# South American electric knifefishes of the genus Archolaemus (Ostariophysi, Gymnotiformes): undetected diversity in a clade of rheophiles 

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#### Abstract

Neotropical electric knifefishes of the family Sternopygidae previously considered to represent a single relatively geographically widespread and morphologically variable species, Archolaemus blax, were analysed and found to represent a complex of five species, four of which are new to science. A fifth undescribed species from the Rio São Francisco basin outside the previous known range of the genus was identified. Recognized species of Archolaemus are: A. blax, previously thought to occur in the Rio Araguari, Rio Branco, Rio Tapajós, Rio Tocantins, and Rio Xingu, but which instead proved to be endemic to the Rio Tocantins; Archolaemus ferreirai sp. nov. from the Rio Mucajaí and Rio Uraricoera in the north-eastern portions of the Amazon basin; Archolaemus janeae sp. nov. of the Rio Xingu and the upper Rio Tapajós, both southern tributaries of the mainstream Amazon; Archolaemus luciae sp. nov. of the Rio Trombetas, Rio Jari, and Rio Tapajós basins of the eastern Amazon, and the independent Rio Araguari draining into the Atlantic Ocean north of the mouth of the Amazon; Archolaemus orientalis sp. nov. of the São Francisco basin in eastern Brazil; and Archolaemus santosi sp. nov. of the Rio Jamari in the south-western portion of the Amazon basin. The phylogenetic placements of Archolaemus and the recently described genus Japigny relative to the other members of the Eigenmanninae are discussed. A series of synapomorphies for Archolaemus are proposed and a hypothesis of the relationships within that genus is advanced. Rheophily of all members of Archolaemus is discussed, with the genus found to be the most specious clade within the Gymnotiformes living primarily in high-energy settings. The reported anterior projection of the dentary teeth in A. blax was found to be a consequence of postmortem displacement.


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## INTRODUCTION

Archolaemus was proposed by Korringa (1970) as a new genus of the Sternopygidae diagnosed from other members of that family, with the exception of Sternopygus, by the possession of a free orbital rim. In his analysis, Korringa cited various differences between

[^0]Archolaemus and Sternopygus, with the externally most obvious being the larger eye of Archolaemus relative to that in any of the species of Sternopygus recognized at that time. Subsequently, Meunier, Jégu \& Keith (2011) described an additional new genus in the Sternopygidae, Japigny, which also differed from Archolaemus in lacking a free orbit rim.

In the original description of Archolaemus, Korringa (1970) assigned a single species to the genus, Archolaemus blax, which was based on three specimens from a
location in the Rio Tocantins. Schwassmann \& Carvalho (1985) subsequently redescribed A. blax from multiple population samples they equated with that species. These series of specimens significantly expanded the range of A. blax, both within the Rio Tocantins and beyond that drainage to four additional river systems. Two of these, the Rio Xingu and Rio Tapajós, arise from the Brazilian Shield, as does the Rio Tocantins. More notably, Schwassmann \& Carvalho documented the presence of Archolaemus north of the mainstream Amazon in two rivers draining from the Guyana Shield: the west-flowing Rio Branco in the north-eastern portion of the Amazon basin and the east-flowing Rio Araguari that drains directly into the Atlantic Ocean. In their study, they documented that the electrogenic A. blax only generates monophasic wave types, a condition subsequently found to be general across the Sternopygidae, and that it produces electric organ discharges at 390-523 Hz (Kirschbaum, 1995: 175; Crampton \& Albert, 2006: 673, fig. 23.9). Bichuette \& Trajano $(2003,2006)$ more recently documented the occurrence of A. blax within limestone caverns of the karstic São Domingos region of the Rio Tocantins basin.

Following Schwassmann \& Carvalho (1985), a monotypic Archolaemus was recognized in the summary treatment of the Gymnotiformes (MagoLeccia, 1994: 18), in species compendia (Albert, 2003; Campos-da-Paz, 2007), in phylogenetic reconstructions (Albert, 2001; Albert \& Crampton, 2005), and in historical biogeographical analyses (Lima \& Ribeiro, 2011; Winemiller \& Willis, 2011). These actions notwithstanding, it is noteworthy that Schwassmann \& Carvalho (1985: fig. 2) reported that A. blax demonstrates pronounced intraspecific variation in the shape of the snout and overall profile of the head, and that observations at locations in the Rio Tocantins and Rio Xingu basins revealed that it is rheophilic an inhabitant of rapid waters. In and of itself the pronounced intraspecific morphological variation is striking. Moreover, such a geographic range across major portions of the Brazilian and Guyana shields is unexpected in light of the reported rheophily of the genus. Rapids in fast-flowing rivers are patchily distributed on both shields, and many species of fishes associated with these widely spaced, high-energy locations demonstrate notable degrees of apparently small-scale endemism. This general pattern of endemicity among rheophilic fishes and the reported range in head morphology in A. blax raises questions about the reported broad geographic range of the species: specifically, is it actually a single morphologically variable species with a disjunct distribution across multiple separated drainages, or does the assumed monotypic Archolaemus instead subsume multiple species?

In this study we re-examined the samples reported on by Schwassmann \& Carvalho (1985) along with a large percentage of the other known material of Archolaemus deposited in research collections. The aims of the analysis are:

1. to evaluate whether the standing hypothesis of the monotypy of the genus is correct;
2. if it is not, to delimit the recognizable species in the genus and provide descriptions of all of them and redefine the genus accordingly;
3. to identify characters pertinent to the hypotheses of the phylogenetic placement and monophyly of Archolaemus and relationships within the genus;
4. to determine whether the phylogeny and distribution of the species within the genus correlated with proposed potential vicariant events in the region of occurrence; and
5. to investigate rheophily across the genus.

## MATERIAL AND METHODS

## Measurements

Gymnotiforms often suffer damage to their caudal elements, most likely as a result of predation (Lundberg et al., 1996: 666), followed by partial regeneration with clearly 'truncated, or abruptly narrowed and/or abruptly depigmented tails' (Mago-Leccia, Lundberg \& Baskin, 1985: 1). These damaged individuals are typically readily recognizable externally, and were excluded from morphometrics involving total length and caudal length, and from counts of anal-fin rays when the damage impinged on the fin. Morphological measurements were point-to-point linear distances, taken using a digital caliper accurate to 0.1 mm . The measurements and abbreviations cited in the text are: anal-fin length, from the origin of the anal fin to the posterior end of the anal-fin base; branchial opening, from the dorsal to the ventral limits of the aperture; caudal filament length (CL), from the posterior margin of the last anal-fin ray to the tip of the caudal filament; depth of caudal filament, between the dorsal and ventral margins of the caudal filament immediately posterior to the base of the last anal-fin ray; greatest body depth, the vertical distance from the origin of the anal fin to the dorsal margin of the body; head length (HL), from the tip of the snout to the posterior margin of the branchial opening; head length at opercle, from the tip of the snout to the posterior margin of the bony opercle; head depth at eye, between the dorsal and ventral margins of the head at the vertical through the eye; head depth at nape, between the dorsal and ventral margins of the head at the vertical through the nape; head width, at the middle of the opercle; internarial
distance, from the posterior margin of the anterior naris to the anterior margin of the posterior naris; internarial width, between the inner margins of the contralateral anterior nares; interorbital distance, the linear distance between the medial margins of the orbits; posterior naris to eye, from the posterior margin of the posterior naris to the anterior margin of the eye; length of mouth, from the tip of the snout to the rictus; snout to origin of anal fin, the distance from the tip of the snout to the base of first anal-fin ray; snout to anus, the distance from the tip of the snout to the anterior margin of the anus; snout to posterior naris, the distance from the anterior margin of the eye to the posterior margin of the posterior naris; length to the end of the anal fin (LEA), the distance from the tip of the snout to the end of the base of the anal fin; orbital diameter, between the anterior and posterior margins of the orbit; pectoralfin length, the distance from the dorsal border of the fin base to the tip of the longest ray; postorbital length, the distance from the posterior margin of the eye to the posterior margin of the opercle; preanal-fin length, the distance from the tip of the snout to the origin of the anal fin; snout length, the distance from the tip of the snout to the anterior margin of the eye; total length (TL), the distance from the tip of the snout to the posterior tip of the caudal filament; and width of mouth, taken across the posterior limit of the mouth.

## MERISTICS

The counts of anal- and pectoral-fin rays were primarily obtained under a microscope using transmitted light, with these data supplemented by counts taken from radiographs. In the pectoral-fin ray counts, the unbranched anterior rays are represented by lower case Roman numerals and the branched rays are represented by Arabic numerals. The branched-ray count includes all rays posterior to the anterior unbranched rays, including the posteriormost ray in the series, which is sometimes unbranched. Analfin ray counts are presented in terms of anterior unbranched and total rays. Values in square brackets are those of the holotype. Osteological observations were made from cleared and stained specimens prepared following Taylor \& Van Dyke (1985), with these cited as CS in the lists of material examined.

The authorship of four of the new species (Archolaemus ferreirai sp. nov., Archolaemus janeae sp. nov., Archolaemus luciae sp. nov., and Archolaemus santo$s i \mathrm{sp}$. nov.) is the same as the authorship of this paper. The fifth new species, Archolaemus orientalis sp. nov., was initially recognized as possibly undescribed by Donald J. Stewart (State University of New York,

College of Environmental Science and Forestry), who is the first author for that species.

## Institutional abbreviations

Institutional abbreviations are: CAS, California Academy of Sciences, San Francisco; FMNH, Field Museum of Natural History, Chicago; INPA, Instituto Nacional de Pesquisas da Amazônia, Manaus; MNRJ, Museu Nacional, Rio de Janeiro; MPEG, Museu Paraense Emílio Goeldi, Belém; MZUSP, Museu de Zoologia da Universidade de São Paulo, São Paulo; and USNM, National Museum of Natural History, Smithsonian Institution, Washington.

