparable to  
191 rates for upland forests modeled from internal CH<sub>4</sub> concentrations. Trees in wetland and  
192 floodplain forests tend to emit CH<sub>4</sub> at rates that are higher than upland forests, but of the  
193 same order of magnitude. A dramatic exception to this generalization is in the Amazon

basin where average stem emissions are 1-2 order of magnitude higher than any other system studied to date (Pangala *et al.*, 2017).

Tree diameter is a measure of size that is often interpreted as a surrogate for age. Emissions from small trees are often different than from large trees, but the direction of the difference varies by ecosystem type. In wetland forests small trees often have higher CH<sub>4</sub> emissions than mature trees (Pangala *et al.*, 2015, 2017), while the opposite is often the case in upland forests (Wang *et al.*, 2017; Pitz *et al.* 2018).

Patterns of CH<sub>4</sub> flux vary by tissue type and position in the tree. Emissions tend to decrease in order main stem > shoots (branches) > leaves (Table 1), a pattern that may be caused by a number of factors such as tissue volume, tissue type (sapwood or hardwood, Wang *et al.*, 2016), or distance from the source. Direct measurements on leaves showed no net flux in mature tropical forest trees *in situ* (Pangala *et al.*, 2017), while the leaves of wetland saplings and seedlings ranged from no net flux to net emissions in mesocosm and microcosm studies (Pangala *et al.*, 2017; Garnet *et al.*, 2004). Tree pneumatophores emit CH<sub>4</sub> in wetland forests (Pulliam 1992; Pangala *et al.*, 2013; Purvaja *et al.*, 2004).

Net CH<sub>4</sub> consumption by leaves or stems occurs in upland and wetland forests. Most measurements on upland trees show stems to be a net source, but there have been reports of net consumption (Machacova *et al.*, 2016; Pitz & Megonigal, 2017; Warner *et al.*, 2017). Wetland tree stem studies report fewer observations of net CH<sub>4</sub> consumption than upland studies, but instances of consumption were observed in a temperate wetland forest (Pitz *et al.*, 2018) and a wet boreal forest (Machacova *et al.*, 2016). Direct measurements on the leaves and stems of several species in an upland forest canopy demonstrated CH<sub>4</sub> consumption *in situ* (Sundqvist *et al.*, 2012), with rates positively related to gross primary production in some cases (Fig. 4). Subsequent laboratory studies found that rates of leaf CH<sub>4</sub> uptake increased with photosynthetically active radiation and stomatal conductance, suggesting that the site of CH<sub>4</sub> consumption was inside the leaf.

### III. Tree Emissions of Soil-Produced CH<sub>4</sub>

A major challenge to explaining spatial and temporal variation in tree CH<sub>4</sub> fluxes is to distinguish between soils versus trees as sites of methanogenesis. The distinction is fundamental for scaling CH<sub>4</sub> emission rates to site-, regional-, and global-scale budgets and models, and it applies equally to upland and wetland forests. It is well established that saturated soils support microbial CH<sub>4</sub> production in wetlands, and that herbaceous plants transport and emit soil-produced CH<sub>4</sub> (Laanbroek, 2010). High CH<sub>4</sub> emission rates from wetland trees is evidence that mature trees also transport soil-produced CH<sub>4</sub> (Table 1). Less well established is the observation that CH<sub>4</sub> is produced in freely drained upland soils in anaerobic microsites (Von Fischer & Hedin, 2002; Brewer *et al.*, 2018). Mature upland trees may transport CH<sub>4</sub> produced in soil microsites or groundwater (Megonigal & Guenther, 2008), but this has not been demonstrated conclusively *in situ*. In principle all trees are capable of transporting and emitting soil-produced CH<sub>4</sub> by diffusion or xylem transport. Aside from transporting soil-produced CH<sub>4</sub>, trees also regulate soil CH<sub>4</sub> fluxes through plant-soil-microbe interactions that control rates of soil CH<sub>4</sub> production and oxidation.

#### 1. Tree support of soil methanogenesis and methanotrophy

Plants regulate the production, oxidation and export of soil-produced CH<sub>4</sub> by acting as electron donors and acceptors that support microbial respiration (Megonigal *et al.*, 2004). Evidence of such regulation by trees is limited, but tree root exudates regulate decomposition in upland soils (Phillips *et al.*, 2011) and are expected to be an important organic carbon source to anaerobic microbial communities in forest soils. Tight coupling between tree carbon metabolism and anaerobic microbial metabolism was demonstrated in a study of *Taxodium distichum* seedlings in which net CH<sub>4</sub> emissions were strongly ( $r^2 \geq 0.87$ ) related to whole-plant photosynthesis (Vann & Megonigal, 2003). Elevated CO<sub>2</sub> increased CH<sub>4</sub> emissions by >60% in the study, suggesting that understanding tree sources of labile carbon to forested wetland microbial communities is an important step in modeling wetland tree CH<sub>4</sub> emissions.

Aerobic methanotrophic bacteria consume CH<sub>4</sub> in the presence of O<sub>2</sub> (Fritz *et al.*, 2011). CH<sub>4</sub> oxidation in wetland soils occurs at the soil surface above the water table, and around roots where plant-transported O<sub>2</sub> diffuses into anaerobic soil (Denier van der Gon & Neue, 1996). In one forested wetland, methanotrophy reduced CH<sub>4</sub> emissions by up to 80% (Megonigal & Schlesinger, 2002). The study did not distinguish between oxidation at the aerobic soil surface versus the rhizosphere, but it is likely that plant-mediated CH<sub>4</sub> oxidation was important because the soils were consistently anaerobic below 6 cm depth. Root O<sub>2</sub> release by wetland trees into anaerobic soils can also indirectly inhibit CH<sub>4</sub> emissions by generating Fe(III) oxides, which then act as competing terminal electron acceptors that suppress methanogenesis (Weiss *et al.*, 2005). Anaerobic CH<sub>4</sub> oxidation occurs in tropical and boreal forest soils (Blazewicz *et al.*, 2012), but nothing is known about the role of plants, if any, in regulating the process. Upland soils tend to support higher rates of atmospheric CH<sub>4</sub> consumption than other terrestrial ecosystems, a pattern that has been linked to the influence of trees on methanotrophy and soil gas diffusivity (Dalal *et al.*, 2007).

## 2. Gas transport through trees

Tree stems can be the dominant pathway for CH<sub>4</sub> egress from forested wetlands, emitting soil-produced CH<sub>4</sub> at higher rates than other ecosystem surfaces (Pangala *et al.*, 2017). A large portion of the volume of a tree stem is gas, estimated at about 25% of the heartwood in angiosperms and 50% in gymnosperms (MacDougal, 1927; Gartner *et al.*, 2004). Connections among gas-filled spaces is one mechanism by which gases such as O<sub>2</sub>, CO<sub>2</sub> and CH<sub>4</sub> pass through trees (MacDougal, 1932). The most well studied tree-mediated pathway for transport of soil-produced CH<sub>4</sub> is aerenchyma, a specialized tissue characterized by enlarged gas spaces that forms in roots and stems following exposure to hypoxic soil conditions (Topa & McLeod, 1986; Drew *et al.*, 2000; Evans, 2004). Aerenchyma tissue allows rapid gas transport between soils and the atmosphere, and it is a ubiquitous adaptation in wetland plants for supplying O<sub>2</sub> to aerobically respiring roots (Jackson & Armstrong, 1999). Wetland trees develop aerenchyma tissue in response to anoxic soil conditions (Topa & McLeod, 1986; Megonigal & Day, 1992), and they show evidence of O<sub>2</sub> transport in the form of oxidized rhizospheres (Huikari, 1954; Armstrong, 1967; Hook *et al.*, 1972; Schröder, 1989). Although trees do not develop aerenchyma tissue under freely drained, upland conditions, they nonetheless transport gases through

284 connected gas-filled porespaces (Armstrong, 1980).

285 Gas flux through trees can proceed by passive or active mechanisms, a potential  
286 source of variation in gas flux rates across species and time. Molecular diffusion occurs  
287 in all trees to some extent, and is a slow, passive process that accounts for ongoing plant-  
288 mediated gas exchange between the soil and atmosphere even when transpiration is near-  
289 zero (Nietch *et al.*, 1999). Rusch & Rennenberg (1998) demonstrated that CH<sub>4</sub> moves by  
290 diffusion alone through stems of the wetland-adapted species *Alnus glutinosa*. Gas can  
291 also be transported in trees by pressurized ventilation, a rapid process that creates mass  
292 flow of O<sub>2</sub> between the atmosphere and soils. Pressurized ventilation of O<sub>2</sub> is driven by  
293 temperature gradients that develop between sunlit tree stems and ambient air (Große &  
294 Schröder, 1984), and has also been shown to occur in *A. glutinosa* (Schröder, 1989). It is  
295 not clear whether pathways of O<sub>2</sub> and CH<sub>4</sub> transport are coupled or independent, but the  
296 absence of diurnal variation in CH<sub>4</sub> emissions in *A. glutinosa* saplings grown under full  
297 sunlight suggests that the dominant pathway for CH<sub>4</sub> *in situ* is diffusive transport despite  
298 the potential for pressurized transport in this species (Pangala *et al.*, 2014). Flux studies  
299 have also detected significant diurnal variation in both upland (Pitz & Megonigal, 2017)  
300 and wetland (Pangala *et al.*, 2015) tree species, suggesting the possibility that hotspots of  
301 tree gas emissions via pressurized ventilation or transpiration-driven mass flow can be  
302 predicted in part from the tree species composition of a forest.

303 Transpiration can also support mass flow of gases between soils or roots and the  
304 atmosphere, and has been shown to be a mechanism for transporting CH<sub>4</sub> dissolved in  
305 soil solutions to the atmosphere through the seedlings of the wetland tree species *Alnus*  
306 *glutinosa* (Rusch & Rennenberg, 1998), *Fraxinus latifolia*, *Populus trichocarpa*, *Salix*  
307 *fluviatilis* (Rice *et al.*, 2010), and *Taxodium distichum* (Garnet *et al.*, 2005). Similar  
308 observations have been made for xylem transport of soil-produced CO<sub>2</sub> (Bloemen *et al.*,  
309 2014). Garnet *et al.* (2005) demonstrated that transpiration-driven CH<sub>4</sub> emissions varies  
310 with CO<sub>2</sub> concentration, humidity, and other variables that affect stomatal conductance.  
311 Diffusive transport of soil-produced CH<sub>4</sub> can continue after tree death, though net CH<sub>4</sub>  
312 consumption on standing dead trees has also been observed (Carmichael *et al.*, 2018).

313 Radial diffusion transports gases from the inside the tree to the atmosphere (CH<sub>4</sub>)  
314 or vice versa (O<sub>2</sub>) and determines both the rate and location of gas exchange at the tree  
315 surface (Teskey *et al.*, 2008). High internal CH<sub>4</sub> concentrations in trees reflect barriers to  
316 transport and diffusion of gases related to wood anatomy or water-filled wood porespace.  
317 Lenticels are specialized tissues that facilitate radial gas exchange across plant surfaces,  
318 and are a particularly important adaptation to flooded environments in trees. Pangala *et*  
319 *al.* (2014) found that lenticel density and porewater CH<sub>4</sub> concentration explained 84% of  
320 the variation in stem CH<sub>4</sub> emissions in flooded *Alnus glutinosa* saplings (Fig. 5).

