doi: 10.1017/jpa.2015.69



A Middle Pennsylvanian macrofloral assemblage from wetland deposits in Indiana (Illinois Basin): a taxonomic contribution with biostratigraphic, paleobiogeographic, and paleoecologic implications

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Abstract.—Taxonomic analysis is provided for a Middle Pennsylvanian macrofloral assemblage collected from clastic wetland deposits in Clay County, Indiana, on the eastern margin of the Illinois Basin. Adpressed plant fossils were recovered from four distinct beds in the lowermost Staunton Formation, positioned above the Minshall Coal (uppermost Brazil Formation), part of a succession deposited near the Atokan-Desmoinesian boundary. The assemblage of 22 fossil-taxa is dominated by pteridosperms (including Neuropteris flexuosa, Macroneuropteris scheuchzeri, Alethopteris densinervosa, Neuropteris ovata, Eusphenopteris neuropteroides, and Neuropteris missouriensis) with lesser cordaitaleans (Cordaites spp. indet.) and sphenopsids (particularly Sphenophyllum cuneifolium). Lycopsids are uncommon, and ferns are rare. In contrast, the microfloral assemblage from the Minshall Coal and overlying clastic units is dominated by lycopsid and tree fern spores. Comparisons with established biozonation schemes yield different ages depending on the regional biostratigraphic framework used: (1) latest Bolsovian (Radiizonates difformis Biozone, American microfloras); (2) latest Bolsovian or earliest Asturian ('Neuropteris' rarinervis Biozone, Appalachian Basin macrofloras); or (3) earliest Asturian (Linopteris obliqua Biozone, European macrofloras). The placement and correlation of the Bolsovian-Asturian and Atokan-Desmoinesian boundaries, which have traditionally been equated by palynology, are evaluated in the context of this discordance. Several revised stratigraphic scenarios are proposed for this interval in the Illinois Basin, which is being increasingly recognized as a time of significant environmental change throughout Euramerica. Homotaxial comparisons with European macrofloral assemblages indicate that, of the 18 biological taxa recorded, between 14 and 17 (78-94%) also are common in coeval wetland deposits in Europe. The similarities exemplify the spatial conservatism and low diversity of wetland plant communities over vast areas of tropical Euramerica, a manifestation of the intrinsically stressful conditions that characterize such habitats, and indicates that neither the Laurentian Shield nor the Appalachian-Variscan Mountains were an insurmountable barrier to plant dispersal during the Middle Pennsylvanian.

Introduction

American paleobotanists have long been criticized by their European contemporaries for a perceived negligence in documenting Pennsylvanian-aged macrofloral assemblages preserved as adpressions (i.e., compression-impression floras). For example, after a reconnaissance fieldtrip to the USA in 1933, W.J. Jongmans summarized the status of Carboniferous paleobotany in America in an unflattering quote (Jongmans and Gothan, 1934, p. 17, translated from German), "If one considers then how poor most of the American illustrations are, how relatively few works are supplied with illustrations—most papers only contain lists with many names, which provide us no clue on which to base a

judgment,—how confused the nomenclature is, because of parallel efforts by rather than collaboration between the American and European paleobotanists—, then it becomes clear to anyone that any attempt [of comparison] without one's own consideration [of the evidence] will be very difficult if not impossible." Thereafter, Jongmans (1937a, p. 364-366) and Bode (1958, p. 217, 218) expressed discontent about correlating between American and European successions, and pointed to the large size of (and distance between) Pennsylvanian basins in the USA, the use of parochial stratigraphic nomenclature in different states, the reliance of American paleobotanists on outdated literature that did not take European taxa into account, and the scarcity of publications with well-illustrated macrofloras. In particular, it was felt that the lack of consultation with the European literature had caused the introduction of many 'new species', creating the illusion that Pennsylvanian vegetation preserved on either side of the Atlantic Ocean differed more than expected.

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Since the mid-20th century, the breakdown of language barriers, better international access to publications, and collaboration between American and European paleobotanists has seen the resolution of many taxonomic issues, as W.C. Darrah foretold in a footnote in Jongmans (1937b, p. 394): "The number of [P]aleozoic plants with wide distributions in both Europe and America will increase rather than diminish with careful study." Furthermore, publications that involve North American adpression assemblages grew substantially after the 1920s (Pfefferkorn and Gillespie, 1980a). Nonetheless, the idea persists that Pennsylvanian macrofloral assemblages lack documentation in the USA (e.g., Cleal et al., 2009, p. 184), thus hindering comparisons and biostratigraphic correlations of strata across the Euramerican paleocontinent. A task of this paper is to demonstrate that such claims are exaggerated.

This contribution focuses on the description and illustration of a Middle Pennsylvanian (middle Moscovian) macrofloral assemblage collected from wetland deposits above the Minshall Coal (lowermost Staunton Formation) of Indiana, situated on the eastern margin of the Illinois Basin. Plant remains were collected at a single locality from four distinct beds, each containing a compositionally unique suite of fossil-taxa (sensu Cleal and Thomas, 2010), although all specimens are treated as belonging to one macrofloral assemblage to simplify biostratigraphic comparisons. The strata accumulated close to the Atokan-Desmoinesian Stage boundary, which has been correlated with the Bolsovian-Asturian Substage boundary of Western Europe (Peppers, 1996; Blake et al., 2002). The microfloral assemblage from the clastic beds and underlying Minshall Coal (uppermost Brazil Formation) also is tabulated, but only touched on in biostratigraphic context. Comprehensive synonymy lists are compiled for fossil-taxa in the macrofloral assemblage based on comparison with a suite of North American literature, which were singled out because they contain well-illustrated records of Middle Pennsylvanian plant remains, with a focus on the Illinois Basin. Homotaxial biostratigraphic comparisons with American and European biozonation schemes are presented, and uncertainties about the placement and correlation of the Atokan-Desmoinesian and Bolsovian-Asturian boundaries are discussed. Evaluating the congruence of these boundaries is timely because they involve an interval that is being increasingly recognized as one of significant environmental change throughout Euramerica (cf., Cecil et al., 1985; Bertier et al., 2008; Van Hoof et al., 2013). Furthermore, nearly all macrofloral taxa identified also are present in contemporaneous deposits of Europe, and the paleobiogeographic and paleoecologic ramifications of this finding are considered in the context of Pennsylvanian wetland ecosystems in tropical Euramerica.

Geologic setting

Illinois Basin.—The Illinois Basin (also called the Eastern Interior Basin) is an interior cratonic basin (Leighton et al., 1991), one of several large depocenters in the USA that contain coal-bearing strata of predominantly Pennsylvanian age (Fig. 1.1). In its present configuration, the basin underlies much of Illinois and parts of western Indiana and western Kentucky, with small outliers in Iowa and Missouri (Fig. 1.2). The Illinois Basin contains a largely siliciclastic succession of Morrowan to Virgilian (Bashkirian to Gzhelian, upper Namurian to upper Stephanian) strata (Nelson et al., 1991, 2013), which accumulated on a low-gradient slope that dipped toward a shallow epeiric sea (Watney et al., 1989; Greb et al., 2003). Glacioeustatic fluctuations from the west resulted in cyclic marine transgressions across the continental platform (Wanless and Shepard, 1935, 1936; Heckel, 1986; Langenheim and Nelson, 1992; Algeo and Heckel, 2008), and in accord with its paleogeographic position, depositional environments in the Illinois Basin tend to be intermediate between the marine-dominated rocks of the Western Interior (Midcontinent) Basin and mainly deltaic to terrestrial strata of the Appalachian Basin to the east (Fig. 1.1; Heckel, 1977, 1980; Nelson et al., 2013). These basins developed in a paleoequatorial setting in the west-central part of Euramerica (Witzke, 1990), and were separated from eastern parts of the paleocontinent (present-day Europe)

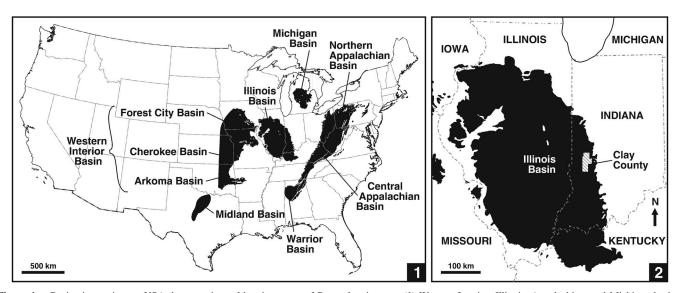


Figure 1. Basins in contiguous USA that contain coal-bearing strata of Pennsylvanian age. (1) Western Interior, Illinois, Appalachian, and Michigan basins, and their components. (2) Illinois Basin, and position of Clay County, Indiana on eastern margin of basin.

by uplands of the Laurentian Shield and, in part, by the Appalachian-Variscan Mountains (Blakey, 2013).

Study locality.—The study area is situated in Clay Country, west-central Indiana, on the eastern margin of the Illinois Basin (Fig. 1.1), where the thin and laterally discontinuous Lower Block, Upper Block, and Minshall coals of the upper Atokan Brazil Formation (Fig. 2) have been exploited due to their low to medium sulfur and ash content (Mastalerz et al., 2000, 2003; Kvale et al., 2004). The Brazil Formation, defined as being from the base of the Lower Block Coal to the top of the Minshall Coal (Hutchison, 1976), is equivalent to the middle part of the Tradewater Formation of Illinois and Kentucky (The Tri-State Committee on Correlation of the Pennsylvanian System in the Illinois Basin, 2001). In Indiana, strata above the Minshall Coal are assigned to the Staunton Formation, which includes the

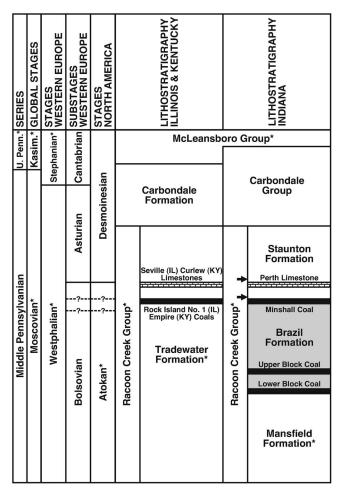


Figure 2. Stratigraphy of Middle Pennsylvanian of Illinois Basin (as currently understood), showing correlation between chronostratigraphic and lithostratigraphic units (* indicates incomplete unit); compiled from Davydov et al. (2010, fig. 2), Falcon-Lang et al. (2011, fig. 1), and Nelson et al. (2013, fig. 2). Brazil Formation (shaded) incorporates strata from base of Lower Block Coal to top of Minshall Coal. Macrofloral assemblage collected from lowermost Staunton Formation (arrows). Atokan-Desmoinesian Stage boundary and Bolsovian-Asturian Substage boundary shown as coeval and equivalent to top of Perth, Seville, and Curlew limestones, after Peppers (1996). Dashed lines denote possible positions of one or both boundaries based on biostratigraphic data presented herein (see Fig. 7; Biostratigraphy and age section).

Perth Limestone, the top of which defines the Atokan-Desmoinesian Stage boundary based on palynology (Peppers, 1996). The Minshall Coal and overlying Perth Limestone correlate with the Rock Island (No. 1) Coal and Seville Limestone of northwestern Illinois and the Empire Coal and Curlew Limestone of southwestern Kentucky (Fig. 2; Tri-State Committee, 2001; Nelson et al., 2013).

The macrofloral assemblage was collected from an open-pit mine north of Indiana Route 46, the now inaccessible Ashboro Pit (SW 1/4, SE 1/4, Section 17, Township 11N, Range 6W) operated by Log Cabin Coal Company. Plant fossils were collected during a reconnaissance in 1980, when the Lower Block and Upper Block coals were being mined, but the pit was idle and partly filled with water during a 1982 visit, leaving the upper highwall inaccessible. The west- and north-facing highwalls of the L-shaped pit are reproduced schematically in Figure 3. About 15 m of the Brazil and overlying Staunton formations were exposed, the lithostratigraphic boundary being the top of the Minshall Coal.

Correlation of limestone and coal members of the Brazil and Staunton formations is challenging due to their lenticular nature and variable lithology, but marker beds in the Ashboro Pit are identified with confidence (Fig. 3). The Lower and Upper Block coals are distinctive in this area, consisting of hard, dullbanded or 'splint' coal that separates into large cubic blocks along widely spaced joint planes. The type area of these coals is 15 km north of the locality, whereas the Minshall Coal and Perth Limestone have type sections five to 15 km farther north. No other marine unit in the Brazil or lower Staunton formations in this part of Indiana is likely to be confused with the Perth Limestone.

Most macrofloral remains were collected from an ~35 cm-thick package of 'roof-shales' immediately above the Minshall Coal on the west-facing highwall (Units 1 to 3; Fig. 3), whereas some adpressions presumably are from a bed (Unit 4) on the north-facing highwall at a stratigraphic position above the Perth Limestone. All sampled beds belong to USNM Locality 38354. Although the strata belong to the lowermost Staunton Formation, as currently understood and depicted in Figure 2, Units 1 to 3 are assigned to the uppermost Atokan and Unit 4 to the lowermost Desmoinesian. Based on new data presented here, the age of the fossiliferous strata and position of the Atokan-Desmoinesian and Bolsovian-Asturian boundaries will be discussed in the Biostratigraphy and age section.

Fossiliferous units

A light-gray, rooted paleosol ('underclay') underlies the 15 cm-thick Minshall Coal (Fig. 3), which is relatively uniform in thickness, but undulatory due to irregularities on the paleosol surface. Following the 'low-stand coal' model (cf., Cecil, 1990; Cecil et al., 2003, 2014; DiMichele, 2014), the Minshall Coal is interpreted to record peat buildup due to paludification and base-level rise under the (per)humid (everwet) conditions that attended a sea-level lowstand at maximum glaciation.

The ~35 cm-thick package of fossiliferous roof-shales above the Minshall Coal is divided into three distinct beds based on lithology and plant-fossil content (Units 1 to 3; Fig. 3). Gradational with the underlying coal, Unit 1 (~10 cm thick) is a

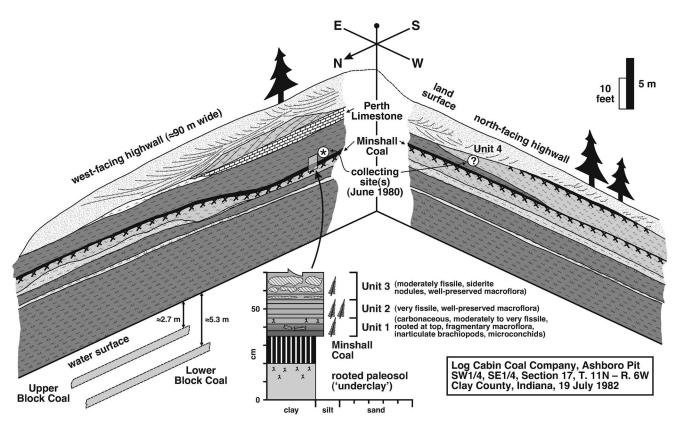


Figure 3. Schematic drawing of succession exposed in Ashboro Pit (now inaccessible open-pit mine of Log Cabin Coal Company; SW 1/4, SE 1/4, Section 17, Township 11N, Range 6W), north of Indiana Route 46, Clay County, Indiana, during visits in June 1980 and July 1982. Brazil Formation incorporates strata from base of Lower Block Coal (under water) to top of Minshall Coal, and overlying strata belong to Staunton Formation. Top of Perth Limestone corresponds to Atokan-Desmoinesian boundary (Peppers, 1996). Macrofloral assemblage collected from roof-shales above Minshall Coal (Units 1 to 3, marked with *) on west-facing highwall, and presumably from claystone-filled hollow (Unit 4, marked with ?) on north-facing highwall. Note that Unit 4 is at stratigraphic position above the Perth Limestone. All sampled beds belong to USNM locality 38354.

medium- to dark-gray, fissile, highly carbonaceous shale that contains abundant but fragmentary and poorly preserved plant remains. Spirorbiform worm-tubes (microconchids) are affixed to some fossil leaves, and inarticulate brachiopods occur sporadically. Fine root structures are present at the top of the bed. Unit 1 is interpreted to record deposition in the shallow, dysaerobic, and at least hyposaline waters of a tidally influenced mudflat or estuary, which developed above the precursor Minshall Coal during initial stages of transgression as deglaciation commenced (cf., Archer et al., 1994, 2016; Mastalerz et al., 1997). Accordingly, plant remains mainly represent the remnants of drowned peat-forming vegetation. Roots at the top of the unit indicate upward shallowing, emergence, and development of a 'clastic swamp' (sensu Gastaldo, 1987; Demko and Gastaldo, 1992). The time required for establishment and colonization of the clastic swamp implies a temporary lull in sea level rise, possibly even a minor regressive event. Elsewhere in Clay County, the Minshall Coal consists of two distinct 'benches' separated by a clastic unit (up to 2.7 m thick) with roots at the top. A similar pattern occurs farther south in Daviess County, where a persistent claystone (~5 cm thick) exists near the middle of the Minshall Coal (incorrectly called the Buffaloville Coal by mine operators). As such, clastic swamp deposits at the top of Unit 1 may coincide with, and/or are a lateral equivalent of, the upper bench (Clay County) or upper part (Daviess County) of the Minshall Coal.

Unit 2 (~10 cm thick) is gradational with Unit 1, and comprises medium-gray, fissile, and well-indurated shale. Adpressed plant fossils are abundant and well preserved. Unit 2 is interpreted to record renewed transgression and further accumulation of tidally influenced mudflat or estuarine deposits. Macrofloral remains in this bed are thought to represent litter shed from clastic swamp vegetation on the periphery of the mudflat or estuary.

Unit 3 (~15 cm thick) is gradational with Unit 2, and consists of maroon, very well-indurated mudstone that contains siderite nodules. Macrofloral remains are well preserved but only moderately abundant. Unit 3 presumably resulted from additional base level rise due to transgression, with the presence of siderite nodules reflecting a change in depositional and/or diagenetic conditions on the mudflat or estuary. Adpressed plant fossils probably were derived from wetland vegetation in local clastic swamp habitats.

Unit 4 (10s of cm thick) consists of pinkish light-gray, well-laminated claystone that contains sparse but well-preserved plant adpressions. The most probable provenance is the fill of a hollow (Fig. 3) that was eroded down to the level of the Minshall Coal during the onset of regression, a consequence of polar ice buildup. If this stratigraphic level is correct, Unit 4 may record the rapid filling of an abandoned channel, either in a fluvial or tidally influenced deltaic setting. Plant debris probably was shed from clastic wetland plant communities alongside the abandoned channel.

Quantification and general characteristics of floral assemblages

Macrofloral assemblage.—Most plant remains are preserved as coalified compressions or impressions (i.e., adpressions; Shute and Cleal, 1987), although a small number of specimens are 'naturally macerated' (cf., Zodrow and Mastalerz, 2009). Proximate analysis of a sample of the underlying Minshall Coal yields a vitrinite reflectance (%R_{omax}) of 0.57, a coal rank of high-volatile C bituminous (Teichmüller and Teichmüller, 1982). Such a low rank indicates that phytoleims (coalified compressions) probably contain epidermal cuticles, although no attempt was made to procure cuticles.

Table 1 presents a quantitative breakdown of the rock slabs and fossil-taxa in Units 1 to 4, and for the macrofloral assemblage as a whole. Of the 191 slabs collected, 326 slab faces bear macrofloral remains (excluding barren faces and counterparts). The quantification technique used is a variant of that described by Pfefferkorn et al. (1975): each slab face was treated as a quadrat, and a fossil-taxon was recorded as 'present' regardless of the number of specimens on the face. Through this process, 688 macrofloral occurrences were recorded on the 326 fossiliferous slab faces. Excluding seeds and some axes, which could not be tied to a particular fossil-plant taxon with confidence (termed 'unidentifiable' macrofloral occurrences in Table 1), there were 449 'identifiable' macrofloral occurrences.

Of the 22 fossil-taxa identified (Table 1), 18 can be considered 'biological' taxa (i.e., a reassembled conceptualization of a whole and once-living fossil plant; sensu Cleal and Thomas, 2010), whereas only four 'non-biological' taxa were recorded (Cyperites bicarinatus Lindley and Hutton, 1832, Lepidostrobophyllum Hirmer, 1927 [sp. indet.], Calamites Brongniart, 1828a [spp. indet.], and a putative conifer cone scale). It is noted, however, that each fossiliferous bed contains a low-diversity assemblage, ranging from six to 12 fossil-taxa and four to nine biological taxa. When relative abundances for the entire macrofloral assemblage are considered, pteridosperms are the predominant (60.8%) and most diverse plant group (nine biological fossil-species). The sphenopsids (11.8%) are relatively diverse (five biological species), whereas all cordaitalean remains (16.7%) are included in *Cordaites* Unger, 1850 (spp. indet). The lycopsids (9.4%) are represented by two biological species, and fern and putative conifer remains are very rare.

Microfloral assemblage.—Samples of the Minshall Coal and four fossiliferous clastic units (~150 g of rock/coal crushed and sieved through –20 mesh) were oxidized using Schulze's Solution (nitric acid saturated with potassium chlorate) to liberate spores and pollen, followed by digestion in 5% potassium hydroxide. The organic fraction was concentrated with zinc chloride solution, and treated with ethylene glycol monoethyl ether (2-ethoxyethanol) and ultrasonic vibration to remove fine particulate matter from the spore/pollen fraction. Slides were prepared by strew-mounting a small amount of spore/pollen residue with polyvinyl alcohol on cover glasses, which were fixed with a synthetic piccolyte resin. Line counts (250 palynomorphs per sample) were performed at 640X magnification to establish

relative abundances, and additional slides were scanned at 250X to document forms of potential biostratigraphic significance that were not encountered in statistical counts. All slides, maceration residues, and unused portions of samples are stored at the Kentucky Geological Survey (KGS), University of Kentucky, Lexington, KY under KGS sample numbers 3264–3268.

Table 2 gives a quantitative breakdown of the plant groups and fossil-taxa found in the Minshall Coal and Units 1 to 4. Forty-nine miospore species were identified, seven of unknown affinity. The beds are dominated by spores of arborescent or sub-arborescent lycopsids (Minshall Coal, Units 1 to 3) or by tree ferns (Unit 4). Non-arborescent fern and calamitalean spores are rare, as is cordaitalean pollen. Although pteridosperm foliage dominates the macrofloral assemblage, it is emphasized that no samples contain pteridosperm pollen, despite targeted searches, even in maceration residues of the $\leq 250 \,\mu m$ size fraction. Thus, although pteridosperm pollen typically is very large (100-600 µm; Taylor, 1978) and may be excluded by traditional preparation techniques (Peppers and Pfefferkorn, 1970), some grains should have been found if present. Because this paper focuses on the macrofloral assemblage, the microfloral data are only used to provide complementary biostratigraphic control.

Biostratigraphy and age

The Minshall Coal and overlying Perth Limestone of western Indiana (Fig. 2), along with their correlates in southern Indiana ('Buffaloville' Coal [see below], Curlew Limestone), western Kentucky (Empire Coal, Curlew Limestone), and western Illinois (Rock Island [No. 1] Coal, Seville Limestone), have figured prominently in the discussion surrounding placement of the contentious Atokan-Desmoinesian Stage boundary in the Illinois Basin (e.g., Shaver, 1984; Peppers, 1996; Heckel, 2013; Nelson et al., 2013). Because a precisely defined Atokan stratotype has yet to be realized (Sutherland and Manger, 1984), in large part because the poorly fossiliferous Atoka Formation (defined in the Arkoma Basin, Oklahoma; Fig. 1.1) is depositionally and tectonically complex, the position of the Atokan-Desmoinesian boundary in the Illinois Basin has shifted depending on the fossil group used for biostratigraphic control. For example, marine faunas in the limestones have indicated either an early Desmoinesian age (ostracodes and fusulinids: Shaver and Smith, 1974, Shaver, 1984; conodonts: Rexroad et al., 1998; fusulinids: Wahlman, 2013) or a late Atokan age (fusulinid foraminifera: Douglass, 1987; conodonts: Barrick et al., 2013). Likewise, the Atokan-Desmoinesian boundary was placed at the top of the Seville, Perth, and Curlew limestones based on the correlation of microfloral assemblages from coal seams (Peppers, 1996; Fig. 2). By extension of these uncertainties, the position of the Minshall and equivalent coals remains uncertain in chronostratigraphic terms, although the seams usually are placed in the uppermost Atokan (Nelson et al., 2013).

Conodonts from the limestone directly overlying the Buffaloville Coal in its type area (southern Indiana) indicate that it is younger than the Perth, Curlew, and Seville limestones (personal communication, P.H. Heckel, 2014; Fig. 2). This result corroborates the palynological study of Peppers (1993),

Table 1. Fossil-taxa in macrofloral assemblage, from Units 1 to 4 above Minshall Coal (Fig. 3), segregated into plant groupings. Right-hand column indicates if fossil-taxon present in Europe. Symbols: (a) total slabs from unit; (b) slab faces with macrofloral remains (i.e., excludes barren faces and counterparts); (c) identifiable macrofloral occurrences + 'unidentifiable' macrofloral occurrences (e.g., axes and seeds); (d) occurrence indicates presence (in any quantity) on slab face; (e) 'biological' fossil-taxa + 'non-biological' fossil-taxa (*Cyperites bicarinatus* Lindley and Hutton, 1832, *Lepidostrobophyllum* Hirmer, 1927 [sp. indet.], *Calamites* Brongniart, 1828a [spp. indet.], and putative conifer cone scale); (f) reassembled conceptualization of once-living, whole fossil plant. See text for details.