## PhYLOGENETIC POSITION AND MONOPHYLY OF JAPIGNY

The recent description of Japigny by Meunier et al. (2011) did not delve into the question of the phylogenetic position of the genus within the Sternopygidae, a factor that complicates the evaluation of the monophyly and intrageneric relationships within Archolaemus. Japigny can be unambiguously diagnosed by the presence of a series of conditions, including: the pattern of alternating dark bands (versus the absence of such pigmentation in other members of the Sternopygidae, with the exception of Sternopygus astrabes); the possession of a distinct single row of teeth at the base of the upper oral valve (versus the absence of such dentition); the ventral curvature of the lateral process of the parapophysis of the second vertebrae (versus a horizontal lateral process of the parapophysis); the distinct separation of the lateral process of the second vertebrae from the parapophysis of the fourth vertebrae (versus the parapophyses of the second and fourth vertebrae in contact); the ventrally curved parapophysis of the fourth vertebrae (versus the horizontal lateral process of parapophysis of the fourth vertebrae); and the angulo-articular not included in the socket of the lower jaw articulating with the quadrate (versus the angulo-articular being included in the socket). A comprehensive analysis of phylogenetic relationships within the Sternopygidae lies outside this study, but the osteological observations associated with the question of the relationships within Archolaemus indicate that Japigny is a component of the Eigenmanninae in the Sternopygidae. Evidence supporting this hypothesis is the possession in Japigny of a scapula with an included scapular foramen, a fusion of the post-temporal and the supracleithrum into a single ossification, with a profile comparable with that of the two elements when separate (e.g. Eigenmannia virescens; MagoLeccia, 1978: fig. 14), and a relatively low number of precaudal vertebrae (12-16).

Alternative hypotheses have been advanced as to the relationships among Archolaemus, Distocyclus, Eigenmannia, and Rhabdolichops (the Eigenmanninae prior to the description of Japigny). Under the hypotheses of Mago-Leccia (1978) and Alves-Gomes (1998), Rhabdolichops is the basalmost genus of the Eigenmanninae. Lundberg \& Mago-Leccia (1986), Albert \& Fink (1996), Albert \& Campos-da-Paz (1998), Albert (2001), and Correia, Crampton \& Albert (2006) conversely proposed that Archolaemus is the basalmost taxon in that subfamily. None of the previous phylogenetic analysis included the recently described Japigny. Three of the examined characters variable within the Sternopygidae were informative for this question, and indicate that Japigny is the sister group to a clade composed of Archolaemus, Distocyclus, Eigenmannia, and Rhabdolichops. These are: (1) the angulo-articular lacking a distinct socket to receive the condyle of the quadrate (versus with a distinct socket); (2) the parapophysis of the second vertebra straight and contacting the parapophysis of the fourth vertebra (versus the parapophysis curved ventrally and clearly separated from the parapophysis of the fourth vertebra); and (3) the parapophysis of the fourth vertebra straight (versus curved ventrally). Thus, Japigny may resolve as the basalmost member of Eigenmanninae, with this resolution providing a foundation for the evaluation of the monophyly of Archolaemus and for character coding within the genus.

## MONOPHYLY AND INTRARELATIONSHIPS of ARCholaemus

Archolaemus shares the following unique combination of characters that define the Sternopygidae: the presence of multiple rows of small needle-like teeth in the form of a villiform band on the dentary; the relatively large eyes; the enlargement of the antorbital and infraorbitals, with expanded bony arches over the laterosensory canal segments; the anterior naris located within the vertical limits of the gape; an anal-fin origin located along the isthmus; and the lack of a caudal fin and dorsal electroreceptive organ (Hulen, Crampton \& Albert, 2005).

Four proposed synapomorphies that delimit Archolaemus as monophyletic were identified in our analysis.

1. A free orbital rim. As reported by several authors (e.g. Nijssen \& Isbrücker, 1972: 174; Mago-Leccia, 1994: 18; Meunier et al., 2011: 48), all species of the Gymnotiformes, except for Archolaemus and Sternopygus, have the orbit covered by skin, with this surface layer attached to tissues bordering the eye. Juveniles of Archolaemus share this apparently primitive condition of an orbit covered
by skin and attached to the surrounding tissues, in some species up to approximately 135 mm TL; however, larger individuals of the genus all have the orbital rim free of the surrounding orbital margin. In the context of present concepts of relationships within the Gymnotiformes, the free orbital rim of Archolaemus and Sternopygus is considered a homoplastic derived character in these two genera.
2. Form of attachment of the teeth to the premaxilla. As in all other sternopygids, the species of Archolaemus bear teeth on the premaxilla (approximately 22-35 teeth, depending on the species). The villiform teeth in sternopygids are typically immobile and attached to the ventral surface of the premaxilla. Conversely, in Archolaemus only the anterobasal margins of the teeth of the first tooth row are attached to the dentigerous surface of the premaxilla. As a consequence, these teeth are variably mobile relative to the premaxilla with the range of movement ranging between a few and 90 degrees: an apparent apomorphy for the members of the genus.
3. Association of the posterior margin of the upper lip and the anterior margin of the premaxilla. The anterior margin of the premaxilla lies close to and contacts, or almost contacts, the posterior margin of the upper lip from a ventral view in all species of the Sternopygidae other than for Archolaemus. In contrast, all species of Archolaemus have a pronounced gap equal to approximately one-half the width of the eye between the anterior margin of the premaxilla and the posterior margin of the upper lip (A. blax in Korringa, 1970: fig. 2A).
4. Morphology of the upper lip. The ventral surface of the upper lip is relatively flat and unelaborated from a ventral view in most members of the Sternopygidae, as well as across the Gymnotiformes. Conversely, the ventral surface of the upper lip is porous and sponge-like with raised papillae and fleshy anteroposteriorly elongate ridges of various extents in all of the species of Archolaemus. Elsewhere within the Sternopygidae this condition was encountered only in some populations of what is now considered to be Eigenmannia trilineata. Within the context of present hypotheses of relationships within the Sternopygidae (see the discussion under character 1 ), the occurrence of this form of upper lip in some specimens of $E$. trilineata and the species of Archolaemus would be homoplastic, but with the attribute a synapomorphy for the members of Archolaemus.

Two characters previously advanced as autapomorphies for what was then thought to be a monotypic

Archolaemus proved not to apply across the expanse of the more species-rich genus in this study. Albert (2001: 71) proposed that a lateral ethmoid contacting four bones was an autapomorphy for A. blax. As such it potentially served as a synapomorphy for the six species of the genus we recognize herein. Our survey shows that the complex lateral ethmoid does not universally contact four bones across the genus. A posterior naris positioned closer to the tip of the snout than to the eye was hypothesized by Albert (2001: 71) to be an autapomorphy for A. blax. Comparisons reveal, however, that the posterior naris is sometimes located approximately in the middle of the snout among several of the previously undescribed species of Archolaemus recognized herein.

Relationships within Archolaemus. The data indicate that $A$. orientalis is the sister group of a clade composed by A. blax, A. ferreirai, A. janeae, A. luciae, and A. santosi. The hypothesis of the monophyly of a clade formed by these five species is supported by three synapomorphies.

1. Presence of a narrow stripe of dark pigmentation along the lateral line. The presence of a narrow stripe of dark pigmentation extending along the lateral line is absent in A. orientalis, and is unique to these five species (see species accounts) among the examined members of the Sternopygidae, other than for also occurring in some species of Eigenmannia such as E. trilineata. What appears as a dark line of pigmentation proximate to the lateral line in A. orientalis is in fact the line of contact between the epaxial and hypaxial musculature.
2. Presence of a band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin. The presence of a broad band of dusky to distinctly dark pigmentation formed of individual vertical bars paralleling the pterygiophores, with the bars sometimes conjoining in darker specimens to form a variably continuous broad stripe, is limited to the members of this clade (see species accounts).
3. Form of posterodorsal portion of the dentary. Whereas the posterodorsal portion of the dentary is ossified in other sternopygids, this region is occupied by cartilage in these five species. Arratia (1992) made the case that given that the dentary is a dermal bone, the cartilage in the area typically occupied by the posterodorsal portion of the dentary, the 'cartilaginous coronoid process' in her terminology, is at least in some catfishes more likely to be a dorsal extension of Meckel's cartilage.

Within the clade of five species supported by characters $1-3$, the evidence supports a hypothesis of two clades, each including two species (A. blax
plus $A$. janeae and A. ferreirai plus A. luciae). Those two clades along with $A$. santosi form an unresolved trichotomy.

Archolaemus blax and A. janeae are hypothesized to be sister species based on their common possession of two derived characters.
4. Relative size of the posterior ceratohyal versus ventral hypohyal. In these species, the posterior ceratohyal is approximately 1.5 times the length of the ventral hypohyal versus the two bones being approximately the same size in their congeners and other sternopygids.
5. Number of tooth rows on the posterior portion of dentary. A single tooth row on the posterior portion of the dentary occurs in both of these species, whereas other congeners have two rows of teeth in this area. The remaining sternopygids with teeth present on the posterior portion of the dentary have two or more rows of teeth in that region.

The remaining pair of sister species, $A$. ferreirai and A.luciae, share one hypothesized derived character.
6. Relative size of the coronomeckelian bone versus Meckel's cartilage. In these two species the elongate coronomeckelian bone extends along a considerable portion of the inner surface of the lower jaw, and corresponds to $50 \%$ or more of the length of Meckel's cartilage. In other congeners and outgroups the more compact coronomeckelian bone is approximately $20 \%$ or less of the length of Meckel's cartilage.

## Historical biogeography

The relatively restricted geographic expanse occupied by the species of Archolaemus on the scale of South America, in conjunction with the long-term stability of both the Guyana and Brazilian shields that underlie most of the areas of occurrence of the members of the genus, result in relatively few major geological events being potentially informative as to the possible minimum ages of the genus and its included clades. One aspect of the phylogeny that is informative as to a minimal age of the genus as a whole is the basal dichotomy between A. orientalis, an endemic of the Rio São Francisco basin of eastern Brazil, versus the clade formed by its five congeners. All congeners of A. orientalis either inhabit rivers within the Amazon basin (Rio Branco, Rio Jari, Rio Tapajós, and Rio Xingu) or occur in drainage systems that empty in areas near the lower portions of that basin (Rio Araguari and Rio Tocantins).