321 High frequency diurnal measurements of CH<sub>4</sub> emissions from mature trees can be  
322 used to assess the relative contributions of diffusion and mass flow to transporting and  
323 emitting tree gases *in situ*. However, it is likely that the pathways interact such that gases  
324 produced in soils or inside trees move by multiple pathways before being emitted from a  
325 tree surface. Macropores in the form of cracks, holes, or wood rot can be preferential  
326 transport pathways, complicating direct gas flux measurements (Teskey *et al.*, 2008).  
327 Unraveling the complex processes that govern CH<sub>4</sub> production, transport, consumption,  
328 and emissions requires detailed mechanistic studies coupled with modeling aimed at



scaling processes to whole-tree scales.

#### IV. Tree-Produced CH<sub>4</sub>

##### 1. Abiotic aerobic methanogenesis in trees

The discovery of a novel aerobic, abiotic pathway of CH<sub>4</sub> production from plant tissue by (Keppler *et al.*, 2006) sparked a new wave of research on CH<sub>4</sub> emissions from plants, and inspired the first sustained investigations of CH<sub>4</sub> emissions from upland trees and forests. Keppler *et al.* (2006) estimated aerobic emissions of 236 Tg CH<sub>4</sub> yr<sup>-1</sup> globally, a flux large enough to explain higher-than-expected atmospheric CH<sub>4</sub> pools over tropical forests (Frankenberg *et al.*, 2005). The study received significant criticism based on three points: (i) a mechanism was not provided, (ii) purported experimental design flaws, and (iii) scaling metrics that significantly overestimated the global source, with significant implications for managing forests for climate protection (Lowe, 2006; Schiermeier, 2006; Evans, 2007). It is now clear that abiotic CH<sub>4</sub> production from plant tissue is real, with several independent groups reporting rates similar to those of Keppler *et al.* (2006) (Bruhn *et al.*, 2012; Liu *et al.*, 2015) (Table S1). However, rigorous scaling exercises have also confirmed that the global impact of aerobic plant CH<sub>4</sub> emissions is far less than initially estimated (Bloom *et al.*, 2010; Fraser *et al.*, 2015).

The precise chemical reaction underlying abiotic aerobic methanogenesis is not clear, but evidence suggests that reactive oxygen species (ROS) commonly produced in response to plant stress are a proximal driver of abiotic emissions. Agents that incite ROS production are associated with abiotic CH<sub>4</sub> production, and those that remove ROS from plant tissues limit production (Messenger *et al.*, 2009). Furthermore, the presence of enzymes that inhibit ROS removal are directly involved in stimulating production (Bruhn *et al.*, 2012; Liu *et al.*, 2015). Reactive oxygen species may initiate non-enzymatic photochemical reactions that foster the breakdown of pectin (Keppler *et al.*, 2006; Bruhn *et al.*, 2009; Messenger *et al.*, 2009), but other structural and non-structural plant compounds such as waxes, lignin, cellulose, MET protein, and ascorbic acid are also potential precursors (Vigano *et al.*, 2008; Keppler *et al.*, 2009; Vigano *et al.*, 2009; Althoff *et al.*, 2010).

Abiotic CH<sub>4</sub> emissions are triggered by a number of physical stressors, with UVB radiation as the most commonly documented inciting agent. UVB triggers abiotic CH<sub>4</sub> production from detached plant parts (Fig. 6) (McLeod *et al.*, 2008; Vigano *et al.*, 2008; Bruhn *et al.*, 2009), from structural components such as pectin (Keppler *et al.*, 2008; Megonigal & Guenther, 2008; Messenger *et al.*, 2009), and whole plants (Qaderi & Reid, 2009). By contrast, exposure to visible light alone does not incite abiotic methanogenesis in plants (Bruhn *et al.*, 2009). In one case abiotic CH<sub>4</sub> emissions were triggered by the lack of light (Martel & Qaderi, 2017). Wang *et al.* (2009) noted that emissions increased with physical wounding of both cuttings and connected stems. High temperature (Keppler *et al.*, 2008; McLeod *et al.*, 2008), drought stress (Qaderi & Reid, 2011), and bacterial infection (Messenger *et al.*, 2009) can also incite abiotic methanogenesis in the presence of O<sub>2</sub>. The highest rates of aerobic CH<sub>4</sub> emissions in lab experiments occur when multiple stress factors interact, suggesting that multi-factor experiments may best reproduce *in situ* rates of abiotic CH<sub>4</sub> emissions (Liu *et al.*, 2015; Abdulmajeed *et al.*, 2017).

Evidence of CH<sub>4</sub> production through an abiotic pathway is increasingly robust in laboratory studies (see reviews by (Keppler *et al.*, 2009; Bruhn *et al.*, 2012; Liu *et al.*, 2015). However, *in situ* evidence of abiotic CH<sub>4</sub> production is weak because the process cannot be effectively isolated from the many potential microbial CH<sub>4</sub> sources (Sanhueza & Donoso, 2006; Cao *et al.*, 2008; Wang, S *et al.*, 2009; Bruhn *et al.*, 2012). One cannot assume that CH<sub>4</sub> emitted *in situ* from plants on freely drained soils has an abiotic source because plants can transport CH<sub>4</sub> from anaerobic microsites in both soils and plant stems. Also, evidence that methanogenic microorganisms can tolerate atmospheric levels of O<sub>2</sub> (Meronigal *et al.*, 2004) suggest that not all aerobic CH<sub>4</sub> production is abiotic. Microbial CH<sub>4</sub> sources may explain why CH<sub>4</sub> emission rates from *in situ* intact foliage are nearly twice those from detached leaves (Qaderi & Reid, 2009). Based on laboratory rates of UVB-irradiated plants and typical Earth surface UVB irradiances, abiotic CH<sub>4</sub> production produces 7 to 50 ng CH<sub>4</sub> g dw<sup>-1</sup> hr<sup>-1</sup> across a temperature range from 25 to 40°C (Vigano *et al.*, 2008).

## 2. Microbial methanogenesis and methanotrophy inside trees

Methanogenesis in living trees has been recognized for nearly five decades, but remains little studied despite the availability of molecular tools for probing anaerobic microbial communities. Zeikus and Ward (1974) observed flammable concentrations of CH<sub>4</sub> inside hardwood trees, and determined that it was produced *in situ* by methanogens. Subsequent authors confirmed an Archaeal CH<sub>4</sub> source (Van Der Kamp *et al.*, 1979; Schink & Ward, 1984; Xu & Leininger, 2001); Archaea have been isolated from trees (Zeikus & Henning, 1975); Archaeal OTUs such as *Methanobacterium* can be dominate (>40% of sequence abundance) in wood microbial communities (Yip *et al.*, 2018); and anaerobic incubations of tree wood cores demonstrate active methanogenesis (Covey *et al.*, 2012; Wang *et al.*, 2016). Archaea in tree stems are accompanied by a variety of decay fungi, non-decay fungi, and bacteria, in competitive, mutualistic, and synergistic relationships (Fig. 7.; Shortle *et al.*, 1978; Schink *et al.*, 1981). Tree methanogenesis is expected to be sensitive to the totality of these interactions that collectively regulate the concentrations of methanogenic substrates.

Degradation of complex biopolymers such as cellulose and pectin to produce CH<sub>4</sub> generally requires the collective activities of fungi, bacteria, and archaeal methanogens operating syntrophically (Wolin & Miller, 1987; Cicerone & Oremland, 1988; Beckmann *et al.*, 2011). The process begins with enzymatic hydrolysis of complex compounds, then fermentation to yield H<sub>2</sub> and low molecular-weight organic acids such as acetate, formate and citrate, all of which occur to varying degrees in tree heartwood (Warshaw *et al.*, 1985; Schmidt, 2006; Worm *et al.*, 2011). Methanogenesis is the terminal step in which the products of fermentation (low molecular weight organic acids and H<sub>2</sub>) are consumed, yielding inorganic gases (CO<sub>2</sub>, CH<sub>4</sub>). Methanogens tend to specialize in one of two respiration pathways, acetate fermentation (CH<sub>3</sub>COOH → CO<sub>2</sub> + CH<sub>4</sub>) or CO<sub>2</sub> reduction (4H<sub>2</sub> + CO<sub>2</sub> → CH<sub>4</sub>), both of which occur in the wood of living trees (Schink *et al.*, 1981; Schink & Ward, 1984). The two pathways yield distinct δ<sup>13</sup>C signatures that can be used to infer mechanisms. Wang *et al.* (2016) reported a δ<sup>13</sup>C-CH<sub>4</sub> of <-70‰ in living *Populus* trees, a highly depleted ratio suggesting that CH<sub>4</sub> production through CO<sub>2</sub> reduction. The δ<sup>13</sup>C of emitted from stems in the Amazon basin ranged from -76.3 to -59.1‰ (Pangala et

al., 2017), indicating possible species- or site-related differences in CH<sub>4</sub> production pathways, though CH<sub>4</sub> oxidation may have also affected the ratios. It is likely that dominance of one pathway over the other varies by internal carbon source. Based on soil studies, we expect that CO<sub>2</sub> reduction dominates when the carbon source is highly aromatic or complex, while acetate fermentation dominates when supported by less complex compounds such as carbohydrates (Conrad & Klose, 1999).

The carbon sources supporting methanogenesis in living trees have important implications for forest CH<sub>4</sub> emission potential. Methanogenesis driven by wood decay must end once the structural wood is consumed, but wood decay (e.g. heart rot) is not a pre-requisite for methanogenesis. Indeed, elevated CH<sub>4</sub> levels are commonly present in trees with no evidence of wood decay (Mukhin & Voronin, 2008; Mukhin & Voronin, 2011; Covey *et al.*, 2012), and such trees have been shown to emit CH<sub>4</sub> through the trunk at high rates (Wang *et al.*, 2016). This pattern along with evidence that CH<sub>4</sub> production from dead wood declines rapidly with decay, is circumstantial evidence that non-structural carbohydrates (NSC) – free sugars and starches stored in wood (Dietze *et al.*, 2014) – are a carbon source to methanogens active in living tree stems (Covey *et al.*, 2016). The few studies of NSC in trees show interspecific patterns that mirror those of CH<sub>4</sub> concentrations in living and dead trees (Covey *et al.*, 2012; Covey *et al.*, 2016; Oberle *et al.*, 2017), such as far higher NSC stem sapwood concentrations in angiosperms than gymnosperms (Hoch *et al.*, 2003), and increasing stem NSC storage in angiosperms with age (Würth *et al.*, 2005). NSCs are a large portion of the total carbon stored in living trees (Würth *et al.*, 2005), rapidly metabolized (Cowling & Merrill, 1966), and continually replenished from newly fixed photosynthates (Richardson *et al.*, 2013; Dietze *et al.*, 2014). Collectively, this suggests an untested mechanism by which living trees could continually produce CH<sub>4</sub> at high rates over their lifetime.

Methanotrophy is a ubiquitous CH<sub>4</sub>-consuming process that is certain to influence the direction and magnitude of CH<sub>4</sub> fluxes across tree surfaces, yet there is little evidence for the process in living trees despite the fact they contain both CH<sub>4</sub> and O<sub>2</sub> (Table 2). Potential methanotrophic species (OTUs) were rare in the heartwood and sapwood of *Populus deltoids* (Yip *et al.*, 2018), and CH<sub>4</sub> oxidation was not detected in incubations of wood from two other temperate forest tree species (Wang *et al.*, 2016). No clear evidence of the *pmoA* gene of methanotrophic bacteria was found in the roots and shoots of boreal forest shrubs (Halmeenmäki *et al.*, 2018). However, the *pmoA* gene is abundant in dead wood where methanotrophs appear to contribute to N<sub>2</sub>-fixation (Mäkipää *et al.*, 2018). Because CH<sub>4</sub> concentrations in living wetland and upland trees vary from ambient to super-ambient, it is expected that trees harbor both high- and low-affinity methanotrophic bacteria. Indeed, net CH<sub>4</sub> uptake from the atmosphere has been observed across living tree surfaces (Table 1 and references therein).