Plant group	Fossil-taxon	Unit 1 (n [%])	Unit 2 (n [%])	Unit 3 (n [%])	Unit 4 (n [%])	Totals (N [%])	In Europe?
LYCOPSIDS		34 [37.36]	6 [3.06]	absent	2 [7.41]	42 [9.35]	
	cf. Asolanus camptotaenia	1 [1.10]	absent	absent	1 [3.70]	2 [0.44]	yes
	Synchysidendron sp. cf. S. andrewsii	1 [1.10]	absent	absent	absent	1 [0.22]	yes
	Cyperites bicarinatus	30 [32.97]	4 [2.04]	absent	absent	34 [7.57]	yes
	Lepidostrobophyllum sp. indet.	2 [2.20]	2 [1.02]	absent	1 [3.70]	5 [1.11]	yes
SPHENOPSIDS		2 [2.20]	12 [6.12]	32 [23.70]	7 [25.93]	53 [11.80]	
	Annularia sp. cf. A. radiata	absent	absent	absent	2 [7.41]	2 [0.44]	yes
	Annularia sphenophylloides	absent	absent	absent	2 [7.41]	2 [0.44]	yes
	Annularia sp. indet.	absent	absent	absent	1 [3.70]	1 [0.22]	yes
	Sphenophyllum cuneifolium	absent	12 [6.12]	32 [23.70]	absent	44 [9.80]	yes
	Sphenophyllum emarginatum	absent	absent	absent	1 [3.70]	1 [0.22]	yes
	<i>Calamites</i> spp. indet.	2 [2.20]	absent	absent	1 [3.70]	3 [0.67]	yes
FERNS	Sphenopteris sp. indet	1 [1.10]	4 [2.04]	absent	absent	5 [1.11]	(?)
PTERIDOSPERMS		absent	152 [77.55]	103 [76.30]	18 [66.67]	273 [60.80]	
	Alethopteris densinervosa	absent	3 [1.53]	25 [18.52]	absent	28 [6.24]	yes
	Macroneuropteris scheuchzeri	absent	51 [26.02]	25 [18.52]	absent	76 [16.93]	yes
	Neuropteris flexuosa	absent	69 [35.20]	25 [18.52]	absent	94 [20.94]	yes
	Neuropteris missouriensis	absent	absent	absent	14 [51.85]	14 [3.12]	Laveineopteris dussartii(?)
	Neuropteris ovata	absent	20 [10.2]	5 [3.70]	1 [3.70]	26 [5.79]	yes
	Neuropteris semireticulata	absent	absent	absent	2 [7.41]	2 [0.44]	yes
	"Mariopteris" anthrapolis	absent	6 [3.06]	absent	absent	6 [1.34]	no
	Eusphenopteris neuropteroides	absent	3 [1.53]	23 [17.04]	absent	26 [5.79]	yes
	Eusphenopteris sp. indet.	absent	absent	absent	1 [3.70]	1 [0.22]	(?)
CORDAITALEANS	Cordaites spp. indet.	54 [59.34]	21 [10.71]	absent	absent	75 [16.70]	yes
CONIFERS	conifer cone scale	absent	1 [0.51]	absent	absent	1 [0.22]	(?)
Slabs $(n)^a$		62	64	43	22	191	
Fossiliferous faces (n) ^b		116	106	70	34	326	
Macrofloral occurrences $(n)^{c}$		200	257	179	52	688	
Identifiable macrofloral occurrences $(n)^d$		91	196	135	27	449	
Fossil-taxa $(n)^e$		7	12	6	11	22	
'Biological' fossil-taxa (n) ^f		4	9	6	9	18	

Table 2. Fossil-taxa in microfloral assemblage, from Minshall Coal and Units 1 to 4 above coal (Fig. 3), segregated into plant groupings/growth forms.

Plant group	Fossil-taxon	Minshall Coal (%)	Unit 1 (%)	Unit 2 (%)	Unit 3 (%)	Unit 4 (%)
ARBORESCENT LYCOPSIDS		6.4	24.0	50.8	75.6	27.6
	Crassispora kosankei	0.0	0.0	0.4	0.0	0.4
	Granasporites medius	0.0	0.4	0.4	1.6	0.0
	Lycospora granulata	5.2	9.2	33.2	36.8	12.8
	Lycospora micropapillata	0.0	0.4	4.8	13.6	2.0
	Lycospora orbicula	0.0	1.6	8.0	15.6	0.8
	Lycospora pellucida	0.0	10.8	1.2	4.4	4.4
	Lycospora pusilla	1.2	1.6	2.8	3.6	7.2
SUB-ARBORESCENT LYCOPSIDS		51.6	28.8	2.4	2.0	0.0
	Anacanthotriletes spinosus	0.8	1.6	0.0	0.0	0.0
	Densosporites lobatus	5.2	1.2	0.0	0.8	0.0
	Densosporites sphaerotriangularis	0.8	0.4	0.0	0.0	0.0
	Endosporites globiformis	0.0	10.0	0.8	0.0	0.0
	Radiizonates difformis Radiizonates rotatus	44.8	15.6	1.6	1.2	0.0
TREE FERNS		26.0	26.0	35.6	7.6	63.6
	Apiculatisporites saetiger	0.4	1.6	2.0	0.4	0.0
	Laevigatosporites globosus	3.6	1.6	0.0	1.2	3.2
	Laevigatosporites minimus	0.0	0.4	0.8	0.0	2.0
	Punctatisporites minutus	21.2	17.2	30.8	4.8	50.0
	Punctatosporites granifer	0.0	3.2	0.0	0.0	0.0
	Punctatosporites minutus	0.8	1.6	2.0	1.2	6.4
	Punctatosporites rotundus	0.0	0.0	0.0	0.0	2.0
	Torispora securis	0.0	0.4	0.0	0.0	0.0
NON-ARBORESCENT FERNS		6.4	4.8	4.0	4.8	2.8
	Acanthotriletes aculeolatus	0.0	0.8	0.0	0.4	0.0
	Acanthotriletes triquetrus	0.4	0.0	0.4	0.0	0.0
	Convolutispora florida	0.0	0.4	0.0	0.0	0.0
	Cyclogranisporites minutus	0.0	0.8	0.0	0.0	0.0
	Granulatisporites adnatoides	0.0	0.8	0.0	0.0	0.8
	Granulatisporites granulatus	0.8	0.0	0.4	0.0	0.0
	Granulatisporites parvus	3.2	0.4	0.4	0.8	0.0
	Granulatisporites pircformis	0.4	0.0	1.6	0.4	0.0
	Leiotriletes priddyi	0.0	0.0	0.4	0.0	0.0
	Leiotriletes subadnatoides	0.0	0.4	0.4	1.6	0.0
	Lophotriletes commissuralis	0.0	0.4	0.0	0.0	0.0
	Lophotriletes granoornatus	0.0	0.4	0.4	1.6	1.2
	Lophotriletes microsaetosus	1.2	0.0	0.0	0.0	0.0
	Punctatisporites pseudolevatus Raistrickia saetosa	0.0 0.4	0.4 0.0	$0.0 \\ 0.0$	$0.0 \\ 0.0$	$0.8 \\ 0.0$
CALAMITALEANS		4.8	4.4	3.6	1.2	2.4
CALLANIA TALLA LIA	Calamospora breviradiata	0.0	2.0	2.8	0.4	0.8
	Calamospora microrugosa	0.0	0.4	0.0	0.0	0.0
	Calamospora pedata	0.0	0.0	0.4	0.0	0.0
	Laevigatosporites minor	4.8	2.0	0.4	0.8	1.6
CORDAITALEANS		4.8	4.8	2.0	8.4	1.6
001111111111111111111111111111111111111	Florinites florini	4.0	2.0	1.6	6.8	1.2
	Florinites mediapudens	0.8	2.8	0.4	1.6	0.4
UNKNOWN AFFINITY		0.0	7.2	1.6	0.4	2.0
	Adelisporites multiplicatus	0.0	0.4	0.0	0.0	0.0
	Echnatispora knoxiae	0.0	0.0	0.0	0.0	0.8
	Spackmanites sp. indet.	0.0	0.8	0.0	0.0	0.0
	Tantillus triquetrus	0.0	0.4	0.0	0.0	0.0
	Triquitrites bransonii	0.0	0.4	0.0	0.0	0.0
	Triquitrites minutus	0.0	0.8	0.0	0.0	0.0
	Triquitrites sculptilis	0.0	4.4	1.6	0.4	1.2

who concluded that the true Buffaloville Coal is younger than the Minshall Coal.

According to Peppers (1996), the Atokan-Desmoinesian Stage boundary of North America coincides with the Bolsovian-Asturian Substage boundary of Western Europe (Figs. 2, 4). If placement of this coupled boundary at the top of the Perth Limestone (and correlates) is accepted, the implication is that the underlying Minshall Coal (and correlates) are late Bolsovian in age. In the following section, the taxonomic composition of the microfloral and macrofloral assemblages from the Minshall Coal and overlying roof-shales (Units 1 to 3) are compared with

established biozonation schemes to determine whether or not this assumption holds. Unit 4, which is presumed to have been at a stratigraphic position above the Perth Limestone (Fig. 3), can be assigned an earliest Asturian age by default because it lies just above the Atokan-Desmoinesian boundary (Fig. 2).

Comparison with American microfloral biozonation.—Peppers (1984, 1996) constructed a microfloral biozonation scheme to facilitate regional correlation between the Illinois Basin and other parts of Euramerica. According to this framework (Fig. 4), the Brazil Formation is in the upper (but not uppermost)

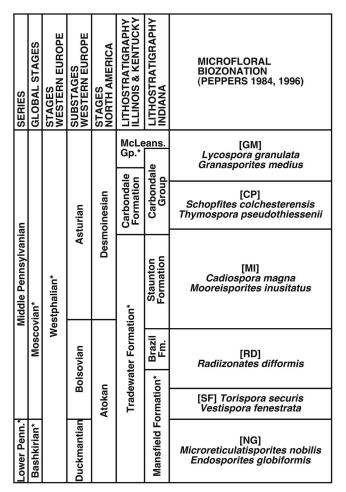


Figure 4. Microfloral biozonation scheme of Peppers (1984, 1996) correlated with chronostratigraphic and lithostratigraphic units (* indicates incomplete unit). Modified from Peppers (1996, pl. 1). Microfloral assemblage corresponds to upper *Radiizonates difformis* (RD) Biozone.

Bolsovian part of the *Radiizonates difformis* (RD) Biozone, with the top of the Minshall Coal corresponding to the top of the biozone (Peppers, 1996).

The composition of the microfloral assemblage (Table 2) supports assignment of the Minshall Coal and overlying roof-shales to the *Radiizonates difformis* (RD) Biozone. Of particular importance is the presence of the diagnostic densospores *Radiizonates difformis* (Kosanke, 1950) Staplin and Jansonius, 1964 and *Radiizonates rotatus* (Kosanke, 1950) Staplin and Jansonius, 1964, which are most abundant in the coal and Unit 1. Further, none of the beds (including Unit 4) contain the namesake taxa of the succeeding *Cadiospora magna-Mooreisporites inusitatus* (MI) Biozone (Fig. 4). *Cadiospora magna* Kosanke, 1950 first appeared at the Bolsovian-Asturian boundary, whereas *Mooreisporites inusitatus* (Kosanke, 1950) Neves, 1958 arose in the latest Bolsovian (Peppers, 1996). These factors collectively point to a late Bolsovian age for the Minshall Coal and overlying clastic beds.

Nonetheless, a microfloral assemblage from a clastic unit directly beneath the paleosol of the Minshall Coal adds uncertainty to this late Bolsovian assignment (these data, incorporated into another publication, are introduced here because they have important ramifications on the age of overlying strata). Most

noteworthy is the statistically significant abundance of *Thymospora* pseudothiessenii (Kosanke, 1950) Wilson and Venkatachala, 1963. The spore is considered a reference for basal Asturian, although it first appears in uppermost Bolsovian strata (Peppers, 1996; Lesnikowska and Willard, 1997) and reaches its epibole in the middle Asturian Schopfites colchesterensis-Thymospora pseudothiessenii (CP) Biozone (Fig. 4). Equally intriguing is the fact that Savitrisporites nux (Butterworth and Williams, 1958) Smith and Butterworth, 1967 occurs (albeit very rarely) beneath the Minshall Coal, but not in the coal itself. Because this spore disappears just below the Bolsovian-Asturian boundary (Peppers, 1996), its absence in the coal does not in itself imply an Asturian age for the Minshall, but it does signify peat accumulation very close to the substage boundary. It is crucial to note that the scheme of Peppers (1984, 1996) is almost exclusively based on palynomorphs from coal, and many taxon ranges would be extended if clastic lithologies had been incorporated. These additional microfloral data signify that an earliest Asturian age for the Minshall Coal cannot be ruled out.

Comparison with American macrofloral biozonation.—Moore et al. (1944) and Read and Mamay (1964) segregated upper Paleozoic strata of the USA into fifteen macrofloral zones defined by the co-occurrence of common or diagnostic taxa. Unfortunately, the absence of range charts and ambiguity of zone boundaries make this biostratigraphic framework difficult to apply. Gillespie and Pfefferkorn (1979) discussed the temporal distribution of key taxa and charted their ranges in the Appalachian Basin. Extending this work, Blake et al. (2002) produced an updated chart that correlates between lithostratigraphic units of the Appalachian Basin, chronostratigraphic units of midcontinental USA and Western Europe, and macrofloral zones of the USA (Read and Mamay, 1964) and paleoequatorial Europe (Wagner, 1984). According to Blake et al. (2002, fig. 1, with updates from B.M. Blake, personal communication, 2014), the Atokan-Desmoinesian boundary, which they equated to the Bolsovian-Asturian boundary following Peppers (1996), is at the Upper Block (No. 5) Coal near the contact between the Kanawha and Allegheny formations (Fig. 5). This level correlates to the middle of Zone 9 ('Neuropteris' rarinervis) of Read and Mamay (1964).

Blake et al. (2002, fig. 2) also provided a biostratigraphic range chart that shows the distribution of important fossil-taxa in the Appalachian Basin. A pragmatic approach to determining the age of the macrofloral assemblage above the Minshall Coal has been to compile the ranges of key fossil-species in the Appalachian Basin (e.g., Read and Mamay, 1964; Darrah, 1969; Gillespie and Pfefferkorn, 1979; Blake et al., 2002; personal communication, B.M. Blake, 2014). The results of this exercise (Fig. 5) are somewhat ambiguous, in part because the first appearance datum (FAD) and last appearance datum (LAD) of Neuropteris semireticulata Josten, 1962, Neuropteris ovata Hoffmann, 1826a, 1826b, and Neuropteris flexuosa Sternberg, 1825 are poorly constrained. Additionally, a short hiatus may exist between the Kanawha and Allegheny formations (personal communication, B.M. Blake, 2014), further confounding age determination. Nonetheless, the compilation indicates that the macrofloral assemblage contains several species that first appear in the middle Bolsovian but extend into and are more typical of the

SERIES	Lower Penn	ısylvanian*		N	Middle Pennsylva	anian*	
GLOBAL STAGES	Bashk	irian*	Moscovian*				
STAGES WESTERN EUROPE			West	phalian*			
SUBSTAGES WESTERN EUROPE	Duckm	Duckmantian Bolsovian Asturian			Asturian		
STAGES NORTH AMERICA		Atokan			D	esmoinesian*	
LITHOSTRATIGRAPHY APPALACHIAN BASIN		Kanawha Formati	on*		Allegheny Formation		
FLORAL ZONES (READ & MAMAY, 1964)	Zone 7 <i>Megalopteris</i> spp.	Zone <i>'Neuropteris'</i>				Zone 10 – Neuropto flexuosa & Pecopteris	eris s spp.
Sphenophyllum cuneifolium Eusphenopteris neuropteroides Macroneuropteris scheuchzeri Annularia sphenophylloides Sphenophyllum emarginatum Neuropteris semireticulata Asolanus camptotaenia Neuropteris ovata Neuropteris flexuosa			?		?		≓

Figure 5. Biostratigraphic ranges of key fossil-species in macrofloral assemblage correlated with chronostratigraphic units, lithostratigraphic units of Central Appalachian Basin, and American macrofloral biozonation scheme of Read and Mamay (1964) (* indicates incomplete unit); compiled from Darrah (1969), Gillespie and Pfefferkorn (1979), Blake et al. (2002), and B.M. Blake (personal communication, 2014). Maximum range overlap occurs in upper Kanawha to lower Allegheny formations (late Atokan to early Desmoinesian, late Bolsovian to early Asturian), equivalent to most of Zone 9 (*'Neuropteris' rarinervis*).

Asturian. Maximum range overlap occurs in a late Bolsovian to earliest Asturian interval, equivalent to much of Zone 9 ('*Neuropteris' rarinervis*) and in accord with the age suggested by the microfloral assemblage.

In contrast to Europe and Atlantic Canada (see below), in the USA *Neuropteris ovata* occurs sparingly in middle Bolsovian assemblages (Zone 8—'*Neuropteris*' tenuifolia), is abundant in (but not characteristic of) late Bolsovian to early Asturian assemblages (Zone 9—'*Neuropteris*' rarinervis), and only becomes more common in younger strata (Jongmans, 1937c; Read and Mamay, 1964; Darrah, 1969; Gillespie and Pfefferkorn, 1979; Blake et al., 2002). Likewise, Read and Mamay (1964) noted rare occurrences of *Neuropteris flexuosa* in Zone 8, but indicated that the species only became abundant in the middle to late Asturian (Zone 10—*Neuropteris flexuosa* and *Pecopteris* spp.); Darrah (1969) placed the FAD in the lower Allegheny Formation (lower Asturian).

That *Neuropteris ovata* and *Neuropteris flexuosa* first appear in middle to upper Bolsovian strata in the USA is supported by re-examination of specimens illustrated by Wood (1963), who documented assemblages from above the middle(?) Bolsovian Lower Block Coal in western Indiana (Fig. 2). Although poorly preserved and fragmentary, both species seem to be present: *N. ovata* was misidentified under *Neuropteris obliqua*? (pl. 9, fig. 10) and *Odontopteris subcuneata* (pl. 10, fig. 5), whereas *N. flexuosa* may have been both correctly identified (pl. 9, fig. 5) and referred to *Neuropteris heterophylla*? (pl. 9, fig. 6). However, all of these taxa are listed as being "not common" (Wood, 1963, p. 32), indicating that they were not characteristic of the assemblages.

This analysis indicates that roof-shales above the Minshall Coal are either late Bolsovian or early Asturian in age. However, the fact that *Neuropteris flexuosa* is the most abundant pteridosperm in Units 2 and 3 (Table 1), and that *Neuropteris ovata* also is a common element in these beds, indicates that an early Asturian age is most probable.

Comparison with European macrofloral biozonation.—Building on previous biostratigraphic schemes (e.g., Dix, 1934; Bell, 1938; Gothan and Remy, 1957; Corsin and Corsin, 1971; Laveine, 1977), a comprehensive framework for Carboniferous plant fossils was assembled by Wagner (1984), who compared the ranges of key taxa and incorporated preexisting floral zones to establish sixteen distinct biozones. The biozonation has been refined to improve its resolution, with several subbiozones now recognized (Cleal, 1991; Cleal and Thomas, 1994). Although primarily based on range data from western and central Europe, Wagner (1984) intended that the scheme would aid biostratigraphic correlations throughout the Euramerican paleoequatorial belt.

Comparison with the European biozonation (Fig. 6) is based on the compilation of taxon ranges as known in Western Europe (e.g., Laveine, 1987; Wagner, 1984; Cleal and Thomas, 1994). Although some taxa are long ranging, there is minimal overlap between the LAD of *Neuropteris semireticulata* and *Alethopteris densinervosa* Wagner, 1968 and the FAD of *Neuropteris flexuosa* and members of the *Neuropteris ovata* complex, the latter group being considered a guide to the basal Asturian (Van Leckwijck, 1964; Laveine, 1977; Wagner, 1984). Accordingly, the age of the macrofloral assemblage can be tightly constrained to the early Asturian, assignable to the *Linopteris obliqua* Biozone. The anomalous presence of *Sphenophyllum cuneifolium* (Sternberg, 1821) Zeiller, 1878-79a, more typical of lower Westphalian assemblages, does not override this outcome.

Stratigraphic ramifications.—These homotaxial comparisons yield slightly different age results for the Minshall Coal and overlying roof-shales depending on the fossil group or biostratigraphic framework used: (1) probably latest Bolsovian (American microfloras; Fig. 4); (2) latest Bolsovian or earliest Asturian (Appalachian Basin macrofloras; Fig. 5); or (3) earliest Asturian (European macrofloras; Fig. 6). Although all analyses clearly convey that both peat and clastic sediment accumulation

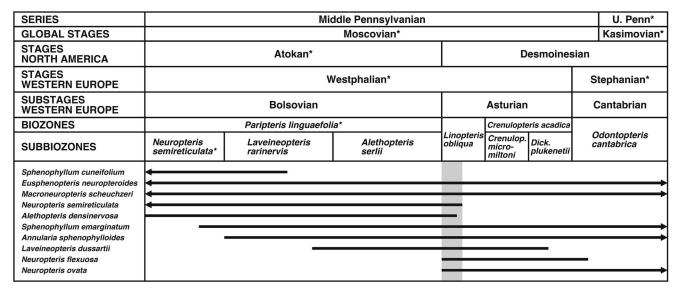


Figure 6. Biostratigraphic ranges of key fossil-species in macrofloral assemblage correlated with chronostratigraphic units and European macrofloral biozonation scheme. Compiled from Wagner (1984), Laveine (1987), Cleal (1991), and Cleal and Thomas (1994). Range of *Laveineopteris dussartii* (Laveine, 1967) Laveine, 2005 included because may be synonymous with *Neuropteris missouriensis* Lesquereux, 1879-80. Maximum range overlap occurs in lower *Linopteris obliqua* Biozone (early Asturian, early Desmoinesian).

occurred close to the Bolsovian-Asturian boundary, three crucial points need to be addressed before attempting to refine the stratigraphic resolution. First, the Minshall Coal and overlying roof-shales need not be the same age; the boundary may have been crossed in the interim between the final phases of peat accumulation and the drowning and burial of the mire during transgression. Second, significant (and perhaps undue) weight has been placed on coincidence of the Bolsovian-Asturian and Atokan-Desmoinesian boundaries, especially since they were defined in widely separated areas based on different parameters. Third, there must have been a temporal lag between the evolution of a new plant taxon and its dispersal to other parts of paleoequatorial Euramerica (cf., Laveine et al., 2000; Blake et al., 2002). These factors are taken into account as four stratigraphic scenarios are explored for the study section, each of which attempts to rectify the slightly disparate ages recorded in the microfloral and macrofloral assemblages.

Figure 7.1 shows the currently accepted correlation scheme in the Indiana portion of the Illinois Basin (with a focus on the Brazil and Staunton formations) and displays the relationships among chronostratigraphic, lithostratigraphic, and biostratigraphic units, at least as they have been understood prior to this analysis. The Bolsovian-Asturian Substage and Atokan-Desmoinesian Stage boundaries are equivalent and occur at the top of the Perth Limestone; the Minshall Coal belongs to the Radiizonates difformis (RD) Biozone and, by implication, the Minshall Coal and its roof-shales (Units 1 to 3) are upper Bolsovian. However, this scenario only is valid if Neuropteris ovata and Neuropteris flexuosa can occur in abundance in late Bolsovian assemblages. As noted above, both taxa make scattered appearances in middle to upper Bolsovian strata in the USA, but do not become common until the Asturian (Fig. 5; N. ovata in Zone 9, N. flexuosa in Zone 10 of Read and Mamay, 1964).

Introduction of the European macrofloral biozonation framework indicates that the roof-shales (Units 1 to 3) above the Minshall Coal (but not necessarily the coal itself) are no

older than earliest Asturian (Fig. 6). At least four scenarios are possible (Fig. 7.2–7.5); all include the Minshall Coal in the *Radiizonates difformis* (RD) Biozone and its roof-shales in the *Linopteris obliqua* Biozone.

Scenario one.—The Bolsovian-Asturian and Atokan-Desmoinesian boundaries could remain coupled and lowered to the top of the Minshall Coal (Fig. 7.2). The implications are that the precursor mire existed during the latest Bolsovian (latest Atokan), and that a chronostratigraphic boundary was crossed before transgression and deposition of the roof-shales in the earliest Asturian (earliest Desmoinesian). As a consequence, the contact between the *R. difformis* (RD) and *Cadiospora magna-Mooreisporites inusitatus* (MI) microfloral biozones would be slightly younger than the late Bolsovian (late Atokan) age proposed by Peppers (1996; Fig. 7.1), and the basalmost Staunton Formation would correspond to the lower Asturian (lower Desmoinesian).

Scenario two.—The coupled Bolsovian-Asturian and Atokan-Desmoinesian boundary could be situated somewhere below the Minshall Coal (Fig. 7. 3), meaning that both the peat and overlying clastics accumulated during the earliest Asturian (earliest Desmoinesian). Consequently, the upper part of the *R. difformis* (RD) Biozone and uppermost Brazil Formation would be younger than currently understood (Fig. 7.1).

Scenario three.—The Bolsovian-Asturian and Atokan-Desmoinesian boundaries could be decoupled, with the former lowered to the top of the Minshall Coal and the latter remaining at the top of the Perth Limestone (Fig. 7.4). The ramifications mostly mirror those for point 1, except that the uppermost Brazil Formation, lowermost Staunton Formation, and lower part of the *C. magna-M. inusitatus* (MI) Biozone would be retained in the uppermost Atokan (although now correlative to the lowermost Asturian).

