Estuaries and Coasts

Feeding behavior of the native mussel Ischadium recurvum and the invasive mussels Mytella charruana and Perna viridis in Florida, USA, across a salinity gradient -- Manuscript Draft--

Manuscript Number:	ESCO-D-17-00358R2				
Full Title:	Feeding behavior of the native mussel lsch. Mytella charruana and Perna viridis in Flori				
Article Type:	Original Paper	da, 30, t, adio55 a sainity gradient			
Keywords:	feeding behavior; bivalve; seston; salinity	St. Johns River			
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Funding Information:	St. Johns River Water Management District (27799)	Not applicable			
Abstract:	The feeding behavior of three species of method were performed along a salinity grawater characteristics (salinity, temperature, related to the feeding behavior. Clearance, rates of I. recurvum were negatively affecte proportion was positively related to salinity absorption rates were positively related to the seston. Total and organic ingestion rates of salinity but positively affected by total particand M. charruana were 13.16 ± 0.64 and 6.0.41 for the native species I. recurvum, indito the environmental conditions in the area. thrive in different water characteristics. Thu occupy the niche of the native mussel in Floritations.	viridis, was studied in an invaded experiments using the biodeposition adient in the St. Johns River. Additionally, dissolved oxygen, and seston loads) were filtration, organic ingestion, and absorption d by salinity. For M. charruana, rejection while total ingestion, organic ingestion, and he percentage of organic matter in the FP. viridis were negatively affected by sulate matter. Condition indices for P. viridis .63 ± 0.43, respectively, compared to 4.82 ± cating that these mussels are well adapted This study indicates that the three species s, the invasive mussels will not totally			



May 14, 2018

Dr. Paul E. Montagna Co-Editor-in-Chief Estuaries and Coasts

Dear Dr. Montagna,

We would like to re-submit the Research Article entitled "Feeding behavior of the native mussel *Ischadium recurvum* and the invasive mussels *Mytella charruana* and *Perna viridis* in Florida, USA, across a salinity gradient" to the journal Estuaries and Coasts.

We submitted a tracked and non-tracked versions of the manuscript yesterday because we thought they both would help understand the changes made in the manuscript. However, as requested, we are now resubmitting just the clean version with no tracked changes.

My coauthors and I have considered all of the comments and have been able to address them. We apologize for any confusion caused by the first revision and the very recent submission and hope that this version of the manuscript will satisfy the Co-Editor-in-Chief and Reviewer 2 hesitations.

We are very thankful for all the comments and suggestions; the article has greatly improved after addressing them. Please find attached a non-tracked version of the manuscript and a document with detailed information provided on how we addressed each suggestion in the revised manuscript.

Best regards,

Dr. Eve Galimany



Co-Editor in Chief

One problem is that the letter with the revision is inadequate. A letter in response to a review should contain an explicit response, i.e., how the text was changed, and where the text was changed (i.e., line numbers). Your responses are more of a narrative explanation that no reader will ever see, and it is not clear that changes were made to the manuscript. You have to tell the editor how you changed the manuscript. Using continuous line numbering makes this easier.

We apologize for the confusion over the changes made to the manuscript; we misunderstood the guide for authors about numbering. We have changed our line numbers to continuous as suggested. We have endeavored to explain the changes made in response to Reviewer 2's comments and more clearly denote the changes made in the manuscript.

Reviewer # 2

Although the revised ms has improved in the sense that essential errors in the previous version have been resolved, the paper still lacks scientific depth. It is interesting to measure physiological parameters of 3 species in a salinity gradient but I do not see how this contributes to explaining the observations, on habitat preferences. Also many more factors determine habitat, and the authors did change the title but are still addressing the habitat preferences. Moreover, the research question as formulated by the authors: "We hypothesized that invasive mussels had higher feeding rates because successful invasive bivalves seem to be very efficient at using the natural resources, often even more efficient than native species" is not testable as it is a circular argument: they have higher feeding rates because they are efficient.

It seems that the salinity gradient determines the distribution of the species whether native or invasive is another issue. This spatial differentiation prevents competition and possible loss of natives, so there is hardly any issue linked to invasion in this case.

We appreciate the Reviewer's concern that we are overstating the results of our study in regards to habitat preferences. We agree that many factors in addition to salinity affect habitat preference, and have endeavored to shift the focus of the paper to the effects of salinity and seston loads on feeding behaviors (Lines 27, 81-85, 292-302, 315-318). Additionally, we have expanded the last paragraph to include discussions of how our results could be influenced by other factors (Lines 331-343).

We disagree with the Reviewer that our hypothesis is a circular argument. We are hypothesizing that invasive species being tested have higher feeding rates than our native species because previous research found that invasive species tend to be more efficient than their native counterparts are. These parameters had not been previously tested and we did not know if *Perna* and

SMITHSONIAN INSTITUTION Smithsonian Marine Station at Fort Pierce 701 Seaway Drive Fort Pierce, FL 34949 *Mytella* were more efficient than *Ischadium*. We have reworded the sentence to clarify our intent (Lines 84-89).

The literature reports all three species do have overlaps in salinity range, a fact we have expanded upon in this version (Lines 66-68, 70-74, 79-81). However, our results suggest that there are salinity preferences, which may allow these species to coexist. We have stated this relationship more clearly (Lines 316-318).

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1 Feeding behavior of the native mussel *Ischadium recurvum* and the invasive mussels *Mytella charruana* 2 and Perna viridis in Florida, USA, across a salinity gradient 3 4 E. Galimany*, J. Lunt, A. Domingos, V. J. Paul 5 Smithsonian Marine Station, 701 Seaway Dr., Fort Pierce, FL 34982, USA 6 7 * galimany@icm.csic.es Phone: +34 93-2309500 8 9 Fax: +34 93-2309555 10 11 12 13 14 **Abstract** 15 The feeding behavior of three species of mussels, the native *Ischadium recurvum* and the 16 17 invasives Mytella charruana and Perna viridis, was studied in an invaded ecosystem in Florida (USA). In 18 situ feeding experiments using the biodeposition method were performed along a salinity gradient in the St. Johns River. Additionally, water characteristics (salinity, temperature, dissolved oxygen, and seston 19 20 loads) were related to the feeding behavior. Clearance, filtration, organic ingestion, and absorption rates 21 of *I. recurvum* were negatively affected by salinity. For *M. charruana*, rejection proportion was positively 22 related to salinity while total ingestion, organic ingestion, and absorption rates were positively related to 23 the percentage of organic matter in the seston. Total and organic ingestion rates of P. viridis were 24 negatively affected by salinity but positively affected by total particulate matter. Condition indices for P. 25 viridis and M. charruana were 13.16 ± 0.64 and 6.63 ± 0.43 , respectively, compared to 4.82 ± 0.41 for the 26 native species *I. recurvum*, indicating that these mussels are well adapted to the environmental conditions 27 in the area. This study indicates that the three species thrive in different water characteristics. Thus, the 28 invasive mussels will not totally occupy the niche of the native mussel in Florida despite overlapping 29 distributions. 30 31 **Key words**: feeding behavior, bivalve, seston, salinity, St. Johns River. 32 33 Introduction 34

Mussels are considered keystone species in ecosystems, contributing to the stabilization of

benthic or intertidal habitats by forming large beds (Meadows et al. 1998). They also have important

ecological roles such as maintaining water quality and participating in the cycling of nutrients from the water column to the benthos (Prins and Smaal 1994; Ostroumov 2005). These benefits are primarily a result of their filter feeding capacity. Mussels filter water through their gills in order to obtain their food source, suspended organic matter mainly in the form of phytoplankton (Gosling 1992). The amount of water filtered may vary according to the amount of suspended particles found in the water. For example, mussels may decrease their clearance rate (i.e.: circulate less water through their gills) when particle concentration is high in the water (Widdows et al. 1979; Babarro et al. 2000). Mussels may also deal with high particle concentrations by selecting the types of particles they want to ingest and rejecting the undesired ones in the form of pseudofeces (Shumway et al. 1985; Ward and Shumway 2004). Understanding the feeding preferences of mussels is of ecological importance, as it may determine the success and competitive advantages of non-native species.

As species arrive in new locations they have the potential to affect terrestrial and aquatic ecosystems on a global scale (Simberloff et al. 2013). The success of species in their new habitat will depend on environmental conditions in relation to their characteristics and preferences (Leung and Mandrak 2007). If individuals succeed in reaching a new site, most will perish from a multitude of physical or biotic agents (Mack 1995). If individuals persist in the new habitat, they may increase in abundance and spread, potentially causing detrimental environmental effects (Elton 1958; Mack et al. 2000). Coastal systems are some of the most highly invaded ecosystems on earth, often as a result of human activities (Grosholz 2002); ships (ballast water, hull fouling), aquaculture imports, and live trade are vectors typically responsible for introducing aquatic invertebrate species (Ruiz and Carlton 2003). Moreover, hard artificial structures in the marine environment may be acting as 'stepping stones' between regions that facilitate the spread of invasive species (Airoldi et al. 2015).