### 3. Regulation of microbial methanogenesis in trees

Archaeal methanogenesis in living trees is likely to be regulated by the same factors that operate in analogous environments such as soils. Molecular oxygen (O<sub>2</sub>) availability is generally the single most important regulator of CH<sub>4</sub> production rates because aerobic microbes outcompete archaeal methanogens for organic compounds, and O<sub>2</sub> is toxic to many, though not all, methanogens (Meganigal *et al.*, 2004). Tree stems

can be hypoxic or anoxic, with stem concentrations of 0.5-19% O<sub>2</sub> that decline from the bark to the heartwood (Table 2). Low O<sub>2</sub> concentrations develop because aerobic plant and microbial respiration consume O<sub>2</sub> faster than it is supplied by physical transport (Soraz & Hietz, 2006; Teskey *et al.*, 2008). Fermentative and methanogenic microbial communities develop under such conditions in systems ranging from wetlands to insects, but have not been a subject of detailed studies in trees.

Stem water content will prove to be a powerful explanatory variable for variation in CH<sub>4</sub> emissions from wetland and upland trees at diurnal, seasonal, and annual scales. Water regulates the O<sub>2</sub> supply by acting as a barrier to gas transport, reducing the O<sub>2</sub> diffusion rate by a factor of 10<sup>4</sup> compared to diffusion in air. Wang *et al.* (2017) observed that stem CH<sub>4</sub> emissions increase dramatically above about 50% stem water content, and continue to increase with water content above this threshold value. High water content also favors high wood CH<sub>4</sub> concentrations as a barrier to CH<sub>4</sub> diffusion out of the tree (Wang *et al.*, 2017), which sets the CH<sub>4</sub> diffusion gradient. If a larger sample of trees validates such relationships, models coupling soil and tree hydraulics should be able to capture temporal and spatial variation in tree CH<sub>4</sub> emissions.

Stem water content may help explain why high CH<sub>4</sub> concentrations and emissions in living upland trees are associated with wetwood, an anatomically distinct and sometimes saturated area of heartwood (Boyce, 1961; Xu & Leininger, 2001; Wang *et al.*, 2016; Wang *et al.*, 2017). Wetwood is also associated with bacterial and fungal infection (Jennings, 1996), and it is known that the wood immediately surrounding fungal colony centers can become highly depleted in O<sub>2</sub> (Schmidt, 2006).

## **V. Trees in Forest CH<sub>4</sub> Budgets**

### **1. Scaling Challenges**

It is no longer sufficient to equate soil fluxes to ecosystem fluxes in forested ecosystems, but quantifying CH<sub>4</sub> budgets is challenging because forests are a complex composite of environments and surfaces that produce, consume, transport, and emit CH<sub>4</sub> (Fig. 1). Eddy covariance flux techniques are promising in wetland forests, but in upland forests the near balance between diffuse sources and sinks is a challenge given the current detection limits of the technology (Saunois *et al.*, 2016).

Phylogeny is an important source of variation in stem CH<sub>4</sub> dynamics in living and dead trees, an observation that applies equally to wetland forests (Pangala *et al.* 2013) and upland forests (Pitz & Megonigal, 2017; Warner *et al.*, 2017). Wang *et al.* (2017) classified 22 upland forest species into three groups based on internal stem CH<sub>4</sub> concentration—consistently high, consistently low, and variable—suggesting an approach to simplify phylogenetic-based scaling through identification of functional groups.

The physiological and anatomical causes of phylogenetic-based variation in CH<sub>4</sub> emissions are not clear, but should differ depending on whether CH<sub>4</sub> sourced from the soil or the stem. For example, CH<sub>4</sub> production rates inside trees should scale positively with factors that regulate the stem's anaerobic volume, such as stem moisture content (Wang *et al.*, 2017). In upland forests, wetwood species are among the most consistent emitters because they maintain high moisture content under a wide range of soil moisture conditions (Wang *et al.*, 2017). Indeed, super-ambient CH<sub>4</sub> concentrations in trees were

509 first discovered in the wetwood genus *Populus* (Bushong, 1907). Anaerobic sites at the  
510 center of stems coincide with the distribution of heartwood, which may explain positive  
511 correlations between stem CH<sub>4</sub> emissions and the ratio of heartwood diameter or total  
512 diameter in upland forests (Wang *et al.*, 2017).

513 Negative relationships between CH<sub>4</sub> emissions and wood density are observed in  
514 both upland (Wang *et al.*, 2017) and wetland trees (Pangala *et al.*, 2013). Relationships  
515 with density and stem moisture content may be useful for scaling, but they are difficult to  
516 interpret mechanistically because of opposing effects on rates of CH<sub>4</sub> production and  
517 diffusion. High wood density and moisture content should enhance CH<sub>4</sub> production by  
518 slowing O<sub>2</sub> diffusion and increasing stem anoxia, but any such effects are masked in the  
519 flux data by the fact that these factors also slow CH<sub>4</sub> diffusion to the stem surface.

520 Methanogenesis in wetland forests occurs primarily in soils rather than inside tree  
521 stems (Pangala *et al.*, 2017), and should produce different scaling relationships compared  
522 to upland forests. CH<sub>4</sub> transport from the soil to the atmosphere through trees is governed  
523 by factors embedded in Fick's law, including the CH<sub>4</sub> concentration gradient, distance  
524 from the source to the atmosphere, and resistance to flux through the stem. Indeed, stem  
525 emissions in wetland forests are related positively to porewater CH<sub>4</sub> concentration  
526 (Pangala *et al.*, 2013, 2014, 2015; Terazawa *et al.*, 2015) and negatively to specific wood  
527 density (Pangala *et al.*, 2013), and emissions decrease rapidly with stem height above the  
528 water table (Pangala *et al.*, 2017). Stem emissions also relate to factors that control the  
529 rate of CH<sub>4</sub> production in soils such as temperature and water table depth (Pangala *et al.*,  
530 2015; Pitz *et al.*, 2018). Similar relationships in upland forests are either weak or absent  
531 (Warner *et al.*, 2017; Pitz *et al.*, 2018).

532 Stem diameter is expected to be a useful scaler for tree CH<sub>4</sub> emissions because it  
533 is a proxy for several factors that should influence CH<sub>4</sub> production or transport. However,  
534 relationships between stem diameter and stem emission vary among studies, with upland  
535 forests showing positive (Wang *et al.*, 2017) or null relationships (Warner *et al.*, 2017;  
536 Pitz *et al.*, 2018), and wetland forests showing negative (Pangala *et al.*, 2015) or null  
537 relationships (Pitz *et al.*, 2018). These contrasting results may reflect sampling limitations  
538 such as a small range of diameters, small sample sizes, or conflation with factors such as  
539 stem age, species, life history, or habitat type. Indeed, the most highly replicated tree CH<sub>4</sub>  
540 emissions study to date found consistently higher emissions from young trees than mature  
541 trees in wetland forests (Pangala *et al.*, 2017).

542 Spatial and temporal variation in stem CH<sub>4</sub> emissions are a significant challenge  
543 to bottom-up scaling. This is especially the case in upland forests where most tree species  
544 are capable of emitting CH<sub>4</sub> at least intermittently, but the distribution of emissions is  
545 highly skewed across individuals and time such that a few individuals or time points  
546 dominate annual emissions ((Maier *et al.*, 2017; Pitz & Megonigal, 2017; Wang *et al.*,  
547 2017; Warner *et al.*, 2017). The sampling challenge in upland forests is even greater to  
548 the extent that small stems (branches), leaves, and deadwood can each emit CH<sub>4</sub> (Covey  
549 *et al.*, 2016; Machacova *et al.*, 2016; Oberle *et al.*, 2017), consume CH<sub>4</sub> (Sundqvist *et al.*,  
550 2012), or have no net flux (Wang *et al.*, 2016; Warner *et al.*, 2017), all of which are  
551 observed. Collectively, these positive and negative fluxes determine the degree to which  
552 upland trees offset or enhance soil CH<sub>4</sub> fluxes, and whether the system is a net source or  
553 sink of atmospheric CH<sub>4</sub>. The same challenges of scaling across large stems, small stems

and leaves apply equally to wetland forests. Wetland forest tree emissions are sensitive to variation water table depth (Pangala *et al.*, 2015; Terazawa *et al.*, 2015; Pitz *et al.*, 2018) and presumably soil factors that regulate the production and oxidation of CH<sub>4</sub> such as Fe(III) and SO<sub>4</sub> content (Megonigal *et al.*, 2014).

## 2. Wetland and Upland Forest CH<sub>4</sub> Budgets

Regional and global estimates of tree contributions to CH<sub>4</sub> budgets to date have been highly speculative (Carmichael *et al.*, 2014; Saunois *et al.*, 2016) and are not a goal of this review. One exception is a comprehensive regional analysis of the contribution of trees to ecosystem CH<sub>4</sub> budgets in the 6.7 x 10<sup>6</sup> km<sup>2</sup> Amazon basin. (Pangala *et al.*, 2017) sampled 13 sites stratified by soil type and geomorphology, 2,357 individual tree stems across a wide range of species and size classes, and hundreds of flux observations using chambers placed on soil-, water-, and floating macrophyte surfaces. Stem surfaces emitted up to 17,000 μmol CH<sub>4</sub> m<sup>-2</sup> h<sup>-1</sup>, with mean rates an order of magnitude higher than any other system measured to date (Table 1). Stem emissions generally decreased with stem height, and emissions from leaves occurred only on younger trees and at far lower rates. Although stem surface rates from small stems were higher than those from large stems, when expressed on a soil area basis, large trees were a far larger CH<sub>4</sub> source at the ecosystem scale (Table 3). In total, 15-21 Tg CH<sub>4</sub> yr<sup>-1</sup> is transported through trees in the Amazon floodplain, which increases the previous bottom-up estimate by 50% and amounts to 3% of the global CH<sub>4</sub> budget (Pangala *et al.* (2017). Their bottom-up estimate for all surfaces (30 to 48 Tg CH<sub>4</sub> yr<sup>-1</sup>) compared well with their top-down estimate derived from a high-resolution air column budget (43±6 Tg CH<sub>4</sub> yr<sup>-1</sup>). Methane production potentials based on incubation of tree cores showed *in situ* production in relatively few stems, establishing soil transport as the primary CH<sub>4</sub> source to the atmosphere. This impressive campaign highlights the exigent need to account for multiple flux pathways in forests across large scales.

Trees are important CH<sub>4</sub> sources in forested wetlands outside the Amazon basin as well (Table 3), such as peatland forests of the Sebangau River catchment in Borneo where trees contribute up to 87% of CH<sub>4</sub> efflux (Pangala *et al.*, 2013). In a temperate forested wetland, emissions from *Alnus glutinosa* and *Betula pubescens* stems accounted for a maximum of 27% of ecosystem CH<sub>4</sub> emissions (Pangala *et al.*, 2015). Because both soils and trees are CH<sub>4</sub> sources, previous studies in forested wetlands appear to have underestimated the CH<sub>4</sub> budget of these ubiquitous ecosystems (Gauci *et al.* 2010, Pitz *et al.* 2018).