Scenario four.—The decoupled Bolsovian-Asturian boundary could be placed at an even lower position below the Minshall Coal (Fig. 7.5). As in point 3, the upper Brazil, lower Staunton, and lower *C. magna-M. inusitatus* (MI) Biozone

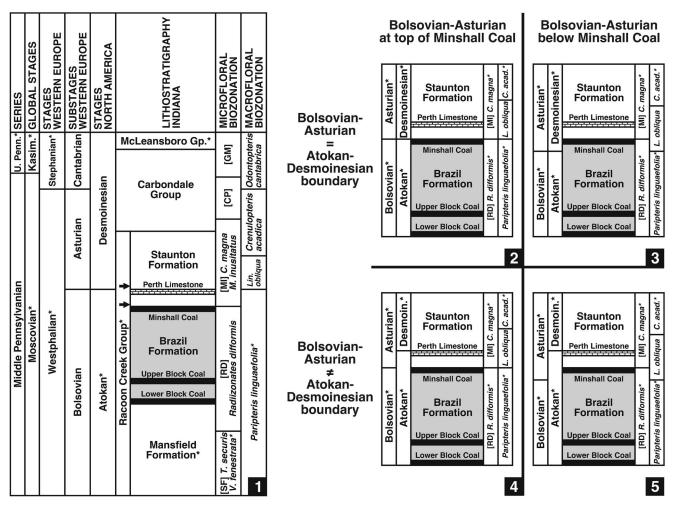


Figure 7. Options for placement of Atokan-Desmoinesian Stage boundary and Bolsovian-Asturian Substage boundary based on biostratigraphic data presented herein. (1) Stratigraphy of Middle Pennsylvanian of Indiana part of Illinois Basin (as currently understood), showing correlation between chronostratigraphic and lithostratigraphic units, and American microfloral and European macrofloral biozonation schemes (* indicates incomplete unit). Compiled from Figures 2, 4, and 6. Macrofloral assemblage collected from lowermost Staunton Formation (arrows). For all four scenarios presented in 7.2–7.5, Minshall Coal retained in *Radiizonates difformis* (RD) Biozone and clastic units bearing macrofloral assemblage (arrows) in *Linopteris obliqua* Biozone. (2) Atokan-Desmoinesian and Bolsovian-Asturian boundaries remain coupled and lowered to top of Minshall Coal. (3) Atokan-Desmoinesian and Bolsovian-Asturian boundaries decoupled, with Atokan-Desmoinesian boundary retained at top of Perth Limestone and Bolsovian-Asturian boundary lowered to below Minshall Coal. (5) Atokan-Desmoinesian and Bolsovian-Asturian boundary lowered to below Minshall Coal. See discussion in text for stratigraphic ramifications of each scenario.

would remain in the upper Atokan, but would correspond to an even younger Asturian age.

All four scenarios have validity, but because this study deals with plant-fossil assemblages from one locality, no further attempt is made to recommend one over another. However, decoupling the Bolsovian-Asturian and Atokan-Desmoinesian boundaries, and lowering the former to the top of (or below) the Minshall Coal (points 3 and 4) is considered the least disruptive to the established stratigraphic framework. Similar studies at other sites in the Illinois Basin and elsewhere are necessary to rectify the position of the Bolsovian-Asturian Substage boundary of western Europe and its relationship to the Atokan-Desmoinesian Stage boundary in the USA.

Paleobiogeographic and paleoecologic implications

The temporal persistence of peat-inhabiting wetland vegetation in Pennsylvanian mires is well established, with evidence pointing to

long periods of ecological stability through recurring glacioeustatic cycles, and the repeated reassembly of communities drawn from a small pool of edaphic specialists when climatic conditions were favorable (DiMichele and Phillips, 1996a, 1996b; DiMichele et al., 1996, 2002; Willard et al., 2007; Falcon-Lang and DiMichele, 2010). Comparable patterns of persistence and compositional recurrence also existed in vegetation from associated clastic wetland habitats (Scott, 1978; Pfefferkorn and Thomson, 1982; DiMichele et al., 1996; Cleal et al., 2012; Tabor et al., 2013). These low-diversity ecosystems were the product of prolonged evolution under the intrinsically stressful conditions that typify wetland settings, which are effectively closed systems with low rates of speciation and turnover, and that experience little taxonomic exchange with terra-firma habitats (Knoll, 1985; DiMichele et al., 1987; Wing and DiMichele, 1995; DiMichele and Phillips, 1996a; Cleal et al., 2012).

Pennsylvanian wetland communities also retained compositional conservatism at a variety of spatial scales. Over tens of

meters to a few kilometers, mire vegetation was markedly heterogeneous yet encompassed a limited species pool (Pfefferkorn, 1979; DiMichele and Nelson, 1989; Pryor and Gastaldo, 2000; Gastaldo et al., 2004, DiMichele et al., 2007, Opluštil et al., 2009a, 2009b), and clastic wetland floras displayed a comparable mosaic of low-diversity patches (Wing and DiMichele, 1995; Bashforth et al., 2010, 2011; Cleal et al., 2012). However, local-scale variability was masked at the landscape or regional scale, and disappeared into the background of the restricted wetland species pool. An expression of this homogeneity is discussed by Moore et al. (2014), who demonstrated that two contemporaneous clastic wetland floras, separated by 1100 km on the American craton, showed convergent dominance-diversity structures and a remarkable degree of compositional similarity.

A key result of this study is confirmation that Pennsylvanian communities from clastic wetland habitats showed spatial conservatism over vast areas of tropical Euramerica. Despite promulgations about the lack of similarity between American and European adpressed macrofloral assemblages, this notion can be tempered, at least for the Middle Pennsylvanian time interval studied. Of the 18 biological fossil-taxa documented (Table 1), at least 14 species (78%; including Cordaites spp. indet.) are common in contemporaneous European assemblages. Assuming that Eusphenopteris Gothan, 1913 ex Simson-Scharold, 1934 (sp. indet.) and Sphenopteris (Brongniart, 1822) Sternberg, 1825 (sp. indet.) also occur in Europe, which they almost certainly do, and considering that the American form Neuropteris missouriensis Lesquereux, 1879-80 may be conspecific with the European form Laveineopteris dussartii (Laveine, 1967) Laveine, 2005, the number of shared species could be as high as 17 (94%). Only the taxon identified as "Mariopteris" anthrapolis Langford, 1958 seems to be unequivocally unique to American floras of this age (Table 1). Hence, it is not hyperbole to state that the macrofloral assemblage could have been collected from a lower Asturian roof-shale in Europe. It is clear, yet unsurprising, that the intrinsic stress inherent to wetland settings existed wherever such environments developed in tropical Euramerica, resulting in low-diversity communities that showed a high degree of taxonomic mutuality over the paleocontinent.

The discovery of *Alethopteris densinervosa* in the macrofloral assemblage, and the realization that it had previously been misidentified as *Alethopteris serlii* (Brongniart, 1833-34) Göppert, 1836 in other American floras (e.g., Basson, 1968; Oleksyshyn, 1982; Moore et al., 2014), provides additional evidence for the spatial congruity in Pennsylvanian clastic wetland vegetation. Formerly only known from the Northern France Coalfield (Wagner, 1968; Laveine, 1987), and perhaps the Sydney Coalfield (Zodrow, 1986; Zodrow and Cleal, 1998), the presence of a relatively obscure fossil-species such as *A. densinervosa* in widely separated areas from strata of similar age (early Bolsovian to middle Asturian) highlights the commonality of Middle Pennsylvanian floras in wetland deposits of Euramerica.

From a paleobiogeographic perspective, these observations are important because west-central and eastern Euramerica were separated by the expansive uplands of the Laurentian Shield to the north and, in part, by the Appalachian-Variscan Mountains (Blakey, 2013). The similarity of wetland communities on either

side of the paleocontinent confirms that, at least during the Middle Pennsylvanian, these elevated areas did not present an insurmountable barrier to dispersal for most wetland plants (cf., White, 1909; Pfefferkorn and Gillespie, 1980b; Blake et al., 2002). In fact, the mountain belt probably comprised a spatially and temporally discontinuous chain of tectonically active centers (Roscher and Schneider, 2006), which rose (and eroded down) earlier in the east than in the west in zipper-like fashion (Hatcher, 2002, 2010), rather than a continuous belt of highelevation that spanned the entire equatorial region of Pangea. Of course, compositional differences and endemic taxa certainly existed in various parts of Euramerica (e.g., Bell, 1944; Pfefferkorn and Gillespie, 1980b; Gastaldo and Boersma, 1983a, 1983b; Cleal, 2002, 2008a, 2008b; Cleal et al., 2009, 2012; Šimůnek and Cleal, 2011), presumably a product of regional landscape partitioning due to elevational, latitudinal, and climatic factors (Blake et al., 2002; Falcon-Lang et al., 2011). Such variables would have determined when and where speciation occurred, and must have impeded the dispersibility of wetland vegetation to some degree. For example, Neuropteris ovata and Neuropteris flexuosa make scattered appearances in middle to upper Bolsovian strata in the USA (Jongmans, 1937c; Wood, 1963; Read and Mamay, 1964; Darrah, 1969; Blake et al., 2002), even though these species are key indicators to identifying the base of the Asturian in Europe and Atlantic Canada (Laveine, 1977; Wagner, 1984; Zodrow and Cleal, 1985; Cleal and Thomas, 1994; Cleal, 2007). The paleobiogeographic ramification of these precocious appearances in the USA is that N. ovata and N. flexuosa originated in the western part of the paleocontinent before synchronously arriving in Europe. Although such paleobiogeographic vagaries may hinder the precision of homotaxial biostratigraphic comparisons between American and European macrofloral assemblages, on the whole, correlations within Euramerica should be possible so long as regional peculiarities are taken into account.

The marked lithological and biological changes that led to the recognition of the Atokan-Desmoinesian Stage boundary in the USA and the Bolsovian-Asturian Substage boundary in Western Europe may record a common climatic shift that occurred throughout equatorial regions of Euramerica (cf., Cecil et al., 1985; Bertier et al., 2008; Van Hoof et al., 2012). At the core of this environmental change was a shift to increasing seasonality of equatorial precipitation at all phases of glacioeustatic cycles (Cecil et al., 1985; Roscher and Schneider, 2006), which corresponded to a progressive decrease in grounded ice in the Southern Hemisphere (Fielding et al., 2008) and the onset of a rise in global sea level (Rygel et al., 2008). Because the change may have begun (or at least initially was most strongly manifest) in western Pangea before rapidly spreading eastward, and given the vast equatorial area over which the modification occurred, the boundaries show a degree of temporal and spatial disjunction. As such, the expression of this major climatic shift in the rock record may present a more realistic model for the timing and effects of large-scale environmental changes than do shortterm, globally contemporaneous catastrophes.

Tectonic activity also played a prominent role during this time interval in North America. In fact, the type Atokan Stage resides in the heart of the Ouachita orogenic belt (southern part of the greater Appalachian-Variscan chain), and the Ouachita orogeny culminated near the end of the Atokan (Houseknecht, 1983). Assuredly, elevation of this new mountain range also affected regional climate.

Materials and methods

A selection of North American publications that provide detailed descriptive and illustrated records of Middle Pennsylvanian adpressions was the main source used for comparison with the macrofloral assemblage (Table 3). The list focuses on the Illinois Basin, and Indiana in particular, but includes records from the Western Interior, Appalachian, and Michigan basins of the USA, and the Maritimes Basin of Atlantic Canada. Almost all fossil-taxa in the macrofloral assemblages (Table 1) are described; the only exceptions are *Calamites* spp. indet. and the putative conifer cone scale. Likewise, most taxa are illustrated in Figures 8–11 (including the conifer cone scale, Fig. 11.7).

Repositories and institutional abbreviations.—Plant fossils are stored at the National Museum of Natural History (NMNH), Smithsonian Institution, Washington, DC, and illustrated

Table 3. Main literature sources used for comparison with macrofloral assemblage, and to compile synonymy lists. All publications provide detailed descriptive and/or illustrated records of Middle Pennsylvanian plant-fossil adpressions in North America, with focus on Illinois Basin (* indicates record from Indiana).

ILLINOIS BASIN Lesquereux (1866) Lesquereux (1870) Jackson (1916)* Noé (1925) Janssen (1940) Langford (1958) Canright (1959)* Langford (1963) Wood (1963)* Darrah (1969) Boneham (1974)* Gastaldo (1977) Janssen (1979) Pheifer (1979)* Jennings (1990) Wittry (2006)

WESTERN INTERIOR BASIN White (1893) White (1899) Sellards (1908)

Basson (1968)

Moore et al. (2014)

APPALACHIAN BASIN Lesquereux (1868) Lesquereux (1879–84) Gillespie et al. (1978) Oleksyshyn (1982) Blake et al. (2002)

MICHIGAN BASIN Arnold (1934) Arnold (1949)

MARITIMES BASIN **Bunbury** (1847) Bell (1938) Bell (1940) Bell (1962)

Zodrow and McCandlish (1980)

Zodrow (1989)

Zodrow and Cleal (1998)

Bashforth (2005)

specimens are reposited in the Plant Type and Illustrated Collection under USNM numbers 558409, 559867-559875, and 594359-594391. Components of the assemblage were the subject of a dissertation (Comer, 1992) that included specimens from several stratigraphic levels and depositional facies not included in this study.

Systematic paleontology

Division Tracheophyta Class Lycopsida Order Isoëtales Bartling, 1830 Family Diaphorodendraceae DiMichele and Bateman, 1992 Genus Synchysidendron DiMichele and Bateman, 1993 Synchysidendron sp. cf. S. andrewsii (Lesquereux, 1879-80) Moore, Wittry, and DiMichele, 2014 Figure 8.1

1879-80 Lepidodendron Andrewsii Lesquereux, p. 389, pl. 64, fig. 6.

1938 Lepidodendron dichotomum Arber var. bretonense Bell, p. 92, pl. 95, figs. 5–9, pl. 96, fig. 1, pl. 97, fig. 4.

1940 Lepidodendron dichotomum; Bell, p. 122, pl. 7, fig. 4.

1944 Lepidodendron dichotomum var. bretonense; Bell, p. 89, pl. 45, fig. 3.

1962 Lepidodendron bretonense Bell, p. 53, pl. 47, figs. 5, 6(?), pl. 48, figs. 4, 6(?).

(?)1963 Lepidodendron Sternberg sp.; Langford, p. 153, figs. 712, 713.

1980 Lepidodendron bretonense: Zodrow and McCandlish. p. 80, pl. 116, fig. 1, pl. 118, figs. 1(?), 2.

1985 Lepidodendron bretonense; Wnuk, p. 169, pl. 2, figs. 7, 8.

2005 Diaphorodendron bretonense (Bell) Bashforth, p. 36, pl. 2, figs. 1-4, 7, 8.

2006 Lepidodendron andrewsii; Wittry, p. 106, figs. 1-3 [holotype: figs. 1, 2].

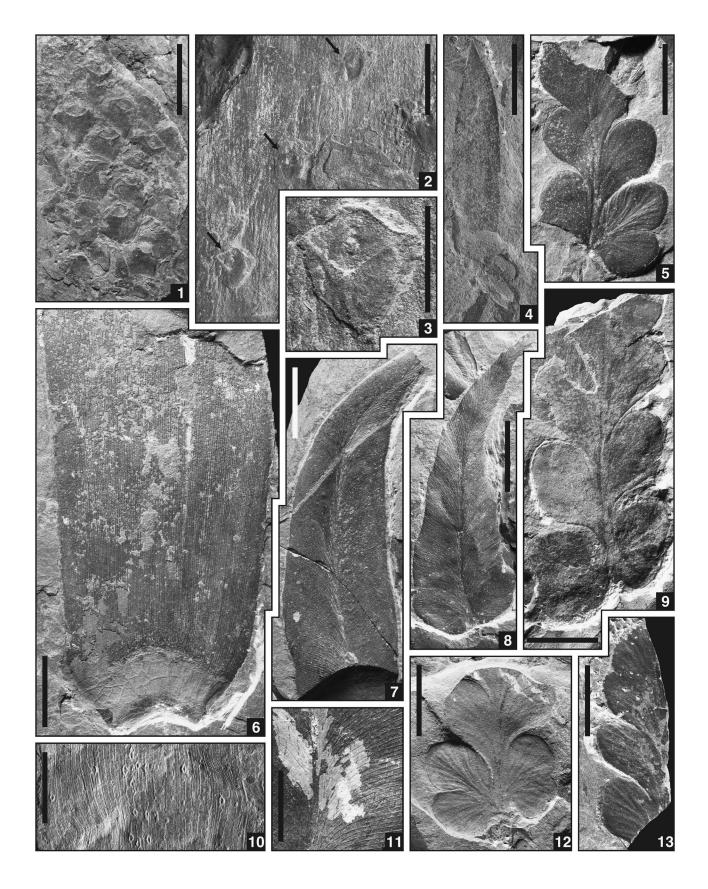
2014 Synchysidendron andrewsii (Lesquereux) Moore, Wittry and DiMichele, p. 28, pl. 1, figs. 1–13.

Holotype.—USNM 15430 (part) and USNM 15431 (counterpart), Plant Type and Illustrated Collection, National Museum of Natural History, Smithsonian Institution, Washington, from Francis Creek Shale, Carbondale Formation (middle Desmoinesian), Illinois Basin, Mazon Creek, Illinois.

Occurrence.—USA and Canada: common throughout Middle Pennsylvanian (Bell, 1938, 1962; Bashforth, 2005; DiMichele et al., 2013a; Moore et al., 2014).

Material examined.—One specimen from Unit 1.

Description.—The axis fragment is 42 mm long and 23 mm wide. Leaf cushions are contiguous, crowded, asymmetrical, 7-9 mm long and 6-7 mm wide (L:W ratio ~1.3), and quadrilateral to napiform with curved to obtusely rounded lateral margins. Upper angles appear obtusely to acutely pointed, whereas lower angles appear acutely pointed (but are



overlapped by the top of the subjacent cushion). Leaf scars are situated in the upper half of a cushion, slightly elevated, $1.2-1.8 \text{ mm} \log, 2.3-4.0 \text{ mm}$ wide (L:W ratio ~ 0.5), with a broadly

convex lower margin, a triangular upper margin with straight sides that converge to an obtusely pointed top, and acutely pointed lateral angles. Cicatricules are not preserved and their position and dimensions are unknown. A gently arched ridge runs from each lateral angle of the leaf scar to the lateral margin of the leaf cushion, separating the cushion into upper and lower fields that lack obvious ornamentation.

Remarks.—In an adpressed state, the absence of infrafoliar parichnos scars beneath the leaf scar is the overriding character that confers assignment to Synchysidendron or Diaphorodendron DiMichele, 1985 of the Diaphorodendraceae (DiMichele, 1985; DiMichele and Bateman, 1992), and distinguishes it from true Lepidodendron Sternberg, 1820 (sensu DiMichele, 1983) of the Lepidodendraceae. Other features, such as the fleshiness of leaf cushions, the presence on large axes of flat interareas between leaf cushions, and the degree of ornamentation on the lower keel of the leaf cushion contribute to generic identification. Segregation of the Diaphorodendraceae from the Lepidodendraceae is more apparent in anatomically preserved examples, as the vegetative anatomy and reproductive organs are conspicuously different (DiMichele, 1979a, 1979b, 1983, 1985; DiMichele and Bateman, 1992). Furthermore, the two families can be differentiated by their growth habits (Wnuk, 1985, DiMichele et al., 2013a). Trunks of Synchysidendron and Diaphorodendron bear two opposite rows of large deciduous lateral branches (or branch scars) that extend into a dichotomously branched crown, whereas Lepidodendron only has a terminal dichotomous crown.

Leaf cushions of the study specimen approach equilateral dimensions (L:W ratio ~1.3), are contiguous to slightly overlapping, and markedly protrude from the stem surface, supporting assignment to *Synchysidendron* rather than *Diaphorodendron*, which tends to have relatively flat and non-protruding leaf cushions with a higher L:W ratio (Wnuk, 1985; DiMichele and Bateman, 1992; DiMichele et al., 2013a; Moore et al., 2014). The axis fragment probably was derived from a deciduous lateral branch, the most common litter elements of the Diaphorodendraceae (DiMichele et al., 2013a).

The specimen resembles adpressed examples of *Synchysidendron andrewsii*, a fossil-species that was documented by Moore et al. (2014) and is the most common form of *Synchysidendron* in the Middle Pennsylvanian of the USA (DiMichele et al., 2013a). The taxon has been recorded in Atlantic Canada under *Lepidodendron dichotomum* var. *bretonense* (Bell, 1938, 1944), *Lepidodendron bretonense* (Bell, 1962; Zodrow and McCandlish, 1980), and incorrectly assigned to *Diaphorodendron bretonense* by Bashforth (2005). Anatomically preserved axes of comparable morphology have been referred to *Synchysidendron*

dicentricum (Felix, 1952) DiMichele and Bateman, 1993 (DiMichele, 1979b; DiMichele and Bateman, 1992, 1993; DiMichele et al., 2013a). Similar to examples of *S. andrewsii* in Moore et al. (2014, pl. 1, figs. 1–13), the specimen has protuberant, napiform to quadrilateral leaf cushions, leaf scars wider than long that are situated in the upper half of the cushion, a gently arched ridge that runs between the scar and cushion margins, and the faint impression of a notch in the upper field just below the upper angle; such notches are exemplified in DiMichele et al. (2013a, fig. 5b). However, *S. andrewsii* tends to have weakly developed upper and lower keels and plications on the lower keel; absence of these features in the specimen makes referral uncertain.

Family Sigillariaceae(?) Unger (in Endlicher), 1842 Genus *Asolanus* Wood, 1860 cf. *Asolanus camptotaenia* Wood, 1860 Figure 8.2, 8.3

- 1860 Asolanus camptotænia Wood, p. 238, pl. 4, fig. 1.
- 1866 *Sigillaria monostigma* Lesquereux, p. 449, pl. 42, figs. 1–5.
- 1869 *Sigillaria camptotænia* (Wood) Wood, p. 342, pl. 9, fig. 3.
- 1870 Sigillaria (Knorria) monostigma; Lesquereux, p. 446, pl. 26, fig. 5.
- 1879-80 *Sigillaria monostigma*; Lesquereux, p. 468, pl. 73, figs. 3–6.
- 1899a *Sigillaria (Asolanus) camptotænia*; White, p. 230, pl. 69, pl. 70, figs. 1, 3(?), 4.
- 1925 Sigillaria camptotaenia; Noé, pl. 11, figs. 1, 3.
- 1938 Asolanus camptotaenia; Bell, p. 102, pl. 104, figs. 1–3.
- 1940 *Asolanus camptotaenia*; Janssen, p. 22, pl. 5, figs. 1, 2, 3(?), pl. 6(?).
- 1949 *Asolanus camptotaena* [sic]; Arnold, p. 179, pl. 11, fig. 3.
- 1958 Asolanus camptotaenia; Langford, p. 108, text-figs. 200–202.
- 1962 Asolanus camptotaenia; Bell, p. 56, pl. 48, figs. 1–3.
- 1968 Asolanus camptotaenia; Basson, p. 40, pl. 1, figs. 1, 2.
- 1969 Asolanus camptotaenia; Darrah, p. 182, pl. 34, fig. 1.
- 1977 Asolanus campotaenia [sic]; Gastaldo, text-fig. 21.
- 1978 Asolanus camptotaenia; Gillespie et al., p. 50, pl. 20, figs. 1, 3.
- 2006 Asolanus camptotaenia; Wittry, p. 111, fig. 1.

Figure 8. Specimens from Units 1 and 2 (Fig. 3) above Minshall Coal, Clay County, Indiana; scale bars = 10 mm (1, 2, 4–10, 12, 13) or 5 mm (3, 11). (1) Synchysidendron sp. cf. S. andrewsii (Lesquereux, 1879-80) Moore, Wittry, and DiMichele, 2014, lycopsid bark, Unit 1, USNM 559868. (2, 3) cf. Asolanus camptotaenia Wood, 1860, lycopsid bark, with arrows (2) indicating position of leaf cushions and details of lowermost leaf cushion (3), Unit 1, USNM 559867. (4) Lepidostrobophyllum sp. indet., lycopsid cone scale, Unit 1, USNM 559870. (5), "Mariopteris" anthrapolis Langford, 1958, lyginopteridalean pteridosperm foliage, ultimate pinna with proximal quadrate pinnules that exhibit strongly incised acroscopic margin and division into rounded lobes, passing apically into strongly decurrent and spatulate to obovate pinnules, part of 12, Unit 2, USNM 559873. (6) Cordaites sp. indet., cordaitalean leaf, with well-preserved leaf base, Unit 2, USNM 594360. (7) Macroneuropteris scheuchzeri (Hoffmann, 1826a) Cleal, Shute, and Zodrow, 1990, medullosalean pteridosperm foliage, subfalcate pinnule, Unit 2, USNM 594361. (8) Macroneuropteris scheuchzeri, subfalcate pinnule exhibiting initial stages of differentiation of pinnule base into two symmetrical lobes, eventually becoming small orbicular pinnules at base of trifoliate pinnule form, Unit 2, USNM 559875. (9) "Mariopteris" anthrapolis, Unit 2, ultimate pinna with large quadrate pinnules near base and spatulate to obovate pinnules near top, USNM 559871. (10) Cordaites sp. indet., with spirorbiform worm-tubes (microconchids) affixed to surface, Unit 1, USNM 559869. (11) Macroneuropteris scheuchzeri, exhibiting hair-like structures characteristic of taxon, Unit 2, USNM 594362. (12) "Mariopteris" anthrapolis, counterpart of 5, Unit 2, USNM 559873. (13) "Mariopteris" anthrapolis, distal end of ultimate pinna with strongly decurrent and spatulate to obovate pinnules that meld into large lobate terminal pinnule with subtly crenulate margin, Unit 2, USNM 559874.