The east coast of Florida, including the St. Johns River and the Indian River Lagoon, has populations of many non-native species including two mussel species. The charru mussel, *Mytella charruana*, is native to Central and South America with a distribution on the Pacific coast from Mexico to Ecuador, including the Galapagos Islands (Keen 1971; Carlton 1992; Szefer et al. 1998; Boehs et al. 2004) and on the south Atlantic coast from Argentina to Uruguay (Lee 1987). The charru mussel was first reported in Florida from a population in an intake valve at the Jacksonville power plant (Lee 1987). However, it was considered extirpated by a freeze in 1987. Presently, the charru mussel is found along the southeastern coast of the US from central Florida to South Carolina (Spinuzzi et al. 2013). This species has a wide salinity range ranging from 2 to 31 ppt with 100% mortality at 0 ppt and 45 ppt (Yuan et al. 2010). The green mussel, *Perna viridis*, is an economically important marine bivalve in the Pacific, native to the tropical Indo-Pacific (Siddall 1980; Vakily 1989; Baker et al. 2007), but has been introduced to Asian and Central American countries and to North America (Baker et al. 2007). The preferred salinity of

P. viridis is 27 to 33 ppt, though it is found in salinities from 19 - 44 ppt (Siddall 1980; Vakily 1989). However, the species is fairly tolerant of large fluctuations in salinity, surviving in the laboratory at salinities as low as 5 ppt (Yuan et al 2016), though salinities lower than ~20 ppt will affect survival (Siddall 1980; Vakily 1989). Within the United States, the invasive range of *P. viridis* is spreading along the Florida coast and northward towards South Carolina (Spinuzzi et al. 2013). These regions are characterized by sedimentary habitats, and the spread of this species has been facilitated by artificial structures (Firth et al. 2016).

Ischadium recurvum is an abundant, intertidal estuarine species found on oyster reefs and other hard substrates in the southeastern United States (Bahr 1981). The hooked mussel is tolerant of a wide salinity range, and has been reported in the field at salinities of 2.5-40 ppt; however, increased mortality is found at salinities lower than 5ppt (Parker 1959, Allen 1960, Castagna and Chanley 1973). Using in situ feeding assays, we assessed the feeding behavior of *I. recurvum* and the two invasive mussel species (Mytella charruana and Perna viridis) under natural conditions across invaded sites within the St. Johns River. We sought to determine the potential competition among these species by comparing feeding behavior throughout their salinity ranges. We hypothesized that the invasive mussels would have higher feeding rates because successful invaders often have more general and/or efficient feeding habits than native species. Moreover, we related the feeding behavior with water characteristics (e.g. salinity and seston content) in multiple sites along a salinity gradient to elucidate potential distribution of the invasive and native mussels and determine any overlapping distributions. This field study is of importance for predicting future potentially invaded areas and warning of possible ecosystem disruption by the invasive mussels.

Materials and Methods

Location and animal collection

Three mussel species were studied: two invasive species, the charru mussel (*Mytella charruana* (d'Orbigny, 1842)) and the green mussel (*Perna viridis* (Linnaeus, 1758)), and the native hooked mussel (*Ischadium recurvum* (Rafinesque, 1820)) (Online Resource 1). Filter feeding experiments were performed during four consecutive days in a different location each day along the St. Johns River in Jacksonville, Florida, during May 2016 (Fig. 1). The sites, Metropolitan Park Marina (30.319, -81.642); Lonnie Wurn Boat Ramp (30.375, -81.585); Browns Creek fish camp (30.417, -81.533); and Mayport Boat Ramp (30.396, -81.428), were chosen to relate the feeding behavior of the three species to different environmental parameters and water characteristics, e.g. salinity, and total and organic matter in the water column. Each

experiment contained six adult mussels of each species (average shell lengths \pm SE: 34.8 \pm 1.4 mm for *I. recurvum*, 32.5 \pm 1.0 mm for *M. charruana*, and 44.4 \pm 1.8 mm for *P. viridis*).

All mussels were collected in Jacksonville, Florida, at two locations. *I. recurvum* and *M. charruana* were collected from the floating dock at Lonnie Wurn Boat Ramp, whereas *P. viridis* individuals were collected from the jetty at Huguenot Memorial Park (Fig. 1). All mussels were cleaned of epiphytes and other encrusting organisms and hung in mesh bags at each site at least two days before experiments to allow them to acclimate to environmental conditions. Controls were made by removing the mussels' meat and gluing their respective shells together (two controls per experiment).

Water characteristics

Physical water characteristics (temperature (°C), salinity (ppt), and dissolved oxygen (mg L⁻¹)) were measured with a YSI meter at the beginning and end of each experiment.

To determine characteristics of the seston, between 80 and 140 ml (enough to clog the filters) of St. Johns River water were collected from the control filter-feeding chambers (explained below) every 15 minutes for 2 hours (n = 7 for each experiment). In the laboratory, all filters were dried at 60 °C for 48 hours and weighed to obtain the total particulate matter (TPM). The filters were then ashed at 450 °C for 4 hours to obtain the particulate inorganic matter (PIM). The particulate organic matter (POM) was calculated as the weight loss between TPM and PIM. Percentage of organic matter in the water was calculated as (POM / TPM) x 100.

Physiological feeding parameters

Two portable, filter feeding, flow-through devices were designed to simulate *in situ* conditions of bivalve feeding (Fig. 2). The devices were slightly modified from Galimany et al. (2011). Each device consisted of a common PVC tank (66 cm length x 30.5 cm width x 14.5 cm height) that received water from an underwater pump. Aeration in the common tank prevented particle settlement. The common tank connected to the 10 PVC chambers (18.5 cm length x 5.7 cm width x 5.7 cm height) through individual rubber tubes. Each chamber contained a single live mussel, except for the control. Individual mussels were positioned near the flow exit tube of the chambers and were attached to the bottom with a piece of plastic hook and loop fastener to avoid movement. The flow of water was maintained at a constant rate of 12 L h⁻¹.

Six mussels (each one considered a replicate) of each species and two controls were placed in the individual chambers of the feeding devices (three individuals of each species and one control in each

feeding device). All experimental mussels were allowed to recover from any stress associated with handling for at least one hour. The individual chambers were cleaned before the beginning of the experiment to remove biodeposits created during the recovery time and any silt that may have accumulated. In each chamber of the feeding devices, feces and pseudofeces were collected with a pipette as soon as they were produced, and kept separately throughout the 2h experimental collection time. All samples of feces and pseudofeces were filtered separately by chamber through preweighed Whatman GF/C filters (25 mm Ø) and rinsed with ammonium formate to dissolve salt from the samples on the filters. In the laboratory, all filters were dried at 60 °C for 48 hours and weighed to determine dry weights of feces and pseudofeces. Then, they were ashed at 450 °C for 4 hours to obtain the ash weight (inorganic matter). The organic matter (ash free dry weight) contained in the feces and pseudofeces was calculated as the difference between dry weight and ash weight.

The physiological parameters of the feeding behavior of the mussels (Table 1) were then calculated according to the biodeposition method (Iglesias et al. 1998). This method is based on using the inorganic matter of the water as a tracer of the ingestion, rejection, and egestion feeding processes. The ingestion processes are those related to the intake of food. Rejection processes are related to the elimination of material captured by the gills but not ingested, whereas the egestion processes are those related to the elimination of undigested matter. All the feeding parameters were then standardized to 1 g of dried bivalve flesh using the following equation:

$$Y_s = Y_e x (1/W_e)^b$$

 Y_s is the standardized physiological rate, Y_e is the experimentally determined rate, and W_e is the dry body mass measured for each bivalve. We used a b value of 0.67, as commonly used in mussel feeding studies (Bayne et al. 1989; Bayne et al. 1993; Hawkins et al. 1997).

To determine when to start feces and pseudofeces collection we calculated the gut transit time (GTT) of all mussel species before each experiment. GTT was calculated using a method adapted from Hawkins et al. (1996). Two mussels of each species were placed individually in beakers in a mixture of water and nutritious phytoplankton (*Tetraselmis* sp.) monoculture. The elapsed time between the ingestion of the mixture and the deposition of green colored feces was considered to be the GTT (min).