Trees and soils in upland forests on freely drained soils typically have opposing effects on forest CH<sub>4</sub> budgets, with soils acting primarily as net sinks and tree stems primarily as net sources than sinks. The consequences of upland trees as CH<sub>4</sub> sources are potentially important because upland soils are the single largest terrestrial CH<sub>4</sub> sink, with net consumption estimated to be 36 Tg CH<sub>4</sub> yr<sup>-1</sup> (Saunois *et al.*, 2016). Ecosystem scaling based on measurements from trunks, branches and shoots in a *Populus*-dominated upland temperate deciduous forest in China suggests that tree emissions may offset up to 63% of the soil CH<sub>4</sub> sink (Wang *et al.*, 2016) (Table 3). This estimate may be an upper boundary to the contribution of trees on freely drained soils with consistently deep (>1 m) water tables because wetwood is common in *Populus*, and stem CH<sub>4</sub> flux estimates from other

species are lower (Table 1). Large offsets of the upland soil CH<sub>4</sub> sink are also suggested in sites where the water table depth fluctuates into the upper meter of the soil profile, a condition that can simultaneously increase soil-derived CH<sub>4</sub> emissions and decrease the soil CH<sub>4</sub> uptake (Pitz & Megonigal, 2017). In a *Fraxinus*-dominated seasonal floodplain in Japan, stem surface fluxes reduced the estimated forest CH<sub>4</sub> sink by more than two thirds (Terazawa *et al.*, 2007), while the stems and shoots of *Pinus sylvestris* offset 35% of the soil sink on a wet upland site in Finland (Machacova *et al.*, 2016). In other studies tree CH<sub>4</sub> emissions have little impact on upland forest CH<sub>4</sub> budgets. Two independent estimates of the soil sink offset in North American temperate forests ranged from 1-6% (Pitz & Megonigal, 2017; Warner *et al.*, 2017), and the soil sink offset in a relatively dry upland *Pinus sylvestris* site was 0.8% (Machacova *et al.*, 2016). Finally, the net effect of trees can also be to increase the soil CH<sub>4</sub> sink. In the only field experiment involving tree stem fluxes published to date, a stand replanted in *Quercus petraea* saplings had two-fold higher CH<sub>4</sub> uptake than an unplanted site (Plain *et al.*, 2018). This result was caused by a combination of very low CH<sub>4</sub> emission rates by the nine year-old trees, and an increase in soil CH<sub>4</sub> uptake caused by unknown factors such as a transpiration-driven decrease in soil water potential or the presence of understory herbaceous plants.

A holistic understanding of the contribution of forests to the global CH<sub>4</sub> budget requires identifying common processes that exist across all forested ecosystems. Water table depth is a master variable that controls rates of CH<sub>4</sub> consumption by upland soils (Topp & Pattey, 1997), CH<sub>4</sub> emission by wetland soils (Turetsky *et al.*, 2014), and CH<sub>4</sub> emissions by both wetland and upland tree stems (Pangala *et al.*, 2015; Terazawa *et al.*, 2015; Pitz *et al.*, 2018). Water table depth was a likely source of spatial variation in an upland boreal forest that appeared to be a net CH<sub>4</sub> sink when tree and soil fluxes were measured by small-scale chambers, but a net source when measured by large-scale micrometeorological methods (Sundqvist *et al.*, 2015). In this case, small areas of wet soils in the tower footprint may have been strong CH<sub>4</sub> sources. In a floodplain forest, stem CH<sub>4</sub> emissions increased sharply during periods when the water table rose into the rooting zone (Terazawa *et al.*, 2015).

Small rates of tree CH<sub>4</sub> emissions may have a role in forests switching between net CH<sub>4</sub> sources and sinks (Shoemaker *et al.*, 2014). For example, a temperate upland forest changed briefly from a net CH<sub>4</sub> sink to a source during a warm, wet period when soil uptake decreased and tree emissions increased simultaneously (Pitz and Megonigal, 2017). This suggests that the global importance of tree emissions is related to the importance of nominally upland systems that periodically emit CH<sub>4</sub>. A rigorous effort to quantify CH<sub>4</sub> emissions from these ecosystems was provided by Spahni *et al.* (2011) who modeled “wet-mineral soil” moisture thresholds ranging from 0.28 to 0.55 fractional water-filled pore space, varying with edaphic factors such as texture. Modeled upland fluxes were about a factor of 10 smaller than wetland fluxes on an aerial basis, but the global area of soils with sufficient soil moisture to periodically emit CH<sub>4</sub> were extensive, yielding a large global emission of ~60 Tg CH<sub>4</sub> yr<sup>-1</sup>. This figure is double the global soil CH<sub>4</sub> sink, 23-36% of global emissions from wetlands, and 10% of all global CH<sub>4</sub> sources (Denman, 2007). Upland ecosystems are analogous to oceans in that a low rate of some biogeochemical process (e.g. NPP) can dramatically influence global cycles because of their large global footprint.

## VI. Conclusions

The growing body of literature on CH<sub>4</sub> dynamics in forest ecosystems shows that they are far more complex biogeochemical environments than previously believed, and that our previous focus on soil processes alone is insufficient for a rigorous understanding of forests greenhouse gas balance and radiative climate forcing. Progress toward this goal will be most effective if we recognize that all CH<sub>4</sub>-generating and consuming processes occur in all forest ecosystems regardless of their classification as upland or wetland. Advances in forest ecosystem CH<sub>4</sub> dynamics require a new focus on the complex interplay between productive and consumptive processes occurring from the top of the canopy to the subsurface ground water, and their implications for generalized scaling. The subject is ripe for collaborations between people with expertise in plant physiology, soil physics, hydrology, geomorphology, and microbial ecology, all of which interact to determine the distribution and activity of microbial communities and abiotic reactions that produce and consume CH<sub>4</sub> as a single coupled process (Megonigal *et al.*, 2004; Liu *et al.*, 2015). Of particular importance is collaborations between experts in biogeochemistry, wood anatomy, and tree physiology because they regulate CH<sub>4</sub> production and exchange across arboreal surfaces. Indeed, a growing research community with diverse interests in tree CH<sub>4</sub> dynamics has developed an agenda for advancing the field (Barba *et al.*, in press).

Further study is needed to refine ecosystem-scale estimates, determine the most appropriate scaling metrics, and resolve the distinctions between the arboreal CH<sub>4</sub> flux pathways. Whole-ecosystem studies currently provide the most robust information for global budgeting efforts, but many studies do not distinguish between the three pathways identified here in order to inform mechanistic numerical models. Laboratory studies can isolate specific pathways of CH<sub>4</sub> production or consumption, but they often fail to capture the substantial temporal and spatial scales of variation that drive *in situ* fluxes. In addition to flux measurements, there is a need for thoughtful integration of existing techniques across sub-disciplinary boundaries. Until additional integrative empirical studies are conducted, and process-based models are developed and tested, the contribution of forests to global CH<sub>4</sub> dynamics will remain poorly resolved.

## Acknowledgements

This contribution was supported by a grant from the US Department of Energy Terrestrial Ecosystem Science Program to JPM (DE-SC0008165) and by a National Science Foundation Award to KRC (NSF DGE-1405135). The authors also wish to acknowledge the Oak Spring Garden Foundation for hosting a symposium on forests and climate, and thoughtful comments on the manuscript from Mark A. Bradford and Paul Brewer. Monte Kawahara and Timothy Terway assisted with graphic design. The image of the flaming tree corer used in figure 2 was captured by Balazs Nyitrai and is used with his permission.

## Author Contributions

KRC and JPM contributed equally in all aspects of the manuscript.



## Citations

- Abdulmajeed AM, Derby SR, Strickland SK, Qaderi MM. 2017.** Interactive effects of temperature and UVB radiation on methane emissions from different organs of pea plants grown in hydroponic system. *Journal of Photochemistry and Photobiology B: Biology* **166**: 193-201.
- Allen G. 2016.** Biogeochemistry: Rebalancing the global methane budget. *Nature* **538**: 46-48.
- Althoff F, Jugold A, Keppler F. 2010.** Methane formation by oxidation of ascorbic acid using iron minerals and hydrogen peroxide. *Chemosphere* **80**: 286-292.
- Anderson B, Bartlett K, Frolking S, Hayhoe K, Jenkins J, Salas W 2010.** Methane and nitrous oxide emissions from natural sources. Office of Atmospheric Programs, US EPA, EPA 430-R-10-001, Washington DC.
- Armstrong W. 1967.** The oxidising activity of roots in waterlogged soils. *Physiologia Plantarum* **20**: 920-926.
- Armstrong W. 1980.** Aeration in Higher Plants. *Advances in Botanical Research* **7**: 225-332.
- Ayin CM, Schlub RL, Yasuhara-Bell J, Alvarez AM. 2015.** Identification and characterization of bacteria associated with decline of ironwood (*Casuarina equisetifolia*) in Guam. *Australasian Plant Pathology* **44**: 225-234.
- Barba J, Bradford MA, Brewer PE, Bruhn D, Covey K, van Haren J, Magonigal JP, Mikkelsen TN, Pangala S, Pihlatie M, Poulter B, Rivas-Ubach A, Schadt CW, Terazawa K, Warner DL, Zhang Z, Vargas R. in press.** Methane emissions from tree stems: a new frontier in the global carbon cycle. *New Phytologist*.
- Beckmann S, Kruger M, Englelen B, Gorbushina A, Cypionka H. 2011.** Role of bacteria, archaea and fungi involved in methane release in abandoned coal mines. *Geomicrobiology journal* **28**: 347-358.
- Blazewicz SJ, Petersen DG, Waldrop MP, Firestone MK. 2012.** Anaerobic oxidation of methane in tropical and boreal soils: Ecological significance in terrestrial methane cycling. *Journal of Geophysical Research: Biogeosciences* **117**. doi: 10.1029/2011JG001864.
- Bloemen J, Agneessens L, Meulebroek L, Aubrey DP, McGuire MA, Teskey RO, Steppe K. 2014.** Stem girdling affects the quantity of CO<sub>2</sub> transported in xylem as well as CO<sub>2</sub> efflux from soil. *New Phytologist* **201**: 897-907.
- Bloom A, Palmer P, Fraser A, Reay D. 2012.** Seasonal variability of tropical wetland CH<sub>4</sub> emissions: the role of the methanogen-available carbon pool. *Biogeosciences* **9**: 2821-2830.
- Bloom AA, Lee-Taylor J, Madronich S, Messenger DJ, Palmer PI, Reay DS, McLeod AR. 2010.** Global methane emission estimates from ultraviolet irradiation of terrestrial plant foliage. *New Phytologist* **187**: 417-425.
- Bonan GB. 2008.** Forests and climate change: Forcings, feedbacks, and the climate benefits of forests. *Science* **320**: 1444-1449.
- Boyce J. 1961.** *Forest Pathology*. New York, New York: McGraw-Hill.
- Brandt FB, Martinson GO, Conrad R. 2017.** Bromeliad tanks are unique habitats for microbial communities involved in methane turnover. *Plant and Soil* **410**: 167-179.
- Brewer PE, Calderón F, Vigil, M, von Fischer JC. 2018.** Impacts of moisture, soil respiration, and agricultural practices on methanogenesis in upland soils as measured with stable isotope pool dilution. *Soil Biology and Biochemistry* **127**: 239-251.

737 **Bruhn D, Mikkelsen TN, Øbro J, Willats WGT, Ambus P. 2009.** Effects of temperature,  
738 ultraviolet radiation and pectin methyl esterase on aerobic methane release from  
739 plant material. *Plant Biology* **11**: 43-48.

740 **Bruhn D, Møller IM, Mikkelsen TN, Ambus P. 2012.** Terrestrial plant methane production  
741 and emission. *Physiologia Plantarum* **144**: 201-209.

742 **Bushong FW. 1907.** Composition of gas from cottonwood trees. *Transactions of the Kansas*  
743 *Academy of Science* **21**: 53.

744 **Canadell JG, Raupach MR. 2008.** Managing forests for climate change mitigation. *Science*  
745 **320**: 1456-1457.

746 **Cao G, Xu X, Long R, Wang Q, Wang C, Du Y, Zhao X. 2008.** Methane emissions by alpine  
747 plant communities in the Qinghai-Tibet Plateau. *Biology Letters* **4**: 681-684.