Holotype.—ASNP 695, Paleobotany Collection, Academy of Natural Sciences of Drexel University, Philadelphia, from "Anthracite Region of Pennsylvania" (Darrah, 1969, p. 183).

Occurrence.—USA: middle(?) Atokan to middle Virgilian, most common in upper Atokan to upper Desmoinesian (Darrah, 1969; Gillespie and Pfefferkorn, 1979; Oleksyshyn, 1982; Blake et al., 2002). UK: lower Duckmantian to lower Cantabrian (Cleal, 2007). Atlantic Canada: middle Asturian (Bell, 1938).

Material examined.—Two specimens from Units 1 and 4.

Description.—The illustrated axis fragment is 130 mm long and 105 mm wide, and involves three preservation states that represent degrees of decortification. All preservation modes exhibit longitudinally corrugated interfoliar areas, expressed as subparallel to slightly flexuous and discontinuous ridges and furrows; corrugations are coarser on the outermost layer, and finer on the innermost layer. Leaf cushions (Fig. 8.3) are spirally arranged and widely spaced, ranging from 14-23 mm apart (measured center to center in the helix), and their morphology is only preserved in the two outermost layers. Cushions protrude slightly from the axis surface, 6-8 mm long, 5-7 mm wide (L:W ratio ~1.1), with rhomboidal to subtriangular outlines (widest at the top). Upper angles are broadly convex, lower angles are obtusely rounded, and lateral angles are obtusely rounded to pointed. Leaf scars are situated at the very top of a cushion, slightly elevated, 2.0-3.3 mm long, 3.5-5.0 mm wide (L:W ratio ~ 0.6), with broadly curved upper and lower margins that converge laterally into acutely rounded to pointed lateral angles. Situated in the center of a leaf scar is a single prominent cicatricule, which is circular, ~1 mm in diameter, and consists of a raised rim around a central hollow; other equivocal pits may occur above and/or below the main cicatricule. Leaf cushions appear smooth and lack obvious ornamentation, although some exhibit a faint and finely pustular texture.

Remarks.—Specimens belong to one of a number of lycopsid genera characterized by widely spaced leaf scars, but identification is ambiguous because they exhibit a blend of features typical of multiple taxa. The relatively coarse longitudinal interfoliar corrugations resemble some fossil-species of Bothrodendron Lindley and Hutton, 1833 (particularly Bothrodendron punctatum Lindley and Hutton, 1833) and Cyclostigma Haughton, 1860, the latter synonymous with Pinakodendron Weiss (in Weiss and Sterzel), 1893 according to Cambier and Renier (1912) and Crookall (1964). However, neither of these fossil-genera have leaf cushions, leaf scars are

oval to almost circular with rounded and inconspicuous lateral angles, leaf scars contain three foliar cicatricules, and a prominent ligule pit occurs above the leaf scar in *Bothrodendron* (absent in *Cyclostigma*). Because all of these morphological traits are incongruent with the material at hand, identity is unlikely.

The specimens most strongly resemble Asolanus (usually reported as Asolanus camptotaenia), which has affinities with Sigillaria Brongniart, 1822 (Wood, 1869; White, 1899a). Like the study examples, this taxon has leaf cushions with a convex upper angle and subtriangular lower angle, leaf scars that are rhomboidal to lenticular with sharp lateral angles, and a centrally positioned cicatricule that is circular and ring-like (Crookall, 1964). Asolanus camptotaenia exhibits a wide range of morphologies depending on preservation state and position on the original plant. However, in its most recognizable form, the species has unique interfoliar ornamentation that consists of straight to slightly flexuous, parallel striations that extend perpendicular from the four edges of the leaf scar, resulting in a rhomboidal pattern on the stem surface. This is inconsistent with the longitudinally corrugated surface on the specimen, so confident assignment to A. camptotaenia is not possible.

Incertae sedis Genus Cyperites Lindley and Hutton, 1832 Cyperites bicarinatus Lindley and Hutton, 1832 emend. Rex, 1983 Figure 9.9

1832 *Cyperites bicarinata* Lindley and Hutton, p. 123, pl. 43, figs. 1, 2.

1944 *Lepidophyllum* Brongniart sp.; Bell, p. 96, pl. 59, fig. 2.

(?)1959 *Lepidophyllum longifolium* Brongniart; Canright, pl. 1, fig. 9.

1963 *Lepidophyllum bicarinatum* (Lindley and Hutton) Langford, p. 159, text-fig. 726.

1963 *Lepidophyllum longifolium*; Langford, p. 159, text-fig. 728.

1963 Lepidophyllum sp.; Langford, p. 159, text-fig. 729.

1963 *Lepidophyllum* sp.; Wood, p. 37, pl. 2, fig. 2.

1967 Lepidophyllum longifolium; Tidwell, p. 22, pl. 1, fig. 3.

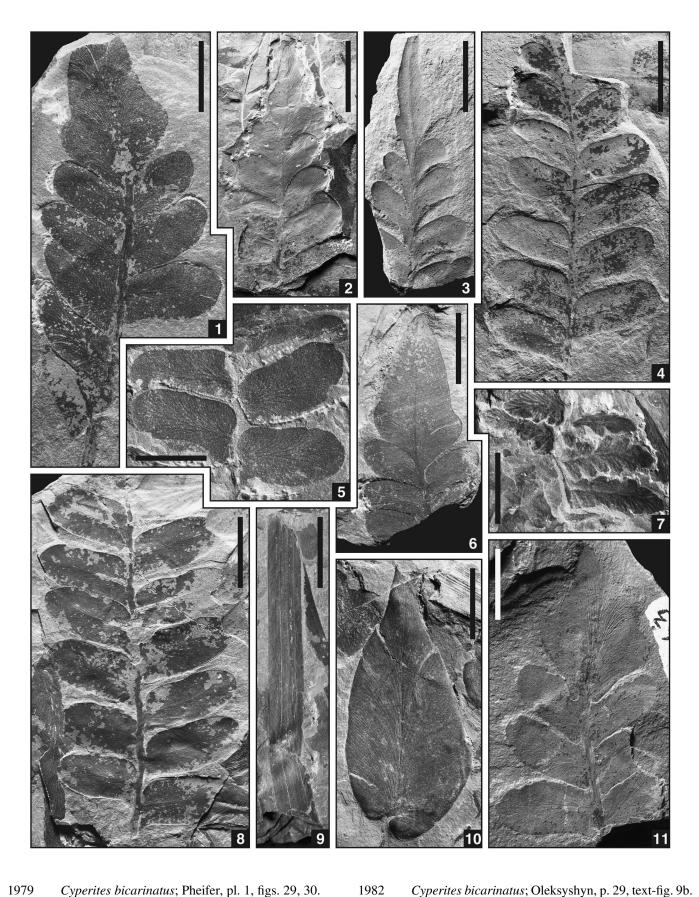
1974 Lepidophyllum longifolium; Boneham, p. 97, pl. 1, fig. 6.

1977 *Lepidophylloides* Snigirevakaya sp.; Gastaldo, text-fig. 18.

1978 Lepidophylloides sp.; Gillespie et al., p. 45, pl. 12, fig. 1, pl. 13, fig. 2.

1979 Lepidophyllum longifolium; Janssen, p. 62, text-fig. 44.

Figure 9. Specimens from Unit 2 (Fig. 3) above Minshall Coal, Clay County, Indiana; all scale bars = 10 mm. (1) Neuropteris flexuosa Sternberg, 1825, medullosalean pteridosperm foliage, pinna with large rhomboid to deltoid terminal pinnule, USNM 594364. (2) Neuropteris ovata Hoffmann, 1826a, 1826b, medullosalean pteridosperm foliage, pinna with small elongate terminal pinnule and 'mixoneurid' habit in distal lateral pinnules, USNM 594366. (3) Neuropteris ovata, pinna with small elongate terminal pinnule and 'mixoneurid' habit in distal lateral pinnules, USNM 594365. (4) Neuropteris ovata, pinna with small elongate terminal pinnule and 'mixoneurid' habit in distal lateral pinnules, USNM 594365. (4) Neuropteris ovata, pinna with lateral pinnules having a strongly incised acroscopic base and a pronounced basiscopic auricle, USNM 594363. (5) Neuropteris flexuosa, showing rounded acroscopic base of lateral pinnules, USNM 59872. (6) Neuropteris flexuosa, pinna with large rhomboid to deltoid terminal pinnule, USNM 594367. (7) Sphenopteris sp. indet., fern foliage of unknown affinity, USNM 594368. (8) Neuropteris ovata, pinna with lateral pinnules having a strongly incised acroscopic base and a pronounced basiscopic auricle, USNM 594359. (9) Cyperites bicarinatus Lindley and Hutton, 1832, lycopsid leaf, USNM 594369. (10) Macroneuropteris scheuchzeri (Hoffmann, 1826a) Cleal, Shute, and Zodrow, 1990, medullosalean pteridosperm foliage, subtriangular pinnule with assiscopic auricle, USNM 594371.



1983

1979 *Cyperites bicarinatus*; Pheifer, pl. 1, figs. 29, 30. (?)1980 *Sigillariophyllum* sp. Grand'Eury; Zodrow and McCandlish, p. 89, pl. 144.

Cyperites bicarinatus; Oleksyshyn, p. 29, text-fig. 9b. *Cyperites bicarinatus* Lindley and Hutton emend. Rex, p. 81, pl. 1, pl. 4 [neotype: pl. 4, fig. 1].

1992	Cyperites	sp.; Tidwell et	al., p. 1018,	text-fig. 4.4.
2005	Cyperites	bicarinatus;	Bashforth,	p. 36, pl.

figs. 1, 2.

2006 *Cyperites* sp.; Wittry, p. 103, figs. 1, 3–5.

Type material.—Syntypes (lost?) from Leebotwood Coal Pit, Halesowen Formation (upper Asturian or lower Cantabrian), Shropshire, UK. Neotype (V.61563), Natural History Museum, London, from roof-shale of No. 10 Coal Seam, Kilmersdon Tip, Farrington Formation (upper Asturian), Radstock Coalfield, Somerset, UK.

Occurrence.—Euramerica: common throughout Carboniferous.

Material examined.—Thirty-four specimens from Units 1 and 2.

Description.—Leaves are linear and taper slightly distally, have entire lateral margins, and are up to 63 mm long (neither bases nor apices preserved) and 2–7 mm wide. A number of ridges and furrows exist on the leaf surface; an elevated or depressed belt (1–2 mm wide) runs along the center of the leaf, and a thin medial furrow (midvein?) may be present.

Remarks.—Isolated, linear lycopsid leaves are assigned to the fossil-genus *Cyperites*. The leaves had a complex 'butterfly' shape in transverse section, and the presence or absence (and number) of ridges and furrows on the exposed leaf surface depends on the path of the fracture plane across the fossil (Rex, 1983). Wider examples of *Cyperites* tend to belong to sigillarians and narrower leaves to lepidodendroids (Bashforth, 2005). Both size fractions exist in the assemblage, suggesting that both groups may be present, although no other macrofloral elements of *Sigillaria* were identified.

Genus *Lepidostrobophyllum* Hirmer, 1927, emend. Allen, 1961 *Lepidostrobophyllum* sp. indet.

Figure 8.4

Occurrence.—Euramerica: fossil-genus common throughout Carboniferous.

Material examined.—Five specimens from Units 1, 2, and 4.

Description.—Sporophyll laminae are 7–11 mm wide, broadest about 1/3 the way up the blade, ~20–45 mm long, and linear-lanceolate with convex, entire lateral margins that converge to an acuminate tip. Lateral angles at the base of sporophyll laminae do not project outwards, but appear flush with the

pedicel top. Midveins (a rib on the adaxial surface) are 0.5-1.0 mm wide and faintly longitudinally striated. Pedicels are ~ 10 mm long and 5 mm wide, widest at the contact with the sporophyll lamina, have straight to slightly convex margins, and may terminate in a sharp base. An oval, thickened rim may be superimposed on the pedicel surface.

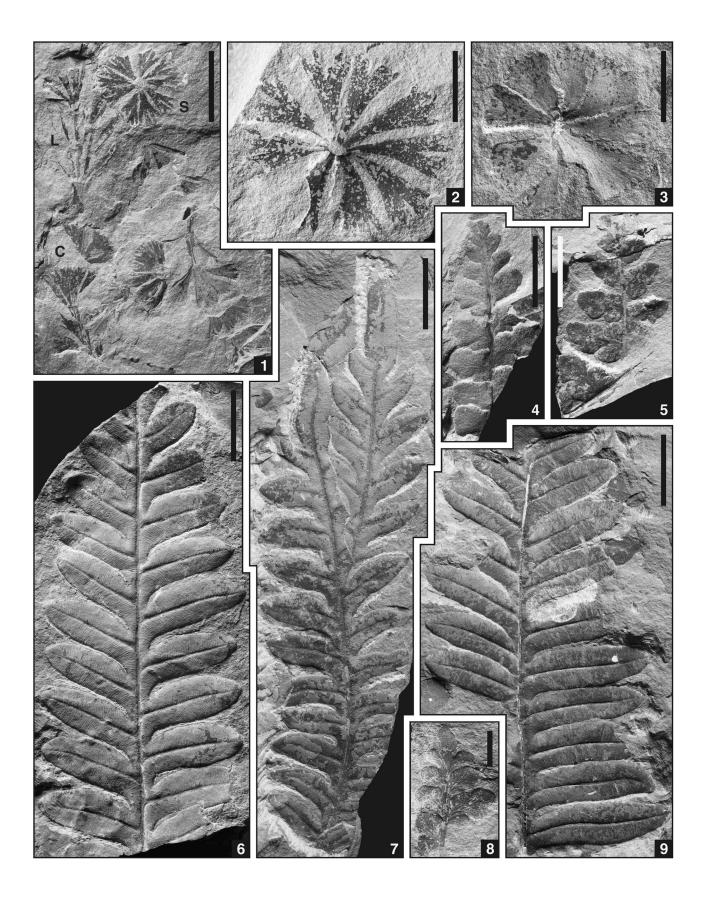
Remarks.—The description is imprecise due to poor preservation. Based on size and shape, the isolated sporophylls disaggregated from a lepidodendroid cone, and thus are assigned to the fossil-genus *Lepidostrobophyllum*. More than one fossil-species may be involved, but the closest comparison is with *Lepidostrobophyllum lanceolatum* (Lindley and Hutton, 1831) Bell, 1938 (e.g., Bell, 1938, pl. 98, figs. 7–11; Crookall, 1966, pl. 99, figs. 4, 5, under *Lepidostrobus lanceolatus*), which Lesquereux (1870, 1879-80) described under the junior synonym *Lepidostrobus oblongifolius*.

Class Sphenopsida Order Sphenophyllales Seward, 1898 Family Bowmanitaceae Meyen, 1978 Genus *Sphenophyllum* Brongniart, 1822

Sphenophyllum cuneifolium (Sternberg, 1821) Zeiller, 1878-79a Figure 10.1–10.3

- 1821 *Rotularia asplenioides* Sternberg, p. 30, pl. 26, figs. 4a, 4b (alternate name).
- 1821 Rotularia cuneifolia Sternberg, p. 33, pl. 26, figs. 4a, 4b.
- 1825 Rotularia saxifragaefolia Sternberg, p. 45, p. XXXII, pl. 55, fig. 4.
- 1831 Sphenophyllum erosum Lindley and Hutton, p. 41, pl. 13.
- 1847 *Sphenophyllum erosum*; Bunbury, p. 430, pl. 23, figs. 3a, 3b.
- 1848 Sphenophyllum saxifragaefolium (Sternberg) Göppert (in Bronn), p. 1166.
- 1868 Sphenophyllum trifoliatum Lesquereux, p. 853, pl. 1, fig. 7.
- 1878-79a Sphenophyllum cuneifolium (Sternberg) Zeiller, p. 30, 31, pl. 161, figs, 1, 2.
- (?)1884 Sphenophyllum saxifragæfolium; Lesquereux, p. 726, pl. 93, figs. 9, 9a.
- 1891 Sphenophyllum erosum; Newberry, p. 215, pl. 19, fig. 1.
- (?)1916 Sphenophyllum cuneifolium; Jackson, p. 411, pl. 1, fig. 6.

Figure 10. Specimens from Unit 3 (Fig. 3) above Minshall Coal, Clay County, Indiana; scale bars = 10 mm (1, 4–7, 9) or 5 mm (2, 3, 8). (1) Sphenophyllum cuneifolium (Sternberg, 1821) Zeiller, 1879, sphenophyll foliage, with whorls of several leaf forms, including (C) cuneate 'normal' leaves (forma typica), (S) cuneate 'divided' leaves (forma saxifragaefolium), and (L) 'deeply divided' to 'linear' leaves, USNM 594376. (2) Sphenophyllum cuneifolium, whorl with cuneate 'divided' leaves (forma saxifragaefolium), USNM 594379. (3) Sphenophyllum cuneifolium, whorl with cuneate 'normal' leaves (forma typica), USNM 594377. (4) Eusphenopteris neuropteroides (Boulay, 1876) Novik, 1947, lyginopteridalean pteridosperm foliage, ultimate pinna with trilobate to bilobate pinnules near base, passing upwards to quadrate or ovate pinnules and topped by small lobate terminal pinnule, USNM 594380. (5) Eusphenopteris neuropteroides, penultimate pinna, from which arises ultimate pinna with trilobate to bilobate pinnules near base and quadrate to year trilobate top, USNM 594378. (6) Alethopteris densinervosa Wagner, 1968, medullosalean pteridosperm foliage, biconvex pinnule form, USNM 594372. (7) Alethopteris densinervosa, subtriangular pinnule form on pinnae topped by a small allantoid terminal pinnule, USNM 594375. (8) Eusphenopteris neuropteroides, strongly decurrent rounded pinnules and small terminal pinnule at distal end of ultimate pinna, USNM 594374. (9) Alethopteris densinervosa, exhibiting transition from biconvex pinnule form at top of pinna to elongate form at base, USNM 594373.



1980

1997

(?)1982

19	934	Sphenophyllum cuneifolium; Arnold, p. 184, pl. 2,
19	934	fig. 4, pl. 4, fig. 6. Sphenophyllum saxifragaefolium; Arnold, p. 184,
		pl. 2, fig. 2, pl. 3, fig. 6, pl. 4, fig. 3.
19	938	<i>Sphenophyllum cuneifolium</i> ; Bell, p. 89, pl. 92, figs. 6–8.
19	944	Sphenophyllum cuneifolium; Bell, p. 105, pl. 75, fig. 6.
19	944	Sphenophyllum cuneifolium forma saxifragaefolium; Bell, p. 105, pl. 75, fig. 5, pl. 76, fig. 10.
19	949	Sphenophyllum cuneifolium; Arnold, p. 185, pl. 18, figs. 1, 3, 6, 9.
19	949	Sphenophyllum saxifragaefolium; Arnold, p. 185, pl. 18, figs. 5, 8.
19	959	Sphenophyllum saxifragaefolium; Canright, pl. 3, fig. 6.
19	963	Sphenophyllum cuneifolium; Wood, p. 46, pl. 5, fig. 6.
19	969	Sphenophyllum cuneifolium; Darrah, p. 178, pl. 44, fig. 2.
19	978	Sphenophyllum cuneifolium; Gillespie et al., p. 72, pl. 28, fig. 4.

Sphenophyllum cuneifolium; Blake et al., pl. 18, figs. 7, 8.Holotype.—E40a and E40b, National Museum, Prague, from

Radnice Member (Bolsovian), Kladno Formation, Central and

Western Bohemian Basin, Vranovice, Czech Republic.

Sphenophyllum

text-figs. 10h, i, 11a, b.

pl. 16, fig. 3 [holotype]

cuneifolium;

Sphenophyllum cuneifolium; Oleksyshyn, p. 43,

Rotularia cuneifolia; Kvaček and Straková, p. 58,

McCandlish, p. 75, pl. 107, fig. 1(?), pl. 109.

Zodrow

and

Occurrence.—USA: lowermost Morrowan to upper Desmoinesian, most common from Atokan to lower Desmoinesian (Abbott, 1958; Darrah, 1969; Gillespie et al., 1978; Gillespie and Pfefferkorn, 1979; Blake et al. 2002). Europe: upper Namurian to upper Bolsovian (Wagner, 1984; Cleal and Thomas, 1994; Cleal, 2007). Atlantic Canada: lower Langsettian to upper Asturian (Bell, 1938, 1944; Zodrow, 1989; Calder, 1998).

Material examined.—Forty-four specimens from Units 2 and 3.

Description.—Whorls comprise leaves of numerous morphologies, depending on the axis order and position on the axis. Axes are 1–5 mm wide, expanded at the nodes, comprise longitudinal ridges and furrows, and may exhibit tiny pits that

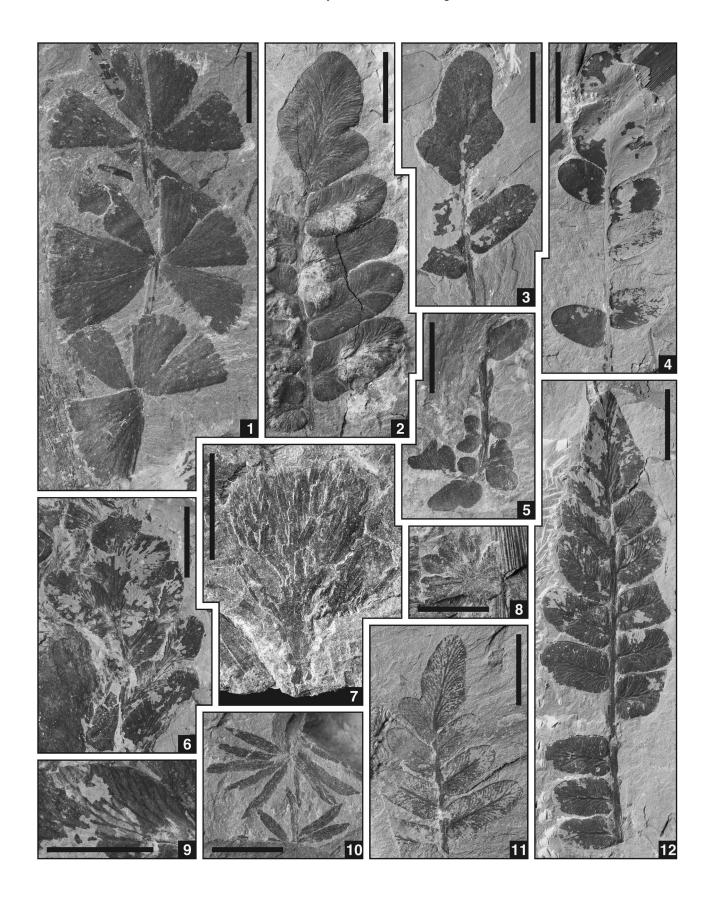
represent trichome bases. Nodes are 2–13 mm long (longest on the widest axes), shortening abruptly near the distal end of an axis (apoxogenesis) and less abruptly near the proximal end (epidogenesis), and each gives rise to a whorl of six to nine leaves. 'Normal' leaves (forma typica; Fig. 10.1, 10.3) are closely spaced, cuneate with straight lateral margins, 5-7 mm long, and 3–4 mm wide at the distal margin, which is slightly convex and comprises seven to nine triangular teeth with acute tips. 'Divided' leaves (forma saxifragaefolium; Fig. 10.1, 10.2) are less closely spaced, cuneate with concave lateral margins, 5–8 mm long, and 2–4 mm wide at the convex distal margin. In less-divided forms, two lobes (with two or three acutely pointed teeth) occur on either side of a central sinus ~1 mm deep; in more-divided forms, four lobes (with one or two teeth) are separated by a 2-3 mm deep central sinus and two shallower secondary sinuses. The most-divided forms (generally situated near the base or top of an axis; Fig. 10.1) comprise two linear lobes that are 7-8 mm long, less than 1 mm wide, and have acuminate tips; solitary linear leaves have the same dimensions. In cuneate leaves, a single vein enters each leaf, branching two to four times before terminating at the end of a tooth or lobe; linear leaves are centered by a single vein.

Remarks.—Much of the assemblage comprises divided leaflets that belong to Sphenophyllum saxifragaefolium, a morphological variant of Sphenophyllum cuneifolium; the designations 'forma typica' and 'forma saxifragaefolium' have sometimes been used (e.g., Bell, 1944; Crookall, 1969; Zodrow, 1989). Sphenophyllum erosum of Lindley and Hutton (1831) and Sphenophyllum trifoliatum of Lesquereux (1868) are junior synonyms of S. cuneifolium. The triangular and acutely pointed teeth on cuneate leaves (forma typica) of S. cuneifolium permit ready distinction from Sphenophyllum emarginatum (Brongniart, 1822) Brongniart, 1828b, which has semicircular to obtusely rounded teeth. Many sphenophylls possessed leaves tipped by stiffened, recurved hooks, and although known for S. cuneifolium (Barthel, 1997, pl. 1, fig. 4), no leaves with 'climber hooks' were observed.

Sphenophyllum emarginatum (Brongniart, 1822) Brongniart, 1828b emend. Batenburg, 1977
Figure 11.1

1822	Sphenophyllites emarginatus Brongniart, p. 234
	pl. 2, figs. 8a, 8b.
1828b	Sphenophyllum emarginatum (Brongniart
	Brongniart, p. 68.
(?)1870	Sphenophyllum cornutum Lesquereux, p. 421
	pl. 19, figs. 1–5.