Condition index

After the feeding behavior determinations, the length of each mussel was measured by an electronic caliper. The dry weight of mussel tissue calculated to standardize the feeding behavior was also used to estimate the condition index (CI) following the equation from Martin et al. (1984):

 $CI = (tissue dry weight (g) \times 100) / shell length (cm)$

Statistical analyses

Seston and St. Johns River water characteristics at each site were compared using a one-way ANOVA. Characteristics compared were temperature, salinity, dissolved oxygen, TPM, POM, PIM and proportion of organic matter of the river water. Proportion of water organic content was arcsine square root transformed prior to analysis. A Tukey's post hoc test determined statistical differences between the different sites.

The feeding behavior parameters studied in the four locations were analyzed using a two-way ANOVA with species and site as factors. Rejection percentage and selection efficiency were arcsine square root transformed prior to analysis. Two-way ANOVA showed significant species x site interactions for most feeding parameters; therefore, to elucidate differences in the feeding behavior among the mussels, the feeding rates were analyzed within each site with a one-way ANOVA comparing the different species. A Tukey's post hoc test determined statistical differences between the species. Non-linear exponential regression analyses were used to relate the water characteristics (salinity, TPM, and proportion of organic matter), with the physiological feeding variables, including clearance rate, filtration rate, rejection percentage, inorganic rejection rate, total ingestion rate, organic ingestion rate, and absorption rate, for each species. Salinity, TPM, and proportion of organic matter were previously checked for multicollinearity and found that they were not correlated ($R^2 < 0.9$). Correlations were examined between rejection parameters (rejection percentage and inorganic rejection rate), and ingestion rates (clearance rate, filtration rate, total ingestion rate, organic ingestion rate, and absorption rate) for each species to further understand the interaction between ingestion and rejection processes.

Condition index was compared between sites and species using a two-way ANOVA. A Tukey's post hoc test determined statistical differences between the species.

The statistical software used was SPSS Statistics 23.0 (Armonk, NY: IBM Corp.).

Results

Water characteristics

Seston characteristics were significantly different among sites (TPM: $F_{3,40} = 49.00$, p < 0.001; POM: $F_{3,40} = 51.37$, p < 0.001; PIM: $F_{3,40} = 48.11$, p < 0.001; Proportion of organic matter: $F_{3,40} = 53.52$, p < 0.001) (Fig. 3a; Online Resource 2). A Tukey's post hoc test showed that Lonnie Wurn had the most TPM, POM and PIM, whereas Metropolitan Park Marina had the lowest TPM and PIM (Fig. 3a).

Temperature and dissolved oxygen were not different among sites ($F_{3,4} = 5.17$, p = 0.073; $F_{3,4} = 0.26$, p = 0.854, respectively) (Online Resource 2). On average (\pm SE), temperatures were 24.58 ± 0.40 °C and dissolved oxygen was 7.42 ± 0.18 mg L⁻¹. In contrast, salinity and percentage of organic matter in the water were different among sites ($F_{3,4} = 96.61$, p < 0.001 and $F_{3,40} = 53.52$, p < 0.001, respectively). Salinity was the highest at the mouth of the river (Browns Creek and Mayport) and the lowest in Metropolitan Park Marina (Fig. 3b; Online Resource 2). Inversely, the percentage of organic matter was highest in Metropolitan Park Marina, whereas Mayport, the river mouth, showed the lowest values.

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Physiological feeding parameters

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There were significant effects of species and site on feeding as well as significant interactions between species and site. The species x site interaction was significant for all feeding parameters except selection efficiency (clearance rate, $F_{5,44} = 7.31$, p < 0.001; filtration rate, $F_{5,44} = 7.27$, p < 0.001; rejection percentage, $F_{5,44} = 7.02$, p < 0.001; inorganic rejection rate, $F_{5,44} = 7.15$, p < 0.001; total ingestion rate, $F_{5,44} = 2.81$, p = 0.028; organic ingestion rate, $F_{5,44} = 5.72$, p < 0.001; absorption rate, $F_{5,44} = 7.22$, p < 0.001) (Online Resource 3). Selection efficiency differed among sites but not among species (Online Resource 3). Results plotting the one-way ANOVA tests are shown in two separated graphs (Fig. 4 and Fig. 5) to better read the values on the y-axis. One-way ANOVA showed no significant differences among the mussels for any feeding parameters at Metropolitan Park Marina (Fig. 4 and Fig. 5, Online Resource 4). At Lonnie Wurn Boat Ramp, the rejection percentage was the only parameter that showed statistical differences ($F_{2,12} = 6.50$, p = 0.012) (Online Resource 4); M. charruana had the lowest value whereas P. viridis had the highest. At Browns Creek, all feeding parameters showed significant differences except for rejection percentage (Online Resource 4). M. charruana had the highest clearance rate $(F_{2,13} = 17.72, p < 0.001)$, filtration rate $(F_{2,13} = 15.10, p < 0.001)$, inorganic rejection rate $(F_{2,13} = 9.84, p < 0.001)$ p = 0.002), and absorption rate ($F_{2.13} = 13.42$, p = 0.001). The total ingestion rate was the lowest for I. recurvum ($F_{2,13} = 13.24$, p = 0.001), whereas the organic ingestion rate was significantly different for the three species ($F_{2,13} = 17.61$, p < 0.001) with *I. recurvum* having the lowest value and *P. viridis* the highest. At Mayport, the three species differed in clearance rate ($F_{2,10} = 5.87$, p = 0.021), filtration rate ($F_{2,10} = 5.87$, p = 0.021), filtration rate ($F_{2,10} = 5.87$, p = 0.021), 5.90, p = 0.020), total ingestion rate ($F_{2,10}$ = 12.24, p = 0.002), and organic ingestion rate ($F_{2,10}$ = 9.24, p = 0.005) (Online Resource 4). In all cases, *P. viridis* had the highest values.

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Relationship between feeding parameters and water characteristics

The relationship between feeding parameters and water characteristics (e.g. salinity, TPM, and proportion of organic matter; Table 2) was different for each of the three mussels. *I. recurvum* had a negative relationship between salinity and clearance rate (F = 14.06, p = 0.002), filtration rate (F = 6.02, p = 0.028), organic ingestion rate (F = 6.98, p = 0.019), and absorption rate (F = 5.12, p = 0.040). For the invasive mussel *M. charruana*, rejection percentage (F = 7.58, p = 0.013) and inorganic rejection rate (F = 5.62, p = 0.029) were positively related to salinity. Moreover, a positive relationship was observed for this same species between total ingestion rate (F = 5.82, p = 0.026), organic ingestion rate (F = 10.15, p = 0.005) and absorption rate (F = 13.13, p = 0.002) and the proportion of organic matter in the water. The invasive mussel *P. viridis* had two different water characteristics related to feeding parameters: total and organic ingestion rates were negatively related to salinity (F = 10.76, p = 0.005; F = 13.23, p = 0.002, respectively) and positively related to total particulate matter (F = 22.70, p < 0.001; F = 11.67, p = 0.004, respectively).

Correlations between feeding parameters

The statistically significant values for the correlations established between rejection percentage and inorganic rejection rate, and the feeding variables (clearance rate, filtration rate, total ingestion rate, organic ingestion rate, and absorption rate) for each species are shown in Table 3. There was a positive relationship between all feeding variables and inorganic rejection rate ($p \le 0.005$) for the native mussel *I. recurvum*; however, none of the feeding variables was related to rejection percentage. For *M. charruana*, total ingestion rate was negatively correlated with rejection percentage (p = 0.032). Additionally, the inorganic rejection rate of *M. charruana* was correlated (p < 0.001) to all ingestion-related variables except for total ingestion rate. The rejection percentage of the invasive *P. viridis* was positively correlated with clearance, filtration, and absorption rates (p < 0.05). All feeding parameters were positively correlated to inorganic rejection rate for *P. viridis* ($p \le 0.01$) (Table 3).

Condition Index

Condition index was different among species ($F_{2,49} = 56.47$, p < 0.001) (Fig. 6). A Tukey's post hoc test indicated that the native mussel had the lowest condition index and the invasive *P. viridis* had the highest. The condition index was not different among sites ($F_{3,49} = 2.32$, p = 0.087), and there was no significant species x site interaction ($F_{5,49} = 0.11$, p = 0.99) (Online Resource 5).

Discussion

The invasive mussel species (*M. charruana* and *P. viridis*) found on the east coast of Florida have a similar feeding performance as the native mussels (*I. recurvum*) and salinity was the main driver of the feeding behavior. *I. recurvum* preferred low salinity, and salinity related to most of the measured feeding parameters. The invasive species preferred high loads of total particulate matter and proportion of organic matter, while salinity was only related to some of their feeding parameters. Understanding feeding behavior and preferred environmental conditions can help to predict invasion potential. Moreover, this understanding may elucidate the role and effects of filter feeding in the ecosystem. The different preferences observed in this study indicate feeding overlap among the species but also limits on their distributions. Thus, *M. charruana* and *P. viridis* represent a potential threat for native mussels in some areas but not others.