748 **Carmichael MJ, Bernhardt ES, Bräuer SL, Smith WK. 2014.** The role of vegetation in  
749 methane flux to the atmosphere: should vegetation be included as a distinct  
750 category in the global methane budget? *Biogeochemistry* **119**: 1-24.

751 **Carmichael MJ, Helton AM, White JC, Smith WK. 2018.** Standing dead trees are a conduit  
752 for the atmospheric flux of CH<sub>4</sub> and CO<sub>2</sub> from wetlands. *Wetlands* **38**: 133-143.

753 **Chase WW. 1934.** The composition, quantity, and physiological significance of gases in tree  
754 stems. Minnesota Agricultural Experiment Station Technical Bulletin 99. St Paul, MN,  
755 USA: University of Minnesota. 5-51.

756 **Cicerone RJ, Oremland RS. 1988.** Biogeochemical aspects of atmospheric methane. *Global*  
757 *Biogeochemical Cycles* **2**: 299-327.

758 **Conrad R, Klose M. 1999.** Anaerobic conversion of carbon dioxide to methane, acetate and  
759 propionate on washed rice roots. *FEMS Microbiology Ecology* **30**: 147-155.

760 **Covey KR, de Mesquita CPB, Oberle B, Maynard DS, Bettigole C, Crowther TW, Duguid**  
761 **MC, Steven B, Zanne AE, Lapin M, et al. 2016.** Greenhouse trace gases in  
762 deadwood. *Biogeochemistry* **130**: 215-226.

763 **Covey KR, Wood SA, Warren RJ, Lee X, Bradford MA. 2012.** Elevated methane  
764 concentrations in trees of an upland forest. *Geophysical Research Letters* **39**: L15705.  
765 doi: 10.1029/2012GL052361.

766 **Cowling EB, Merrill W. 1966.** Nitrogen in wood and its role in wood deterioration.  
767 *Canadian Journal of Botany* **44**: 1539-1554.

768 **Crowther TW, Glick HB, Covey KR, Bettigole C, Maynard DS, Thomas SM, Smith JR,**  
769 **Hintler G, Duguid MC, Amatulli G, et al. 2015.** Mapping tree density at a global  
770 scale. *Nature* **525**: 201-205.

771 **Dalal RC, Allen DE, Livesley, SJ, Richards G. 2007.** Magnitude and biophysical regulators  
772 of methane emission and consumption in the Australian agricultural, forest, and  
773 submerged landscapes: a review. *Plant and Soil* **309**: 43-76. doi:10.1007/s11104-  
774 007-9446-7

775 **del Hierro AM, Kronberger W, Hietz P, Offenthaler I, Richter H. 2002.** A new method to  
776 determine the oxygen concentration inside the sapwood of trees. *Journal of*  
777 *Experimental Botany* **53**: 559-563.

778 **Denier van der Gon HAC, Neue H-U. 1996.** Oxidation of methane in the rhizosphere of rice  
779 plants. *Biology & Fertility of Soils* **22**: 359-366.

780 **Denman KL, G. Brasseur, A. Chidthaisong, P. Ciais, P.M. Cox, R.E. Dickinson, D.**  
781 **Hauglustaine, C. Heinze, E. Holland, D. Jacob, U. Lohmann, S. Ramachandran,**  
782 **P.L. da Silva Dias, S.C. Wofsy, and X. Zhang. 2007.** Couplings between changes in  
783 the climate system and biogeochemistry In S. Solomon DQ, M. Manning, Z. Chen, M.  
784 Marquis, K.B. Averyt, M. Tignor, and H.L. Miller *Climate Change 2007: The Physical*

785 *Basis. Contribution of Working Group I to the Fourth Assessment Report of the*  
786 *Intergovernmental Panel on Climate Change* Cambridge, UK, and New York, NY:  
787 Cambridge University Press. 499-587.

788 **Dietze MC, Sala A, Carbone MS, Czimczik CI, Mantooth JA, Richardson AD, Vargas R.**  
789 **2014.** Nonstructural Carbon in Woody Plants. *Annual Review of Plant Biology* **65**:  
790 667-687.

791 **Dlugokencky EJ, Nisbet EG, Fisher R, Lowry D. 2011.** Global atmospheric methane:  
792 budget, changes and dangers. *Philosophical Transactions of the Royal Society A:*  
793 *Mathematical, Physical and Engineering Sciences* **369**: 2058-2072.

794 **do Carmo JB, Keller M, Dias JD, Camargo PBd, Crill P. 2006.** A source of methane from  
795 upland forests in the Brazilian Amazon. *Geophysical research letters* **33**. doi:  
796 10.1029/2005GL025436.

797 **Drew MC, He C-J, Morgan PW. 2000.** Programmed cell death and aerenchyma formation in  
798 roots. *Trends in Plant Science* **5**: 123-127.

799 **Eklund L. 1990.** Endogenous levels of oxygen, carbon dioxide and ethylene in stems of  
800 Norway spruce trees during one growing season. *Trees - Structure and Function* **4**:  
801 150-154.

802 **Eklund L. 1993.** Seasonal variations of O<sub>2</sub>, CO<sub>2</sub>, and ethylene in oak and maple stems.  
803 *Canadian journal of forest research* **23**: 2608-2610.

804 **Eklund L. 2000.** Internal oxygen levels decrease during the growing season and with  
805 increasing stem height. *Trees - Structure and Function* **14**: 177-180.

806 **Evans DE. 2004.** Aerenchyma Formation. *New Phytologist* **161**: 35-49.

807 **Evans JR. 2007.** Resolving methane fluxes. *New Phytologist* **175**: 1-4.

808 **Ferretti DF. 2007.** Stable isotopes provide revised global limits of aerobic methane  
809 emissions from plants. *Atmospheric Chemistry & Physics* **7**: 237.

810 **Frankenberg C, Bergamaschi P, Butz A, Houweling S, Meirink JF, Notholt J, Petersen**  
811 **AK, Schrijver H, Warneke T, Aben I. 2008.** Tropical methane emissions: A revised  
812 view from SCIAMACHY onboard ENVISAT. *Geophysical Research Letters* **35**. doi:  
813 10.1029/2008GL034300

814 **Frankenberg C, Meirink JF, Weele Mv, Platt U, Wagner T. 2005.** Assessing Methane  
815 Emissions from Global Space-Borne Observations. *Science* **308**: 1010-1014.

816 **Fraser WT, Blei E, Fry SC, Newman MF, Reay DS, Smith KA, McLeod AR. 2015.** Emission  
817 of methane, carbon monoxide, carbon dioxide and short-chain hydrocarbons from  
818 vegetation foliage under ultraviolet irradiation. *Plant, Cell & Environment* **38**: 980-  
819 989.

820 **Fritz C, Pancotto VA, Elzenga JTM, Visser EJW, Grootjans AP, Pol A, Iturraspe R,**  
821 **Roelofs JGM, Smolders AJP. 2011.** Zero methane emission bogs: extreme  
822 rhizosphere oxygenation by cushion plants in Patagonia. *New Phytologist* **190**: 398-  
823 408.

824 **Gansert D, Burgdorf M, Lösch R. 2001.** A novel approach to the in situ measurement of  
825 oxygen concentrations in the sapwood of woody plants. *Plant, Cell & Environment*  
826 **24**: 1055-1064.

827 **Garnet KN, Megonigal JP, Litchfield C, Taylor Jr GE. 2005.** Physiological control of leaf  
828 methane emission from wetland plants. *Aquatic Botany* **81**: 141-155.

829 **Gartner BL, Moore JR, Gardiner BA. 2004.** Gas in stems: abundance and potential  
830 consequences for tree biomechanics. *Tree Physiology* **24**: 1239-1250.

831 **Gauci V, Gowing DJ, Hornibrook ERC, Davis JM, Dise NB. 2010.** Woody stem methane  
832 emission in mature wetland alder trees. *Atmospheric Environment* **44**: 2157-2160.

- 833 **Goffredi SK, Jang G, Woodside WT, Ussler III W. 2011.** Bromeliad catchments as habitats  
834 for methanogenesis in tropical rainforest canopies. *Frontiers in Microbiology* **2**: 256.
- 835 **Große W, Schröder P. 1984.** Oxygen supply of roots by gas transport in alder-trees.  
836 *Zeitschrift für Naturforschung C* **39**: 1186-1188.
- 837 **Halmeenmäki E, Heinonsalo J, Putkinen A, Santalahti M, Fritze H, Pihlatie M. 2018.**  
838 Above- and belowground fluxes of methane from boreal dwarf shrubs and *Pinus*  
839 *sylvestris* seedlings. *Plant & Soil* **420**: 361-373.
- 840 **Hansen MC, Potapov PV, Moore R, Hancher M, Turubanova S, Tyukavina A, Thau D,**  
841 **Stehman S, Goetz S, Loveland T. 2013.** High-resolution global maps of 21st-  
842 century forest cover change. *Science* **342**: 850-853.
- 843 **Hietala A, Dörsch P, Kvaalen H, Solheim H. 2015.** Carbon dioxide and methane  
844 formationMethane formation in Norway spruce stems infected by white-rot fungi.  
845 *Forests* **6**: 3304-3325.
- 846 **Hoch G, Richter A, KÖRner C. 2003.** Non-structural carbon compounds in temperate  
847 forest trees. *Plant, Cell & Environment* **26**: 1067-1081.
- 848 **Hook DD, Brown CL, Wetmore RH. 1972.** Aeration in Trees. *Botanical Gazette* **133**: 443-  
849 454.
- 850 **Hu S, Niu Z, Chen Y, Li L, Zhang H. 2017.** Global wetlands: Potential distribution, wetland  
851 loss, and status. *Science of the Total Environment* **586**: 319-327.
- 852 **Huikari, O. (1954).** Experiments on the effect of anaerobic media upon birch, pine and  
853 spruce seedlings. *Communicationes Instituti Forestalis Fenniae*  
854 **42**: 13.
- 855 **Jackson M, Armstrong W. 1999.** Formation of aerenchyma and the processes of plant  
856 ventilation in relation to soil flooding and submergence. *Plant Biology* **1**: 274-287.
- 857 **Jennings DH. 1996.** *Fungal biology: understanding the fungal lifestyle*. BIOS Scientific  
858 Publishers, Oxford.
- 859 **Jensen KF. 1967.** Oxygen and carbon dioxide affect the growth of wood-decaying fungi.  
860 *Forest Science* **13**: 384-389.
- 861 **Keppler F, Boros M, Frankenberg C, Lelieveld J, McLeod A, Pirttilä M, Rockmann T,**  
862 **Schnitzler J-P. 2009.** Methane formation in aerobic environments. *Environmental*  
863 *chemistry* **6**: 459-465.
- 864 **Keppler F, Hamilton JTG, Braß M, Rockmann T. 2006.** Methane emissions from  
865 terrestrial plants under aerobic conditions. *Nature* **439**: 187-191.
- 866 **Keppler F, Hamilton JTG, McRoberts WC, Vigano I, Braß M, Röckmann T. 2008.**  
867 Methoxyl groups of plant pectin as a precursor of atmospheric methane: evidence  
868 from deuterium labelling studies. *New Phytologist* **178**: 808-814.
- 869 **Laanbroek HJ. 2010.** Methane emission from natural wetlands: interplay between  
870 emergent macrophytes and soil microbial processes. A mini-review. *Annals of*  
871 *Botany* **105**: 141-153.
- 872 **Le Mer J, Roger P. 2001.** Production, oxidation, emission and consumption of methane by  
873 soils: a review. *European Journal of Soil Biology* **37**: 25-50.
- 874 **Lenhart K, Bunge M, Ratering S, Neu TR, Schüttmann I, Greule M, Kammann C, Schnell**  
875 **S, Müller C, Zorn H, et al. 2012.** Evidence for methane production by saprotrophic  
876 fungi. *Nature Communications* **3**. doi: 10.1038/ncomms2049.
- 877 **Liu J, Chen H, Zhu Q, Shen Y, Wang X, Wang M, Peng C. 2015.** A novel pathway of direct  
878 methane production and emission by eukaryotes including plants, animals and  
879 fungi: An overview. *Atmospheric Environment* **115**: 26-35.
- 880 **Lowe DC. 2006.** Global change: A green source of surprise. *Nature* **439**: 148-149.