Figure 11. Specimens from Units 2 and 4 (Fig. 3) above Minshall Coal, Clay County, Indiana; scale bars = 10 mm (2–6, 11, 12) or 5 mm (1, 7–10). (1) Sphenophyllum emarginatum (Brongniart, 1822) Brongniart, 1828b, sphenophyll foliage, with whorls displaying cuneate leaves, Unit 4, USNM 594381. (2) Neuropteris missouriensis Lesquereux, 1879-80, medullosalean pteridosperm foliage, pinna with large obtusely rounded terminal pinnule, Unit 4, USNM 594387. (4) Neuropteris missouriensis, pinna with large obtusely rounded terminal pinnule and small ovate lateral pinnules, Unit 4, USNM 594387. (4) Neuropteris missouriensis, pinna with large obtusely rounded terminal pinnule and small ovate lateral pinnules, Unit 4, USNM 594390. (5) Eusphenopteris sp. indet., lyginopteridalean pteridosperm foliage, Unit 4, USNM 594385. (6, 9) Neuropteris semireticulata Josten, 1962, medullosalean pteridosperm foliage, with details of flexuous to pseudoreticulate venation characteristic of taxon in 11.9, Unit 4, USNM 594382. (7) Possible cone scale of primitive conifer, Unit 2, USNM 558409. (8) Annularia sphenophylloides (Zenker, 1833) Gutbier, 1837, calamitalean foliage, Unit 4, USNM 594383. (10) Annularia sp. cf. A. radiata (Brongniart, 1822) Sternberg, 1825, calamitalean foliage, Unit 4, USNM 594384. (11) Neuropteris missouriensis, top of pinna with small terminal and elongate lateral pinnules, somewhat atypical of taxon, and prominent compression margin on some pinnules, Unit 4, USNM 594386. (12) Neuropteris missouriensis, large pinna with smaller, less obtusely rounded terminal pinnule and lateral pinnules that display prominent compression margin, Unit 4, USNM 594389.



- (?)1879-80 *Sphenophyllum cornutum*; Lesquereux, p. 56, pl. 56, figs. 5, 5a, 5b.
- (?)1879-80 *Sphenophyllum schlotheimii* Brongniart; Lesquereux, p. 52, pl. 2, figs. 6, 7.
- 1899a *Sphenophyllum emarginatum*; White, p. 177, pl. 59, fig. 1d.
- 1925 *Sphenophyllum emarginatum*; Noé, pl. 6, figs. 3, 4, 5(?), 6(?).
- 1938 Sphenophyllum emarginatum; Bell, p. 89, pl. 93, figs. 1, 2, 3(?).
- (?)1940 Sphenophyllum cornutum; Janssen, p. 12, pl. 1, fig. 2b.
- 1949 Sphenophyllum emarginatum; Arnold, p. 185, pl. 18, fig. 4.
- 1958 Sphenophyllum emarginatum; Abbott, p. 339, pl. 44, fig. 66a, pl. 45, fig. 72.
- 1958 Sphenophyllum cornutum; Abbott, p. 334, pl. 44, figs. 66b, 68(?).
- 1958 *Sphenophyllum emarginatum*; Langford, p. 55, text-fig. 78.
- 1959 *Sphenophyllum emarginatum*; Canright, pl. 3, fig. 5.
- 1962 *Sphenophyllum emarginatum*; Bell, p. 46, pl. 44, figs. 1–4, pl. 56, fig. 1.
- 1963 Sphenophyllum emarginatum; Wood, p. 47, pl. 5, fig. 7.
- 1968 Sphenophyllum emarginatum; Basson, p. 64, pl. 6, fig. 4.
- 1969 *Sphenophyllum emarginatum*; Darrah, p. 177, pl. 55, fig. 1.
- 1974 Sphenophyllum emarginatum; Boneham, p. 101, pl. 3, fig. 2.
- 1977 Sphenophyllum emarginatum (Brongniart) Brongniart emend. Batenburg, p. 81, pl. 2, figs. 3–6, pls. 3–6.
- (?)1977 *Sphenophyllum* cf. *emarginatum*; Gastaldo, text-fig. 31.
- 1977 *Sphenophyllum emarginatum*; Remy and Remy, p. 386, text-fig. 237a, b [holotype].
- 1978 Sphenophyllum emarginatum; Gillespie et al., p. 72, pl. 28, fig. 8, pl. 29, fig. 1.
- 1979 *Sphenophyllum emarginatum*; Janssen, p. 92, text-fig. 80.
- 1979 *Sphenophyllum emarginatum*; Pheifer, pl. 4, fig. 52.
- 1980 *Sphenophyllum emarginatum*; Storch, p. 172, pl. 1, fig. 1 [holotype].
- 1980 Sphenophyllum emarginatum; Zodrow and McCandlish, p. 75, pl. 110.
- 1982 *Sphenophyllum emarginatum*; Oleksyshyn, p. 45, fig. 11c–f.
- 1989 Sphenophyllum emarginatum; Zodrow, p. 323, pl. 3, fig. 2, pl. 6.
- 1997 *Sphenophyllum emarginatum*; Wagner and Lyons, pl. 1, fig. 6.
- 2002 *Sphenophyllum emarginatum*; Blake et al., pl. 18, figs. 9, 10.

- 2005 *Sphenophyllum emarginatum*; Bashforth, p. 41, pl. 3, pl. 4, figs. 1, 2, 5, 8, 9.
- 2006 Sphenophyllum emarginatum; Wittry, p. 96, fig. 1.

Holotype.—No. 3519, Muséum national d'Histoire naturelle, Paris, from upper Westphalian (Asturian?), Saar Basin, Germany.

Occurrence.—USA: upper Atokan to lower Virgilian, most common in Desmoinesian (Abbott, 1958; Darrah, 1969; Gillespie and Pfefferkorn, 1979; Oleksyshyn, 1982; Blake et al., 2002). Europe: uppermost Duckmantian to middle Barruelian (Storch, 1980; Wagner, 1984; Laveine 1987). UK: lowermost Bolsovian to lower Cantabrian (Crookall, 1969; Cleal and Thomas, 1994). Atlantic Canada: lowermost Asturian to lower Cantabrian (Bell, 1938; Zodrow, 1989).

Material examined.—One specimen from Unit 4.

Description.—Axes are ~1 mm wide, expanded at the nodes, and comprise pronounced longitudinal ridges and furrows. Nodes are 9–10 mm long and give rise to a whorl of six leaves. Leaves are closely spaced, cuneate with straight lateral margins, 6–8 mm long, and 4–5 mm wide at the distal margin, which is straight to convex, comprises 10 to 12 obtusely rounded teeth, and contains a shallow (0.5–1 mm deep) central sinus. A single vein enters each leaf and branches two to four times before terminating in a tooth.

Remarks.—Sphenophyllum emarginatum can be readily distinguished from undivided cuneate forms of Sphenophyllum cuneifolium (i.e., forma typica), which has smaller leaves with triangular to acutely pointed teeth. Like most sphenophylls, S. emarginatum is remarkably heterophyllous (Batenburg, 1977); the example represents undivided cuneate leaves from last-order branches. Isolated, linear leaves in the assemblage (not described) may record disaggregated whorls from firstorder branches. Not present are deeply laciniate leaves that Lesquereux (1870) erected as Sphenophyllum cornutum, a probable morphological variant of S. emarginatum (Batenburg, 1977). Abbott (1958, pl. 44, fig. 66b) figured S. cornutum-type leaves arising from a plant bearing typical S. emarginatum whorls, but many sphenophyll species produced deeply divided leaves with acuminate lobes that conform to the S. cornutum concept. According to White (1899a), much of what Lesquereux included in Sphenophyllum schlotheimii Brongniart, 1828b probably belongs to S. emarginatum.

> Order Equisetales Du Mortier, 1829 Family Calamitaceae Unger, 1840 Genus *Annularia* Sternberg, 1821

Annularia sp. cf. A. radiata (Brongniart, 1822) Sternberg, 1825 Figure 11.10

- 1822 Asterophyllites radiatus Brongniart, p. 239, pl. 2, figs. 7a, 7b.
- 1825 Annularia radiata (Brongniart) Sternberg, p. XXXI.
- 1925 Annularia radiata; Noé, pl. 4, fig. 2, pl. 5, fig. 1.
- 1938 Annularia radiata; Bell, p. 85, pl. 88, fig. 2.
- 1949 Annularia radiata; Arnold, p. 183, pl. 17, fig. 3.

1958	Annularia radiata; Abbott, p. 317, pl. 41, fig. 56.
1958	Annularia radiata; Langford, p. 40, text-fig. 41.
1959	Annularia radiata; Canright, pl. 3, fig. 1.
(?)1962	Annularia pseudostellata Potonié; Bell, p. 50, pl. 45
	figs. 2, 5.
1963	Annularia radiata; Wood, p. 40, pl. 3, fig. 8.
1974	Annularia radiata; Boneham, p. 99, pl. 2, fig. 6.
(?)1977	Annularia radiata; Gastaldo, text-fig. 29.
1978	Annularia radiata; Gillespie et al., p. 70, pl. 26, fig. 8
	pl. 27, figs. 1, 2(?), 6(?).
(?)1979	Annularia radiata; Pheifer, pl. 3, fig. 42.
(?)1980	Annularia radiata; Zodrow and McCandlish, p. 68
	pl. 91.
2006	Annularia radiata; Wittry, p. 90, figs. 1–3.

Holotype.—Repository and locality details not known.

Occurrence.—USA: upper Morrowan to Missourian, rare above uppermost Atokan (Abbott, 1958; Darrah 1969; Gillespie et al., 1978; Gillespie and Pfefferkorn, 1979; Blake et al., 2002). UK: upper Namurian to upper Duckmantian (Cleal and Thomas, 1994). Atlantic Canada: upper Bolsovian to upper Asturian (Bell, 1938).

Material examined.—Two specimens from Unit 4.

Description.—Axes are ~0.4 mm wide and longitudinally striated, with internodes ~5 mm apart. Whorls are probably symmetrical, slightly overlap the subjacent whorl, and comprise at least eight loosely spaced leaves of similar length. Leaves are linear to linear-lanceolate with acuminate tips, 3.5–11.0 mm long and 0.5–1.0 mm wide (L:W ratio ~7), and broadest near the midpoint but have parallel lateral margins through most of their length. Midveins are 0.1–0.2 mm wide, longitudinally striated, and protrude beyond the distal margin to form a blunt mucro.

Remarks.—The taxonomy of Annularia (calamitalean foliage) remains in disarray despite efforts to stabilize the nomenclature (Walton, 1936; Abbott, 1958; Crookall, 1969; Barthel, 2004). The morphological plasticity of the foliage is particularly troublesome because several fossil-species of Annularia may occur on one plant. None of the specimens examined in this study comprises a complete whorl, therefore the degree of symmetry and number of leaves in a whorl are equivocal; such features are important for specific determination. However, the examples compare best with what Abbott (1958) included in Annularia radiata, although at the low end of the size range. A transfer preparation of a specimen that Walton (1936, pl. 31, figs. 16, 17) determined as A. radiata is remarkably similar, showing leaves that end in a blunt mucro. According to Abbott (1958), specimens referred to Annularia ramosa Weiss, 1881 by White (1893, 1899a) belong to A. radiata.

Annularia sphenophylloides (Zenker, 1833) Gutbier, 1837 Figure 11.8

1833 Galium sphenophylloides Zenker, p. 398, pl. 5, figs. 6–9.
 1837 Annularia sphenophylloides (Zenker) Gutbier, p. 436.

- 1868 Annularia sphenophylloides; Lesquereux, p. 852, pl. 1, figs. 5, 5a.
- 1879-80 *Annularia sphenophylloides*; Lesquereux, p. 48, pl. 2, figs. 8, 9.
- (?)1908 Annularia sphenophylloides; Sellards, p. 425, pl. 53, fig. 5.
- 1925 Annularia sphenophylloides; Noé, pl. 5, figs. 2, 3.
- 1938 Annularia sphenophylloides; Bell, p. 84, pl. 85, fig. 3, pl. 87, fig. 1.
- 1958 Annularia sphenophylloides; Abbott, p. 319, pl. 41, fig. 55.
- 1958 *Annularia sphenophylloides*; Langford, p. 40, text-fig. 44.
- 1962 Annularia sphenophylloides; Bell, p. 51, pl. 44, fig. 7.
- 1963 Annularia sphenophylloides; Wood, p. 41, pl. 3, figs. 7(?), 9.
- 1968 *Annularia sphenophylloides*; Basson, p. 57, pl. 4, fig. 3.
- 1969 Annularia sphenophylloides; Darrah, p. 172, pl. 33, figs. 1, 2, pl. 43, fig. 7.
- 1978 *Annularia sphenophylloides*; Gillespie et al., p. 71, pl. 26, figs. 3(?), 6, 7.
- 1979 Annularia sphenophylloides; Janssen, p. 86, text-fig. 69.
- 1980 *Annularia sphenophylloides*; Zodrow and McCandlish, p. 69, pls. 92, 93.
- 1992 *Annularia sphenophylloides*; Mamay and Mapes, text-fig. 2g.
- 2005 *Annularia sphenophylloides*; Bashforth, p. 43, pl. 5, figs. 2, 4–7, 10.
- 2006 Annularia sphenophylloides; Wittry, p. 91, figs. 1, 2.
- 2011 *Annularia sphenophylloides*; Blake and Gillespie, pl. 5, fig. 3.
- 2013b *Annularia sphenophylloides*; DiMichele et al., p. 291, text-fig. 4.4.
- 2014 Annularia sphenophylloides; Moore et al., p. 34, pl. 3, figs. 6, 7.

Holotype.—Private collection of Dr. Schüler of Jena, Thüringen, Germany, from Carboniferous of Zittau, Sachsen, Germany (Crookall, 1969, p. 736).

Occurrence.—USA: upper Atokan to upper Virgilian, most common above Atokan (Abbott, 1958; Darrah, 1969; Gillespie and Pfefferkorn, 1979; Blake et al., 2002). UK and Atlantic Canada: lower Bolsovian to lower Cantabrian (Bell, 1938; Cleal and Thomas, 1994; Cleal, 1997). Europe: extends to top of Stephanian (Wagner, 1984).

Material examined.—Two specimens from Unit 4.

Description.—Whorls are symmetrical and comprise \sim 12 crowded or partly overlapping leaves of similar length. Leaves are markedly spatulate with obtusely rounded tips (because the mucronate tips are embedded in the matrix), 3–7 mm long and 1–2 mm wide (L:W ratio \sim 3), and broadest close to the distal margin. Midveins are \sim 0.2 mm wide, longitudinally striated, and

flare at the distal margin (terminal expansion) before protruding as an acuminate mucro (not observed).

Remarks.—Annularia sphenophylloides is a common and easily recognized species in Middle to Upper Pennsylvanian strata of Euramerica. The small spatulate leaves are especially characteristic, and the embedding of the mucronate tip in the matrix is common for the taxon.

Annularia sp. indet.

Material examined.—One specimen from Unit 4.

Description.—Leaves are at least 18 mm long (but presumably much longer) and ~ 2 mm wide, parallel-sided for most of their length but tapering towards the base and top, and centered by a midvein ~ 0.2 mm wide. Leaf surfaces are villous, with short trichomes oriented obliquely to the lateral margins.

Remarks.—The specimen is not illustrated given its incompleteness, and identification of these leaves is equivocal. However, the dimensions, presumed shape, and densely villous nature of the leaves point either to Annularia carinata Gutbier, 1849, an earlier synonym of Annularia mucronata Schenk, 1883 (cf., Barthel, 1976; Kerp, 1984), or Annularia spinulosa Sternberg, 1821, the proper name for what is commonly called Annularia stellata (Schlotheim ex Sternberg, 1825) Wood, 1860 (cf., Barthel, 2000). Assignment to A. spinulosa is most tenable given that complete leaves probably were oblanceolate (cf., Abbott, 1958). Annularia stellata auct. has commonly been recorded in North America under Annularia longifolia Brongniart, 1828b and Annularia inflata Lesquereux, 1870 (cf., Janssen, 1940; Abbott, 1958; Darrah, 1969).

Class Filicopsida
Incerti ordinis
Incertae familiae
Genus Sphenopteris (Brongniart, 1822) Sternberg, 1825
Sphenopteris sp. indet.
Figure 9.7

Occurrence.—Euramerica: fossil-genus common throughout Carboniferous.

Material examined.—Five specimens from Units 1 and 2.

Description.—Foliar elements are polymorphic. Penultimate pinnae have a thin (~0.5 mm wide), winged rachis, and give rise to alternate and obliquely inserted ultimate pinnae centered by a weakly to moderately decurrent and somewhat flexuous rachis. Ultimate pinnae are linear, taper to an obtusely rounded tip, and range from 13–30 mm long and 5–12 mm wide. In distal positions, ultimate pinnae grade into pinnatifid pinnules that are linguaeform to subtriangular with an obtusely rounded apex, may overlap their neighbor, and are 8–11 mm long and 4–6 mm wide. Entire pinnules are obliquely inserted (more so distally), confluent, linguaeform to subtriangular, and 3–6 mm long and 1.5–3.0 mm wide. Pinnule midveins are thin and strongly decurrent, and give rise to thin, sparse lateral veins that branch

once (twice in largest pinnules) as they arch slightly to reach the lateral margin at a very oblique angle. Sporangia appear to be situated near the end of larger pinnules, but their dimensions, morphology and arrangement are equivocal.

Remarks.—The rarity and fragmentary nature of specimens, and ambiguity about sporangial organization, precludes taxonomic assignment. Although the fully developed, entire pinnules are linguaeform and thus resemble *Pecopteris* (Brongniart, 1822) Sternberg, 1825, more distal pinnae and pinnatifid pinnules can be subtriangular and appear more 'sphenopteroid.' Thus, inclusion in the fossil-genus *Sphenopteris* is tentative.

Class Spermatopsida Order Medullosales Corsin, 1960 Family Alethopteridaceae Corsin, 1960 emend. Cleal and Shute, 2003

Genus Alethopteris Sternberg, 1825 emend. Wagner, 1968 Alethopteris densinervosa Wagner, 1968 Figures 10.6, 10.7, 10.9, 13.1

1932 *Alethopteris Serli* (Brongniart) Göppert; Corsin, pl. 29, figs. 1, 1a, 1b [holotype].

1961 Alethopteris Serli; Buisine, p. 74, pl. 6, figs. 1, 1a [holotype]

1968 Alethopteris densinervosa Wagner, p. 59, pl. 15, figs. 43–44a. [holotype: fig. 43]

1968 Alethopteris serlii; Basson, p. 71, pl. 11, fig. 1.

(?)1980 "Alethopteris serli morphogroups 1, 2, 3"; Scheihing and Pfefferkorn, pl. 1, figs. 1–6.

1982 Alethopteris serlii; Oleksyshyn, p. 99, text-fig. 20d. 2014 Alethopteris serlii; Moore et al., p. 40, pl. 7, figs. 5, 6.

Holotype.—USTL 349, Collection Paléobotanique, Université des Sciences et Technologies de Lille, Lille, from lower Faisceau d'Ernestine (middle Bolsovian), Northern France Coalfield, France.

Occurrence.—USA: rare in lower to middle Desmoinesian (Basson, 1968; Oleksyshyn, 1982; Moore et al., 2014). Northern France Coalfield: base of Bolsovian to lower Asturian (Buisine, 1961; Wagner, 1968; Laveine, 1987). Atlantic Canada (?): Asturian (Zodrow, 1986; Zodrow and Cleal, 1998).

Material examined.—Twenty-eight specimens from Units 2 and 3.

Description.—Ultimate pinnae are linear, but narrow slightly near the base and taper rapidly to an acutely rounded apex topped by a small, allantoid, individualized terminal pinnule (Fig. 10.7). Lateral pinnules are extremely variable in habit and size (Table 4; Fig. 12). Most pinnules are inserted obliquely (more so distally, perpendicular in larger forms), closely spaced and occasionally overlapping, moderately vaulted, and distinctly confluent with a marked acroscopic incision and a strongly decurrent basiscopic side, resulting in an asymmetrical shape and a 'wing' of lamina about the rachis. Average pinnules (Fig. 10.6, 10.9) have biconvex lateral margins and obtusely to acutely rounded tips, are widest in the middle, 12–15 mm long and 5–6 mm wide (L:W ratio ~2.4), and squatter in distal positions. Fully developed (elongate) pinnules (Figs. 10.9, 13.1)

Table 4. Morphometric data (length, width, L:W ratio, and venation density) from specimens identified as Alethopteris densinervosa Wagner, 1968. To facilitate comparison and validate identification (see Fig. 14), venation density data also are presented from type specimens and other previously illustrated examples of A. densinervosa, Alethopteris missouriensis (White, 1899a) Wagner, 1968, and Alethopteris serlii (Brongniart, 1833-34) Göppert, 1836. Data sources (measurements from actual specimens denoted by *): A. densinervosa (holotype)—Buisine (1961, pl. 6, fig. 1a); A. densinervosa (North American examples)—Basson (1968, pl. 11, fig. 1), Oleksyshyn (1982, text-fig. 20d), and Moore et al. (2014, pl. 7, fig. 6); A. missouriensis (syntypes)—White (1899a, pl. 37, fig. 2*, USNM E3594b; pl. 42, fig. 5, USNM E3591b); A. missouriensis (other North American examples)—unillustrated pinnules on syntype slab of White (1899a, pl. 37, fig. 2*, USNM E3594b) and Wittry (2006, p. 71, fig. 1a); A. serlii (syntypes)—Wagner (1968, pl. 56, figs. 162 [lectotype], 163); A. serlii (North American examples)—Lesquereux (1879-80, pl. 29, figs. 2–4*, USNM 12098–12100), Noé (1925, pl. 38, fig. 1), Langford (1958, text-fig. 436), Bell (1962, pl. 41, fig. 4), Wagner (1968, pl. 57, fig. 167a), Darrah (1969, pl. 50, fig. 1), Scheihing and Pfefferkorn (1980, pl. 1, figs. 1–6), Oleksyshyn (1982, text-fig. 20c), and Blake et al. (2002, pl. 8, fig. 1; pl. 28, fig. 4).

Pinnule shape	Parameter	Range	Percentiles	Mean	N
Alethopteris densinervosa					
Biconvex	Length	8.4–17.3	12.3–14.9	13.6	44
	Width	4.1–7.2	5.2-6.2	5.7	44
	Length:Width	1.7–2.8	2.2–2.5	2.4	44
-	Venation density	38–56	45–52	48	49
Elongate	Length	12.8–30.0	14.6–22.0	18.9	12
	Width	5.1–8.0	5.4–7.0	6.2	15
	Length:Width	2.4–4.3	2.6–3.3	3.0	12
C1-4	Venation density	37–50	41–46	44 9.0	18
Subtriangular	Length Width	4.3–12.1 2.5–5.0	8.2–10.0 2.9–4.0	9.0 3.6	30 30
		2.5-3.0 1.4-3.4	2.9-4.0	3.6 2.5	30
	Length: Width	1.4–3.4 39–55	2.2–2.9 44–50	2.3 47	
Terminal	Venation density	39–33 13.7–14.3		14.0	21
Terminai	Length Width	4.5–4.7	n/a	4.6	2 2 2 4
	Length:Width	3.0	n/a	3.0	2
		3.0 45–53	n/a 47–50	3.0 49	2
All minnules	Venation density	4.3–33.0	9.5–14.9	12.8	88
All pinnules	Length Width	4.5–30.0 2.5–8.0	4.0–6.0	5.0	91
	Length:Width	1.4–4.3	2.3–2.8	2.5	88
	Venation density	37–56	2.3–2.8 44–50	2.3 47	92
	·	37–30	44-30	47	92
Alethopteris densinervosa					
Biconvex	Venation density	42–54	47–53	49	11
Alethopteris densinervosa	(NORTH AMERICAN EXAMPLES				
Variable	Venation density	42–53	44–48	46	12
ATT Aladamental density	COMPINED				
ALL Alethopteris densiner Variable		37–56	44–50	47	115
variable	Venation density	37–30	44–30	47	113
Alethopteris missouriensis	(SYNTYPES)				
Elongate	Length	22.3–29.0	25.0–28.2	26.3	15
	Width	4.2–7.0	5.0-6.0	5.4	17
	Length:Width	4.1–6.3	4.6–5.0	5.0	14
	Venation density	32–48	35–42	39	28
Alethonteris missouriensis	(OTHER NORTH AMERICAN EX	AMPLES)			
Variable	Venation density	32–46	36–41	38	24
	•	32 10	30 11	30	2.
ALL Alethopteris missouri					
Variable	Venation density	32–48	36–41	39	52
Alethopteris serlii (SYNTY	VPFS)				
Biconvex	Venation density	29-38	32–35	33	19
	·	2, 30	32 33	55	17
	H AMERICAN EXAMPLES)				
Variable	Venation density	26–41	29–35	32	82
ALL Alethopteris serlii CC	OMBINED				
Variable	Venation density	26-41	30–35	32	101

reach 30 mm long and 8 mm wide (L:W ratio \sim 3.0), are parallel-sided but narrower near the base, and symmetrical due to reduction of the acroscopic incision and basiscopic wing. Less typical are widely spaced, subtriangular pinnules (Fig. 10.7) \sim 8–10 mm long and 3–4 mm wide (L:W ratio \sim 2.5) with a more acute top. Midveins are non-decurrent, depressed in a distinct furrow, moderately strong, straight to slightly down-curved, and extend to just below the pinnule apex. Lateral veins are thin and dense (typically 44–50 veins per cm, mean 47; Table 4; Figs. 13.1, 14), and originate obliquely but immediately curve and travel straight to reach the lateral margin at \sim 90°. Most veins branch close to the midvein and some bifurcate again on approach to the margin; unbranched veins are rare. Subsidiary veins extend from the rachis into the pinnule base on the basiscopic and acroscopic sides.