Water characteristics may affect the feeding physiology of bivalves by decreasing their feeding under stressful environmental conditions (Gosling 2003). As the invasive mussels were found within the native habitat of *I. recurvum*, it is evident that the water characteristics, i.e. temperature, salinity and dissolved oxygen, were well within the tolerance limits of the three species. However, salinity was one of the most important environmental factors influencing the feeding behavior of the three studied species. When salinity decreased to 8.7 ppt, *P. viridis* stopped feeding and did not produce any biodeposits. Salinities under 19 ppt are stressful for this species, though survival has been observed at salinities as low as 5 ppt in the laboratory (Siddall 1980; Vakily 1989). Similarly, the invasive Indo-Pacific mussel Brachidontes pharaonic in the Mediterranean Sea decreased its clearance rate when salinity and temperature reached their lower thresholds (Sarà et al. 2008). However, salinity was negatively related to total and organic ingestion rates indicating that the highest salinities are not ideal for *P. viridis* either. Salinity was also inversely related to the feeding behavior of *I. recurvum*. Under laboratory conditions and salinity at 10ppt, I. recurvum has been reported to have maximum clearance rates of 4.6 L h⁻¹ at 26°C and minimum clearance rates, close to 0, below 10°C (Gedan et al. 2014). Conversely, higher salinities enhanced M. charruana feeding. These data indicate that while these species can exploit a wide range of salinities their preferred ranges may allow for niche partitioning that precludes the native species from being eliminated.

Bivalve feeding behavior is also affected by the seston in the water as bivalves regulate their feeding according to seston concentrations (Hawkins et al. 1998a; Ward and Shumway 2004). In our study, *I. recurvum* was not affected by the seston levels, while *P. viridis* and *M. charruana* were. Clearance rate for *P. viridis* was the highest compared to the other species at Mayport, where the percentage of organic matter in the water was the lowest. This is in keeping with previous research demonstrating that *P. viridis* had clearance rates above average when the organic particulate matter was <

1 mg L⁻¹ (Hawkins et al. 1998b). For *M. charruana*, higher organic matter led to higher total, organic ingestion, and absorption rates, suggesting that this species thrives in high organic environments. These strategies maximize the food resources and suggest that both invasive species can tolerate high seston loads. Another way for bivalves to enhance food resources is through pre-ingestive organic selection. Mussels may preferentially reject inorganic particles in the form of pseudofeces to increase the amount of organic matter ingested and promote higher absorption (Bayne et al. 1993; Newell and Shumway 1993; Hawkins et al. 1996). The invasive mussels were also more efficient at pseudofeces production, leading to *I. recurvum* having lower organic ingestion. The overlap in feeding behaviors and increased efficiency of the invasive mussels suggest interspecific competition for food resources is possible; however, the three species have different preferences, which may allow them to coexist within separate microhabitats.

Condition indices (CI) among mussel species can be compared at similar gonadal stages, as the gonad develops within the mantle tissue and increases tissue weight (Gosling 1992). Though we did not study their gonadal cycles, the three mussel species had very well developed mantles when dissected, indicating gonadal maturation. In fact, both invasive mussels have been reported to reproduce the whole year round in Florida (Stenyakina et al. 2010; McFarland et al. 2016). Therefore, the differences found in the CI of the mussels are most likely an indication that the invasive mussels seem to be well adapted to the environmental conditions in the area. Moreover, invasive bivalves have been reported to have a wider tolerance range and higher survival rates under stressful environmental situations (Lenz et al. 2011), which is one reason why invasive bivalves succeed in new environments.

Understanding the distribution of invasive species and identifying potential invaded areas are essential for conservation and management purposes. The invasive mussels in Florida do not seem to be predominant in the ecosystems, but there is no official record of their abundance nor predictions on their invasive potential. Our study indicates that in the St. Johns River the invasive mussels do not share the same environmental preferences with the native *I. recurvum*; thus, the species do not fully overlap. Within the St. Johns River, salinity seems to be the main water characteristic influencing the feeding responses. This is surprising since they are all found in similar salinity ranges. However, salinity is not the only important factor in determining a species range. For example, temperature can alter salinity tolerance. *M. charruana* can survive salinities 5-44 ppt at 20°C, but their salinity range narrows considerably when the temperature fluctuates (Yuan et al 2016). In this study, we focused on comparing behaviors along a salinity gradient at similar temperatures. This was to investigate how these species might compete within local systems and determine if *I. recurvum* could potentially be outcompeted by these recent invaders. While the species do overlap, different preferences in salinity and water characteristics indicate that the three species may coexist by habitat partitioning within ecosystems. However, the efficiency, and wide

342 ranges tolerated by M. charruana and P. viridis represent a potential threat to other, untested native filter 343 feeders. 344 345 Acknowledgements 346 347 We would like to thank St. Johns River Water Management District for funding the project (Contract 348 27799) and the staff of the Smithsonian Marine Station for logistical support. We are very thankful to Dr. 349 Yvette Garner and the students from the University of West Georgia for their help making this project 350 happen. We also want to acknowledge everyone who allowed us to work in Jacksonville: Mr. Jim Suber, the dock master of the city of Jacksonville, Browns Creek Fish Camp, and Morningstar Marinas. We 351 thank Dr. Quinton White from the Jacksonville University for letting us use his laboratory. We appreciate 352 353 the reviewers and the co-editor in chief for improving this article with their suggestions. This is 354 Smithsonian Marine Station contribution #xxx. 355 356 References 357 Airoldi, L., X. Turon, S. Perkol-Finkel, and M. Rius. 2015. Corridors for aliens but not for natives: effects 358 359 of marine urban sprawl at a regional scale. Diversity and Distributions 21: 755-768. 360 Babarro, J.M.F., M.J. Fernández-Reiriz, and U. Labarta. 2000. Feeding behavior of seed mussel Mytilus 361 gallorprovincialis: environmental parameters and seed origin. Journal of Shellfish Research 19: 362 195-201. Baker, P., J.S. Fajans, W.S. Arnold, D.A. Ingrao, D.C. Marelli, and S.M. Baker. 2007. Range and 363 364 dispersal of a tropical marine invader, the Asian green mussel, Perna viridis, in subtropical 365 waters of the southeastern United States. Journal of Shellfish Research 26: 345–355. 366 Bayne, B.L., A.J.S. Hawkins, E. Navarro, and J.I.P. Iglesias. 1989. Effects of seston concentration on feeding, digestion and growth in the mussel, Mytilus edulis. Marine Ecology Progress Series 55: 367 368 47-54. 369 Bayne, B.L., J.I.P. Iglesias, A.J.S. Hawkins, E. Navarro, M. Heral, and J.M. Deslous Paoli. 1993. Feeding 370 behaviour of the mussel, Mytilus edulis: responses to variations in quantity and organic content of 371 the seston. Journal of the Marine Biological Association of the United Kingdom 73: 813–829. Boehs, G., T. Absher, and A. Da Cruz-Kaled. 2004. Composition and distribution of benthic mollusks on 372 373 intertidal flats of Paranagua Bay (Parana, Brazil). Scientia Marina 68: 537-543. 374 Carlton, J.T. 1992. Introduced marine and estuarine mollusks of North America: an end-of-the-20th-375 century perspective. Journal of Shellfish Research 11: 489–505.