Various sister-group relationships between fish taxa living in the São Francisco basin versus Rio Tocantins
and river systems further to the west have been documented in other groups of fishes (see Costa, 2003, 2010). It has been proposed that disruption of the connection between these drainages is a consequence of uplift events that date back approximately 60-40 Myr (Campos \& Dardenne, 1997). Presuming that the basal division between $A$. orientalis versus the clade formed by A.blax, A.ferreirai, A.janeae, A. luciae, and A. santosi is a result of that geological event, then Archolaemus extends back at least 40 Myr . This estimated age is admittedly distinctly older than the oldest known fossil gymnotiform from the Yecua Formation in Bolivia, which dates back about 10 Myr (Gayet \& Meunier, 1991), but in so far as the specimen in question represents the only identified fossil of the order, fossil information is at best minimally informative as to the age of the order and the included subunits. Evidence from molecular studies (AlvesGomes, 1999; Lovejoy et al., 2010) indicates that the order and included major taxa date to periods prior to the $40-\mathrm{Myr}$ minimum possible date for Archolaemus. Thus, the uplift sequence that led to a separation of the São Francisco systems from drainages to the west temporally falls within the realm of possibility as a basal vicariance event within that genus.

Biogeographical analyses within the clade sister to A. orientalis are limited by the incomplete resolution of the scheme of relationships of the five species that resolve as a trichotomy. The units within the trichotomy at this level are A. santosi and two sisterspecies pairs: A. blax plus A. janeae on the one hand and A.ferreirai plus A.luciae on the other. These species pairs have largely disjunct distributions on either side of the mainstream Amazon. Archolaemus blax is limited to the Rio Tocantins and A. janeae is an inhabitant of the Rio Xingu and Rio Tapajós basins: all river systems to the south of the mainstream Amazon. Within the second species pair, A. ferreirai is limited to the upper Rio Branco and A. luciae is found primarily in the Rio Trombetas and Rio Araguari, all of which lie north of the Amazon, but with some populations of A. luciae in the Rio Tapajós south of the Amazon. This general pattern is congruent with the hypothesis of a division of Archolaemus populations at the level of the ancestor of A. blax, A. ferreirai, A. janeae, A. luciae, and A. santosi, by either marine transgressions into what is now the lower Amazon (Bloom \& Lovejoy, 2011) or by the development of the present lower Amazon system following the breaching of the Purus Arch in the late Miocene (approximately 8 Mya ; Lundberg et al., 1998). Either of these events could have separated the ancestors of these two species pairs, with subsequent speciation events in each unit followed by a dispersal of A. luciae into the Rio Tapajós to the south of the main channel.

## Rheophily of Archolaemus

In their multifaceted ecological and taxonomic analysis of what they considered to be populations of A. blax, Schwassmann \& Carvalho (1985: 236) reported that individuals of the species took refuge during the day 'inside crevices and between rocks at places of high current velocities' at localities in the Rio Xingu (Altamira and Belo Monte) and the Rio Tocantins (Rio Itacaiunas) basins. Our results demonstrate that Schwassmann \& Carvalho's life-history observations were actually of two species, A.janeae (Rio Xingu localities) and A. blax (Rio Tocantins), both of which are clearly rheophiles (those authors did not make life observation in the Rio Araguari system, the home of the third species included in their concept of $A$. blax). Archolaemus blax also inhabits high water flow subterranean settings within karstic formations in the São Domingos region of the upper Rio Tocantins system (Bichuette \& Trajano, 2003). Ferreira, dos Santos \& Jégu (1988: 345) reported swift water habitat preferences for A. ferreirai (identified as A. blax in that study) in the rapids of the Rio Mucajaí of the Rio Branco system. Ferreira (1995: 51), in turn, indicated that A. luciae (identified as A.blax in that study) is resident in swiftly moving waters within the Rio Trombetas. Observations of A. luciae in the Rio Tapajós system, A. orientalis at its type locality in the Rio São Francisco, and information on the type locality of A. santosi in the Rio Madeira basin (see Habitat in the accounts for these species) indicate that these three species similarly dwell in high-energy settings. Habitat preference information is unavailable for many of the lots examined of Archolaemus in this study, but it is noteworthy that many of these samples originated at, or proximate to, localities identified as falls (cachoeiras), attesting to the rapid water conditions in those localities. A preference for swiftly flowing waters is clearly an attribute that is general across the species of Archolaemus.

Recent advances in the knowledge on the alphataxonomy, habitat preferences, and phylogenetic relationships among Neotropical electric fishes demonstrate that life in shallow waters, including swift water conditions, evolved several times independently across the Gymnotiformes (de Santana \& Vari, 2009, 2010a, b). Crampton (2011) noted that there have been multiple transitions within the Gymnotiformes from deep river habitats to life in high-energy systems, such as rapids, with no apparent reversals in habitat preferences. It is interesting that only two genera in the Gymnotiformes, Archolaemus and Megadontognathus, are composed entirely of rheophiles: species specializing in life in a high-energy aquatic environment. Megadontognathus includes only two species (Campos-da-Paz, 1999), which makes Archolaemus, with six
species, by far the most species-rich clade of exclusively rheophlic species in the Gymnotiformes.

Rheophily, the behaviour of inhabiting high-energy aquatic environments, such as occurs in all species of Archolaemus, is a phenomenon that is widespread among freshwater fishes, albeit limited to a small subset of the total number of species in the geographically extensive ichthyofaunas encompassing such swift water specialist species (Kullander, 1988; Lima \& Zuanon, 2004; de Santana \& Vari, 2010a). Adaptations for rheophily are myriad, but in some instances are apparently limited to behavioural strategies, or at least would seem to be so restricted based on externally obvious morphological adaptations. The observations by Schwassmann \& Carvalho (1985: 237) document that behaviour undoubtedly contributes to the utilization of rapid flowing waters by A. blax in the Rio Itacaiunas and A. janeae in the Rio Xingu. Behavioural adaptations are supplemented in many rheophilic species by, sometimes manifold, morphological specializations for life in rapids and proximate to waterfalls. One of these specializations, the possession of a subconical downwardly oriented snout (Kullander, 1988), characterizes the species of Archolaemus, but many of the other specializations common in other rheophiles (e.g. dorsoventrally flattened head and body) are not feasible within the body plan of the Sternopygidae.

## TAXONOMIC ACCOUNTS

## Archolaemus Korringa, 1970

Archolaemus Korringa, 1970: 267 (Type species Archolaemus blax Korringa, 1970; by original designation).

Diagnosis: Archolaemus is diagnosed by the synapomorphies described and discussed above, and distinguished from all remaining genera of the Sternopygidae by a pronounced gap between the anterior margin of the premaxilla and the posterior margin of the upper lip, with this separation equal to approximately one-half the width of the eye. It is further separated from Distocyclus, Eigenmannia, Japigny, and Rhabdolichops by the possession of a free orbital rim in adults (a feature uniquely shared with Sternopygus within the Gymnotiformes). Archolaemus is distinguished from Sternopygus by various features, with those of particular note being the possession of a postcleithrum (versus the absence of that ossification), the presence of a foramen in the scapula (versus the absence of that aperture), and the presence of some branched anal-fin rays (versus all anal-fin rays unbranched). One of the attributes that Korringa (1970) used to delimit Archolaemus from Sternopygus was the relative orbital diameter. The subsequent
description of S. astrabes by Mago-Leccia (1994), with a relatively large eye ( $13.8-19.5 \% \mathrm{HL}$; Hulen et al., 2005), resulted in an overlap of the range of orbital diameters in the species of Sternopygus, with values for this feature sitting among the range of values found in species of Archolaemus (Tables 1 and 2).

Dentition: Jaw dentition of all species of Archolaemus consists of small, needle-like teeth embedded in the fleshy jaws. Consequently, details of the number of teeth and their exact distribution in each jaw are difficult to ascertain other than in cleared and counterstained specimens. Korringa (1970: fig. 2) provides a good illustration of the general dentition pattern (see, however, our comment concerning the dentary dentition in the next paragraph), with the dentary dentition consisting of bands of small teeth extending for varying distances posteriorly along the dorsal margin of the bone. Premaxillary dentition consists of two rounded to anteroposteriorly elongate patches that correspond to the form of the overlying premaxillae, and are separated from each other medially.

Korringa (1970: 267) reported that larger specimens of A. blax have 'a number of dentary teeth outside the mouth and project forward in larger specimens.' Schwassmann \& Carvalho (1985: 232) were unable to confirm this condition: an observation that is in line with our results. Some larger specimens of Archolaemus can have the dentary dentition somewhat more prominent anteriorly, but not projecting forwards.

Distribution: Species of Archolaemus are known from the Rio Branco, Rio Madeira, Rio Tapajós, and Rio Xingu catchments within the Amazon basin, and outside that system in the Rio Tocantins, the easterly flowing Rio Araguari in the state of Amapá, Brazil, and the Rio São Francisco system of eastern Brazil (see species accounts).

Secondary sexual dimorphism: Secondary sexual dimorphism is manifest in several features among the species of Archolaemus. These include the distinctly darker overall pigmentation of the head and body in mature males, relative to the condition in females and juveniles, in A. ferreirai, the pronounced elongation of the snout in mature males of A. luciae, versus the condition in females and immatures, and the presence of teeth on the upper lip of large males of A.ferreirai, compared with the absence of such dentition in mature females. Limited samples of many species and the absence of mature males for others make it impossible to determine the generality of these features across the genus. This limitation notwithstanding, these modifications represent the first reported instances of secondary sexual dimorphism in the Sternopygidae.
Table 1. Morphometrics for examined specimens of Archolaemus blax, Archolaemus ferreirai sp. nov., and Archolaemus janeae sp. nov.