- 881 **MacDougal DT. 1927.** Composition of gases in trunks of trees. *Carnegie Institution of*  
882 *Wahsingont Year Book* **26**: 162-163.
- 883 **MacDougal DT. 1932.** The pneumatic system of trees. *Proceedings of the American*  
884 *Philosophical Society* **71**: 299-307.
- 885 **Machacova K, Bäck J, Vanhatalo A, Halmeenmäki E, Kolari P, Mammarella I,**  
886 **Pumpanen J, Acosta M, Urban O, Pihlatie M. 2016.** *Pinus sylvestris* as a missing  
887 source of nitrous oxide and methane in boreal forest. *Scientific Reports* **6**: 23410.  
888 doi: 10.1038/srep23410
- 889 **Mäkipää R, Leppänen SM, Munoz SS, Smolander A, Tirolab M, Tuomivirta T,**  
890 **Fritze H. 2018.** Methanotrophs are core members of the diazotroph community in  
891 decaying Norway spruce logs. *Soil Biology and Biochemistry* **120**: 230–232.
- 892 **Maier M, Machacova K, Lang F, Svobodova K, Urban O. 2017.** Combining soil and tree-  
893 stem flux measurements and soil gas profiles to understand CH<sub>4</sub> pathways in *Fagus*  
894 *sylvatica* forests. *Journal of Plant Nutrition and Soil Science* **181**: 31–35. doi:  
895 10.1002/jpln.201600405
- 896 **Mancuso S, Marras AM. 2003.** Different pathways of the oxygen supply in the sapwood of  
897 young *Olea europaea* trees. *Planta* **216**: 1028-1033.
- 898 **Martel AB, Qaderi MM. 2017.** Light quality and quantity regulate aerobic methane  
899 emissions from plants. *Physiologia Plantarum* **159**: 313-328.
- 900 **Martinson GO, Werner FA, Scherber C, Conrad R, Corre MD, Flessa H, Wolf K, Klose M,**  
901 **Gradstein SR, Veldkamp E. 2010.** Methane emissions from tank bromeliads in  
902 neotropical forests. *Nature Geoscience* **3**: 766-769.
- 903 **Matthews E, Fung I. 1987.** Methane emission from natural wetlands: Global distribution,  
904 area, and environmental characteristics of sources. *Global Biogeochem. Cycles* **1**: 61-  
905 86.
- 906 **McLeod AR, Fry SC, Loake GJ, Messenger DJ, Reay DS, Smith KA, Yun B-W. 2008.**  
907 Ultraviolet radiation drives methane emissions from terrestrial plant pectins. *New*  
908 *Phytologist* **180**: 124-132.
- 909 **Megonigal JP, Day FP. 1992.** Effects of flooding on root and shoot production of bald  
910 cypress in large experimental enclosures. *Ecology* **73**: 1182-1193.
- 911 **Megonigal JP, Guenther AB. 2008.** Methane emissions from upland forest soils and  
912 vegetation. *Tree Physiology* **28**: 491-498.
- 913 **Megonigal JP, Hines ME, Visscher PT 2004.** Anaerobic metabolism: Linkages to trace  
914 gases and aerobic processes. In: Schlesinger WH ed. *Biogeochemistry*. Oxford, UK:  
915 Elsevier-Pergamon, 317-424.
- 916 **Megonigal JP, Patrick W, Faulkner S. 1993.** Wetland identification in seasonally flooded  
917 forest soils: soil morphology and redox dynamics. *Soil Science Society of America*  
918 *Journal* **57**: 140-149.
- 919 **Megonigal JP, Schlesinger WH. 2002.** Methane-limited methanotrophy in tidal freshwater  
920 swamps. *Global biogeochemical cycles* **16**: 1088.
- 921 **Messenger DJ, McLeod A, Fry SC. 2009.** The role of ultraviolet radiation, photosensitizers,  
922 reactive oxygen species and ester groups in mechanisms of methane formation from  
923 pectin. *Plant, cell and environment* **32**: 1-9.
- 924 **Moya R, Muñoz F, Jeremic D, Berrocal A. 2009.** Visual identification, physical properties,  
925 ash composition, and water diffusion of wetwood in *Gmelina arborea*. *Canadian*  
926 *journal of forest research* **39**: 537-545.
- 927 **Mukhin V, Voronin P. 2007.** Methane emission during wood fungal decomposition.  
928 *Doklady Biological Sciences* **413**: 159-160.

- 929 **Mukhin V, Voronin P. 2008.** A new source of methane in boreal forests. *Applied*  
930 *Biochemistry and Microbiology* **44**: 297-299.
- 931 **Mukhin V, Voronin P. 2011.** Methane emission from living tree wood. *Russian Journal of*  
932 *Plant Physiology* **58**: 344-350.
- 933 **Myhre G, Shindell D, Bréon F-M, Collins W, Fuglestedt J, Huang J, Koch D, Lamarque J-**  
934 **F, Lee D, Mendoza B, et al. 2013.** Anthropogenic and Natural Radiative Forcing. In:  
935 Stocker TF, Qin D, Plattner G-K, Tignor M, Allen SK, Boschung J, Nauels A, Xia Y, Bex  
936 V, Midgley PM eds. *Climate Change 2013: The Physical Science Basis. Contribution of*  
937 *Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on*  
938 *Climate Change*. Cambridge, United Kingdom and New York, NY, USA: Cambridge  
939 University Press, 659–740.
- 940 **Neubauer SC, Megonigal JP. 2015.** Moving beyond global warming potentials to quantify  
941 the climatic role of ecosystems. *Ecosystems* **18**: 1000-1013.
- 942 **Nietch CT, Morris JT, Vroblesky DA. 1999.** Biophysical mechanisms of trichloroethene  
943 uptake and loss in baldcypress growing in shallow contaminated groundwater.  
944 *Environmental Science & Technology* **33**: 2899-2904.
- 945 **Oberle B, Covey KR, Dunham KM, Hernandez EJ, Walton ML, Young DF, Zanne AE.**  
946 **2017.** Dissecting the effects of diameter on wood decay emphasizes the importance  
947 of cross-stem conductivity in *Fraxinus americana*. *Ecosystems* **21**: 85-97.
- 948 **Pan Y, Birdsey RA, Fang J, Houghton R, Kauppi PE, Kurz WA, Phillips OL, Shvidenko A,**  
949 **Lewis SL, Canadell JG, et al. 2011.** A Large and persistent carbon sink in the  
950 world's forests. *Science* **333**: 988-993.
- 951 **Pangala SR, Enrich-Prast A, Basso LS, Peixoto RB, Bastviken D, Hornibrook ERC, Gatti**  
952 **LV, Marotta H, Calazans LSB, Sakuragui CM, et al. 2017.** Large emissions from  
953 floodplain trees close the Amazon methane budget. *Nature* **552**: 230-234.
- 954 **Pangala SR, Gowing DJ, Hornibrook ERC, Gauci V. 2014.** Controls on methane emissions  
955 from *Alnus glutinosa* saplings. *New Phytologist* **201**: 887-896.
- 956 **Pangala SR, Hornibrook ERC, Gowing DJ, Gauci V. 2015.** The contribution of trees to  
957 ecosystem methane emissions in a temperate forested wetland. *Global Change*  
958 *Biology* **21**: 2642-2654.
- 959 **Pangala SR, Moore S, Hornibrook ERC, Gauci V. 2013.** Trees are major conduits for  
960 methane egress from tropical forested wetlands. *New Phytologist* **197**: 524-531.
- 961 **Phillips RP, Finzi AC, Bernhardt ES. 2011.** Enhanced root exudation induces microbial  
962 feedbacks to N cycling in a pine forest under long-term CO<sub>2</sub> fumigation. *Ecology*  
963 *Letters* **14**: 187-194.
- 964 **Plain C, Ndiaye F, Bonnaud P, Ranger J, Epron D. 2018.** Impact of vegetation on the  
965 methane budget of a temperate forest. *New Phytologist* doi: 10.1111/nph.15452
- 966 **Pitz S, Megonigal JP. 2017.** Temperate forest methane sink diminished by tree emissions.  
967 *New Phytologist*. **214**: 1432-1439.
- 968 **Pruyn ML, Gartner BL, Harmon ME. 2002.** Within-stem variation of respiration in  
969 *Pseudotsuga menziesii* (Douglas-fir) trees. *New Phytologist* **154**: 359-372.
- 970 **Pitz SL, Megonigal JP, Chang C-H, Szlavecz K. 2018.** Methane fluxes from tree stems and  
971 soils along a habitat gradient. *Biogeochemistry* **137**: 307-320.
- 972 **Pulliam WM. 1992.** Methane emissions from cypress knees in a southeastern floodplain  
973 swamp. *Oecologia* **91**: 126–128.
- 974 **Purvaja R, Ramesh R, Frenzel P. 2004.** Plant-mediated methane emission from an Indian  
975 mangrove. *Global Change Biology* **10**: 1825-1834.
- 976 **Prigent C, Papa F, Aires F, Rossow WB, Matthews E. 2007.** Global inundation dynamics

977           inferred from multiple satellite observations, 1993–2000. *Journal of Geophysical*  
978 *Research: Atmospheres* **112**. doi: 10.1029/2006JD007847.

979 **Qaderi MM, Reid DM. 2009.** Methane emissions from six crop species exposed to three  
980 components of global climate change: temperature, ultraviolet-B radiation and  
981 water stress. *Physiologia Plantarum* **137**: 139-147.

982 **Qaderi MM, Reid DM. 2011.** Stressed crops emit more methane despite the mitigating  
983 effects of elevated carbon dioxide. *Functional Plant Biology* **38**: 97-105.

984 **Rice AL, Butenhoff CL, Shearer MJ, Teama D, Rosenstiel TN, Khalil MAK. 2010.**  
985 Emissions of anaerobically produced methane by trees. *Geophysical Research*  
986 *Letters*. **37**. doi: 10.1029/2009GL041565.

987 **Richardson AD, Carbone MS, Keenan TF, Czimczik CI, Hollinger DY, Murakami P,**  
988 **Schaberg PG, Xu X. 2013.** Seasonal dynamics and age of stemwood nonstructural  
989 carbohydrates in temperate forest trees. *New Phytologist* **197**: 850-861.

990 **Rusch H, Rennenberg H. 1998.** Black alder (*Alnus Glutinosa* (L.) Gaertn.) trees mediate  
991 methane and nitrous oxide emission from the soil to the atmosphere. *Plant & Soil*  
992 **201**: 1-7.

993 **Sanhueza E, Donoso L. 2006.** Methane emission from tropical savanna *Trachypogon* sp.  
994 grasses. *Atmospheric Chemistry & Physics Discussion* **6**: 5215-5319.

995 **Sauniois M, Bousquet P, Poulter B, Peregon A, Ciais P, Canadell JG, Dlugokencky EJ,**  
996 **Etiopie G, Bastviken D, Houweling S, et al. 2016.** The Global Methane Budget:  
997 2000-2012. *Earth System Science Data Discussion*. **2016**: 1-79.