Remarks.—Specimens belong to a group of Middle Pennsylvanian alethopterids that Wagner (1968) segregated into five distinct species based on subtle differences in pinnule shape, size, and venation density: Alethopteris serlii, Alethopteris missouriensis (White, 1899a) Wagner, 1968, Alethopteris lonchitifolia Bertrand, 1932, Alethopteris densinervosa, and Alethopteris westphalensis Wagner, 1968. With the exception of A. serlii, excluded from the complex based on its somewhat flexuous venation, Cleal (1978) observed that examples of the other four taxa are present in large assemblages and represent morphological variants of one species (the earliest legitimate synonym being A. missouriensis). Scheihing and Pfefferkorn (1980) corroborated the opinion that these taxa (but including A. serlii) formed a morphological continuum, which they termed the

"Alethopteris serli group." Again excluding A. serlii, Zodrow and Cleal (1998) synonymized the other four taxa under Alethopteris lonchitica Schlotheim ex Sternberg, 1825. Wagner and Álvarez-Vázquez (2008) rejected identification with A. lonchitica (and we agree with their opinion), and maintained that A. missouriensis,

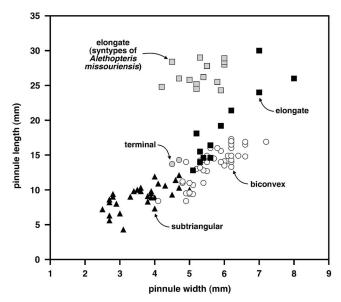


Figure 12. Bivariate (length, width) plot of several pinnule types of *Alethopteris densinervosa* Wagner, 1968 in the macrofloral assemblage ('study specimens' in Table 4), and comparison with elongate pinnules of syntypes of *Alethopteris missouriensis* (White, 1899a) Wagner, 1968 from White (1899a, pl. 37, fig. 2, USNM E3594b; pl. 42, fig. 5, USNM E3591b).

A. lonchitifolia, A. densinervosa, and A. westphalensis are different species with partly overlapping characters. Resolving how many biological species are in this complex will require a detailed morphometric and cuticular analysis of the types and associated specimens, and although some morphometric data are presented here, a full study is outside the scope of this contribution.

Given the morphological similarities among taxa in the aforementioned alethopterid complex, Wagner (1968) considered venation density as a key distinguishing criterion. The venation density in various pinnule forms (biconvex, subtriangular, elongate, and terminal) in the assemblage is provided in Figure 14 and Table 4. The range of all pinnule types is 37–56 veins per cm on the lateral margin, with most pinnules having 44-50 (mean 47) veins per cm. Based on the descriptions of Wagner (1968), only two species in the complex—Alethopteris missouriensis and Alethopteris densinervosa—have a similarly dense venation, both ranging from 40-50 veins per cm. Of the two, the material compares most closely with A. densinervosa. The full range of pinnule morphologies can be accommodated in the species, which was best illustrated by Buisine (1961, pls. 1–7; pl. 8, figs. 1, 1a; pl. 10, figs. 1, 1a, 3, 4) under Alethopteris Serli. Despite the likeness, however, referral of the study specimens to A. densinervosa initially presented a conundrum. In North America, Middle Pennsylvanian alethopterids with this morphology are almost invariably included in serlii (rarely in A. missouriensis), and given that A. densinervosa has to date never been confirmed outside of the Northern France Coalfield, identity with the latter would make this the first record from North America.

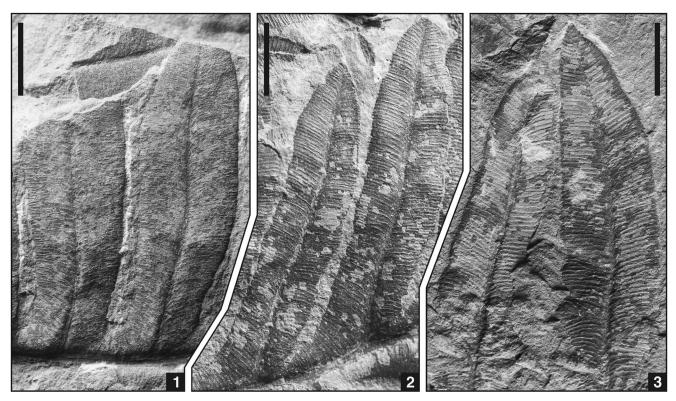


Figure 13. Comparison of elongate pinnule types of *Alethopteris* Sternberg, 1825, showing differences in venation density measured directly from illustrated pinnules; scale bars = 5 mm. (1) *Alethopteris densinervosa* Wagner, 1968, Unit 3, USNM 594391 (43–50 veins per cm). (2) Syntype of *Alethopteris missouriensis* (White, 1899a) Wagner, 1968, illustrated by White (1899a, pl. 37, fig. 2), USNM E3594b (40–44 veins per cm). (3) Specimen of *Alethopteris serlii* (Brongniart, 1833-34) Göppert, 1836, illustrated by Lesquereux (1879, pl. 29, fig. 4), USNM 12100 (32–36 veins per cm).

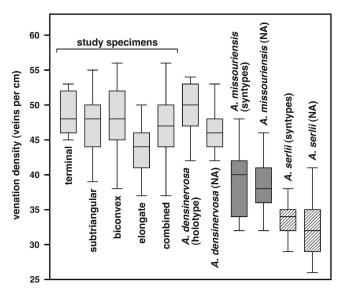


Figure 14. Box and whisker plots showing range, 25th and 75th percentiles, and median of venation densities of several pinnule types from specimens identified herein as *Alethopteris densinervosa* Wagner, 1968. Comparison with venation density data from type specimens and other previously illustrated examples of *A. densinervosa*, *Alethopteris missouriensis* (White, 1899a) Wagner, 1968, and *Alethopteris serlii* (Brongniart, 1833-34) Göppert, 1836 confirm identification of study specimens as *A. densinervosa*. See Table 4 for data sources.

To facilitate identification, venation density data were gathered from illustrations of North American material referred to *Alethopteris serlii* or *Alethopteris missouriensis* (see Table 4 for list of sources), augmented by measurement of actual specimens where possible (e.g., a syntype of *A. missouriensis* [White, 1899a], reillustrated here [Fig. 13.2] and three specimens of *A. serlii* recorded by Lesquereux [1879-80], one reillustrated here [Fig. 13.3]). Venation densities also were measured from photographs of the syntypes of *A. serlii* (Wagner, 1968), a syntype of *A. missouriensis* (White, 1899a), and the holotype of *A. densinervosa* (Buisine, 1961).

These venation density data (Fig. 14; Table 4) reveal that at least three distinct fossil-species (Alethopteris serlii, Alethopteris missouriensis, and Alethopteris densinervosa), united in similarity of pinnule morphology but differentiated by venation density, are present in Middle Pennsylvanian strata of North America. The lateral veins of A. densinervosa are extremely dense (most 44-50, mean 47 veins per cm; Fig. 13.1), comparable to the diagnosis of 40-50 veins per cm (Wagner, 1968), whereas those of A. serlii are more widely spaced (most 30-35, mean 32 veins per cm; Fig. 13.3), faithful to the emended diagnosis of 30–35 veins per cm (Wagner, 1968). The foliar elements of these species are otherwise nearly indistinguishable. The venation density of A. missouriensis is intermediate (most 36-41, mean 39 veins per cm; Fig. 13.2), equivalent to the 35-40 veins per cm claimed by Wittry (2006), but somewhat lower than the 28-42 veins per cm given in the diagnosis (White, 1899a), although Wagner (1968, p. 108) noted that the lower part of this range only applies to what White (1899a, p. 117, pl. 37, figs. 1, 1a) considered the non-varietal form of A. serlii. Alethopteris missouriensis is characterized by more elongate pinnules (Figs. 12, 13.2), and Figure 14 and Table 4 demonstrate that specimens in the assemblage have L:W ratios that largely range from ~2.5 (subtriangular and biconvex forms) to ~3.0 (elongate forms), whereas elongate pinnules of the syntypes of A. missouriensis are narrower (L:W ratio ~5.0). To verify the separation of A. serlii, A. missouriensis, and A. densinervosa based on venation density, 'post-hoc' pairwise comparisons after a Kruskal-Wallis test (H=253.1, p[same] <0.0001) confirm that there are statistically significant differences among the three species groupings (p[same] ≤ 0.003), and that there are no statistically significant differences between the type material of each species and other measured specimens identified as such. These results support the assertion that the three taxa are separate species (Wagner, 1968; Wagner and Álvarez-Vázquez, 2008).

This exercise also demonstrates that Alethopteris densinervosa is not only present in the study population in the Illinois Basin, but has previously been documented in the Western Interior and Appalachian basins under Alethopteris serlii (see synonymy list) by Basson (1968), Oleksyshyn (1982), and Moore et al. (2014), the latter noting comparison with A. densinervosa. Venation density data provided by Zodrow and Cleal (1998, table 1, fig. 7) for specimens included in their concepts of Alethopteris lonchitica and A. serlii suggest that some examples in the Sydney Coalfield probably also belong to A. densinervosa, as Zodrow (1986) initially proposed. In contrast, venation densities recorded for the "Alethopteris serli group" (morphogroups 1 to 3) of Scheihing and Pfefferkorn (1980, table 3, means of 42-43 veins per cm) imply that A. densinervosa was represented, but such high-density values could not be duplicated based on measurements of their illustrated specimens; thus, the material documented by Scheihing and Pfefferkorn (1980) probably belongs to A. serlii.

Family Neurodontopteridaceae Cleal and Shute, 2003 Genus *Macroneuropteris* Cleal, Shute, and Zodrow, 1990 *Macroneuropteris scheuchzeri* (Hoffmann, 1826a) Cleal, Shute, and Zodrow, 1990

Figures 8.7, 8.8, 8.11, 9.10

1826a	Neuropteris Scheuchzeri Hoffmann, p. 157, pl. 1b,
	figs. 1–4.
1847	Neuropteris cordata Brongniart; Bunbury, p. 423,
	nl 21 figs 1s o f

pl. 21, figs. 1a, c–f.

1847 *Neuropteris cordata* Brongniart var. *angustifolia* Bunbury, p. 424, pl. 21, fig. 1b.

1847 *Odontopteris subcuneata* Bunbury, p. 427, pl. 23, figs. 1a, 1b.

Neuropteris hirsuta Lesquereux, p. 417.

1866 Odontopteris Wortheni Lesquereux, p. 432, pl. 36, figs. 1, 1b.

1866 *Odontopteris heterophylla* Lesquereux, p. 433, pl. 38, figs. 2–5.

1868 *Neuropteris hirsuta*; Lesquereux, p. 857, pl. 3, fig. 6, pl. 4, figs. 1–16.

1870 *Neuropteris fasciculata* Lesquereux, p. 381, pl. 5, figs. 1–4.

1870 *Odontopteris subcuneata*; Lesquereux, p. 390, pl. 8, fig. 10, 10b.

1879-80 *Neuropteris hirsuta*; Lesquereux, p. 88, pl. 8, figs. 1, 4, 5, 7, 9.

1969

1974

1977

1977

Neuropteris decipiens; Darrah, p. 94, pl. 1,

figs. 1-4, pl. 2, fig. 3, pl. 5, figs. 1, 2, pl. 6, fig.

Neuropteris scheuchzeri; Boneham, p. 109, pl. 5,

Neuropteris scheuchzeri Hoffman [sic] forma

Neuropteris scheuchzeri Hoffman [sic] forma

2, pl. 8, figs. 1-3, pl. 46, fig. 3, pl. 76, fig. 1.

scheuchzeri Gastaldo, p. 138, text-fig. 50.

decipiens Gastaldo, p. 138, text-fig. 51.

1879-80	Neuropteris angustifolia Brongniart; Lesquereux, p. 89, pl. 8, figs. 2, 3, 6, 8, 10, 11.	1978	Neuropteris scheuchzeri; Gillespie et al., p. 104, pl. 42, figs. 1–3, 5–9, pl. 46, fig. 8.
1879-80	Neuropteris fasciculata; Lesquereux, p. 93, pl. 24, figs. 5, 6.	1979	Neuropteris scheuchzeri; Janssen, p. 149, text-figs. 8, 137.
(?)1879-80	Odontopteris cornuta Lesquereux, p. 128, pl. 22, figs. 7–9.	1979	Odontopteris worthenii; Janssen, p. 161, text-fig. 154.
1879-80	Odontopteris heterophylla; Lesquereux, p. 129,	1979	Neuropteris scheuchzeri; Pfeiffer, pl. 6, figs. 65, 66.
	pl. 22, fig. 6.	1980	Neuropteris scheuchzeri; Zodrow and
1879-80	<i>Odontopteris worthenii</i> ; Lesquereux, p. 130, pl. 22, fig. 1.		McCandlish, p. 47, pl. 37, figs. 2, 3, pls. 38–42, pl. 43, fig. 1.
(?)1879-80	<i>Odontopteris subcuneata</i> ; Lesquereux, p. 134, pl. 22, figs. 4, 5.	1980	Odontopteris subcuneata; Zodrow and McCandlish, p. 53, pl. 53, figs. 1, 2, pl. 54.
1880	Neuropteris decipiens Lesquereux, p. 93.	1982	Neuropteris decipiens; Oleksyshyn, p. 105, text-
1884	Neuropteris decipiens; Lesquereux, p. 733, pl. 94,		fig. 24a.
1000	figs. 1, 2.	1982	Neuropteris scheuchzeri; Oleksyshyn, p. 119,
1899a	Odontopteris? bradleyi Lesquereux; White,	1000	text-figs. 24b–f, 25a, b.
1899a	p. 125, pl. 42, fig. 2, 2a. Neuropteris scheuchzeri; White, p. 132, pl. 37,	1990	<i>Macroneuropteris scheuchzeri</i> (Hoffmann) Cleal, Shute, and Zodrow, p. 488.
1099a	fig. 4, pl. 42, figs. 3, 3a, pl. 64, fig. d.	1999	Macroneuropteris scheuchzeri; Tidwell et al.,
1908	Neuropteris gilmani Sellards, p. 403, pl. 58, fig. 4.	1777	p. 301, text-fig. 2d.
1925	Neuropteris decipiens; Noé, pl. 24, figs. 2–4, pls. 25, 26, pl. 27, figs. 1, 2.	2002	Macroneuropteris scheuchzeri; Blake et al., pl. 24, figs. 1–6.
1925	Odontopteris subcuneata; Noé, pl. 37, fig. 1.	2003	Macroneuropteris scheuchzeri; Zodrow, figs. 2,
1925	Odontopteris worthenii; Noé, pl. 37, fig. 4.		3, 6–13.
1938	Neuropteris scheuchzeri; Bell, p. 57, pl. 51, figs. 1–4, pl. 52, figs. 1, 2(?).	2005	<i>Macroneuropteris scheuchzeri</i> ; Bashforth, p. 61, pl. 13, figs. 6, 7.
1938	Neuropteris scheuchzeri Hoffmann forma angustifolia Bell, p. 57, pl. 50, figs. 3, 4, pl. 51, fig. 5.	2005	<i>Macroneuropteris scheuchzeri</i> ; DiMichele and Chaney, text-fig. 3.3.
1938	<i>Odontopteris subcuneata</i> ; Bell, p. 62, pl. 57, figs. 1–8, pl. 58, figs. 1–3.	2005	Macroneuropteris scheuchzeri; DiMichele et al., text-fig. 4.
1940	Neuropteris scheuchzeri; Janssen, p. 45, pl. 13, figs. 2, 3.	2006	Macroneuropteris scheuchzeri (Hoffmann) Cleal, Shute, and Zodrow forma decipens [sic] Wittry,
1940	Odontopteris subcuneata; Janssen, p. 50, pl. 12,		p. 57, figs. 2–4.
	figs. 1(?), 2, 4, 5.	2006	Odontopteris subcuneata; Wittry, p. 64, fig. 1.
1949	Neuropteris scheuchzeri; Arnold, p. 189, pl. 20.	2006	Odontopteris worthenii; Wittry, p. 65, figs. 1–3. Macroneuropteris scheuchzeri; Blake and
1958	<i>Neuropteris scheuchzeri</i> ; Langford, p. 189, text-figs. 327, 328.	2011	Gillespie, p. 110, pl. 6, figs. 1–6.
1958	Odontopteris subcuneata; Langford, p. 238, text-figs. 428–432.	2012	<i>Macroneuropteris scheuchzeri</i> ; Stull et al., text-figs. 2, 3, 5.
1959	Neuropteris scheuchzeri; Canright, pl. 4, figs. 8, 9.	2013	Macroneuropteris scheuchzeri; Stull et al., text-
1962	Neuropteris scheuchzeri forma angustifolia; Bell,		figs. 2.3, 2.4.
	p. 44, pl. 37, figs. 1, 2, 5.	2013b	Macroneuropteris scheuchzeri; DiMichele et al.,
1963	Neuropteris scheuchzeri; Langford, p. 222, text-	2014	p. 298, text-figs. 10.1, 10.2.
	fig. 845.	2014	<i>Macroneuropteris scheuchzeri</i> ; Moore et al., p. 38, pl. 6, pl. 7, figs. 1, 2.
1963	Neuropteris scheuchzeri; Wood, p. 60, pl. 10, fig. 1.		p. 50, pr. 0, pr. 7, 11gs. 1, 2.
1968	Neuropteris scheuchzeri; Basson, 100, pl. 24,	Holotyne.—	-Geologisch-Paläontologische Sammlungen, Mar-
10.00	fig. 1.	tin-Luther-U	

Holotype.—Geologisch-Paläontologische Sammlungen, Martin-Luther-Universität Halle-Wittenberg, Halle, from Osnabrück(?) Formation (basal Asturian), Piesberg Coalfield (Osnabrück Highlands), Germany.

Occurrence.—USA (assuming synonymy with Neuropteris decipiens): lower Atokan to middle Wolfcampian, common above Atokan (Darrah, 1969; Gillespie and Pfefferkorn, 1979; Oleksyshyn, 1982; Wagner, 1984; Blake et al., 2002; Stull et al., 2012). Europe: Langsettian to uppermost Cantabrian, common from upper Duckmantian to middle Cantabrian

(Wagner, 1984; Cleal and Shute, 1995; Laveine and Belhis, 2007). Atlantic Canada: Bolsovian to lower Cantabrian (Bell, 1938, 1962; Zodrow, 2003).

Material examined.—Seventy-six specimens from Units 2 and 3.

Description.—Pinnules exhibit a wide range of size and morphology. Typical pinnules are conspicuously large, with complete specimens 27-74 mm long and 9-23 mm wide (Table 5), linear-lanceolate to subtriangular (and frequently subfalcate; Fig. 8.7, 8.8) with an acutely rounded or acuminate tip, and pedicellate with a cordate to oblique (asymmetric) base (Fig. 9.10). Pinnules are rarely trifoliate (Fig. 8.8), with the pinnule base first differentiating into two symmetric lobes that, as development proceeds, become small orbicular to subtriangular pinnules that are 12-19 mm long and 7-8 mm wide (Table 5); such orbicular pinnules frequently are isolated from the main pinnule. Rare subulate pinnules are >95 mm long and ~17 mm wide and taper to an acuminate tip, whereas laciniate pinnules with deeply divided lateral margins only are represented by small fragments. Midveins are set in a furrow, are initially robust but narrow distally, and extend close to the pinnule apex. Lateral veins are thin, arise at an acute angle, branch twice close to the midvein, and then abruptly curve before extending nearly straight to reach the lateral margin at ~65-80°, branching once or twice more along their course; veins may curve upwards at the margin. Lateral veins are dense, with the venation density typically 29–37 veins per cm (range: 24–46, mean: 33; Table 5). Venation density broadly correlates with pinnule size, although lateral veins are more widely spaced and reach the lateral margin more obliquely in apical parts of large pinnules. Pinnule surfaces may display sparsely and randomly distributed hair-like structures (Fig. 8.11) that are ~1 mm long, bristle-like, straight to slightly curved, and generally oriented apically such that they cross-cut lateral veins, imparting a conspicuous yet falsely reticulate venation pattern.

Remarks.—Macroneuropteris is typified by large pinnules (generally over 2 cm long) with a narrow basal attachment to the rachis, a non-decurrent midvein that extends for much of the pinnule length, and unique epidermal characteristics (Cleal and Zodrow, 1989; Cleal et al., 1990; Cleal and Shute, 1995, 2012). The frond architecture of Macroneuropteris is less divided than in most neuropteroids (e.g., Neuropteris [Brongniart, 1822] Sternberg, 1825 or Laveineopteris Cleal, Shute, and Zodrow, 1990), resulting in comparatively large pinnules that are homologous to ultimate pinnae of other genera (Laveine, 1967; Cleal et al., 1996, 1998; Laveine and Belhis, 2007).

Macroneuropteris scheuchzeri is characterized by conspicuously large pinnules that often are isolated, trifoliate, and exhibit prominent hair-like features on the lower surface, and by laciniate pinnules of myriad morphologies (Laveine, 1967; Zodrow, 2003; Laveine and Belhis, 2007). The stiff and bristle-like structures on the abaxial surface of most pinnules support assignment to M. scheuchzeri. Zodrow et al. (2014) reinterpreted the hair-like structures as self-organizing epicuticular chemical deposits, but Laveine and Oudoire (2015) determined that they represent subepidermal resin-filled canals. The frequent occurrence of isolated pinnules points to a caducous plant, an

Table 5. Morphometric data (length, width, L:W ratio, and venation density) from specimens identified as *Neuropteris flexuosa* Sternberg, 1825, *Neuropteris ovata* Hoffmann, 1826a, 1826b, *Neuropteris missouriensis* Lesquereux, 1879-80, *Neuropteris semireticulata* Josten, 1962, and *Macroneuropteris scheuchzeri* (Hoffmann, 1826a) Cleal, Shute, and Zodrow, 1990. To facilitate comparison between *N. flexuosa* in the assemblage and elsewhere (see Fig. 16), venation density data also are presented from the holotype and other previously illustrated specimens. Data sources: holotype—Stockmans (1933, pl. 4, fig. 3); American examples—Noé (1925, pl. 20, fig. 1, pl. 23, fig. 3) and Jongmans (1937c, pl. 40, figs. 18, 19, pl. 41, fig. 25); Canadian examples—Bell (1938, pl. 46, figs. 6, 7, pl. 47, figs. 1, 2, 4, pl. 48, fig. 1) and Cleal and Zodrow (1989, text-figs. 13a, 13c); UK examples—Crookall (1959, pl. 37, figs. 1, 3, pl. 38, figs. 1–3, pl. 50, fig. 4).

Pinnule type	Parameter	Range	Percentiles	Mean	N
Neuropteris flexuosa (S'	TUDY SPECIMEN				
Lateral	Length	7.0–35.0	10.0-15.0	12.7	163
	Width	4.0 - 14.0	5.4–7.4	6.6	163
	Length:Width	1.4-2.5	1.7-2.1	1.9	163
	Venation density	21–40	26–31	28	73
Terminal	Length	19.3–46.0	22.0-24.0	25.5	13
	Width	10.4–18.5	11.3–15.7	13.4	18
Neuropteris flexuosa (H Lateral	OLOTYPE) Venation density	20–32	24–28	26	12
Neuropteris flexuosa (A	MERICAN EXAN	(IPLES)			
Lateral	Venation density	21–37	24-31	27	27
Neuropteris flexuosa (C	ΔΝΔΟΙΔΝ ΕΧΔΙ	(DI ES)			
Lateral	Venation density	18–43	23-32	28	36
	•	10 .6	20 02	-0	
Neuropteris flexuosa (U Lateral	K EXAMPLES) Venation density	24–50	28–36	33	44
Neuropteris ovata (STU	DY SPECIMENS)			
Lateral	Length	5.0–14.2	7.5 - 12.6	9.8	50
	Width	3.0 - 7.0	4.8 - 6.2	5.4	50
	Length: Width	1.2 - 2.3	1.6 - 2.0	1.8	50
	Venation density	29-44	35-40	37	31
Terminal	Length	13.0-23.0	na	19.0	3
	Width	5.0-6.2	na	5.6	3
	Length:Width	2.6 - 3.8	na	3.4	3
Neuropteris missourien.	sis (STUDY SPEC	IMENS)			
Lateral	Length	7.7–23.7	8.6-13.9	11.6	29
	Width	3.8–10.0	5.2-6.9	6.0	29
	Length:Width	1.4–2.6	1.6-2.1	1.9	29
	Venation density	20-32	23-26	25	34
Terminal	Length	16.0-20.0	16.2-20.0	18.4	5
	Width	9.2 - 13.6	9.4 - 12.7	11.5	5
	Length:Width	1.5 - 1.7	1.5 - 1.7	1.6	5
Neuropteris semireticul	ata (STUDY SPFC	IMENS)			
Lateral	Length	12.7–17.0	14.0-15.9	14.9	6
Euterur	Width	5.3–8.1	6.5–7.0	6.7	7
	Length:Width	1.7–3.2	1.9–2.3	2.2	6
	Venation density	28–36	30–33	32.0	9
Terminal	Length	>13.0	na	na	1
	Width	14.9	na	na	1
Macroneuropteris schei	ıchzeri (STUDY S	PECIMENS)		
Elongate	Length	~27–95	na	na	22
	Width	9.0–23.0	13.5–17.9	15.6	22
	Length:Width	~2.2–5.6	na	na	22
	Venation density	24–46	29–37	33.0	20
Orbicular/subtriangular		11.5-19.0	11.8–15.5	14.2	3
3	Width	6.8-8.0	7.3-7.6	7.4	4
	Length:Width	1.5 - 2.4	1.6-2.1	1.9	3
	Venation density	28-34	28-30	30.0	4
	<u> </u>				

established trait of the species. Although trifoliate pinnules are rare in the assemblage, the scattered presence of small orbicular pinnules, originally situated at the base of trifoliate forms, attests to pinnule disaggregation. Laciniate pinnules are equally rare and fragmentary; well-preserved examples from elsewhere are divided into spatulate or cuneate lobes with an 'odontopteroid' attachment to the rachis. Such forms were borne near the main frond bifurcation (Laveine, 1967; Laveine and Belhis, 2007), and have been assigned a variety of names, including *Odontopteris lindleyana* Sternberg, 1833 in the UK (Crookall, 1959),

Odontopteris subcuneata in Atlantic Canada (Bell, 1938; Zodrow, 2003), and O. subcuneata, Odontopteris heterophylla, Odontopteris worthenii, and Odontopteris cornuta in the USA (Lesquereux, 1866, 1879-80; White, 1899a). All of the aforementioned features help differentiate M. scheuchzeri from Macroneuropteris macrophylla (Brongniart, 1831) Cleal, Shute, and Zodrow, 1990, the pinnules of which are smaller, lack obvious hair-like structures, have an asymmetric cordate base with a prominent basiscopic auricle but no lobate pinnules, and which usually are affixed to the rachis (Bell, 1938; Crookall, 1959; Cleal et al., 1996, 1998). The American species Neuropteris clarksonii Lesquereux, 1854 is a junior synonym of M. macrophylla (Kidston, 1888; Bell, 1938; Crookall, 1959; Cleal et al., 1996; Wittry, 2006).