- Elton, C.S. 1958. *The ecology of invasions by animals and plants*. London, UK.
- Firth, L.B., A.M. Knights, D. Bridger, A.J. Evans, N. Mieszkowska, P.J. Moore, N.E. O'Connor, E.V.
- 378 Sheehan, R.C. Thompson, and S.J. Hawkins. 2016. Ocean sprawl: challenges and opportunities
- for biodiversity management in a changing world. *Oceanography and Marine Biology: An*
- 380 *Annual Review* 54: 193–269.
- Galimany, E., M. Ramón, and I. Ibarrola. 2011. Feeding behavior of the mussel *Mytilus galloprovincialis*
- 382 (L.) in a Mediterranean estuary: A field study. *Aquaculture* 314: 236–243.
- Gedan, K.B., L. Kellogg, and D.L. Breitburg. 2014. Accounting for multiple foundation species in oyster
- reef restoration benefits. *Restoration Ecology* 22: 517–524.
- Gosling, E. 1992. *The mussel Mytilus: ecology, physiology, genetics and culture*. Amsterdam: Elsevier.
- Gosling, E. 2003. *Bivalve molluscs: biology, ecology, and culture*. Oxford (UK): Blackwell Publishing.
- 387 Grosholz, E. 2002. Ecological and evolutionary consequences of coastal invasions. *Trends in Ecology &*
- 388 Evolution 17: 22–27.
- Hawkins, A.J.S., B.L. Bayne, S. Bougrier, M. Héral, J.I.P. Iglesias, E. Navarro, R.F.M. Smith, and M.B.
- 390 Urrutia. 1998a. Some general relationships in comparing the feeding physiology of suspension-
- feeding bivalve molluscs. *Journal of Experimental Marine Biology and Ecology* 219: 87–103.
- Hawkins, A.J.S., R.F.M. Smith, B.L. Bayne, and M. Héral. 1996. Novel observations underlying the fast
- growth of suspension-feeding shellfish in turbid environments: *Mytilus edulis. Marine Ecology*
- 394 *Progress Series* 131: 179–190.
- Hawkins, A.J.S., R.F.M. Smith, S. Bougrier, B.L. Bayne, and M. Héral. 1997. Manipulation of dietary
- conditions for maximal growth in mussels, *Mytilus edulis* L., from the Marennes-Oléron, France.
- 397 *Aquatic Living Resources* 10: 13–22.
- Hawkins, A.J.S., R.F.M. Smith, S.H. Tan, and Z.B. Yasin. 1998b. Suspension-feeding behaviour in
- tropical bivalve molluscs: Perna viridis, Crassostrea belcheri, Crassostrea iradelei, Saccostrea
- 400 cucculata and Pinctada margarifera. Marine Ecology Progress Series 166: 173–185.
- 401 Iglesias, J.I.P., M.B. Urrutia, E. Navarro, and I. Ibarrola. 1998. Measuring feeding and absorption in
- suspension-feeding bivalves: an appraisal of the biodeposition method. *Journal of Experimental*
- 403 *Marine Biology and Ecology* 219: 71–86.
- 404 Keen, A.M. 1971. Sea shells of tropical West America: marine mollusks from Baja California to Peru.
- 405 Stanford, CA: Stanford University Press.
- 406 Lee, H.G. 1987. Immigrant mussel settles in Northside generator. *The Shell-O-Gram* 28: 7–9.
- Lenz, M., B.A.P. DaGama, N.V. Gerner, J. Gobin, F. Gröner, A. Harry, S.R. Jenkins, P. Kraufvelin, C.
- 408 Mummelthei, J. Sareyka, E.A. Xavier, and M. Wahl. 2011. Non-native marine invertebrates are

409	more tolerant towards environmental stress than taxonomically related native species: Results
410	from a globally replicated study. Environmental Research 111: 943-952.
411	Leung, B., and N.E. Mandrak. 2007. The risk of establishment of aquatic invasive species: joining
412	invasibility and propagule pressure. Proceedings of the Royal Societty B 274: 2603-2609.
413	Mack, R.N. 1995. Understanding the processes of weed invasions: the influence of environmental
414	stochasticity. In Weeds in a changing world, ed. C. Stirton, 65-74. Brighton, UK: British Crop
415	Protection Council, Symposium Proceedings No. 64.
416	Mack, R.N., D. Simberloff, W.M. Lonsdale, H. Evans, M. Clout, and F.A. Bazzaz. 2000. Biotic
417	invasions: causes, epidemiology, globalconsequences, and control. Ecological Applications 10:
418	689–710.
419	Martin, M., G. Ichikawa, J. Goetzl, M. de los Reyes, and M.D. Stephenson. 1984. Relationships between
420	physiological stress and trace toxic substances in the bay mussel, Mytilus edulis, from San
421	Francisco bay, California. Marine Environmental Research 11: 91-110.
422	McFarland, K., P. Soudant, F. Jean, and A.K. Volety. 2016. Reproductive strategy of the invasive green
423	mussel may result in increased competition with native fauna in the southeastern United States.
424	Aquatic Invasions 11: 411-423.
425	Meadows, P.S., A. Meadows, F.J.C. West, P.S. Shand, and M.A. Shaikh. 1998. Mussels and mussel beds
426	(Mytilus edulis) as stabilizers of sedimentary environments in the intertidal zone. In Sedimentary
427	Processes in the Intertidal Zone, ed. K.S. Black, D.M. Paterson and A. Cramp, 331–347. London:
428	Geological Society, Special Publications.
429	Newell, R.C., and S.E. Shumway. 1993. Grazing of natural particulates by bivalve molluscs: a spatial and
430	temporal perspective. In Bivalve filter feeders in estuarine and coastal ecosystem processes, ed.
431	R.F. Dame, 85–148. Berlin: Springer-Verlag.
432	Ostroumov, S.A. 2005. Some aspects of water filtering activity of filter-feeders. <i>Hydrobiologia</i> 542: 275–
433	286.
434	Prins, T.C., and A.C. Smaal. 1994. The role of the blue mussel Mytilus edulis in the cycling of nutrients in
435	the Oosterschelde estuary (The Netherlands). <i>Hydrobiologia</i> 282/283: 413–429.
436	Ruiz, G.M., and J.T. Carlton. 2003. Invasive species: Vectors and management strategies. Washington:
437	Island Press.
438	Sarà, G., C. Romano, J. Widdows, and F.J. Staff. 2008. Effect of salinity and temperature on feeding
439	physiology and scope for growth of an invasive species (Brachidontes pharaonis - Mollusca:
440	Bilvalvia) within the Mediterranean sea. Journal of Experimental Marine Biology and Ecology
441	363: 130–136.

442	Shumway, S.E., T.L. Cucci, R.C. Newell, and C.M. Yentsch. 1985. Particle selection, ingestion, and
443	absorption in filter-feeding bivalves. Journal of Experimental Marine Biology and Ecology 91:
444	77–92.
445	Siddall, S.E. 1980. A clarification of the genus Perna (Mytilidae). Bulletin of Marine Science 30: 858–
446	870.
447	Simberloff, D., JL. Martin, P. Genovesi, V. Maris, D.A. Wardle, J. Aronson, F. Courchamp, B. Galil, E.
448	García-Berthou, M. Pascal, P. Pyšek, R. Sousa, E. Tabacchi, and M. Vilà. 2013. Impacts of
449	biological invasions: what's what and the way forward. Trends in Ecology & Evolution 28: 58-
450	66.
451	Spinuzzi, S.S., K.R. Schneider, L.J. Walters, W.S. Yuan, and E.A. Hoffman. 2013. Tracking the
452	distribution of non-native marine invertebrates (Mytella charruana, Perna viridis, and
453	Megabalanus coccopoma) along the southeastern USA. Marine Biodiversity Records 6: 55-67.
454	Stenyakina, A., L.J. Walters, E.A. Hoffman, and C. Calestani. 2010. Food availability and sex reversal in
455	Mytella charruana, an introduced bivalve in the southeastern United States. Molecular
456	Reproduction and Development 77: 222–230.
457	Szefer, P., J. Geldom, A. Ali, F. Páez-Osuna, A. Ruiz-Fernandes, and S. Galvan. 1998. Distribution and
458	association of trace metals in soft tissue and byssus of Mytella strigata and other benthal
459	organisms from Mazatlan Harbour, mangrove lagoon of the northwest coast of Mexico.
460	Environmental International 24: 359–374.
461	Vakily, J.M. 1989. The biology and culture of mussels of the genus <i>Perna</i> . In International Center for
462	living aquatic resources management and Deutsche Gesellschaft flr Technische Zusammenarbeit
463	(GTLZ), Iclarm studies and reviews 17, 63. Manila, Philippines.
464	Ward, J.E., and S.E. Shumway. 2004. Separating the grain from the chaff: particle selection in
465	suspension- and deposit-feeding bivalves. Journal of Experimental Marine Biology and Ecology
466	300: 83–130.
467	Widdows, J., P. Fieth, and C.M. Worrall. 1979. Relationships between seston, available food and feeding
468	activity in the common mussel Mytilus edulis. Marine Biology 50: 195-207.
469	Yuan, W., L.J. Walters, K.R. Schneider, and E.A. Hoffman. 2010. Exploring the survival threshold: A
470	study of salinity tolerance of the nonnative mussel Mytella charruana. Journal of Shellfish
471	Research 29: 415–422.

Fig. 1: Detailed map of the mussel collection and sampling sites. Black arrows indicate mussel collection sites, from left to right: Lonnie Wurn Boat Ramp (collection site for *I. recurvum* and *M. charruana*); Huguenot Memorial Park (collection site for *P. viridis*). White stars indicate sites of feeding experiments, from left to right: Metropolitan Park Marina; Lonnie Wurn Boat Ramp; Browns Creek fish camp; and Mayport Boat Ramp. The top left square image shows the SE of the USA with a black square locating the Jacksonville area.