|  | Archolaemus blax |  |  | Archolaemus ferreirai sp. nov. |  |  |  | Archolaemus janeae sp. nov. |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Range | $N$ | Mean | H | Range | $N$ | Mean | H | Range | $N$ | Mean |
| Total length (mm) | 176-382 | 17 | - | 183 | 68.8-342 | 50 | - | 160 | 110-224 | 13 | - |
| Length to end of anal fin (mm) | 140-300 | 17 | - | 142 | 67.4-286 | 40 | - | 127 | 118-171 | 8 | - |
| Caudal filament length (mm) | 36.8-77.5 | 12 | - | 34.3 | 16.8-67.3 | 32 | - | 32.6 | 21.3-42.5 | 6 | - |
| Head length (mm) | 20.1-45.3 | 16 | - | 23.2 | 12.0-40.2 | 50 | - | 19.8 | 16.9-26.6 | 12 | - |
| Percentage of length to end of anal fin |  |  |  |  |  |  |  |  |  |  |  |
| Anal-fin length | 63.1-94.1 | 17 | 82.4 | 82.3 | 78.3-86.8 | 39 | 83.0 | 82.6 | 82.6-87.4 | 9 | 84.7 |
| Snout to anus | 6.4-10.7 | 17 | 8.6 | 9.0 | 7.4-13.3 | 40 | 9.5 | 13.1 | 8.7-13.1 | 9 | 10.7 |
| Greatest body depth | 10.6-15.9 | 17 | 13.3 | 13.2 | 10.7-16.0 | 39 | 13.4 | 12.3 | 11.4-13.7 | 9 | 12.5 |
| Preanal-fin distance | 13.1-18.6 | 17 | 16.8 | 17.7 | 14.8-26.3 | 39 | 18.1 | 17.8 | 15.2-17.8 | 9 | 16.3 |
| Pectoral-fin length | 9.0-14.2 | 16 | 11.3 | 11.2 | 8.0-15.3 | 40 | 10.6 | 10.7 | 8.2-11.8 | 9 | 10.0 |
| Caudal filament length | 22.9-38.1 | 12 | 30.4 | 24.2 | 18.1-36.6 | 31 | 26.6 | 25.7 | 16.0-25.7 | 7 | 20.5 |
| Head length | 13.5-17.0 | 15 | 14.8 | 16.3 | 14.0-18.6 | 39 | 16.1 | 15.6 | 13.8-15.8 | 9 | 14.8 |
| Head length at opercle | 13.2-16.3 | 16 | 14.2 | 15.8 | 13.9-20.9 | 39 | 15.9 | 14.3 | 13.5-14.7 | 9 | 14.1 |
| Percentage of head length |  |  |  |  |  |  |  |  |  |  |  |
| Head depth at eye | 39.6-48.0 | 17 | 44.2 | 46.9 | 38.2-58.0 | 50 | 48.2 | 49.4 | 39.9-53.5 | 15 | 46.2 |
| Head depth at nape | 59.3-72.1 | 16 | 65.2 | 61.8 | 57.9-75.8 | 50 | 66.8 | 62.5 | 57.2-72.8 | 15 | 65.3 |
| Head width | 39.6-59.6 | 17 | 46.0 | 43.5 | 37.7-54.4 | 50 | 46.7 | 43.9 | 39.8-55.6 | 15 | 44.8 |
| Orbital diameter | 12.1-18.5 | 16 | 15.0 | 15.8 | 9.2-18.9 | 50 | 14.2 | 18.2 | 14.0-22.0 | 15 | 16.7 |
| Interorbital distance | 17.6-25.2 | 17 | 21.7 | 21.8 | 16.4-29.0 | 50 | 22.2 | 22.2 | 19.2-23.9 | 15 | 21.9 |
| Internarial distance | 7.2-9.6 | 17 | 8.4 | 8.4 | 6.0-9.4 | 50 | 7.9 | 8.3 | 7.1-9.7 | 15 | 8.2 |
| Internarial width | 9.2-11.5 | 17 | 10.2 | 9.4 | 6.5-13.5 | 50 | 9.8 | 14.3 | 8.5-14.3 | 15 | 10.5 |
| Snout length | 39.8-47.4 | 16 | 43.2 | 44.6 | 34.9-47.6 | 50 | 42.2 | 39.0 | 35.8-43.0 | 15 | 39.3 |
| Snout to posterior naris | 17.1-22.1 | 17 | 20.0 | 23.7 | 17.5-23.7 | 50 | 20.6 | 20.7 | 18.8-23.6 | 15 | 21.1 |
| Posterior naris to eye | 15.7-24.1 | 17 | 20.1 | 18.7 | 12.9-24.5 | 50 | 19.1 | 13.6 | 12.5-18.1 | 15 | 14.7 |
| Mouth length | 18.5-29.6 | 17 | 23.8 | 25.2 | 19.3-30.0 | 50 | 22.8 | 24.4 | 19.5-30.5 | 15 | 23.7 |
| Mouth width | 12.5-10.7 | 17 | 15.9 | 16.6 | 11.1-19.9 | 50 | 15.9 | 16.1 | 11.0-19.1 | 15 | 15.7 |
| Branchial opening | 26.6-31.0 | 17 | 28.5 | 26.4 | 23.2-31.1 | 50 | 27.3 | 28.0 | 22.8-31.1 | 15 | 26.2 |
| Postorbital length | 41.0-44.5 | 14 | 43.1 | 39.8 | 39.8-51.6 | 50 | 44.6 | 45.8 | 38.6-48.2 | 15 | 44.2 |
| Percentage of caudal filament |  |  |  |  |  |  |  |  |  |  |  |
| Caudal filament depth | 5.0-6.5 | 12 | 5.7 | 6.1 | 3.6-9.2 | 32 | 6.2 | 5.9 | 5.6-11.2 | 7 | 7.3 |

$N$, number of specimens; H, holotype; range includes holotype of species.

Table 2. Morphometrics for examined specimens of Archolaemus luciae sp. nov., Archolaemus orientalis sp. nov., and Archolaemus santosi sp. nov.

|  | Archolaemus luciae sp. nov. |  |  |  | Archolaemus orientalis sp. nov. |  |  |  | Archolaemus santosi sp. nov. |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | H | Range | $N$ | Mean | H | Range | $N$ | Mean | H | Range | $N$ | Mean |
| Total length (mm) | 267 | 96-284 | 29 | - | 156 | 150.8-181 | 3 | - | 197 | 73-212 | 12 | - |
| Length to end of anal fin (mm) | 192 | 76-207 | 25 | - | 110.7 | 110.7-155 | 3 | - | 159 | 62-198 | 11 | - |
| Caudal filament length (mm) | 73.3 | 23.7-89.9 | 8 | - | 44.4 | 36.7-44.4 | 2 | - | 38.1 | 11.1-38.1 | 8 | - |
| Head length (mm) | 32.8 | 10.7-42.1 | 29 | - | 15.8 | 15.3-19.9 | 3 | - | 27.0 | 10.6-32.1 | 12 | - |
| Percentage of length to end of anal fin |  |  |  |  |  |  |  |  |  |  |  |  |
| Anal-fin length | 89.9 | 79.8-88.9 | 25 | 82.7 | 83.0 | 70.3-83.0 | 3 | 78.5 | 80.5 | 79.7-85.3 | 11 | 82.2 |
| Snout to anus | 7.1 | 7.1-12.8 | 25 | 9.5 | 7.0 | $6.5-7.0$ | 3 | 6.8 | 9.0 | 7.9-14.0 | 11 | 10.7 |
| Greatest body depth | 13.5 | 11.5-14.3 | 25 | 12.9 | 15.3 | 13.0-15.3 | 3 | 14.5 | 14.6 | 12.7-14.7 | 11 | 13.8 |
| Preanal-fin distance | 17.5 | 15.0-20.7 | 25 | 17.7 | 17.0 | 15.0-17.6 | 3 | 16.3 | 17.4 | 16.6-19.6 | 11 | 17.9 |
| Pectoral-fin length | 12.7 | 8.5-17.8 | 25 | 10.9 | 10.0 | 9.4-10.0 | 3 | 9.6 | 10.0 | 9.2-11.9 | 11 | 10.4 |
| Caudal filament length | 46.1 | 28.6-46.1 | 7 | 35.4 | 40.1 | 32.1-40.1 | 2 | 36.1 | 19.3 | 14.4-27.0 | 7 | 20.6 |
| Head length | 16.8 | 13.8-20.4 | 25 | 16.0 | 14.3 | 12.8-14.3 | 3 | 13.5 | 17.0 | 15.5-17.1 | 11 | 16.4 |
| Head length at opercle | 16.4 | 13.2-19.4 | 25 | 15.0 | 13.2 | 11.7-13.2 | 3 | 12.5 | 16.0 | 15.0-16.3 | 11 | 15.6 |
| Percentage of head length |  |  |  |  |  |  |  |  |  |  |  |  |
| Head depth at eye | 43.3 | 37.6-57.9 | 28 | 43.7 | 48.7 | 47.9-49.0 | 3 | 48.5 | 43.6 | 41.4-48.4 | 12 | 45.2 |
| Head depth at nape | 60.6 | 53.1-70.0 | 29 | 62.8 | 77.6 | 73.3-79.8 | 3 | 76.9 | 65.5 | 63.9-70.9 | 12 | 65.5 |
| Head width | 41.5 | 38.1-61.7 | 29 | 43.7 | 50.7 | 50.7-53.3 | 2 | 52.0 | 43.5 | 42.6-48.2 | 12 | 44.8 |
| Orbital diameter | 12.2 | 10.5-17.8 | 29 | 14.1 | 15.8 | 15.8-17.2 | 3 | 16.4 | 11.7 | 8.9-16.6 | 12 | 14.1 |
| Interorbital distance | 16.1 | 14.1-27.5 | 29 | 20.7 | 27.3 | 20.8-27.3 | 3 | 24.9 | 19.7 | 19.7-30.7 | 12 | 24.1 |
| Internarial distance | 8.6 | 5.4-10.5 | 28 | 7.8 | 9.0 | 8.2-9.0 | 2 | 8.6 | 8.3 | 7.0-10.3 | 12 | 8.4 |
| Internarial width | 9.7 | 6.6-12.4 | 29 | 8.8 | 13.3 | 13.3-15.3 | 2 | 13.3 | 14.3 | 7.6-14.3 | 12 | 10.7 |
| Snout length | 48.4 | 38.4-50.2 | 29 | 44.4 | 34.9 | 34.6-38.4 | 3 | 36.0 | 38.6 | 35.3-46.5 | 12 | 39.1 |
| Snout to posterior naris | 20.9 | 16.7-25.6 | 29 | 20.4 | 21.0 | 21.0-21.2 | 3 | 21.1 | 20.7 | 11.7-24.7 | 12 | 21.4 |
| Posterior naris to eye | 26.3 | 10.2-26.3 | 29 | 19.5 | 11.5 | 11.5-12.8 | 2 | 12.1 | 16.7 | 6.4-18.9 | 12 | 13.7 |
| Mouth length | 26.3 | 15.1-32.6 | 29 | 20.3 | 25.2 | 22.4-25.2 | 2 | 23.8 | 19.3 | 14.3-26.6 | 12 | 21.8 |
| Mouth width | 15.2 | 9.3-20.9 | 29 | 13.3 | 19.0 | 19.0-20.6 | 2 | 19.8 | 15.0 | 7.0-15.0 | 12 | 8.4 |
| Branchial opening | 29.6 | 19.0-31.6 | 29 | 24.9 | 28.4 | 27.6-28.4 | 3 | 28.0 | 24.5 | 23.1-29.2 | 12 | 26.7 |
| Postorbital length | 43.6 | 41.3-59.1 | 29 | 45.4 | 54.0 | 49.7-54.0 | 3 | 51.8 | 43.4 | 42.1-49.8 | 12 | 45.6 |
| Percentage of caudal filament |  |  |  |  |  |  |  |  |  |  |  |  |
| Caudal filament depth | 3.3 | 3.3-4.8 | 8 | 3.9 | 3.3 | 3.3-4.4 | 2 | 3.9 | 7.6 | 6.7-9.1 | 7 | 8.2 |