Previous authors seem unanimous in the opinion that the American species Neuropteris hirsuta is identical to Macroneuropteris scheuchzeri (White, 1899a; Crookall, 1959; Laveine, 1967; Darrah, 1969; Gastaldo, 1977). In contrast, there has been longstanding debate about the relationship between M. scheuchzeri and Neuropteris decipiens (cf., Laveine and Belhis, 2007), the latter found almost exclusively in the Mazon Creek Flora (Carbondale Formation) of Illinois, where the former is rare (Wittry, 2006). Some authors considered N. decipiens to be a junior synonym or variety/form of M. scheuchzeri (Crookall, 1959; Laveine, 1967; Gastaldo, 1977; Schabilion and Reihman, 1985; Wittry, 2006; Stull et al., 2012), whereas others maintained the taxa as different species (Lesquereux, 1880, 1884; White, 1899a; Darrah, 1969; Oleksyshyn, 1982), in part because of the highly restricted distribution of N. decipiens. As summarized by Gastaldo (1977), pinnules of both taxa share many morphological features (large size, abaxial hair-like structures, trifoliate and laciniate forms, an extended midvein, and similarly arching lateral veins), although *N. decipiens* generally has larger, more linguaeform pinnules with an obtusely rounded apex and somewhat sparser lateral veins. All study specimens have acutely rounded to acuminate tips, and fall within the limits of M. scheuchzeri sensu stricto. However, illustrated examples that conform to N. decipiens have been included in the synonymy list because the issue of identity will not be resolved without a detailed morphometric and cuticular analysis. Given the extremely long stratigraphic range and wide paleobiogeographic distribution of M. scheuchzeri sensu lato, as well as morphological changes through time (White, 1899a; Bell, 1938; Darrah, 1969; Gillespie et al., 1978), it is probable that more than one species is involved (Laveine and Belhis, 2007).

Genus *Neuropteris* (Brongniart, 1822) Sternberg, 1825 emend. Cleal, Shute, and Zodrow, 1990 *Neuropteris flexuosa* Sternberg, 1825 Figure 9.1, 9.5, 9.6, 9.11

- 1823 Osmunda gigantea var. β Sternberg, p. 36, pl. 32, fig. 2.
- 1825 Neuropteris flexuosa Sternberg, p. XVI.
- 1925 *Neuropteris flexuosa*; Noé, pl. 20, fig. 1, pl. 23, fig. 3.
- 1933 *Neuropteris tenuifolia* Schlotheim; Stockmans, p. 14, pl. 4, fig. 3 [holotype].
- 1937c *Neuropteris ovata* sensu ampl.; Jongmans, p. 418, pl. 40, figs. 17–19, 21, pl. 41, figs. 24, 25.

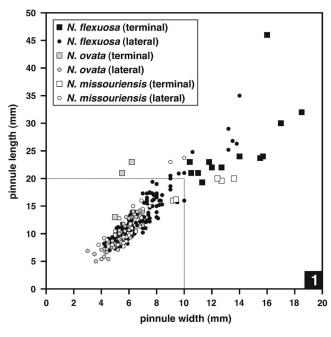
- 1938 *Neuropteris (Mixoneura) flexuosa*; Bell, p. 55, pl. 46, figs. 6, 7, pl. 47, figs. 1–3.
- 1938 Neuropteris flexuosa forma magna Bell, p. 55, pl. 47, fig. 4, pl. 48, figs. 1, 2.
- 1959 *Neuropteris ovata* Hoffmann forma *flexuosa* Sternberg; Crookall, p. 158, text-fig. 52 [holotype].
- (?)1958 *Neuropteris gigantea* Sternberg; Langford, p. 201, text-figs. 357, 359.
- (?)1959 Mixoneura Weiss sp.; Canright, pl. 4, fig. 12.
- 1962 Neuropteris (Mixoneura) flexuosa; Bell, p. 42, pl. 34, fig. 2, pl. 37, fig. 3(?)
- (?)1963 Neuropteris flexuosa; Wood, p. 57, pl. 9, fig. 5.
- 1963 *Neuropteris heterophylla*? (Brongniart) Sternberg; Wood, p. 57, pl. 9, figs. 6, 7(?)
- (?)1969 *Neuropteris ovata* forma *typica*; Darrah, p. 105, pl. 10, fig. 2.
- (?)1969 Neuropteris ovata forma flexuosa; Darrah, p. 106, pl. 73, fig. 1.
- (?)1977 Neuropteris ovata forma flexuosa; Gastaldo, text-fig. 48.
- (?)1978 Neuropteris ovata; Gillespie et al., p. 104, pl. 43, figs. 1, 2.
- (?)1979 Neuropteris flexuosa; Janssen, p. 150, text-fig. 139.
- 1979 *Neuropteris plicata* Sternberg; Janssen, p. 150, text-fig. 140.
- 1980 Neuropteris (Mixoneura) flexuosa; Zodrow and McCandlish, p. 40, pl. 22, fig. 2(?), pls. 23, 24.
- (?)1980 *Neuropteris flexuosa* forma *magna*; Zodrow and McCandlish, p. 42, pl. 25.
- 1989 *Neuropteris flexuosa*; Cleal and Zodrow, p. 856, text-fig. 13.
- 1997 *Neuropteris flexuosa*; Wagner and Lyons, pl. 1, figs. 1, 2.
- (?)1997 *Neuropteris* cf. *ovata*; Wagner and Lyons, pl. 4, fig. 1, pl. 5, fig. 3.
- (?)2005 Neuropteris ovata; DiMichele et al., text-fig. 6b.

Holotype.—V.9405, Natural History Museum, London, from Radstock Formation (upper Asturian), Bristol-Somerset Coalfield, UK.

Occurrence.—USA: upper Atokan to upper Virgilian, rare below lower Desmoinesian, common in upper Desmoinesian to middle Virgilian (Wood, 1963; Read and Mamay, 1964; Darrah, 1969). UK: basal Asturian to lower Cantabrian (Cleal and Thomas, 1994; Cleal and Shute, 1995; Cleal, 2007). Atlantic Canada: upper Bolsovian to lower Cantabrian, very rare below Asturian (Bell, 1938, 1962; Zodrow and Cleal, 1985; Zodrow, 1986).

Material examined.—Ninety-four specimens from Units 2 and 3.

Description.—Ultimate pinnae are linear and taper to an obtusely rounded apex. Terminal pinnules (Fig. 9.1, 9.6, 9.11) are large, typically 22–24 mm (but up to 46 mm) long and 11–16 mm wide (L:W ratio ~ 1.9; Table 5; Fig. 15), rhomboid to deltoid and asymmetric with a basal lobe on one or both sides,



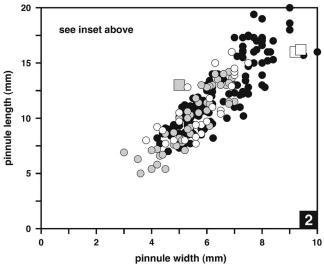


Figure 15. Bivariate (length, width) plot of terminal and lateral pinnules of *Neuropteris flexuosa* Sternberg, 1825, *Neuropteris ovata* Hoffmann, 1826a, 1826b, and *Neuropteris missouriensis* Lesquereux, 1879-80, in the macrofloral assemblage (study specimens in Table 5). (1) Complete dataset. (2) Inset magnifying lower left corner of plot.

and have an obtusely to acutely rounded apex. Lateral pinnules are variable in size, with most 10–15 mm long and 5–7 mm wide (L:W ratio ~1.9), and large forms reaching 35 mm long and 14 mm wide (Table 5; Fig. 15). Pinnules are inserted slightly obliquely, generally crowded and often overlapping, mainly linguaeform to rectangular with an obtusely rounded apex (becoming more elongate to slightly falcate in the largest forms), and are characterized by a pronounced basiscopic auricle (Fig. 9.11) and a rounded acroscopic base (Fig. 9.5) that may overlap the pinna rachis. Lateral pinnules are almost exclusively pedicellate (only the most distal pinnules have a broadly attached, 'mixoneurid' base), although the point of attachment is relatively wide because the midvein may be accompanied above and below by subsidiary veins derived from the rachis. Midveins are slightly decurrent, more so apically, depressed in a

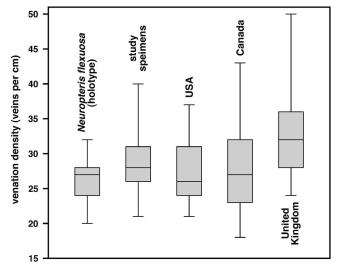


Figure 16. Box and whisker plots showing range, 25th and 75th percentiles, and median of venation densities of lateral pinnules of *Neuropteris flexuosa* Sternberg, 1825, showing comparison between study specimens, holotype, and examples from USA, Canada, and UK. See Table 5 for data sources.

shallow furrow, somewhat flexuous, thin, and extend to the pinnule midpoint before dissipating. Lateral veins are thin, arise at an acute angle, branch close to the midvein and two to three more times on approach to the margin, which they reach at ~60–80°. Veins extend straight between successive forks, imparting a subtly flexuous appearance, and are sparse, with the venation density typically 26–31 veins per cm (range: 21–40, mean: 28; Table 5; Fig. 16). In the largest pinnules, midveins extend farther, and lateral veins reach the margin at ~90° and may be more dense due to an additional dichotomy.

Remarks.—Specimens are assigned to Neuropteris flexuosa, one of a group of neuropterids closely allied to Neuropteris ovata that are typified by epidermal characters, lateral pinnules that can be partially fused to the rachis in distal parts of pinnae ('mixoneurid' habit), and thin decurrent midveins that extend no more than two-thirds the pinnule length (Wagner, 1963; Cleal and Zodrow, 1989; Cleal et al., 1990; Cleal and Shute, 1995, 2012). Distinguishing features of N. flexuosa include the combination of a large, asymmetric, deltoid terminal pinnule, and relatively large lateral pinnules with a rounded acroscopic base, auriculate basiscopic side, and a relatively coarse and slightly flexuous venation; broadly attached pinnules only occur at the extreme top of ultimate pinnae. The degree of uncertainty expressed in the synonymy list reflects the challenge of separating species in this neuropterid complex, particularly when relying on photographs.

We initially were reluctant to assign specimens to *Neuropteris flexuosa* because the average venation density has previously been reported as ~30–45 veins per cm (e.g., Bell, 1938; Crookall, 1959; Wagner, 1963; Darrah, 1969; Cleal and Zodrow, 1989). Although the study specimens range from 21–40 veins per cm, most have a sparser venation (26–31 veins per cm, mean 28; Table 5). To investigate this disparity, veins were counted on illustrated specimens of *N. flexuosa* from elsewhere in the USA, Canada, and the UK, including the British holotype (see Table 5 for list of sources). Comparison

indicates that venation densities of the holotype and examples from the USA and Canada are similar to those measured in the study specimens (and perhaps more importantly, lower than reported), whereas British specimens tend to have a denser venation (Fig. 16). 'Post-hoc' pairwise comparisons after a Kruskal-Wallis test (H = 32.26, p[same] < 0.0001) confirm that the venation density in the British specimens (excluding the holotype) is significantly different from N. flexuosa in the assemblage and from the broader suite of North American specimens (p[same] ≤ 0.0004). One explanation for the inconsistency between reported and actual venation density in some publications may be a tendency to overemphasize larger pinnules, which have a denser venation. It also is possible that differences reflect an actual species distinction, as suspected for the Macroneuropteris scheuchzeri-Neuropteris decipiens complex.

Due to their morphological similarities, some authors considered *Neuropteris flexuosa* to be synonymous with, or a form of, *Neuropteris ovata* (Jongmans, 1937c; Crookall, 1959; Darrah, 1969). However, in addition to epidermal differences (Cleal and Zodrow, 1989), comparisons based on specimens in the assemblage (Table 5; Figs. 9, 15) reveal consistent differences that permit segregation of these two species. In particular, *N. flexuosa* can be distinguished from *N. ovata* by having overall larger lateral pinnules, a large deltoid terminal pinnule, lower venation density, and by the fact that broadly attached lateral pinnules only occur (if at all) at the extreme top of an ultimate pinna.

A closely related American fossil-species is Neuropteris vermicularis Lesquereux, 1861, which Crookall (1959) and Jongmans (1937c) considered identical to (or a form of) Neuropteris ovata. However, Lesquereux (1861, 1879-80), Darrah (1969), and Wittry (2006) maintained that N. vermicularis was a distinct taxon characterized by larger and more symmetrical terminal pinnules (sometimes with a lobate margin), a coarser venation, and lateral pinnules with a more constricted or cordate base. Examination of N. vermicularis specimens in the Lacoe Collection (NMNH), most of which were determined by Lesquereux or White, adds to the ambiguity. Large pinnules with a dense venation are virtually indistinguishable from equally large forms of Neuropteris flexuosa, although with a less prominent basiscopic auricle (Lesquereux, 1861). In contrast, smaller and/or more distal pinnules of N. vermicularis have a coarser venation and are markedly elongate-linguaeform (L:W ratio ~2-3, compared with $\sim 1.5-2.0$ in equivalent pinnules of *N. flexuosa*). As such, *N*. vermicularis may be a unique species, and has been excluded from the synonymy list. Representative examples believed to conform to Lesquereux' concept of N. vermicularis were illustrated (under a variety of names, including N. flexuosa) by Noé (1925, pl. 20, fig. 3, pl. 24, fig. 1), Jongmans (1937c, pl. 41, figs. 22, 23), Janssen (1940, pl. 25, fig. 2; 1979, text-fig. 142), Darrah (1969, pl. 2, figs. 1, 2), Boneham (1974, pl. 5, fig. 4), and Wittry (2006, p. 59, 60, figs. 1, 2, 4, 5).

According to White (1899a), *Neuropteris missouriensis* also resembles *N. flexuosa*, but differs in having very large, oblong-ovate to oblong-deltoid terminal pinnules in the smallest pinnae, and oblong to ovate lateral pinnules typified by an equally rounded (i.e., cordate or non-auriculate) base and a more distinct midvein.

Neuropteris missouriensis Lesquereux, 1879-80 Figures 11.2–11.4, 12

- 1879-80 *Neuropteris Missouriensis* Lesquereux, p. 104, pl. 7, figs. 5, 6, 6a.
- (?)1893 Neuropteris flexuosa Sternberg; White, p. 91, pl. 5, figs. 1, 2.
- 1899a *Neuropteris missouriensis*; White, p. 130, pl. 41, figs. 4, 5, pl. 42, figs. 4, 4a, pl. 45, fig. 3.
- (?)1963 Neuropteris flexuosa; Wood, p. 57, pl. 9, fig. 5.
- (?)1967 Neuropteris dussarti Laveine, p. 191, pl. 48.
- 1968 Neuropteris missouriensis; Basson, p. 97, pl. 22.
- (?)1991 Neuropteris dussarti; Josten, p. 322, pls. 186, 187.
- (?)2005 Laveineopteris dussartii (Laveine) Laveine, p. 464.

Type material.—One syntype (USNM 30861; Lesquereux, 1879-80, pl. 7, fig. 6, 6a) in Plant Type and Illustrated Collection, National Museum of Natural History, Smithsonian Institution, Washington, from Drywood(?) Formation (lower Desmoinesian), Western Interior Basin, Clinton, Missouri.

Occurrence.—USA: upper Atokan to lower Desmoinesian (White, 1899a; Sellards, 1908; Darrah, 1969). Laveineopteris dussartii (possible synonym), in UK: upper Bolsovian to upper Asturian (Cleal, 1978; Cleal and Shute, 1995); in northern France and northwestern Germany: middle(?) Asturian (Laveine, 1967, 1987; Josten and Laveine, 1984; Josten, 1991).

Material examined.—Fourteen specimens from Unit 4.

Description.—Ultimate pinnae are linear and parallel-sided for most of their length, but can be slightly narrower at the base and taper to an obtusely rounded apex. Pinna axes are conspicuously robust. Terminal pinnules are large, 16-20 mm long and 9–14 mm wide (L:W ratio ~ 1.6; Table 5; Fig. 15), oblong-ovate to oblong-deltoid and asymmetric with a basal lobe on one or both sides, and have a very obtusely rounded apex; terminal pinnules are larger, more oblong, and more obtusely rounded in small pinnae (compare Fig. 11.2–11.4 with 11.11, 11.12). Lateral pinnules are 8-24 mm long and 4-10 mm wide (L:W ratio ~1.9; Table 5; Fig. 15). Pinnules are inserted slightly obliquely, closely spaced to slightly overlapping, ovate in the most distal positions (Fig. 11.4), but generally oblong and linguaeform with an obtusely rounded apex, and typically are bilaterally symmetrical, but somewhat subfalcate in the largest examples. Pinnule shape in distal parts of pinnae depends on the degree of development: ovate pinnules occur on short pinnae, and linguaeform pinnules occur on long pinnae. Pinnules are always sessile (resting on the rachis without an elongate petiolule), attached by a broad midvein (even in the smallest forms), have nearly equally rounded acroscopic and basiscopic bases (cordate), and are characterized by a narrow but thick compression margin (Fig. 11.11, 11.12). Midveins are slightly decurrent, very prominent and set within a deep furrow, extend about two-thirds of the pinnule length, and generally are straight, but can appear flexuous due to the wide angle of departure of lateral veins. Lateral veins are thin but depressed in the thick lamina, branch close to the midvein and one to three more times before reaching the lateral margin at $\sim 65-80^{\circ}$, and are sparse, with venation density typically 23–26 veins per cm (range: 20–32, mean: 25; Table 5). Near the midvein, lateral veins may extend straight between successive forks, conferring a somewhat flexuous appearance to the venation.

Remarks.—Specimens compare closely with the syntypes of Neuropteris missouriensis illustrated by Lesquereux (1879-80), although the fossil-species is otherwise rare and poorly documented. Distinguishing features include the robust pinnae axes and pinnule midveins, large obtusely rounded and oblong terminal pinnules, and sparse venation on lateral pinnules that invariably are sessile with a cordate base. Sellards (1908) emphasized the prominent compression margin on the pinnules, implying that margins were inrolled. The fact that lateral pinnules at the distal end of a pinna are attached only by the pinnule petiolule rather than being partly fused to the rachis, and that midveins are prominent and extend beyond the pinnule midpoint, suggests that N. missouriensis belongs in the fossil-genus Laveineopteris rather than Neuropteris (Cleal et al., 1990). White (1899a) illustrated a wide range of morphological variety in N. missouriensis, and demonstrated that the terminal pinnule in longer (more proximal) pinnae is smaller, less oblong, and less obtusely rounded than originally envisioned by Lesquereux (1879-80). However, the assertion by White (1899a) that terminal pinnules only have a single lobe is incorrect; some specimens have two lobes, although one is better developed.

White (1893, 1899a) stated that *Neuropteris missouriensis* most resembles *Neuropteris flexuosa*, and Darrah (1969) also implied that the species might belong to the '*Neuropteris ovata* complex'. However, *N. flexuosa* has larger terminal pinnules that are rhomboid to deltoid, lateral pinnules that are broadly attached (i.e., 'mixoneurid') in extreme distal positions and have a prominent basiscopic auricle, thin midveins that extend only to the midpoint of a pinnule, and a greater venation density (Table 5). Crookall (1959, pl. 53, figs. 1–3) reported *N. missouriensis* in the UK, but these examples may involve multiple species, including *Laveineopteris loshii* (Brongniart, 1831) Cleal, Shute, and Zodrow, 1990 and *Laveineopteris hollandica* (Stockmans, 1933) Cleal and Shute, 1995.

In many respects, particularly in the distinct shape of terminal pinnules, venation density, and cordate pinnule bases, *N. missouriensis* compares closely with *Laveineopteris dussartii* (cf., Laveine, 1967; Josten and Laveine, 1984; Josten, 1991), which only is known from the Bolsovian to Asturian of Western Europe. Nonetheless, *L. dussartii* differs in having less-robust pinnae axes and midveins. If synonymous, *N. missouriensis* has nomenclatural priority.

Neuropteris ovata Hoffmann, 1826a, b Figure 9.2–9.4, 9.8

1826b Neuropteris ovata Hoffmann, p. 266.
1826a Neuropteris ovata Hoffmann, p. 158, pl. 1b
figs. 5–7 [lectotype: fig. 6].
(?)1879-80 Neuropteris plicata Sternberg; Lesquereux, p. 96
pl. 10, figs. 1–4.
(?)1879-80 Neuropteris loschii [sic] Brongniart; Lesquereux
p. 98, pl. 11.

- 1908 *Neuropteris ovata*; Sellards, p. 408, pl. 53, fig. 2, pl. 56, fig. 6(?).
- (?)1925 Neuropteris ovata; Noé, pl. 21, fig. 4.
- (?)1925 *Neuropteris capitata* Lesquereux; Noé, pl. 22, fig. 2. 1937c *Neuropteris ovata* sensu ampl.; Jongmans, p. 417, pls. 37–39, 42.
- 1938 Neuropteris (Mixoneura) ovata; Bell, p. 60, pl. 52, fig. 4, pl. 53, fig. 3, pl. 54, figs. 1–3.
- (?)1958 Neuropteris plicata; Langford, p. 207, text-fig. 361.
- (?)1958 *Neuropteris capitata*; Langford, p. 207, text-fig. 362.
- 1958 Neuropteris (Mixoneura) ovata; Langford, p. 218, text-figs. 382, 383.
- (?)1958 Neuropteris (Mixoneura) jenneyi White; Langford, p. 218, text-fig. 384.
- 1963 *Neuropteris obliqua*? (Brongniart) Zeiller; Wood, p. 59, pl. 9, fig. 10.
- 1963 *Odontopteris subcuneata* Bunbury; Wood, p. 61, pl. 10, fig. 5.
- 1968 Neuropteris ovata Hoffmann forma flexuosa Crookall; Basson, p. 98, pl. 23, figs. 1, 2(?).
- 1969 *Neuropteris ovata* Hoffmann forma *typica* Crookall; Darrah, p. 105, pl. 9., fig. 2, pl. 10, fig. 1, 2(?).
- 1969 *Imparipteris ovata* (Hoffmann) Gothan; Saltzwedel, pl. 24, figs. 1–3 [lectotype].
- 1977 *Neuropteris ovata* forma *typica*; Gastaldo, text-fig. 47.
- (?)1978 *Neuropteris ovata*; Gillespie et al., p. 104, pl. 43, figs. 1, 2.
- 1979 *Mixoneura ovata* (Hoffmann) Zalessky; Janssen, p. 157, text-fig. 150.
- (?)1979 *Neuropteris heterophylla* (Brongniart) Sternberg; Pfeiffer, pl. 6, fig. 63.
- (?)1979 Neuropteris ovata; Pfeiffer, pl. 6, fig. 64.
- (?)1979 *Neuropteris obliqua* (Brongniart) Zeiller; Pfeiffer, pl. 7, fig. 70.
- 1980 Neuropteris (Mixoneura) ovata; Zodrow and McCandlish, p. 44, pl. 28, fig. 3, pls. 29–32, pl. 33, fig. 1.
- 1988 Neuropteris ovata; Zodrow and Cleal, pls. 2, 3, pl. 4, fig. 4.
- 1982 *Neuropteris ovata* forma *typica*; Oleksyshyn, p. 111, text-figs. 21g, h, 22a–e.
- 1982 *Neuropteris ovata* forma *flexuosa*; Oleksyshyn, p. 114, text-fig. 23a, b.
- 1992 *Neuropteris ovata*; Mamay and Mapes, text-fig. 4a, c, e.
- 1997 *Neuropteris ovata*; Wagner and Lyons, pl. 8, fig. 5.
- Neuropteris cf. ovata; Wagner and Lyons, pl. 3, figs. 1, 2.
- Neuropteris ovata; Blake et al., pl. 21, fig. 7.
- (?)2005 *Neuropteris ovata*; DiMichele et al., text-fig. 6b.
- (?)2006 *Neuropteris flexuosa* Sternberg; Wittry, p. 49, figs. 1, 2.
- 2006 Neuropteris ovata; Wittry, p. 55, fig. 1.