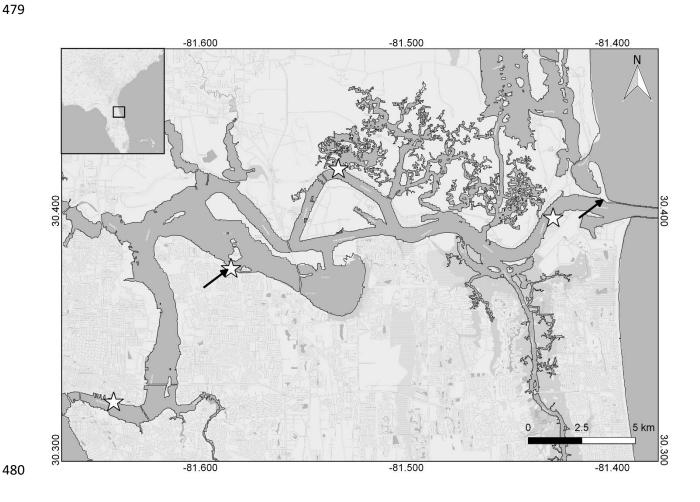


Fig. 2: Image detail of a portable, flow-through device used for the filter feeding experiments. 1: tube transporting water pumped from the environment to the device; 2: PVC "reservoir" tank; 3: plastic tubes with valves to regulate flow connecting the "reservoir" tank with each individual chamber; 4: individual chambers that each hold a single bivalve. White arrows show water flow direction.

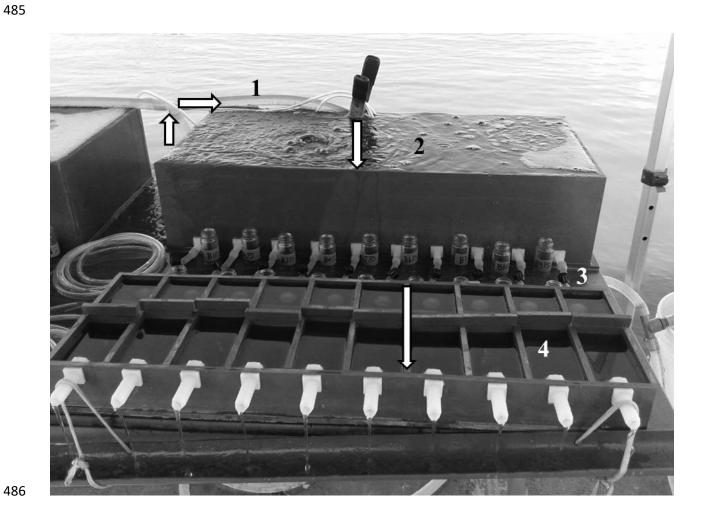
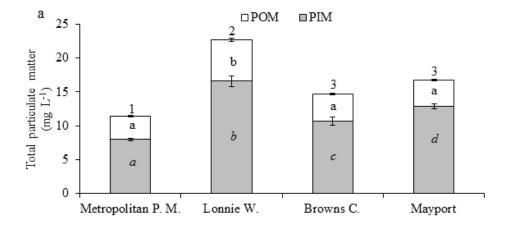


Fig. 3: Average (\pm SE) values for seston and water characteristics at each site. a: Column bars denote values for the total particulate matter (TPM) which are subdivided into particulate organic matter (POM) and particulate inorganic matter (PIM). Error bars on the top of the column are for POM whereas lower errors bars, on top of the grey bar, are for PIM. Numbers denote significant differences for TPM, letters denote significant differences for POM and letters in italics denote significant differences for PIM. b: Percentage of organic matter in the water (Organic) on the left Y axis (black line) and Salinity on the right Y axis (gray line). Letters denote significant differences for organic and letters in italics denote significant differences for salinity.



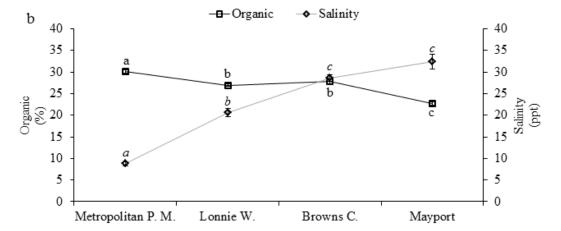


Fig. 4: Results of the one-way ANOVA tests for the feeding parameters clearance rate (CR; L h⁻¹), organic ingestion rate (OIR; mg h⁻¹), and absorption rate (AR; mg h⁻¹). Column bars denote average values (\pm SE) at each sampling site for the three mussel species: *I. recurvum*, *M. charruana*, and *P. viridis*. *P. viridis* bars are absent from the Metropolitan Park Marina site because none of the animals fed. Letters on top of each column denote statistical differences by Tukey's tests (p < 0.05); no letters denote no significant differences.

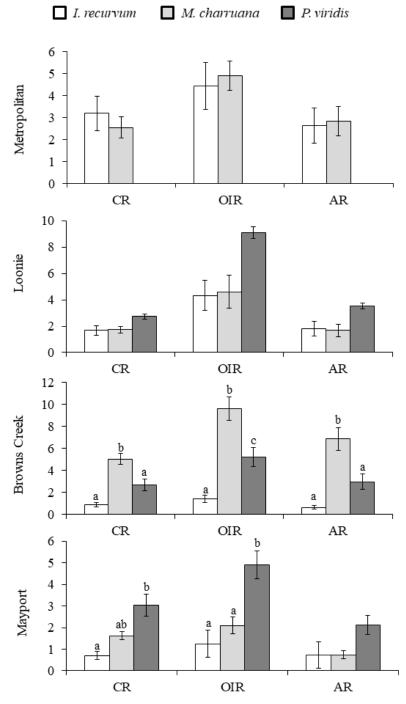


Fig. 5: Results of the one-way ANOVA tests for the feeding parameters filtration rate (FR; mg h⁻¹), rejection percent (RP; %), inorganic rejection rate (IRR; mg h⁻¹), and total ingestion rate (TIR; mg h⁻¹). Column bars denote average values (\pm SE) at each sampling site for the three mussel species: *I. recurvum*, *M. charruana*, and *P. viridis*. *P. viridis* bars are absent from the Metropolitan Park Marina site because none of the animals fed. Letters on top of each column denote statistical differences by Tukey's tests (p < 0.05); no letters denote no significant differences.

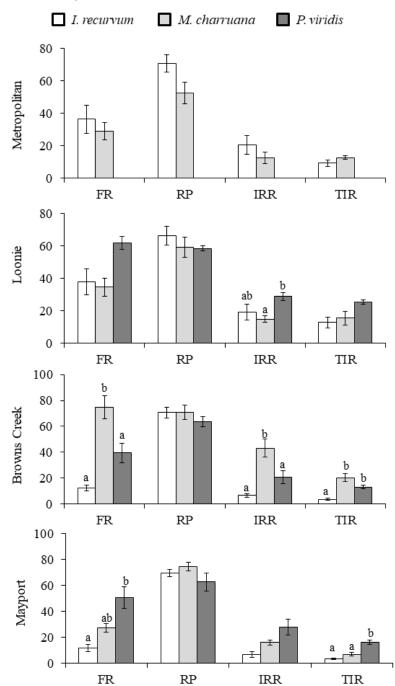


Fig. 6: Average (\pm SE) of the condition index for the different mussel species. Letters denote significant differences by Tukey's test.



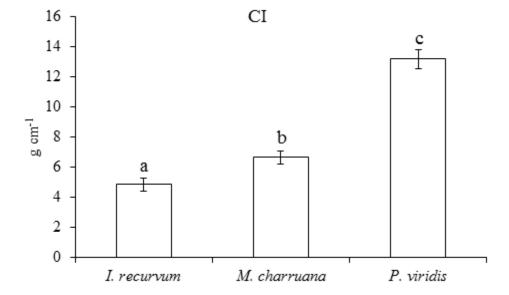


Table 1: Description of the different variables of the feeding behavior of the mussels. TPM: total particulate matter from the water (mg L^{-1}); PIM: particulate inorganic matter from the water (mg L^{-1}); POM: particulate organic matter from the water (mg L^{-1}).