998 **Sauniois M, Bousquet P, Poulter B, Peregon A, Ciais P, Canadell JG, Dlugokencky EJ,**  
999 **Etiopie G, Bastviken D, Houweling S, et al. 2017.** Variability and quasi-decadal  
1000 changes in the methane budget over the period 2000–2012. *Atmospheric Chemistry*  
1001 *& Physics*. **17**: 11135-11161.

1002 **Schiermeier Q. 2006.** Methane finding baffles scientists. *Nature* **439**: 128-128.

1003 **Schink B, Ward J. 1984.** Microaerobic and anaerobic bacterial activities involved in  
1004 formation of wetwood and discoloured wood. *International Association of Wood*  
1005 *Anatomist Bulletin* **5**: 105-109.

1006 **Schink G, Ward JC, Zeikus JG. 1981.** Microbiology of Wetwood: Role of Anaerobic Bacterial  
1007 Populations in Living Trees. *Journal of General Microbiology* **123**: 313-322.

1008 **Schlesinger WH, Jasechko S. 2014.** Transpiration in the global water cycle. *Agricultural*  
1009 *and Forest Meteorology* **189**: 115-117.

1010 **Schmidt O. 2006.** *Wood and Tree Fungi: Biology, Damage, Protection, and Use*. Berlin,  
1011 Germany: Springer-Verlag.

1012 **Schröder P. 1989.** Aeration of the root system in *Alnus glutinosa* L. Gaertn. *Annales des*  
1013 *sciences forestières* **46**(Supplement): 310s-314s.

1014 **Shoemaker JK, Keenan TF, Hollinger DY, Richardson AD. 2014.** Forest ecosystem  
1015 changes from annual methane source to sink depending on late summer water  
1016 balance. *Geophysical research letters* **41**: 673-679.

1017 **Shortle WC, Menge JA, Cowling EB. 1978.** Interaction of bacteria, decay fungi, and live  
1018 sapwood in discoloration and decay of trees1. *European Journal of Forest Pathology*  
1019 **8**: 293-300.

1020 **Siegenthaler A, Welch B, Pangala SR, Peacock M, Gauci V. 2016.** Technical Note: Semi-  
1021 rigid chambers for methane gas flux measurements on tree stems. *Biogeosciences*  
1022 **13**: 1197-1207.

1023 **Sorz J, Hietz P. 2006.** Gas diffusion through wood: implications for oxygen supply. *Trees -*  
1024 *Structure and Function* **20**: 34-41.

- 1025 **Spahni R, Wania R, Neef L, van Weele M, Pison I, Bousquet P, Frankenberg C, Foster**  
 1026 **PN, Joos F, Prentice IC, et al. 2011.** Constraining global methane emissions and  
 1027 uptake by ecosystems. *Biogeosciences* **8**: 1643-1665.
- 1028 **Spicer R, Holbrook NM. 2005.** Within-stem oxygen concentration and sap flow in four  
 1029 temperate tree species: does long-lived xylem parenchyma experience hypoxia?  
 1030 *Plant, Cell & Environment* **28**: 192-201.
- 1031 **Sundqvist E, Crill P, Mölder M, Vestin P, Lindroth A. 2012.** Atmospheric methane  
 1032 removal by boreal plants. *Geophysical Research Letters* **39**. doi:  
 1033 10.1029/2012GL053592.
- 1034 **Sundqvist E, Mölder M, Crill P, Kljun N, Lindroth A. 2015.** Methane exchange in a boreal  
 1035 forest estimated by gradient method. *Tellus B: Chemical and Physical Meteorology*  
 1036 **67**. doi: 10.3402/tellusb.v67.26688.
- 1037 **Terazawa K, Ishizuka S, Sakata T, Yamada K, Takahashi M. 2007.** Methane emissions  
 1038 from stems of *Fraxinus mandshurica* var. *japonica* trees in a floodplain forest. *Soil*  
 1039 *Biology & Biochemistry* **39**: 2689-2692.
- 1040 **Terazawa K, Yamada K, Ohno Y, Sakata T, Ishizuka S. 2015.** Spatial and temporal  
 1041 variability in methane emissions from tree stems of *Fraxinus mandshurica* in a cool-  
 1042 temperate floodplain forest. *Biogeochemistry* **123**: 349-362.
- 1043 **Teskey RO, McGuire MA. 2007.** Measurement of stem respiration of sycamore (*Platanus*  
 1044 *occidentalis* L.) trees involves internal and external fluxes of CO<sub>2</sub> and possible  
 1045 transport of CO<sub>2</sub> from roots. *Plant, Cell & Environment* **30**: 570-579.
- 1046 **Teskey RO, Saveyn A, Steppe K, McGuire MA. 2008.** Origin, fate and significance of CO<sub>2</sub> in  
 1047 tree stems. *New Phytologist* **177**: 17-32.
- 1048 **Topa MA, McLeod KW. 1986.** Aerenchyma and lenticel formation in pine seedlings: a  
 1049 possible avoidance mechanism to anaerobic growth conditions. *Physiologia*  
 1050 *Plantarum* **68**: 540-550.
- 1051 **Turetsky MR, Kotowska A, Bubier J, Dise NB, Crill P, Hornibrook ER, Minkinen K,**  
 1052 **Moore TR, Myers-Smith IH, Nykänen H. 2014.** A synthesis of methane emissions  
 1053 from 71 northern, temperate, and subtropical wetlands. *Global Change Biology* **20**:  
 1054 2183-2197.
- 1055 **UNFCCC. 2016.** The Paris Agreement. [https://unfccc.int/process-and-meetings/the-paris-](https://unfccc.int/process-and-meetings/the-paris-agreement/the-paris-agreement)  
 1056 [agreement/the-paris-agreement](https://unfccc.int/process-and-meetings/the-paris-agreement/the-paris-agreement)
- 1057 **Van Der Kamp BJ, Gokhale AA, Smith RS. 1979.** Decay resistance owing to near-anaerobic  
 1058 conditions in black cottonwood wetwood. *Canadian Journal of Forest Research* **9**: 39-  
 1059 44.
- 1060 **Vann CD, Megonigal JP. 2003.** Elevated CO<sub>2</sub> and water depth regulation of methane  
 1061 emissions: Comparison of woody and non-woody wetland plant species.  
 1062 *Biogeochemistry* **63**: 117-134.
- 1063 **Vigano I, Röckmann T, Holzinger R, van Dijk A, Keppler F, Greule M, Brand WA,**  
 1064 **Geilmann H, van Weelden H. 2009.** The stable isotope signature of methane  
 1065 emitted from plant material under UV irradiation. *Atmospheric Environment* **43**:  
 1066 5637-5646.
- 1067 **Vigano I, van Weelden H, Holzinger R, Keppler F, Rockmann T. 2008.** Effect of UV  
 1068 radiation and temperature on the emission of methane from plant biomass and  
 1069 structural components. *Biogeosciences* **5**: 937-947.
- 1070 **Von Fischer JC, Hedin LO. 2002.** Separating methane production and consumption with a  
 1071 field-based isotope pool dilution technique. *Global Biogeochemical Cycles* **16**: 1034.
- 1072 **Wang S, Yang X, Lin X, Hu Y, Luo C, Xu G, Zhang Z, Su A, Chang X, Chao Z, et al. 2009.**



- 1073 Methane emission by plant communities in an alpine meadow on the Qinghai-  
 1074 Tibetan Plateau: a new experimental study of alpine meadows and oat pasture.  
 1075 *Biology Letters* **5**: 535-538.
- 1076 **Wang Z, Gullledge J, Zheng JQ, Liu W, Li LH, Han XG. 2009.** Physical injury stimulates  
 1077 aerobic methane emissions from terrestrial plants. *Biogeosciences* **6**: 615-621.
- 1078 **Wang Z-P, Gu Q, Deng F-D, Huang J-H, Megonigal JP, Yu Q, Lü X-T, Li L-H, Chang S,**  
 1079 **Zhang Y-H, et al. 2016.** Methane emissions from the trunks of living trees on  
 1080 upland soils. *New Phytologist* **211**: 429-439.
- 1081 **Wang Z-P, Han S-J, Li H-L, Deng F-D, Zheng Y-H, Liu H-F, Han X-G. 2017.** Methane  
 1082 production explained largely by water content in the heartwood of living trees in  
 1083 upland forests. *Journal of Geophysical Research: Biogeosciences* **122**. doi:  
 1084 10.1002/2017JG003991.
- 1085 **Warner DL, Villarreal S, McWilliams K, Inamdar S, Vargas R. 2017.** Carbon dioxide and  
 1086 methane fluxes from tree stems, coarse woody debris, and soils in an upland  
 1087 temperate forest. *Ecosystems* **20**: 1205-1216.
- 1088 **Warshaw JE, Leschine SB, Canale-Parola E. 1985.** Anaerobic cellulolytic bacteria from  
 1089 wetwood of living trees. *Applied & Environmental Microbiology* **50**: 807-811.
- 1090 **Wolin MJ, Miller TL. 1987.** Bioconversion of organic carbon to CH<sub>4</sub> and CO<sub>2</sub>.  
 1091 *Geomicrobiology Journal* **5**: 239-259.
- 1092 **Worm P, Müller N, Plugge C, Stams A, Schink B 2011.** Syntrophy in methanogenic  
 1093 degradation. In: Hackstein JHP ed. *(Endo)symbiotic Methanogenic Archaea*. Berlin:  
 1094 Springer Berlin Heidelberg, 143-173.
- 1095 **Würth MR, Peláez-Riedl S, Wright SJ, Körner C. 2005.** Non-structural carbohydrate pools  
 1096 in a tropical forest. *Oecologia* **143**: 11-24.
- 1097 **Xu Z, Leininger T. 2001.** Chemical properties associated with bacterial wetwood in red  
 1098 oaks. *Wood and Fiber Science* **33**: 76-83.
- 1099 **Yip DZ, Veach AM, Yang ZK, Cregger MA, Schadt CW. 2018.** Methanogenic Archaea  
 1100 dominate mature hardwood habitats of Eastern Cottonwood (*Populus deltoides*).  
 1101 *New Phytologist*. doi: 10.1111/nph.15346.
- 1102 **Zeikus J, Henning D. 1975.** *Methanobacterium arbophilicum* sp. nov. An obligate anaerobe  
 1103 isolated from wetwood of living trees. *Antonie van Leeuwenhoek* **41**: 543-552.
- 1104 **Zeikus J, Ward JC. 1974.** Methane formation in living trees: a microbial origin. *Science* **184**:  
 1105 1181.

## 1108 Supporting Information

1109 Additional Supporting Information may be found online in the Supporting Information  
 1110 tab for this article:

1111

1112 **Table S1** Compilation of primary publications on methane as it relates to trees or forests.

**Table 1.** Methane emission rates from tree stems. All fluxes were directly measured using static chambers and are expressed as per area of stem or knee. Measures of central tendency and errors are in the original forms reported in the citation. When no statistical information is reported the data are means with standard errors in parentheses; exceptions are reported in the "emission surface" column as median for central tendency, or standard deviations (SD) or 95% confidence intervals (CI) for errors. Not all studies reported errors.