[^1]Remarks: In their description of Japigny, Meunier et al. (2011) differentiated that genus from Archolaemus on the basis of pigmentation patterns and the total number of anal-fin rays. Their concept of Archolaemus was based on samples from the Rio Xingu, which presumably represent what we consider to be A. janeae, the only member of the genus known from that river system. Archolaemus orientalis described in this paper, however, has a range of total anal-fin rays (164-186) overlapping, albeit barely, with the values in Japigny (132-164), and lacks the pattern of longitudinal dark and light pigmentation common to the other species of Archolaemus. As such, the characters originally proposed to distinguish the two genera are no longer applicable. Archolaemus and Japigny differ in the association of the eye with the surrounding regions (free versus attached) and pigmentation pattern (dark pigmentation either absent or, when present, in the form of a dark stripe along the lateral line and a broad dusky to dark band over the basal pterygiophores of the anal fin versus wide alternatively dark and light vertical bars along the body).

In their analysis of Archolaemus blax, Schwassmann \& Carvalho (1985) detailed the apparent pronounced variation in head form between populations within what they considered to be that species. In part, this variation was a consequence of ontogenetic modifications in snout and overall head morphology, as reflected in differences in the head profile from a lateral view. Supplementing these intraspecific ontogenetic alterations are alternative degrees of snout development across the species in Archolaemus, four of which (A. blax, A. ferreirai, A. janeae, and A. luciae) were encompassed within A. blax as delimited by Schwassmann \& Carvalho (1985). Of note is the par-
ticularly elongate and broader snout of A. luciae, more so in mature males (Fig. 7). This factor contributes significantly to the range of snout profiles within what those authors considered to be A. blax.

Conservation status: The rapids and waterfalls patchily distributed across the gradients of the Brazilian and Guiana shields (Lima \& Ribeiro, 2011; Lujan \& Armbruster, 2011) are home to a number of specialized communities of plants and animals, including rheophilic fishes. Such high-energy settings significantly hinder the sampling of these communities, and as such our knowledge of the ichthyofaunas in these settings is often deficient, even within the context of the still inadequate overall understanding of the Neotropical freshwater fish fauna (Vari \& Malabarba, 1998). At the same time the high-energy conditions that complicate the thorough sampling of these habitats provide the attributes that are ideal for hydroelectric facilities. The development of hydroelectric generating capacity is accelerating across the Neotropics, with the consequent submergence under reservoirs of multiple habits preferentially occupied by rheophiles. Moreover, proposed impoundments (i.e. the Belo Monte project on the lower Rio Xingu; Fearnside, 2006) threaten many as yet undeveloped high-energy settings. It is likely that various rheophilic species have been extirpated from their type localities by hydroelectric developments. Within the Gymnotiformes, this eventuality probably applies both in Archolaemus (e.g. A. santosi from the Rio Jamari at Usina Hidroelétrica Samuel in Rondônia) and Sternarchorhynchus (e.g. Sternarchorhynchus britskii in the upper Rio Paraná; Campos-da-Paz, 2005: 399). The same is likely to befall many of the locations inhabited by the species of Archolaemus and other rheophiles.

## Key to the species of Archolaemus

1a. 18-22 total pectoral-fin rays; 190 to 228 total anal-fin rays; dark narrow stripe running along lateral line...... 2 1b. 16 total pectoral-fin rays; 164-186 total anal-fin rays; no dark narrow stripe running along lateral line
.Archolaemus orientalis sp. nov. (upper Rio São Francisco basin, Brazil)
2a. Mouth with posterior limit of opening extending posteriorly beyond vertical through posterior naris
. .3
2b. Mouth with posterior limit of opening falling short of vertical through posterior naris.
3a. Caudal-filament depth 3.3-4.8\% of caudal filament length
.Archolaemus luciae sp. nov. (Rio Jari, Rio Trombetas and Rio Tapajós basins in eastern Amazon; independent Rio Araguari, Brazil)
3b. Caudal-filament depth $5.0-6.5 \%$ of caudal filament length.........Archolaemus blax (Rio Tocantins basin, Brazil)
4a. 10-14 scales above lateral line at midbody ( 14 scales present only in three of 50 examined specimens.
.Archolaemus ferreirai sp. nov. (Rio Mucajaí and Rio Uraricoera, Amazon basin, Brazil)
4b. 14-17 scales above lateral line at midbody ( 14 scales present in none of the examined specimens of $A$. janeae sp. nov. and in only one of 12 examined specimens of $A$. santosi sp. nov).
.. 5
5a. Head length to rear of opercle $13.5-14.7 \%$ of length to the end of the anal fin; mouth width $11.0-19.1 \%$ of head length.
.Archolaemus janeae sp. nov. (Rio Xingu and upper Rio Tapajós, Amazon basin, Brazil)
5b. Head length to rear of opercle $15.0-16.3 \%$ of length to the end of the anal fin; mouth width $7.0-10.3 \%$ of head length.
.Archolaemus santosi sp. nov. (Rio Jamari, Amazon basin, Brazil)

## Archolaemus blax Korringa, 1970

(Figs 1, 2; TABLE 1)
Archolaemus blax Korringa, 1970: 267, fig. 1 (type locality: Brazil, Goiás, Porto Nacional, Rio Tocantins, $10^{\circ} 40^{\prime} \mathrm{S}, 048^{\circ} 30^{\prime} \mathrm{W}$; type species of Archolaemus by original designation). Mago-Leccia, 1994: 115, fig. 1A and B (Brazil, Rio Tocantins, Jatobal; drawing of head of holotype; based on information in Korringa, 1970; assignment to Eigenmanninae). Schwassmann \& Carvalho, 1985: 231 (only cited specimens from the Rio Tocantins system; no samples from Rio Branco, Rio Cupixi, Rio Tapajós, or Rio Xingu; not fig. 1; reports of rheophily, behaviour, and diet of populations in the Rio Itacaiunas and Rio Tocantins basins). Albert \& Fink, 1996: 87 (phylogenetic relationships). Albert \& Campos-da-Paz, 1998: 423 (phylogenetic relationships). Alves-Gomes, 1998: 447 (phylogenetic relationships). Albert, 2003: 487 (in listing of members
of Sternopygidae). Bichuette \& Trajano, 2003: 1109 (subterranean habits, São Domingos karst, upper Rio Tocantins basin). Bichuette \& Trajano, 2006: 100 (occurrence in caves, São Domingos karst, upper Rio Tocantins basin). Campos-da-Paz, 2007: 121 (Brazil, Amazon basin). Lima \& Caires, 2011: 10 (Brazil, upper Rio Tocantins basin, Rio Novo/Rio do Sono basin). Lucinda et al., 2007: 82 (Brazil, Rio Tocantins basin, Lajeado Reservoir region).

Diagnosis: Archolaemus blax is diagnosed from $A$. ferreirai by the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris, versus falling short of that line), the length of the coronomeckelian bone (less than $20 \%$ the length of Meckel's cartilage versus $50 \%$ or more of the length of Meckel's cartilage), and the length of the posterior ceratohyal ( 1.5 times the length of the ventral hypohyal


Figure 1. Archolaemus blax, 288 mm total length; MNRJ 12158, female, Brazil, Goiás, Município de Minaçu/Cavalcante, Rio Tocantins, at site of Usina Hidroelétrica Serra da Mesa (approximately $13^{\circ} 44^{\prime} \mathrm{S}, 048^{\circ} 08^{\prime} \mathrm{W}$ ).


Figure 2. Map of Rio Tocantins and adjoining areas, showing geographical distribution of Archolaemus blax ( $1=$ type locality; some symbols represent more than one locality and/or lot of specimens).
versus approximately the same size as the ventral hypohyal). Archolaemus blax is diagnosed from A. janeae by the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris versus falling short of that line), the number and arrangement of teeth on the internal surface of the endopterygoid (approximately six irregularly distributed teeth versus two or three teeth arranged in a single row), the form of the premaxilla (the anteroposterior length greater than the transverse width versus the anteroposterior length equal to the transverse width), and the distribution of teeth on the dentary (dentition restricted to the anterior half or slightly more of the dorsal margin versus occupying most of the dorsal region). Archolaemus blax is diagnosed from A. luciae by the depth of the caudal filament (5.0-6.5\% CL versus $3.3-4.8 \% \mathrm{CL}$ ), the length of the coronomeckelian bone (less than $20 \%$ the length of Meckel's cartilage versus $50 \%$ or more of the length of Meckel's cartilage), and the length of the posterior ceratohyal (1.5 times the length of the ventral hypohyal versus approximately the same length as the ventral hypohyal). Archolaemus blax is diagnosed from $A$. orientalis by the pigmentation pattern (the presence of a narrow dark stripe along the lateral line and a broad dusky to dark band of pigmentation overlying the basal pterygiophores of the anal fin, versus the lack of such dark pigmentation), the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris, versus falling short of the vertical through the posterior margin of the posterior naris), the total number of pectoral-fin rays (19-22 versus 16), the total number of anal-fin rays (193-222 versus 164-186), the form of the premaxilla (the anteroposterior length greater than the transverse width, versus the anteroposterior length equal to the transverse width), and the length of the posterior ceratohyal ( 1.5 times the length of the ventral hypohyal versus approximately the same length as the ventral hypohyal). Archolaemus blax is diagnosed from A. santosi by the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris versus falling short of that line), the depth of the caudal filament (5.0-6.5\% CL versus 6.7-9.1\% CL), the form of the premaxilla (the anteroposterior length greater than the transverse width versus the anteroposterior length equal to the transverse width) and the length of the posterior ceratohyal (1.5 times the length of the ventral hypohyal versus approximately the same length as of the ventral hypohyal).