Parameter (units)	Description	Calculation
Clearance rate	Volume of seawater passing through the gills per	(mg inorganic matter from both feces and pseudofeces per unit of
$(L h^{-1})$	unit of time	time) / (PIM water)
Filtration rate	Total particulate matter from the seawater retained	Clearance rate x TPM water
$(mg h^{-1})$	in the gills per unit of time	Clearance rate x 1 F W water
Rejection percentage	Total particulate matter that has been retained in	(mg organic and inorganic matter from pseudofeces per unit of
(%)	the gills but rejected prior to ingestion	time / filtration rate) x 100
Inorganic rejection rate	Particulate inorganic matter that has been retained	mg inorganic matter from pseudofeces per unit of time
$(mg h^{-1})$	in the gills but rejected prior to ingestion	ing morganic matter from pseudoreces per unit of time
Selection efficiency	Pre-ingestive selection of organic matter through	[1 – ((mg organic matter from pseudofeces per unit of time / mg
•	pseudofeces production	organic and inorganic matter from pseudofeces per unit of time) /
(%)	pseudoreces production	(organic matter in the water))] x 100
Total ingestion rate (mg	Total particulate matter retained in the gills and	Filtration rate - mg organic and inorganic matter from
h ⁻¹)	ingested by the bivalve per unit of time	pseudofeces per unit of time
Organic ingestion rate	Particulate organic matter retained in the gills and	(Clearance rate x POM water) – (mg organic matter from
$(mg h^{-1})$	ingested by the bivalve per unit of time	pseudofeces per unit of time)
Absorption rate	Particulate organic matter ingested by the bivalve	Organic ingestion rate - (mg organic matter from feces per unit of
$(mg h^{-1})$	and not egested as feces per unit of time	time)

Table 2: Exponential regression analyses results between the feeding physiological parameters and the water characteristics, i.e. salinity, total particulate matter (TPM), and proportion of organic matter (organic). NC: no correlations.

Feeding variable (acronym)	I. recurvum	M. charruana	P. viridis	
Clearance rate	$CR = 4.47 \ (\pm \ 1.50) \ e^{(-0.06 \pm 0.02) \ x \ salinity}$	NC	NC	
(CR)	$F = 14.06; p = 0.002; R^2 = 0.47$	NC	IVC	
Filtration rate	$FR = 49.71 (\pm 18.51) e^{(-0.04 \pm 0.02) x \text{ salinity}}$	NC	NC	
(FR)	$F = 6.02$; $p = 0.028$; $R^2 = 0.25$	INC	NC	
Rejection percentage	NC	$RP = 0.67 \; (\pm \; 0.07) \; e^{\; (0.01 \; \pm \; 0.01) \; x \; salinity}$	NC	
(RP)	NC	$F = 7.58; p = 0.013; R^2 = 0.25$	NC	
Inorganic rejection rate	NG	IRR = 7.33 (\pm 2.89) e $^{(0.04 \pm 0.02) \text{ x salinity}}$	NG	
(IRR)	NC	$F = 5.62; p = 0.029; R^2 = 0.19$	NC	
_			TIR = 62.55 (\pm 25.08) e (-0.05 \pm 0.01) x salinity	
Total ingestion rate	NG	TIR = 0.04 (± 0.10) $e^{(10.54 \pm 4.37) \text{ x organic}}$	$F = 10.76$; $p = 0.005$; $R^2 = 0.37$	
(TIR)	NC	NC $F = 5.82; p = 0.026; R^2 = 0.19$		
			$F=22.70; p<0.001; R^2=0.56$	
			OIR = $30.29 \ (\pm \ 13.88) \ e^{(-0.06 \pm 0.02) \ x \ salinity}$	
Organic ingestion rate	OIR = 6.53 (\pm 2.71) e (-0.05 \pm 0.02) x salinity	OIR = 0.01 (\pm 0.01) e $^{(14.66\pm4.60)x}$ organic	$F = 13.23; p = 0.002; R^2 = 0.42$	
(OIR)	$F = 6.98$; $p = 0.019$; $R^2 = 0.28$	$F = 10.15; p = 0.005; R^2 = 0.31$	OIR = 1.33 (\pm 0.59) e $^{(0.08 \pm 0.02) \text{ x TPM}}$	
			$F = 11.67$; $p = 0.004$; $R^2 = 0.39$	
Absorption rate	$AR = 3.47 \ (\pm \ 2.08) \ e^{(-0.06 \ \pm \ 0.03) \ x \ salinity}$	$AR = 9.97 \text{ x } 10^{-6} (\pm 0.00) \text{ e}^{(22.57 \pm 6.23) \text{ x organic}}$	NC	
(AR)	$F = 5.12$; $p = 0.040$; $R^2 = 0.22$	$F = 13.13; p = 0.002; R^2 = 0.38$	INC	

Table 3: Pearson correlation (r) values and statistical significance (p) for the relationships between feeding variables for the three mussel species. Pearson r value indicates if the correlation is positive or negative. RP: Rejection percentage arcsine square root transformed for analysis; IRR: Inorganic rejection rate; CR: Clearance rate; FR: Filtration rate; TIR: Total ingestion rate; OIR: Organic ingestion rate; AR: Absorption rate. Asterisks denote significant differences.

		RP			IRR		
		I. recurvum M	1. charruana	P. viridis	I. recurvum N	1. charruana	P. viridis
CR	r	0.334	0.340	0.837*	0.911*	0.926*	0.937*
	p	0.207	0.131	< 0.001	< 0.001	< 0.001	< 0.001
FR	r	0.234	0.267	0.632*	0.977*	0.929*	0.960*
	p	0.383	0.241	0.005	< 0.001	< 0.001	< 0.001
TIR	r	-0.308	-0.468*	0.099	0.668*	0.307	0.590*
	p	0.246	0.032	0.697	0.005	0.176	0.010
OIR	r	-0.040	-0.127	0.229	0.848*	0.700*	0.643*
	p	0.882	0.583	0.360	< 0.001	< 0.001	0.004
AR	r	0.189	0.205	0.501*	0.898*	0.893*	0.712*
	p	0.484	0.373	0.034	< 0.001	< 0.001	0.001

Online Resource 1: Example of each species of mussel used for the feeding study. 1. *Perna viridis*; 2. *Ischadium recurvum*; 3. *Mytella charruana*.



Online Resource 2: One-way ANOVA comparing seston and river water characteristics at each site. TPM: total particulate matter; POM: particulate organic matter; PIM: particulate inorganic matter; Organic: proportion of organic matter arcsine square root transformed for analysis; DO: dissolved oxygen.

		Sum of Squares	df	Mean Square	F	Sig.
TPM	Between Groups	674.313	3	224.771	49.005	< 0.001
	Within Groups	183.466	40	4.587		
	Total	857.779	43			
POM	Between Groups	43.981	3	14.660	51.367	< 0.001
	Within Groups	11.416	40	0.285		
	Total	55.397	43			
PIM	Between Groups	401.710	3	133.903	48.115	< 0.001
	Within Groups	111.321	40	2.783		
	Total	513.031	43			
Organic	Between Groups	0.045	3	0.015	53.524	< 0.001
	Within Groups	0.011	40	0.000		
	Total	0.057	43			
Temperature	Between Groups	7.245	3	2.415	5.166	0.073
	Within Groups	1.870	4	0.468		
	Total	9.115	7			
Salinity	Between Groups	654.355	3	218.118	96.608	< 0.001
	Within Groups	9.031	4	2.258		
	Total	663.386	7			
DO	Between Groups	0.307	3	0.102	0.256	0.854
	Within Groups	1.600	4	0.400		
	Total	1.907	7			

Online Resource 3: Two-way ANOVA table for the feeding behavior with site and species as factors. CR: Clearance rate; FR: Filtration rate; RP: Rejection percentage arcsine square root transformed for analysis; IRR: Inorganic rejection rate; SE: Selection efficiency arcsine square root transformed for analysis; TIR: Total ingestion rate; OIR: Organic ingestion rate; AR: Absorption rate.