Tree Condition	Ecosystem Type	Forest Type	Plant Community	Site	Emission Surface	Rate ( $\mu\text{mol CH}_4 \text{ m}^{-2} \text{ stem h}^{-1}$ )	Citation
Living Trees	Wetland	Temperate Floodplain	<i>Taxodium distichum</i>		Taxodium knees	2.34 (0.78)†	Pulliam 1992
		Temperate Floodplain	<i>Fraxinus mandshurica</i>		Stem at 15 cm	11	Terazawa <i>et al.</i> 2007
		Temperate Floodplain	<i>Alnus glutinosa</i>		Stem at 30 cm	0.26 to 6.31	Gauci <i>et al.</i> 2010
		Tropical Peatland	Diverse angiosperms		Stem at 20-50 cm	1.06 (0.9) to 11.56 (0.44)	Pangala <i>et al.</i> 2013
		Temperate Floodplain	<i>Fraxinus mandshurica</i>		Stem at 15 cm	5.1 to 81.6	Terazawa <i>et al.</i> 2015
		Temperate Peatland	<i>Alnus glutinosa</i>		Young stem at 20-50 cm	140.42 (18.43)	Pangala <i>et al.</i> 2015
					Mature stem at 20-50 cm	9.79 (1.29)	
					Young stem at 20-50 cm	51.47 (8.18)	
		Tropical Floodplain	Diverse angiosperms	Negro River	Mature stem at 20-50 cm	6.71 (0.83)	Pangala <i>et al.</i> 2017
					Large stems	1,887.50 (1,293.75)	
					Small stems	3,687.5 (1,762.5)	
		Tropical Floodplain	Diverse angiosperms	Madeira River	Small leaves	1 (2.5)	
					Large stems	2,075 (1625)	
					Small stems	3,137.5 (2,056.25)	
		Tropical Floodplain	Diverse angiosperms	Amazon River	Small leaves	1.19 (2.5)	
					Large stems	2,900 (2,106.25)	
					Small stems	6,437.5 (2,806.25)	
		Tropical Floodplain	Diverse angiosperms	Solimoes River	Small leaves	2.38 (4.38)	
					Large stems	5,200 (2,675)	
					Small stems	9,375 (4,212.5)	
		Tropical Floodplain	Diverse angiosperms	Tapajos River	Small leaves	3.19 (5.63)	
					Large stems	8,812.5 (4,462.5)	
					Small stems	16,937.5 (6,812.5)	
		Temperate Floodplain	Diverse angiosperms	Wetland	Small leaves	5.63 (6.88)	Pitz <i>et al.</i> 2018
					Stem at 30-60 cm	35.49 (10.91)	

**Table 1.** Continued.

Living Trees	Upland	Boreal Evergreen	<i>Pinus sylvestris</i>	Dry Plot	Stem chamber <sup>‡</sup> (median)	0.0008	Machacova <i>et al.</i> 2016
				Wet Plot		0.0063	
				Dry Plot	Shoot (median)	0.0031	
		Temperate Deciduous	<i>Populus</i>	Dry Plot	Stems (median)	0.00031	Wang <i>et al.</i> 2016
				Upper Plot	Stem at 30 cm	5.33	
				Lower Plot		6.44	
		Temperate Deciduous	<i>Fagus sylvatica</i>	Stina Site	Stem at 40-200 cm (SD)	1.87 (3.31)	Maier <i>et al.</i> 2017
				Conventwald Site		0	
				Temperate Deciduous	Diverse angiosperms	Stem at 130 cm (CI)	
		Temperate Deciduous	Diverse angiosperms		Stem at 30-60 cm (CI)	0.396 (0.180)	Warner <i>et al.</i> 2017
						1.59 (0.88)	
		Temperate Deciduous	<i>Populus</i>	Upper Site	Stem at 30 cm	12.63	Pitz and Megonigal 2017
				Middle Site		20.72	
				Lower Site		17.05	
		Temperate Deciduous	Diverse angiosperms	Highest Elevation	Stem at 30-60 cm	4.3 (0.81)	Pitz <i>et al.</i> 2018
				Wetland Adjacent	Stem at 30-60 cm	11.29 (3.45)	
				Vegetated Plots	Stem at 25-45 cm (CI)	0.032 (0.022)	
Dead Trees	Wetland	Temperate Deciduous	<i>Quercus petrea</i>		Standing dead trees	25 (6.25)	Carmichael <i>et al.</i> 2017
					Standing dead trees	-37.5 (18.75)	
Dead Trees	Upland	Temperate Deciduous	Diverse angiosperms		Coarse wood debris (CI)	-1.15 (0.94)	Warner <i>et al.</i> 2007
					Fresh woody debris	1.15 (2.05)	
					Decayed debris	-1.44 (0.72)	

<sup>†</sup>Flux units are  $\mu\text{mol knee}^{-1} \text{h}^{-1}$

<sup>‡</sup>Chamber height not reported

**Table 2. Oxygen concentrations in living tree stems. All studies are from upland ecosystems.**

Forest Type	Plant Genera or Species	[O <sub>2</sub> ]	Study
Not specified	<i>Unspecified Populous sp.</i>	1.2%	Bushong 1907
Temperate Forest	<i>Quercus rubra</i> , <i>Quercus macrocarpa</i> , <i>Ulmus</i>	0.02-	Chase 1937
Temperate Hardwood Forest	<i>Quercus rubra</i>	5.5-7.5%	Jensen 1967
Temperate Hardwood Forest	<i>Populus delectans</i>	0.02-2.9	Van Der Kamp <i>et al.</i> 1979
Conifer Forest	<i>Picea abies</i>	5.6-15.1%	Eklund 1990
Hardwood Forest	<i>Quercus robur</i> , <i>Acer platanoides</i>	5-19%	Eklund 1993
Upland Conifer Forest	<i>Picea abies</i>	0.5-21%	Eklund 2000
Hardwood Forest	<i>Betula pendula</i>	1-5%	Gansert <i>et al.</i> 2001
Greenhouse Study	<i>Olea europaea</i>	1-5%	Mancuso and Marras 2003
Temperate Arboretum	<i>Fagus orientalis</i> , <i>Carya ovata</i> , <i>Larix sibirica</i> ,	13-20%	del Hierro <i>et al.</i> 2002
Conifer Forest	<i>Pseudotsuga menziesii</i>	0.5-20%	Pruyn <i>et al.</i> 2002
Hardwood Forest	<i>Acer rubrum</i> , <i>Fraxinus americana</i> , <i>Tsuga</i>	3-20%	Spicer and Holbrook 2004

**Table 3.** Tree methane emissions scaled to forest area.

Ecosystem Type	Forest Type	Plant Community	Site	Emission Surface	Description	Portion of Ecosystem flux†	Tree Flux (g CH <sub>4</sub> ha <sup>-1</sup> d <sup>-1</sup> )‡	Other Surfaces	Other Surface Flux (g CH <sub>4</sub> ha <sup>-1</sup> d <sup>-1</sup> )	Study
Wetland	Tropical Peatland	Diverse Angiosperm		Tree stems	Scaled to 15 m	87.0%	28.5 (3.4)	Soil hollows	3.9 (1.0)	Pangala <i>et al.</i> 2013
				Tree stems	Scaled to 3 m	62.0%	6.7 (0.7)			
	Temperate Peatland			Tree stems	Summer	13.5%	13.2 (1.34)	Soil hollows	37.3 (10.2)	Pangala <i>et al.</i> 2015
								Soil hummocks	2.51 (2.03)	
				Tree stems	Winter	24.6%	5.65 (0.9)	Soil hollows	11.3 (3.57)	
								Soil hummocks	0.09 (0.1)	
	Tropical Floodplain	Diverse Angiosperms	Negro River	Large stems		58.3%	47.4 (11)	Soil	67.7 (56)	Pangala <i>et al.</i> 2017
				Small stems		5.8%	67.7 (56)	Aquatic surface	219 (544)	
				Small leaves		0.5%	3.86 (4.6)			
			Madeira River	Large stems		58.3%	47.4 (11)	Soil	251 (289)	
				Small stems		5.2%	251 (289)	Aquatic surface	423 (148)	
				Small leaves		0.3%	5.07 (4.8)			
			Amazon River	Large stems		43.6%	50.3 (13.3)	Soil	49 (179)	
				Small stems		2.7%	49 (179)	Aquatic surface	768 (1,792)	
				Small leaves		0.3%	5.93 (7.3)	Floating macrophytes	190 (745)	
			Solimoes River	Large stems		53.0%	157 (40.5)	Soil	88.6 (108)	
				Small stems		4.4%	88.6 (108)	Aquatic surface	1,269 (1,111)	
				Small leaves		5.8%	67.7 (56)	Floating macrophytes	134 (261)	
			Tapajos River	Large stems		41.5%	181 (56.1)	Soil	456 (564)	
				Small stems		2.6%	456 (564)	Aquatic surface	2,426 (2,898)	
				Small leaves		0.2%	17.3 (15.7)	Floating macrophytes	966 (2105)	
Upland	Temperate Floodplain	<i>Fraxinus mandshurica</i>	Dry plot	Tree Stems	Scaled 5-80 cm	69%	0.17	Soil	-0.24	Terazawa <i>et al.</i> 2007
				Stems		0.02%	0.00072	Soil	-3.43	Machacova <i>et al.</i> 2016
		<i>Pinus sylvestris</i>	Wet plot	Shoots		0.8%	0.0264			
				Stems		0.6%	0.01	Soil	-1.7	
		Diverse angiosperms		Shoots		33.5%	0.57			
				Stems (SD)		3.5%	0.75 (2.21)	Soil (SD)	-3.92 (0.25)	Warner <i>et al.</i> 2017
		Diverse angiosperms						Deadwood (SD)	-0.37 (0.25)	
				Tree Stems (95% CI)	Scaled to 3 m	4.5%	0.79 (0.44)	Soil (95% CI)	-17.4 (2.5)	Pitz and Megonigal 2017
		<i>Quercus petrea</i>	Vegetated Plots	Tree Stems (95% CI)	Scaled to full height	0.3%	0.07 (0.02)	Soil (95% CI)	-220 (45.5)	Plain <i>et al.</i> 2018

†Represents the contribution of trees to total ecosystem efflux in wetlands studies. In upland studies it is the fraction of soil oxidation that is offset by tree emissions.

‡Values and errors are in the original units reported in the citation. When no statistical information is reported the data are means with standard errors in parentheses; exceptions are reported in the "emission surface" column as either standard deviations (SD) or 95% confidence intervals (CI). Not all studies reported errors.

## Figure Legends

Figure 1. The complex variety of CH<sub>4</sub> sources and sinks in upland and wetland forests. Red arrows are CH<sub>4</sub> sources and blue arrows are sinks. See Carmichael *et al.* (2014) for a treatment of the role of vegetation in CH<sub>4</sub> dynamics across a variety of terrestrial ecosystems.

Figure 2. Flammable concentrations of CH<sub>4</sub> in the heartwood of living trees are common even on upland sites, such as this *Quercus cerris* tree in Hungary that was cored and the pressurized stem gas ignited. Photo by Balazs Nyitrai.

Figure 3. Methane emissions from a *Liriodendron tulipifera* (closed circles) and a *Fagus grandifolia* (open circles) at 75 cm above the soil surface. Note that the y-axes for the two gases are scaled differently. From Pitz & Megonigal, 2017.

Figure 4. Correlation between CH<sub>4</sub> exchange and GPP for a Spruce tree. The best-fit line to the points has a correlation coefficient is 0.57. Notice that a negative GPP means uptake from the atmosphere. From Sundqvist *et al.*, 2012.

Figure 5. Stem CH<sub>4</sub> emissions and stem lenticel density at a height of 2–12 cm above the soil surface are strongly related in *Alnus glutinosa* saplings. From Pangela *et al.* (2014).

Figure 6. Snapshot of raw CH<sub>4</sub> concentration data over 24 hours. Peak values (~2.01 ppm) are a response to UV irradiation, while troughs (~1.95 ppm) are the result of UV lamps that shut down every 105 minutes to monitor instrument function. From Vigano *et al.*, 2008.

Figure 7. Mean relative abundance of dominant phyla (bacteria and archaea) and subphyla (Proteobacteria) across wood tissue types in *Populus deltoids*. From Yip *et al.*, 2018.