Description: Morphometric data for the examined specimens of A. blax are presented in Table 1.

Body elongate and laterally compressed. Greatest body depth located along abdominal cavity or slightly posterior to that region. Body profile gently convex from rear of head for approximately anterior one-
third of body, and then gradually angled posteroventrally along progressively tapering body and caudal filament. Ventral profile of body distinctly convex anteriorly along anterior half of abdominal cavity, and then gradually angled posterodorsally along base of anal fin and caudal filament. Anteriormost perforated lateral line scale located at vertical approximately at origin of pectoral fin. Lateral line continuous, extending from immediately after vertical through origin of pectoral fin to end of caudal filament.

Head laterally compressed, widest at opercular region and deepest at nape. Head profile varying from nearly straight to slightly concave along antorbital region, and then straight to slightly convex from that point to rear of head. Ventral profile of head straight and posteroventrally aligned. Snout subconical and anteroventrally directed. Eye small and located laterally on dorsal half of head. Orbital rim partially attached to surface tissues of adjoining region of head in specimen of approximately 105 mm TL, but rim totally free and without membranous attachment in larger examined individuals. Anterior naris located at end of short tube and positioned short distance posterior of tip of snout, but distant from anterior margin of eye. Posterior naris elliptical, without tube, and located nearer to tip of snout than to anterior margin of eye in adults. Mouth distinctly inferior, with upper jaw clearly longer than and overlapping lower jaw. Gape relatively long and rictus in adults positioned posterior of vertical through posterior margin of posterior naris. Branchial opening moderately elongate, located along posterior margin of opercle and immediately anterior to anterior margin of pectoral-fin origin. Branchial membranes joined at isthmus and extending posteriorly to under pectoral-fin base. Anus proximate to elongate urogenital papilla, and both structures positioned approximately ventral of orbit. Position of anus and urogenital papilla shifting ontogenetically from posterior of vertical through posterior margin of orbit to more anterior position.

Scales small, cycloid, and present from immediately posterior of head to end of caudal filament. Scales along lateral line 131 to 152 ( 137 reported for the holotype by Korringa, 1970: 268) ( $N=20$ ). Scales above lateral line at midbody 12 to $14(N=30)$. Scales absent on head.

Pectoral fin long, approximately two-thirds of head length, broad, and distally pointed with lateral rays longer. Pectoral-fin rays ii, 19 to ii, 22 (19 reported for the holotype by Korringa, 1970: 268) ( $N=30$ ). Analfin elongate, extending from vertical through base of pectoral-fin base along most of length of body. Fin margin gently convex anteriorly and then straight for much of length of fin, but with rays becoming progressively shorter posteriorly. Anal fin with 23-27 unbranched rays ( $N=25$ ) and $193-222$ total fin rays
(218 reported for the holotype by Korringa, 1970: 268) ( $N=20$ ). Caudal filament of moderate length.

Coloration in alcohol: Overall body coloration ranging from tan to brown. Lightly coloured specimens lacking, or with faint indications of, pigmentation pattern characteristic of darker individuals. More intensely pigmented individuals with overall brown pigmentation further developed dorsally, and with narrow band of dark pigmentation extending along lateral line; band extending from vertical through tip of pectoral fin or slightly posterior of that line to end of caudal filament. Basal pterygiophores of anal fin outlined by very narrow dark bars situated on body surface. Bars cumulatively form dark, broad, band running along ventral portion of body, with height of band gradually decreasing posteriorly. Dark pigmentation along centre of band irregularly less intense in many individuals, resulting in darker dorsal and ventral components in band. Dark stripe along lateral line distinctly separated in most specimens from dark band overlying anal-fin basal pterygiophores by more lightly pigmented, broad, midlateral stripe. Lighter midlateral stripe less obvious in some overall more darkly pigmented individuals. Head ranging from tan to dark brown. Tan specimens with upper two-thirds of head slightly dusky, but otherwise without distinct pigmentation pattern. Darkly pigmented specimens sometimes with ventral surface of head tan, but most often with head dark over nearly all surfaces. Dark pigmentation often particularly intense on opercle and around mouth, but with lips lightly coloured. Pigmentation of pectoral and anal fins variable, ranging from tan in lightly coloured specimens to quite dusky with dark pigmentation overlying fin rays in more intensely pigmented individuals.

Distribution: Archolaemus blax is known from multiple locations within the Rio Tocantins basin (Fig. 2).

Ecology: The concept of A. blax described by Schwassmann \& Carvalho (1985) encompassed what has now proven to be a species complex. Analysis of a sample from the Rio Itacaiunas, in the Rio Tocantins basin, reported on by those authors (MZUSP 24158) confirmed that this population is A. blax. At that locality the species preferred fast-flowing waters, with individuals sheltering during the day between rocks and within crevices in locations with high-velocity currents (Schwassmann \& Carvalho, 1985: 237). They observed individuals emerge from sheltered locations in the evening, but their nocturnal activities could not be tracked. Stomach content analysis showed that at that location, A. blax feeds most often on insect larvae, primarily of the Chironomidae, with the stomachs also containing some sand and filamentous
algae, presumably ingested incidentally with the capture of the target prey. Individuals of both sexes had mature or maturing gonads and well-developed genital papillae during the period (October) of Schwassmann \& Carvalho's study, with mature males and females being of approximately the same size. The species has also been reported from subterranean habitats in the São Domingos karst formation in the upper Rio Tocantins basin, with these settings characterized by moderate to very strong water currents, particularly during the rainy season (Bichuette \& Trajano, 2003: 1111).

Remarks: Multiple researchers after Schwassmann \& Carvalho (1985) followed those authors in treating all samples of Archolaemus from a series of major river systems in the Amazon and adjoining catchments as A. blax. Results herein demonstrate that Schwassmann \& Carvalho's concept of A. blax encompassed a species complex. Based on the analysis of the bulk of the specimens reported on by those authors, supplemented on the examination of numerous other samples from across the range of the genus, we consider citations of A. blax from the Rio Xingu to be based on A. janeae, those from the Rio Cupixi to be A. luciae, and that from the Rio Branco to be A. ferreirai.

## Material examined

BRAZIL. Goiás: Porto Nacional, Rio Tocantins (approximately $\quad 10^{\circ} 40^{\prime} \mathrm{S}, \quad 048^{\circ} 30^{\prime} \mathrm{W}$ ); CAS 24743 (435; holotype). Município de Minaçu/Cavalcante, Rio Tocantins, at site of Usina Hidroelétrica Serra da Mesa (approximately $13^{\circ} 44^{\prime} \mathrm{S}, 048^{\circ} 08^{\prime} \mathrm{W}$ ); MNRJ 12158, 22 (90-382; 4 CS). Rio Bezerra, right-bank tributary of Rio Paranã, 2 km above Fazenda Monte Alegre de Goiás (approximately $13^{\circ} 16^{\prime} \mathrm{S}, 047^{\circ} 30^{\prime} 10^{\prime \prime} \mathrm{W}$ ); MZUSP 40686, 4 (65-193). Rio Tocantizinho and tributary, Serra da Mesa, Minaçu (approximately $13^{\circ} 48^{\prime} \mathrm{S}$, $048^{\circ} 18^{\prime} 10^{\prime} \mathrm{W}$ ); MZUSP 54080, 3 (215-285). Rio São Vicente, São Domingos in Lapa São Vicente II (approximately $13^{\circ} 30^{\prime} \mathrm{S}, 046^{\circ} 18^{\prime} \mathrm{W}$ ); MZUSP 55987, 1 (139). Rio Angélica, Lapa do Angélica, São Domingos (approximately $13^{\circ} 31^{\prime} \mathrm{S}, 046^{\circ} 23^{\prime} \mathrm{W}$ ); MZUSP 86112, 1 (not measured). Maranhão: Rio Tocantins, estreito do Tocantins (approximately $06^{\circ} 08^{\prime} 58^{\prime} \mathrm{S}, 047^{\circ} 29^{\prime} 15^{\prime} \mathrm{W}$ ); MZUSP 4991, 1 (212). Pará: Rio Tocantins, Capuerana, upriver from Tucurui dam (approximately $03^{\circ} 51^{\prime} \mathrm{S}, 049^{\circ} 41^{\prime} \mathrm{W}$ ); INPA 6421, 1 (313). Rio Tocantins, above Tucurui dam ( $\left.03^{\circ} 45^{\prime} 58^{\prime \prime} \mathrm{S}, \quad 049^{\circ} 40^{\prime} 21^{\prime \prime} \mathrm{W}\right)$; INPA 4828, 11 (45-340); INPA 6424, 23 (87-292; 4 CS; one Sternopygus intermingled with material); INPA 9071, 1 (280). Breu Branco (approximately $04^{\circ} 04^{\prime} \mathrm{S}$, $\left.049^{\circ} 38^{\prime} \mathrm{W}\right)$; INPA 20963, 1 (325). Rio Tocantins, Jatobal, sandy lagoon in front of Jatobal $\left(04^{\circ} 32^{\prime} \mathrm{S}\right.$, $\left.049^{\circ} 32^{\prime} \mathrm{W}\right)$; MZUSP 24158, 2 (92-133); MZUSP 24129, 24 (37-390). Rio Anapu, Estuário ( $01^{\circ} 48^{\prime} 25^{\prime} \mathrm{S}$,
$\left.051^{\circ} 21^{\prime} 19^{\prime \prime} \mathrm{W}\right)$; MPEG 8596, 1 (195). Tocantins: Rio do Sono, Cachoeira da Velha, Rio Novo basin ( $09^{\circ} 20^{\prime} \mathrm{S}$, $047^{\circ} 44^{\prime} \mathrm{W}$ ); MZUSP 97943, 1 (286).

## ARCHOLAEMUS FERREIRAI SP. NOV.

 (Figs 3, 4; TABLE 1)Archolaemus blax, Schwassmann \& Carvalho, 1985: 233 (only specimens from Brazil, Roraima, Rio Branco, Cachoeira de Bem Querer; not details of species redescription or biological information). Ferreira et al., 1988: 345 (Brazil, Roraima, Rio Mucajaí, vicinity of Ilha Paredão; carnivorous diet). Ferreira et al., 2007: 184 (Brazil, Roraima, Rio Branco basin).