2013b *Neuropteris ovata*; DiMichele et al., p. 298, text-figs. 12, 12, 13(?).

Lectotype.—Geologisch-Paläontologische Sammlungen, Martin-Luther-Universität Halle-Wittenberg, Halle, from Dickenburg Coal, Osnabrück Formation (basal Asturian), Ibbenbüren Coalfield (Osnabrück Highlands), Germany.

Occurrence.—USA: upper Atokan to upper Virgilian, common in lower Desmoinesian to Missourian (Jongmans, 1937c; Wood, 1963; Read and Mamay, 1964; Darrah, 1969; Gillespie and Pfefferkorn, 1979; Oleksyshyn, 1982; Blake et al., 2002). Europe: rare in upper Bolsovian (Cleal, 1978), common from basal Asturian to Cantabrian (Cleal, 1978, 1997; Laveine, 1977, 1987, 1989; Josten, 1991; Cleal and Shute, 1995), some varieties extend up to Stephanian C (Wagner, 1963, 1984). Atlantic Canada: middle Asturian to lower Cantabrian (Bell, 1938; Zodrow and Cleal, 1985; Zodrow, 1986).

Material examined.—Twenty-six specimens from Units 2 to 4.

Description.—Ultimate pinnae are linear but rapidly taper to an acutely rounded apex. Terminal pinnules (Fig. 9.2, 9.3) are small, 13-23 mm long and 5-6 mm wide (L:W ratio ~3.4; Table 5; Fig. 15), elongate-rhomboid to lanceolate and asymmetric with a basal lobe on one or both sides, and have an acutely rounded or acuminate summit. Lateral pinnules are 5-14 mm long and 3-7 mm wide (L:W ratio ~1.8; Table 5; Fig. 15). Pinnules are inserted slightly obliquely (markedly so in distal positions, where pinnules are strongly decurrent), generally closely spaced, linguaeform to ovate with an obtusely rounded apex, and are characterized by a strongly incised, rounded acroscopic base and a pronounced, acutely rounded basiscopic auricle (Fig. 9.4, 9.8). Lateral pinnules are narrowly attached by a petiolule in proximal parts of pinnae, but in distal parts much of the base is affixed to the rachis ('mixoneurid' habit), particularly on the basiscopic side (Fig. 9.2-4), from whence subsidiary veins enter the pinnule. Midveins may comprise multiple thin strands, and are decurrent (more so apically) and depressed in a shallow crease; the central strand extends straight to near the pinnule midpoint before dissipating. Lateral veins are thin, arise at an acute angle, branch close to the midvein and generally two to three more times, broadly arch to reach the lateral margin at $\sim 40-70^{\circ}$, and are dense, with the venation density typically 35–40 veins per cm (range: 29-44, mean: 37; Table 5).

Remarks.—Although Neuropteris ovata is a common Middle to Late Pennsylvanian medullosalean pteridosperm, its widespread distribution and polymorphic habit have resulted in a complex and unresolved taxonomic history (Crookall, 1959; Wagner, 1963; Laveine, 1967). Laveine (1989, pls. 47, 48) illustrated the array of morphological variation that can occur in the species concept from a single basin. Given the wide range of pinnule forms, previous workers have split the species into several taxa, based on differences in regional distribution (Bertrand, 1930), stratigraphic distribution (Wagner, 1963), or a combination of these factors together with differences in epidermal anatomy (Cleal and Zodrow, 1989). Although some of the resulting 'varieties' may represent distinct species (Cleal and Shute, 1995),

no attempt has been made to assign specimens in the assemblage to a variety because cuticles were not procured. The material at hand compares well with the lectotype of *N. ovata*, in addition to other specimens from the type area (Saltzwedel, 1969).

According to Darrah (1969, p. 105), in the USA the majority of specimens that Lesquereux (1879-80) assigned to Neuropteris loschii [sic] Brongniart belong to Neuropteris ovata. As discussed in the Neuropteris flexuosa section above, Jongmans' (1937c) study of N. ovata from American strata included three species (N. ovata, N. flexuosa, and Neuropteris vermicularis). Specimens in the assemblage belong to what Crookall (1959) and Darrah (1969) termed N. ovata forma typica. Based on illustrations in the literature and comparison between N. ovata and N. flexuosa in the study population (Table 5; Figs. 9, 15), the former has overall smaller lateral pinnules, a small elongate terminal pinnule with an acutely rounded tip, more-dense and non-flexuous venation, and broadly attached lateral pinnules that normally occur in distal and medial parts of an ultimate pinna. Small forms of N. vermicularis may be confused with N. ovata (e.g., Noé, 1925, pl. 20, fig. 3), but the former has larger and more symmetrical terminal pinnules with an obtusely rounded apex, a coarser venation, and more elongate lateral pinnules that may have only a slight basiscopic auricle. A single cyclopteroid pinnule with a fimbriate margin, referable to Cyclopteris fimbriata Lesquereux, 1854, in the assemblage presumably was part of the N. ovata frond, but other North American examples are not included in the synonymy list.

Neuropteris semireticulata Josten, 1962 Figure 11.6, 11.9

- (?)1938 Neuropteris heterophylla (Brongniart) Sternberg; Bell, p. 56, pl. 50, fig. 2.
- (?)1938 *Linopteris muensteri* (Eichwald) Potonié; Bell, p. 65, pl. 59, figs. 2, 3.
- 1962 *Neuropteris semireticulata* Josten, p. 39, pl. 3, figs. 2–5 [holotype: figs. 3, 3a].
- (?)1962 *Neuropteris heterophylla*; Bell, p. 43, pl. 33, fig. 2, pl. 34, fig. 1.
- 2002 *Neuropteris semireticulata*; Blake et al., pl. 19, fig. 1, pl. 22, fig. 1.
- 2005 Neuropteris semireticulata; Bashforth, p. 62, pl. 16, pl. 17, figs. 1–11.

Holotype.—No. 927, Type Collection, Geologischen Landesamtes, Krefeld, from Dorsten Formation (Bolsovian), Ruhr Coalfield, near Bottrop, North Rhine-Westphalia, Germany.

Occurrence.—USA: upper Atokan to lower Desmoinesian (Blake et al., 2002). UK: middle Duckmantian to lower Asturian (Cleal, 1978, 2007; Cleal and Thomas, 1994). France: Bolsovian (Laveine, 1967, 1987). Germany: upper Duckmantian to upper Bolsovian (Josten, 1962, 1991). Atlantic Canada: upper Bolsovian to lower Asturian (Bell, 1938, 1962; Bashforth, 2005).

Material examined.—Two specimens from Unit 4.

Description.—Terminal pinnules are ~15 mm wide and of indeterminate length, and have a lobe on one side. Lateral

pinnules are 13-17 mm long and 5-8 mm wide (L:W ratio ~ 2.2). Pinnules are inserted obliquely, crowded to overlapping, subovate to linguaeform (more elongate proximally) with a broadly rounded top, and are characterized by a rounded acroscopic base that overlaps the pinna rachis. Lateral pinnules generally are cordate, but in distal positions the base becomes partly fused with the rachis and the midvein is accompanied above and below by subsidiary veins. Midveins are decurrent (more so apically), robust, flexuous, and extend beyond the pinnule midpoint before dissipating. Lateral veins are strong, arise at an open angle, branch close to the midvein and generally one to two more times on approach to the margin, which they reach at 70-80°. Lateral veins are markedly flexuous to pseudoreticulate (Fig. 11.9), closely approach or run alongside (but do not join) adjacent veins, and are relatively sparse, the venation density typically being 30–33 veins per cm (range: 28–36, mean: 32; Table 5). Midveins extend farther and lateral veins are denser in the longest pinnules.

Remarks.-Neuropteris semireticulata can be distinguished from other neuropterids by its flexuous to pseudoreticulate venation, and although easily recognized, the taxon has rarely been recorded in Euramerica. Whereas this may reflect genuine scarcity, it is more probable that the species has been incorrectly identified as Neuropteris heterophylla (Brongniart, 1822) Sternberg, 1825 or Reticulopteris muensteri (Eichwald, 1840) Gothan, 1941 at many localities. In West Virginia, for example, some specimens that Gillespie et al. (1978) referred to the latter species probably represent N. semireticulata, as Blake et al. (2002) also illustrated examples from the state. Similar misidentifications are probable in Atlantic Canada (see synonymy list), although N. semireticulata was documented in upper Bolsovian strata of Newfoundland (Bashforth, 2005). The American species Dictyopteris rubella Lesquereux, 1870 is a junior synonym of R. muensteri according to Janssen (1940).

Order Lyginopteridales Corsin, 1960 Family Lyginopteridaceae Potonié, 1900 Genus *Eusphenopteris* Gothan, 1913 ex Simson-Scharold, 1934 *Eusphenopteris neuropteroides* (Boulay, 1876) Novik, 1947 Figure 10.4, 10.5, 10.8

- 1854 Sphenopteris squamosa Lesquereux, p. 420.
- (?)1868 *Sphenopteris lesquereuxii* Newberry; Lesquereux, p. 862, pl. 10, fig. 1.
- 1868 Sphenopteris squamosa Lesquereux, p. 862, pl. 10, fig. 3.
- 1876 *Pecopteris neuropteroides* Boulay, p. 32, pl. 2, figs. 6, 6bis [lectotype: fig. 6].
- 1879-80 *Pseudopecopteris anceps* Lesquereux, p. 207, pl. 38, figs. 1, 1a, 2(?)
- 1883 Sphenopteris nevropteroides (Boulay) Zeiller, p. 186.
- 1899b *Pseudopecopteris squamosa* (Lesquereux) White, p. 474.
- 1938 *Sphenopteris neuropteroides*; Bell, p. 23, pl. 5, fig. 4(?), pl. 6, pl. 7, pl. 8, fig. 1.
- (?)1943 Diplothmema aldrichi White var. anthracitica White, p. C98, pl. 33, figs. 6, 9, pl. 34, fig. 2.

- 1947 Eusphenopteris neuropteroides (Boulay) Novik, p. 277.
- (?)1952 Eusphenopteris neuropteroides; Novik, p. 312, pl. 52, figs. 9, 10.
- 1975 Eusphenopteris neuropteroides; Van Amerom, p. 54, pl. 28, figs. 1, 2 [lectotype: fig. 1].
- (?)1978 Eusphenopteris neuropteroides; Gillespie et al., p. 110, pl. 52, figs. 2, 3.
- (?)1978 *Eusphenopteris* Gothan ex Simson-Scharold sp.; Gillespie et al., pl. 52, fig. 5.
- 1980 Sphenopteris neuropteroides; Zodrow and McCandlish, p. 54, pl. 56, figs. 2–4, pls. 57, 58.
- 1982 Eusphenopteris neuropteroides; Oleksyshyn, p. 58, text-fig. 13b(?), c(?), d, e, f(?).
- (?)2002 Eusphenopteris neuropteroides; Blake et al., pl. 21, figs. 2, 4.
- (?)2014 cf. Eusphenopteris neuropteroides; Moore et al., p. 44, pl. 7, fig. 16.

Lectotype.—Université Catholique de Lille, Lille, probably from the Faisceau de Six-Sillons or d'Ernestine (Bolsovian), Northern France Coalfield, France.

Occurrence.—USA: lower Atokan to lower Virgilian(?), common in Desmoinesian (Darrah, 1969; Oleksyshyn, 1982; Blake et al., 2002). UK: middle Langsettian to lower Cantabrian (Cleal and Thomas, 1994). Europe: extends into Stephanian C (Wagner, 1984). Atlantic Canada: Asturian (Bell, 1938).

Material examined.—Twenty-six specimens from Units 2 and 3.

Description.—Penultimate pinnae are linear, taper to an obtusely rounded apex topped by a small, lobate terminal pinnule, reach at least 50 mm long and 35 mm wide, and comprise alternate ultimate pinnae (becoming pinnatifid and then entire pinnules apically) that arise from a thick, straight, and longitudinally striated rachis with faint transverse ridges. Ultimate pinnae are perpendicular to slightly oblique, generally linear with a blunt apex, but can be subtriangular with a slightly more acutely rounded tip, typically 10-30 mm long and 5-15 mm wide, and have a pronounced, winged rachis that gives rise to 2 to 8 pairs of lateral pinnules. The shape and size of lateral pinnules are highly variable. In more proximal positions of a pinna (Fig. 10.4, 10.5), trilobate to bilobate pinnules (4–8 mm long, 4-7 mm wide) stand approximately perpendicular to the rachis, are attached by a wide footstalk, and pass apically into more or less entire pinnules (4-6 mm long, 3-5 mm wide) that are quadrate to ovate and affixed by most of their base. The most distal pinnules (2-6 mm long, 2-5 mm wide) are strongly decurrent, confluent, round, and fuse upwards with a small, lobate terminal pinnule (Fig. 10.4, 10.5, 10.8). Midveins are decurrent (less so in quadrate to lobate pinnules), thick, and divide into several branches shortly after entering the pinnule; in round, distal pinnules, one branch ascends nearly parallel to the pinna rachis before dichotomizing.

Remarks.—Despite having polymorphous foliage, Eusphenopteris neuropteroides is readily distinguished by having large quadrate pinnules that superficially resemble the habit and

venation of neuropteroid foliage. Van Amerom (1975) documented the full range of variation. In the USA, E. neuropteroides has been published under various names. Lesquereux (1854, 1868) introduced the earlier synonym Sphenopteris squamosa. Subsequently, Lesquereux (1879-80) erected Pseudopecopteris anceps, although he explicitly included S. squamosa as a synonym. However, the only syntype of P. anceps in the NMNH collections (Lesquereux, 1879-80, pl. 38, figs. 3, 4; USNM 13401) is not representative and may belong to Eusphenopteris nummularia (Gutbier, 1835) Novik, 1947 or Eusphenopteris obtusiloba (Brongniart, 1830-31) Novik, 1947. Zeiller (1883, 1886-88) recognized the identity between these two American taxa and the European form he referred to Sphenopteris nevropteroides, and White (1899b) synonymized all three taxa under Pseudopecopteris squamosa. However, Bell (1938) suggested that the epithet neuropteroides be retained because the original drawing and descriptions of S. squamosa were insufficient. Darrah (1969) supported this assertion and noted that the holotype of S. squamosa is missing. According to van Amerom (1975), Diplothmema aldrichi var. anthracitica of White (1943) could be accommodated in E. neuropteroides.

Although specimens illustrated by Gillespie et al. (1978) and Blake et al. (2002) exhibit the general characters of *Eusphenopteris neuropteroides*, they are included in the synonymy list with reservation because they have overall larger dimensions than is typical. These examples perhaps should be referred to the superficially similar fossil species "*Mariopteris*" anthrapolis.

Eusphenopteris sp. indet. Figure 11.5

Material examined.—One specimen from Unit 4.

Description.—Penultimate pinnae reach 26 mm long, and comprise alternate to subopposite ultimate pinnae (becoming pinnatifid and then entire pinnules apically). The rachis is stiff and straight, bears faint longitudinal striations with no transverse ridges, and has a pronounced central groove flanked by elevated lateral margins. Ultimate pinnae reach 11 mm long and 7 mm wide, and have a winged rachis that gives rise to at least 3 pairs of vaulted, orbicular to obovate pinnules. In distal parts of the penultimate pinna, trilobate to bilobate(?) pinnules (at least 8 mm long and 5-8 mm wide) stand slightly obliquely, are attached by a wide and decurrent footstalk, and consist of one or two small orbicular basal lobes (larger on the basiscopic side) and a large apical lobe. Near the pinna apex, entire pinnules (~7 mm long and 5 mm wide) are ovate and affixed by most of their base. Midveins occupy a shallow groove, and lateral veins are difficult to discern, but appear to be relatively sparse.

Remarks.—Although the single penultimate pinna fragment clearly belongs to the fossil-genus Eusphenopteris, the specimen does not display enough morphological attributes to permit specific identification. The best comparison may be with Eusphenopteris nummularia, which is characterized by strongly vaulted foliage and essentially indiscernible venation (Van Amerom, 1975).

Incertae sedis
"Mariopteris" anthrapolis Langford, 1958
Figure 8.5, 8.9, 8.12, 8.13

- 1958 *Mariopteris anthrapolis* Langford, p. 268, text-figs. 488, 489(?), 490(?), 493.
- (?)1969 Mariopteris thompsonii Darrah, p. 123, pl. 25, fig. 1.
- (?)1978 Eusphenopteris neuropteroides (Boulay) Novik; Gillespie et al., p. 110, pl. 52, figs. 2, 3.
- (?)1978 *Eusphenopteris* Gothan ex Simson-Scharold sp.; Gillespie et al., pl. 52, fig. 5.
- (?)1982 *Mariopteris* cf. *inflata* White; Oleksyshyn, p. 63, text-fig. 13i.
- (?)2002 Eusphenopteris neuropteroides; Blake et al., pl. 21, figs. 2, 4.
- 2006 *Mariopteris anthrapolis*; Wittry, p. 80, figs. 1, 2 [syntype: fig. 1].

Syntypes.—FMNH P31072 and P31113, Paleobotany Collections, The Field Museum, Chicago and Nos. 15722 and 15204, Illinois State Museum, Springfield, from Francis Creek Shale, Carbondale Formation (middle Desmoinesian), Illinois Basin, Wilmington Township, Will County, Illinois.

Occurrence.—Illinois: middle Desmoinesian (Langford, 1958; Wittry, 2006). If synonymy list accurate, in West Virginia: lower Atokan (Gillespie et al., 1978; Blake et al., 2002); and Pennsylvania: upper Atokan (Oleksyshyn, 1982).

Material examined.—Six specimens from Unit 2.

Description.—The foliage appears coriaceous. Ultimate pinnae are linear with a broadly rounded apex, 47 mm long and 20 mm wide (based on incomplete specimens), and are centered by an inflated and winged rachis (Fig. 8.9), within which sits a thin (~1 mm wide) and depressed vascular strand with faint longitudinal striations. At least four pairs of subopposite to alternate, strongly vaulted lateral pinnules are present on an ultimate pinna. In proximal parts (Fig. 8.5, 8.9, 8.12), markedly quadrate pinnules (9-14 mm long, 11-14 mm wide) are affixed to the rachis near their basiscopic margin, have a strongly incised acroscopic margin that runs parallel to the rachis, and exhibit a shallow incision on the outermost margin, manifest as two rounded lobes on the lowest pinnules. Toward the pinna apex, pinnules (10-14 mm long, 6-9 mm wide) are increasingly decurrent, more broadly attached, closely spaced, and spatulate to obovate, eventually merging with a large, lobate terminal pinnule (Fig. 8.13) that is ~15 mm long and 17 mm wide. Midveins are decurrent (particularly in distal positions), very short, and divide into a number of branches shortly after entering the pinnule; the uppermost branch may ascend nearly parallel to the pinna rachis before reaching the lateral margin. Lateral veins are sparse, gently arch toward the margin, and are depressed within the conspicuously vaulted lamina, resulting in a subtly crenulate lateral margin (Fig. 8.13).

Remarks.—Specimens are fragmentary, precluding confident assignment. The pinnule habit is distinctly 'eusphenopteroid,' although of unusually large size for the fossil-genus. Comparison initially was made with *Eusphenopteris scribanii* Van Amerom, 1975, which has the largest foliar elements known in the genus (Van Amerom, 1975). However, as exhibited in the

holotype and paratype illustrated by Kidston (1923, pl. 3, fig. 2, pl. 4, fig. 4, under *Sphenopteris obtusiloba* Brongniart, 1830-31), pinnules of *E. scribanii* have a denser venation that can be partly obscured by a finely striated surface; these features are inconsistent with the material at hand. Specimens documented by Moore et al. (2014, p. 44, pls. 8, 9, under *Eusphenopteris* sp.), which also are atypically large for the genus, share some similarities with the study examples, including an inflated winged rachis with a sunken vascular strand, a thick lamina, and subtly undulatory pinnule margins that correspond to lateral vein endings. However, the taxon described by Moore et al. (2014) has an overall more lax or loosely spaced appearance and fan-shaped pinnules that may be deeply incised into two or three lobes, rather than quadrate pinnules with only two lobes resulting from shallow incision.

The best comparison is with "Mariopteris" anthrapolis, particularly with two syntypes that have large, vaulted, quadrate to obovate pinnules, probably from proximal parts of a frond (Langford, 1958, text-figs. 488, 493). Wittry (2006, p. 80, fig. 1) reillustrated one syntype, in addition to an example (fig. 2) with a pinnule habit, subtly crenulate margin, and venation scheme identical to the study specimens. Two other syntypes of "M." anthrapolis (Langford, 1958, text-figs. 489, 490), presumably from more distal parts of a frond, have a more 'mariopteroid' appearance, including subtriangular pinnules that show less resemblance to the present examples. Darrah (1969, p. 123) also noted the morphological differences among Langford's syntypes, and posited that they may include more than one species. Unfortunately, the full gamut of variability is not available in our assemblage for comparison, and it is possible that "M." anthrapolis is highly polymorphic. Darrah (1969) erected Mariopteris thompsonii for a species he considered closely related to "M." anthrapolis, although he did so hesitantly because his specimens came from the same stratigraphic unit and location as the syntypes of the latter. The slightly denser venation and trilobate pinnules of M. thompsonii resemble those of Eusphenopteris scribanii. The specimen Oleksyshyn (1982) identified as Mariopteris cf. inflata White, 1943 appears more similar to "M." anthrapolis than what White (1943) probably envisioned.

The genus and systematic position of "Mariopteris" anthrapolis are ambiguous, hence the quotation marks and placement under Incertae sedis. The species could be assigned to Eusphenopteris, and Langford (1958) and Darrah (1969) noted that it could easily be mistaken for a large form of Eusphenopteris neuropteroides. A case in point may be specimens illustrated by Gillespie et al. (1978) and Blake et al. (2002), which are markedly larger than is typical of E. neuropteroides. Two syntypes (Langford, 1958, text-figs. 489, 490) may indeed belong to Mariopteris Zeiller, 1879b, or to the related genus Karinopteris Boersma, 1972 as speculated by Wittry (2006). Hence, more than one genus may be included in the type material, but the specimens on hand probably belong to Eusphenopteris.

Order Cordaitanthales Meyen, 1984
Family Cordaitanthaceae Meyen, 1984
Genus *Cordaites* Unger, 1850 *Cordaites* spp. indet.
Figure 8.6, 8.10

Occurrence.—Euramerica: fossil-genus common throughout Carboniferous and Permian.

Material examined.—Seventy-five specimens in Units 1 and 2.

Description.—Leaves are thick, coriaceous, linear-lanceolate, and reach 150 mm long (although some probably an order of magnitude longer in life) and 8-60 mm wide (full width not always preserved). Lateral margins are entire, subparallel, and taper towards the base and apex, presumably resulting in a strapshaped leaf. The basal attachment structure (Fig. 8.6) is thicker than the lamina, has a conspicuous wrinkled or cracked texture, and has a convex distal edge (where it adjoins the lamina) and a concave proximal edge (where it attached to the branch). Leaf surfaces comprise parallel, longitudinal ridges and furrows, representing primary veins and intervening lamina. Primary veins rarely bifurcate at a very low angle, and venation densities range from 11–30 veins per cm in central positions, becoming very dense near lateral margins. The trace of intermediate sclerotic strands, exhibited as faint longitudinal lines between primary veins, may be present. Some examples from Unit 1 have spirorbiform worm-tubes (microconchids) affixed to the surface (Fig. 8.10).

Remarks.—Attempts to classify Cordaites based on the size, shape, and venation pattern of leaves is recognized as wholly artificial (cf., Šimůnek, 2006). Different leaf types can be found on a single branch (Césari and Hünicken, 2013), and venation schemes are not diagnostic because the spacing of primary veins and the presence or absence (and number) of sclerotic strands between the veins can differ depending on placement on a branch, maturity of the plant, which leaf surface is exposed, and even position on a single leaf (Harms and Leisman, 1961; Crookall, 1970; Barthel, 1976; Bashforth, 2005). Furthermore, although Cordaites shows a relatively conservative morphology, cuticular studies highlight the diversity in the group (Zodrow et al., 2000; Šimůnek, 2006). Because no cuticles were procured from the study specimens, all leaves are assigned to Cordaites spp. indet.

During analysis, it was difficult to differentiate between *Cordaites* and axes with densely spaced longitudinal striations, which probably represent naked pteridosperm petioles. The fact that no apices and only rare bases of *Cordaites* were observed further compounded the problem. We emphasize that only specimens that could confidently be assigned to *Cordaites* were described and measured.

Acknowledgments

A Postdoctoral Fellowship from the Natural Sciences and Engineering Research Council of Canada to ARB is acknowledged. Hans-Dieter Sues provided translations of German text. Appreciation is offered to the following for correspondence about systematics, repositories, type localities, and stratigraphic information: B.M. Blake, C.J. Cleal, N. Gilmore, J.-P. Laveine, R. Rößler, J. Schneider, and D. Uhl. The reviews of H.W. Pfefferkorn and E.L. Zodrow were beneficial, as was the assistance of the editorial team (B.R. Pratt, S. Davis, K. Huber).

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