Source	Dependent Variable	Type III Sum of Squares	df	Mean Square	F	Sig.
Corrected Model	CR	70.127 ^a	10	7.013	5.718	< 0.001
	FR	16813.399 ^b	10	1681.340	5.905	< 0.001
	RP	8616.422°	10	861.642	4.576	< 0.001
	IRR	5616.841 ^d	10	561.684	4.734	< 0.001
	SE	0.323 ^e	10	0.032	2.442	0.020
	TIR	2400.766 ^f	10	240.077	6.389	< 0.001
	OIR	359.013 ^g	10	35.901	7.302	< 0.001
	AR	158.538 ^h	10	15.854	6.989	< 0.001
Intercept	CR	303.796	1	303.796	247.719	< 0.001
	FR	78157.777	1	78157.777	274.494	< 0.001
	RP	33242.380	1	33242.380	176.558	< 0.001
	IRR	20549.889	1	20549.889	173.184	< 0.001
	SE	8.940	1	8.940	676.863	< 0.001
	TIR	9456.044	1	9456.044	251.637	< 0.001
	OIR	1287.988	1	1287.988	261.977	< 0.001
	AR	328.399	1	328.399	144.763	< 0.001
Site	CR	13.747	3	4.582	3.736	0.018
	FR	1788.232	3	596.077	2.093	0.115
	RP	498.418	3	166.139	0.882	0.458
	IRR	296.808	3	98.936	0.834	0.483
	SE	0.145	3	0.048	3.653	0.019
	TIR	842.159	3	280.720	7.470	< 0.001
	OIR	92.474	3	30.825	6.270	0.001
	AR	33.473	3	11.158	4.918	0.005
Species	CR	15.708	2	7.854	6.404	0.004
	FR	4722.271	2	2361.135	8.292	0.001
	RP	1559.971	2	779.986	4.143	0.022
	IRR	1036.605	2	518.302	4.368	0.019
	SE	0.055	2	0.028	2.096	0.135
	_ TIR	862.969	2	431.485	11.482	< 0.001

ļ	OIR	102.593	2	51.297	10.434	< 0.001
	AR	27.221	2	13.610	6.000	0.005
Site x Species	CR	44.818	5	8.964	7.309	< 0.001
	FR	10347.166	5	2069.433	7.268	< 0.001
	RP	6606.750	5	1321.350	7.018	< 0.001
	IRR	4242.652	5	848.530	7.151	< 0.001
	SE	0.100	5	0.020	1.508	0.207
	TIR	527.566	5	105.513	2.808	0.028
	OIR	140.730	5	28.146	5.725	< 0.001
	AR	81.930	5	16.386	7.223	< 0.001
Error	CR	53.960	44	1.226		
	FR	12528.294	44	284.734		
	RP	8284.309	44	188.280		
	IRR	5221.004	44	118.659		
	SE	0.581	44	0.013		
	TIR	1653.435	44	37.578		
	OIR	216.322	44	4.916		
	AR	99.815	44	2.269		
Total	CR	486.854	55			
	FR	123946.859	55			
	RP	57452.620	55			
	IRR	35952.116	55			
	SE	10.614	55			
	TIR	15333.603	55			
	OIR	2128.132	55			
	AR	663.964	55			
Corrected Total	CR	124.088	54			
	FR	29341.692	54			
	RP	16900.730	54			
	IRR	10837.845	54			
	SE	0.904	54			
	TIR	4054.201	54			1
	OIR	575.335	54			
i e e e e e e e e e e e e e e e e e e e						

a. $R^2 = 0.565$ (Adjusted $R^2 = 0.466$)

b. $R^2 = 0.573$ (Adjusted $R^2 = 0.476$)

c. $R^2 = 0.510$ (Adjusted $R^2 = 0.398$)

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d. R^2 = 0.518 (Adjusted R^2 = 0.409)
e. R^2 = 0.357 (Adjusted R^2 = 0.211)
f. R^2 = 0.592 (Adjusted R^2 = 0.499)
g. R^2 = 0.624 (Adjusted R^2 = 0.539)
h. R^2 = 0.614 (Adjusted R^2 = 0.526)
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Online Resource 4: One-way ANOVA table for the feeding behavior of the three species at the four different studied locations. CR: Clearance rate; FR: Filtration rate; RP: Rejection percentage arcsine square root transformed for analysis; IRR: Inorganic rejection rate; SE: Selection efficiency arcsine square root transformed for analysis; TIR: Total ingestion rate; OIR: Organic ingestion rate; AR: Absorption rate.

Metropolitan Park Marina									
Dependent variable		Sum of Squares	df	Mean Square	F	Sig.			
CR	Between Groups	1.147	1	1.147	0.444	0.522			
	Within Groups	23.264	9	2.585					
	Total	24.411	10						
FR	Between Groups	149.224	1	149.224	0.461	0.514			
	Within Groups	2912.673	9	323.630					
	Total	3061.897	10						
RP	Between Groups	0.103	1	0.103	4.760	0.057			
	Within Groups	0.195	9	0.022					
	Total	0.299	10						
IRR	Between Groups	175.082	1	175.082	1.311	0.282			
	Within Groups	1202.221	9	133.580					
	Total	1377.303	10						
TIR	Between Groups	29.781	1	29.781	1.938	0.197			
	Within Groups	138.326	9	15.370					
	Total	168.106	10						
OIR	Between Groups	0.581	1	0.581	0.123	0.733			
	Within Groups	42.393	9	4.710					

	Total	42.974	10			
AR	Between Groups	0.119	1	0.119	0.038	0.850
	Within Groups	28.051	9	3.117		
	Total	28.170	10			

	Lonnie Wurn Boat Ramp								
Dependen	t variable	Sum of Squares	df	Mean Square	F	Sig.			
CR	Between Groups	3.761	2	1.880	3.817	0.052			
	Within Groups	5.912	12	0.493					
	Total	9.672	14						
FR	Between Groups	1936.367	2	968.183	3.849	0.051			
	Within Groups	3018.602	12	251.550					
	Total	4954.969	14						
RP	Between Groups	0.089	2	0.044	3.263	0.074			
	Within Groups	0.163	12	0.014					
	Total	0.252	14						
IRR	Between Groups	679.191	2	339.596	6.500	0.012			
	Within Groups	626.957	12	52.246					
	Total	1306.148	14						
TIR	Between Groups	390.097	2	195.049	2.251	0.148			
	Within Groups	1039.656	12	86.638					
	Total	1429.753	14						
OIR	Between Groups	55.894	2	27.947	3.503	0.063			
	Within Groups	95.746	12	7.979					

	Total	151.640	14			
AR	Between Groups	7.806	2	3.903	2.977	0.089
	Within Groups	15.732	12	1.311		
	Total	23.538	14			

Browns Creek								
Dependent variable		Sum of Squares df		Mean Square	F	Sig.		
CR	Between Groups	43.517	2	21.759	17.723	< 0.001		
	Within Groups	15.961	13	1.228				
	Total	59.478	15					
FR	Between Groups	9672.955	2	4836.478	15.097	< 0.001		
	Within Groups	4164.747	13	320.365				
	Total	13837.702	15					
RP	Between Groups	0.024	2	0.012	0.853	0.449		
	Within Groups	0.184	13	0.014				
	Total	0.208	15					
IRR	Between Groups	3432.076	2	1716.038	9.845	0.002		
	Within Groups	2266.013	13	174.309				
	Total	5698.089	15					
TIR	Between Groups	672.110	2	336.055	13.238	0.001		
	Within Groups	330.017	13	25.386				
	Total	1002.127	15					
OIR	Between Groups	166.388	2	83.194	17.615	< 0.001		
	Within Groups	61.398	13	4.723				

	Total	227.786	15			
AR	Between Groups	100.458	2	50.229	13.424	0.001
	Within Groups	48.641	13	3.742		
	Total	149.100	15			

Mayport								
Dependent variable		Sum of Squares	df	Mean Square	F	Sig.		
CR	Between Groups	10.352	2	5.176	5.866	0.021		
	Within Groups	8.824	10	0.882				
	Total	19.176	12					
FR	Between Groups	2868.411	2	1434.206	5.897	0.020		
	Within Groups	2432.272	10	243.227				
	Total	5300.683	12					
RP	Between Groups	0.046	2	0.023	1.236	0.331		
	Within Groups	0.185	10	0.019				
	Total	0.231	12					
IRR	Between Groups	794.476	2	397.238	3.528	0.069		
	Within Groups	1125.813	10	112.581				
	Total	1920.289	12					
TIR	Between Groups	356.151	2	178.075	12.244	0.002		
	Within Groups	145.436	10	14.544				
	Total	501.587	12					
OIR	Between Groups	31.017	2	15.509	9.239	0.005		
	Within Groups	16.786	10	1.679				

	Total	47.803	12			
AR	Between Groups	6.330	2	3.165	4.282	0.051
	Within Groups	7.391	10	0.739		
	Total	13.721	12			

Online Resource 5: Two-way ANOVA table for the condition index with site and species as factors.

Source	Type III Sum of Squares	df	Mean Square	F	Sig.
Corrected Model	750.853a	10	75.085	15.194	< 0.001
Intercept	3738.439	1	3738.439	756.522	< 0.001
Site	34.339	3	11.446	2.316	0.087
Species	558.121	2	279.060	56.472	< 0.001
Site x Species	2.715	5	0.543	0.110	0.990
Error	242.139	49	4.942		
Total	4845.009	60			
Corrected Total	992.992	59			

a. $R^2 = 0.756$ (Adjusted $R^2 = 0.706$)