Diagnosis: Archolaemus ferreirai is diagnosed from A. blax by the length of the gape (the rictus falling short of the vertical through the posterior naris versus extending beyond that line), the length of the coronomeckelian bone ( $50 \%$ or more the length of Meckel's cartilage versus less than $20 \%$ the length of Meckel's cartilage) and the length of the posterior ceratohyal (approximately the same length as the ventral hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus ferreirai is diagnosed from A.janeae in the number of scales above the lateral line at the midbody ( $10-14$ versus $15-17$ ), by the form of the premaxilla (the anteroposterior length greater than the transverse width versus the anteroposterior length equal to the transverse width), the


Figure 3. A, Archolaemus ferreirai sp. nov., holotype, female, 183 mm total length, INPA 3757; Brazil, Roraima, Rio Mucajaí, Cachoeira Paredão 2 (approximately $02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}$ ); B, Archolaemus ferreirai sp. nov., paratype, male, 342 mm total length, INPA 36379; Brazil, Roraima, Rio Mucajaí, Cachoeira Paredão 2 (approximately $02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}$ ).


Figure 4. Map of upper Rio Branco and adjoining areas showing geographical distribution of Archolaemus ferreirai sp. nov. (1, holotype locality; some symbols represent more than one locality and/or lot of specimens).
distribution of the teeth on the dentary (dentition restricted to anterior one-half or slightly more of the dorsal margin versus occupying most of the dorsal margin), the length of the coronomeckelian bone ( $50 \%$ or more the length of Meckel's cartilage versus less than 20\% the length of Meckel's cartilage), and the length of the posterior ceratohyal (approximately the same length as the ventral hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus ferreirai is diagnosed from A. luciae by the length of the gape (the rictus not extending posteriorly beyond the vertical through the posterior naris versus extending beyond that line) and the distribution of the teeth on the internal surface of the endopterygoid (five or six teeth arranged in single row versus between seven and ten irregularly distributed teeth). Archolaemus ferreirai is diagnosed from A. orientalis in the pigmentation pattern (the presence of a narrow dark stripe along the lateral line and a broad band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin, versus the lack of such dark pigmentation), the total number of pectoral-fin rays ( $18-21$ versus 16 ), the total number of anal-fin rays (190-222 versus 164-186), the form of the premaxilla (the anteroposterior length greater than the transverse width versus the anteroposterior length equal to the transverse width), and the length of the coronomeckelian bone ( $50 \%$ or more of the length of Meckel's cartilage versus less than $20 \%$ the length of Meckel's cartilage). Archolaemus ferreirai is diag-
nosed from A. santosi in the form of the premaxilla (the anteroposterior length greater than the transverse width versus the anteroposterior length equal to the transverse width) and the length of the coronomeckelian bone ( $50 \%$ or more the length of Meckel's cartilage versus less than $20 \%$ the length of Meckel's cartilage).

Description: Morphometric data for the examined specimens of $A$. ferreirai are presented in Table 1.

Body elongate and laterally compressed. Greatest body depth located along abdominal cavity. Body profile gently convex from rear of head for approximately anterior one-third of body, and then gradually angled posteroventrally along progressively tapering body and caudal filament. Ventral profile of body slightly convex anteriorly along anterior half of abdominal cavity, and then gradually angled posterodorsally along base of anal fin and caudal filament. Anterior most perforated lateral line scale located at vertical approximately at origin of pectoral fin. Lateral line continuous, extending from immediately after vertical through origin of pectoral fin to end of caudal filament.

Head laterally compressed, widest at opercular region and deepest at nape. Head profile nearly straight dorsally. Ventral profile of head straight and posteroventrally aligned. Snout subconical and anteroventrally directed. Eye small and located laterally on dorsal half of head. Orbital rim covered by
membrane and partially free of surface tissues of adjoining region of head in specimens of up to approximately 119 mm TL, but rim totally free and without membranous attachment in larger examined individuals. Anterior naris located at end of short tube and positioned short distance posterior of tip of snout, but distant from anterior margin of eye. Posterior naris elliptical, without tube and located nearer tip of snout than to anterior margin of eye. Mouth distinctly inferior, with upper jaw clearly longer than, and overlapping, lower jaw. Gape relatively short and in adults falling short posteriorly of vertical through posterior margin of posterior naris. Branchial opening moderately elongate; located along posterior margin of opercle and immediately anterior to anterior margin of pectoral-fin origin. Branchial membranes joined at isthmus and extending posteriorly to under pectoral-fin base. Anus proximate to elongate urogenital papilla, and both structures positioned approximately ventral of orbit. Position of anus and urogenital papilla shifting anteriorly ontogenetically from posterior of vertical through posterior margin of orbit to position along, or anterior of, vertical through anterior margin of orbit in larger specimens.

Scales small, cycloid, and present from immediately posterior of head to end of caudal filament. Scales along lateral line 136 to 150 (145) ( $N=25$ ). Scales above lateral line at midbody 10 to $14(12)(N=50)$. Scales absent on head

Pectoral fin long, approximately two-thirds of head length, broad, and distally pointed with lateral rays longer. Pectoral-fin rays between ii, 16 and ii, 19 (18) ( $N=50$ ). Anal-fin elongate, extending from vertical through base of pectoral-fin base along most of length of body. Fin margin gently convex anteriorly and then straight, but with rays becoming progressively shorter posteriorly. Anal fin with 24-33 (28) unbranched rays $(N=30)$ and $190-222$ (197) total fin rays $(N=30)$. Caudal filament of moderate length.

Coloration in alcohol: Overall body coloration ranging from tan to light brown, with lightly coloured specimens demonstrating a less intense version of pigmentation pattern characteristic of darker individuals. Overall brown pigmentation more developed dorsally, with narrow dark stripe extending along lateral line from vertical through tip of pectoral fin, or slightly posterior of that line to end of caudal filament. Band less apparent, but still obvious in very darkly pigmented adult male specimens. Skin overlying basal pterygiophores of anal fin with pterygiophores outlined by very narrow dark bars on body surface. Bars cumulatively form dusky to dark, broad, band running along ventral portion of body, with height of band gradually decreasing posteriorly. Dark pigmentation along centre of band often less intense, with
resultant darker dorsal and ventral components to band. Dark stripe along lateral line and ventral band above anal fin separated by more lightly pigmented, broad, midlateral stripe. Lighter midlateral stripe less obvious in some overall more darkly pigmented individuals, and obscured to significant degree by overall dark pigmentation in the single examined large male specimen. Head ranging from tan ventrally and dark dorsally to dark brown over all surfaces, other than for narrow lightly coloured midventral stripe. Lips of darkly pigmented specimens distinctly lighter than proximate areas. Region anteroventral and posteroventral of pectoral-fin base more darkly pigmented than adjoining regions. Pigmentation of pectoral and anal fins ranging from tan in more lightly coloured specimens to quite dusky with dark pigmentation overlying fin rays in more intensely pigmented individuals.

Distribution: Archolaemus ferreirai is known from north-eastern portions of the Amazon basin in the Rio Mucajaí at Cachoeira Paredão 2 and the Rio Uraricoera at Ilha de Maracá, both in Roraima, Brazil (Fig. 4).

Secondary sexual dimorphism: The only examined mature male of Archolaemus ferreirai (INPA 3757) has an overall body coloration (Fig. 3B) much darker than in the juveniles and females (Fig. 3A) captured at the same time. This darker pigmentation largely obscures the lighter longitudinal band present in mature females and juveniles between the dark stripe along the lateral line and the dark band overlying the basal pterygiophores of the anal fin. The upper lip of this large male is also notable in bearing some teeth, contrary to the absence of such dentition in mature females and juveniles.

Habitat: Ferreira et al. (1988: 342) reported the type region of Archolaemus ferreirai, the Rio Mucajaí at Cachoeira Paredão 2, as characterized by rapidly flowing waters and a 20 m high waterfall. The sample of A. ferreirai (reported as A. blax in that paper) was captured above these falls, in an area with many rapids and no still backwaters.

Etymology: The species name, Archolaemus ferreirai sp. nov., is in honor of Efrem Ferreira of the Instituto Nacional de Pesquisas da Amazônia, one of the collectors of the type series, in recognition of his many contributions to our understanding of the Amazonian fish fauna.

## Material examined

Holotype: BRAZIL. Roraima: Rio Mucajaí, Cachoeira Paredão 2 (approximately $02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}$ ), collected by E. G. Ferreira \& M. Jégu, 19 February 1987; INPA 3757 (183).

Paratypes: BRAZIL. Roraima: collected with holotype; INPA 36379, 21 (119-342; 1 CS); MPEG 21683, 1 (286); MZUSP 110183, 1 (150); USNM 404354, 1 (172). Rio Mucajaí, mouth of Igarapé Traira (approximately $02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}$ ), collected by E. G. Ferreira \& M. Jégu, 21 February 1987; INPA 6422, 12 (131-269; 4 CS).
Non-type specimens: BRAZIL. Roraima: Rio Uraricoera, Ilha Maracá, Furo Santa Rosa (approximately $03^{\circ} 22^{\prime} \mathrm{N}, 061^{\circ} 22^{\prime} \mathrm{W}$ ); INPA 2038, 6 (122-176). Rio Uraricoera, Ilha Maracá, below Furo Santa Rosa (approximately $03^{\circ} 22^{\prime} \mathrm{N}, 061^{\circ} 22^{\prime} \mathrm{W}$ ); INPA 2054, 4 (135-184). Rio Uraricoera, Ilha Maracá (approximately $03^{\circ} 22^{\prime} \mathrm{N}, 061^{\circ} 22^{\prime} \mathrm{W}$ ); INPA 2675, 1 (54). Rio Uraricoera, Ilha Maracá, below Furo Santa Rosa (approximately $03^{\circ} 22^{\prime} \mathrm{N}, 061^{\circ} 22^{\prime} \mathrm{W}$ ); INPA 3747, 1 (60). Rio Mucajaí, station Fé-Esperança; INPA 3758, 3 (83145). Rio Mucajaí, 2 km above Cachoeira Paredão (approximately $02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}$ ); INPA 6423, 1 (338). Rio Mucajaí, 2 km above Cachoeira Paredão, mouth of igarapé (approximately $02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}$ ); INPA 6495, 1 (136); INPA 6500, 7 (115-223). Rio Mucajaí, near Cachoeira Paredão (approximately $\left.02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}\right)$; INPA 6496, 11 (69-196). Rio Mucajaí, above Cachoeira Paredão (approximately $\left.02^{\circ} 57^{\prime} \mathrm{N}, 061^{\circ} 27^{\prime} \mathrm{W}\right)$; INPA 6499, 1 (192).

## Archolaemus Janeae sp. nov.

(Figs 5, 6; TABLE 1)
Archolaemus blax, Schwassmann \& Carvalho, 1985: 233, fig. 1 (in part, specimens from Rio Xingu, Belo Monte, Pará, Brazil; not details in species redescription or biological information). Moller, 1995: 451, 455, 453 (illustration, feeding habits). Camargo, Giarrizzo \& Isaac, 2004: 139 (Brazil, lower and middle Rio Xingu). Montag, Freitas, Wosiacki, \& Barthem, 2008: 20 (Brazil, Para, Rio Xingu basin, Floresta Nacional de Caxiuanã).

Diagnosis: Archolaemus janeae is diagnosed from A. blax in the length of the gape (the rictus falling short posteriorly of the vertical through the posterior naris versus extending beyond that line), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), the distribution of the teeth on the dentary (dentition occupying most of the dorsal margin versus restricted to the anterior one-half or slightly more of the margin), and the number and arrangement of teeth on the internal surface of the endopterygoid (two or three teeth arranged in a single row versus six irregularly distributed teeth). Archolaemus janeae is diagnosed from A. ferreirai by the number of scales above the lateral line at the midbody ( $15-17$ versus $10-14$ ), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), the distribution of the teeth on the dentary (dentition occupying most of the dorsal margin versus restricted to the anterior one-half or slightly more of the margin), the length of the coronomeckelian bone (less than $20 \%$ the length of Meckel's cartilage versus $50 \%$ or more of length of Meckel's cartilage), and the length of the posterior ceratohyal (1.5 times the length of the ventral hypohyal versus approximately the same size as the ventral hypohyal). Archolaemus janeae is diagnosed from A. luciae in the length of the gape (the rictus falling short posteriorly of the vertical through the posterior naris versus extending beyond that line), the length of the caudal filament (16.0$25.7 \%$ LEA versus $28.6-46.1 \%$ LEA), the depth of the caudal filament (5.6-11.2\% CL versus 3.3-4.8\% CL), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), the distribution of the teeth on the dentary (dentition occupying most of the dorsal margin versus restricted


Figure 5. Archolaemus janeae sp. nov., holotype, 160 mm total length, INPA 30832; Brazil, Pará, Rio Iriri, just upriver of its mouth into Rio Xingu, Município de Altamira ( $03^{\circ} 48^{\prime} 54^{\prime \prime} \mathrm{S}$, $052^{\circ} 37^{\prime} 09^{\prime \prime} \mathrm{W}$ ).


Figure 6. Map of lower portion of Amazon basin and adjoining areas, showing geographical distribution of Archolaemus janeae sp. nov. (dots; 1, holotype locality) and Archolaemus luciae sp. nov. (squares; 2, holotype locality) (some symbols represent more than one locality and/or lot of specimens).
to the anterior one-half or slightly more of the margin), the length of the coronomeckelian bone (less than $20 \%$ the length of Meckel's cartilage versus $50 \%$ or more of the length of Meckel's cartilage), and the length of the posterior ceratohyal (1.5 times the length of the ventral hypohyal versus approximately the same size as the ventral hypohyal). Archolaemus janeae is diagnosed from A. orientalis by the pigmentation pattern (the presence of a narrow dark stripe along the lateral line and a broad band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin, versus the lack of such dark pigmentation), the total number of pectoral-fin rays (19-21 versus 16), the total number of anal-fin rays (205-228 versus $164-186)$, the distribution of the teeth on the dentary (dentition occupying most of the dorsal margin versus restricted to the anterior one-half or slightly more of the margin), and the length of the posterior ceratohyal ( 1.5 times the length of the ventral hypohyal versus approximately the same size as the ventral hypohyal). Archolaemus janeae is diagnosed from A. santosi by the pigmentation pattern (the presence of a narrow dark stripe along the lateral line and a broad band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin, versus the lack of such dark pigmentation), the head length at the opercle ( $13.5-14.7 \%$ LEA versus $15.0-$ $16.3 \% \mathrm{LEA}$ ), the distribution of the teeth on the dentary (dentition occupying most of the dorsal margin versus restricted to the anterior one-half or
slightly more of the margin), and the length of the posterior ceratohyal ( 1.5 times the length of the ventral hypohyal versus approximately the same size as the ventral hypohyal).

Description: Morphometric data for examined specimens of A. janeae are presented in Table 1.

Body elongate and laterally compressed. Greatest body depth located along abdominal cavity or slightly posterior to that region. Body profile very gently convex from rear of head for approximately anterior one-third of body, and then gradually angled posteroventrally along progressively tapering body and caudal filament. Ventral profile of body distinctly convex anteriorly below anterior one-half to twothirds of abdominal cavity, more so in specimens with distended abdomens, and then gradually angled posterodorsally along base of anal fin and caudal filament. Anteriormost perforated lateral line scale located at vertical situated approximately at origin of pectoral fin. Lateral line continuous, extending from immediately after vertical through origin of pectoral fin to end of caudal filament.

Head laterally compressed, widest at opercular region and deepest at nape. Head profile ranging from nearly straight to slightly concave from tip of snout to vertical through orbit, and then straight from that area to rear of head. Ventral profile straight and posteroventrally aligned. Snout subconical, anteroventrally directed and moderately developed.

Eye small and located laterally on dorsal one-half of head. Orbital rim partially attached to surface tissues of adjoining region of head in examined specimen of approximately 105 mm TL, but rim totally free and without membrane in larger individuals. Anterior naris located at end of short tube and positioned short distance posterior of tip of snout, but distant from anterior margin of eye. Posterior naris elliptical, without tube and located nearer to tip of snout than to anterior margin of eye. Mouth distinctly inferior, with upper jaw clearly longer than, and overlapping, lower jaw. Gape relatively long, and extending posteriorly beyond vertical through posterior margin of posterior naris. Branchial opening moderately elongate; located along posterior margin of opercle and immediately anterior to anterior margin of pectoralfin origin. Branchial membranes joined at isthmus and extending posteriorly to under pectoral-fin base. Anus proximate to elongate urogenital papilla, and both structures positioned approximately ventral of orbit. Position of anus and urogenital papilla shifting ontogenetically from posterior of vertical through posterior margin of orbit to more anterior position.

Scales small, cycloid, and present from immediately posterior of head to end of caudal filament. Scales along lateral line 146 to 161 (155) $(N=15)$. Scales above lateral line at midbody 15 to 17 (15) $(N=15)$. Scales absent on head.

Pectoral fin long, approximately two-thirds of head length, broad, and distally pointed with lateral rays longer. Pectoral-fin rays ii, 19 to ii, 21 (19) rays ( $N=15$ ). Anal-fin elongate, extending from vertical through base of pectoral-fin base along most of length of body. Fin margin gently convex anteriorly and then straight for length of fin, but with rays becoming progressively shorter posteriorly. Anal fin with 22-33 (33) unbranched rays $(N=15)$ and $205-228$ total fin rays (220) ( $N=9$ ). Caudal filament of moderate length.

Coloration in alcohol: Overall ground body coloration ranging from tan to dark brown. More lightly coloured specimens with pigmentation pattern characteristic of darker specimens less pronounced. More intensely pigmented individuals with overall brown pigmentation more developed dorsally, and narrow dark stripe extending over lateral line from vertical through tip of pectoral fin or slightly posterior of that line to end of caudal filament. Stripe particularly dark in intensely pigmented specimens, such that it stands out even against an overall dark anterior portion of body. Area overlying anal-fin basal pterygiophores with ossifications outlined by variably dark bars on body surface. Bars narrow, distinct, and dusky in overall more lightly coloured specimens, in which they cumula-
tively form dusky band along ventral portion of body with height of band gradually decreasing posteriorly. More darkly pigmented band in dark specimens quite intense, and individual narrow bars sometimes merge into continuous dark broad band running along ventral portion of body. Dark stripe along lateral line and ventral band overlying basal pterygiophores separated in most specimens by more lightly pigmented, broad midlateral stripe. Lighter stripe less obvious in overall very lightly coloured specimens, and obscured anteriorly in some overall more darkly pigmented individuals. Head ranging from tan ventrally and dusky dorsally in lightly coloured individuals to totally dark brown in large, dark specimens. Smaller darkly pigmented specimens sometimes with lightly coloured area on ventral surface of head. Fleshy covering of branchial opening lightly pigmented, even in dark specimens. Larger specimens in some samples with dark pigmentation patch present immediately anterior to base of pectoral fin. Pectoral and anal fins ranging from hyaline in smaller individuals to dusky in larger specimens.

Coloration in life: (Based on photograph of recently captured specimen from São Felix do Xingu along the Rio Xingu.) Dark coloration overall quite intense, with head, portion of body dorsal of lateral line, and region overlying basal pterygiophores of anal fin nearly black. Rays of anal fin overlain by black pigmentation. Pectoral fin black. Midlateral more lightly pigmented stripe present in most preserved specimens, obscured anteriorly by darker pigmentation that gradually becomes more diffuse posteriorly.

Distribution: Archolaemus janeae is known from the upper and lower portions of the Rio Xingu basin at the Rio Iriri, the Cachoeira de Kaituká at Município de Altamira, and localities in the Rio Culuene, as well as the upper Rio Tapajós basin, where it was collected in the Rio Jamanxim at the Município do Novo Progresso, Brazil (Fig. 6).

Remarks: Two specimens of A. janeae from the Rio Xingu listed by Schwassmann \& Carvalho (1985: 233; cited therein as A. blax) as MPEG 1370 are now registered as MZUSP 31491.

Etymology: The species name, Archolaemus janeae, is in honor of Jane Mertens of Humboldt Universität zu Berlin for her assistance to the second author.

Habitat: The holotype and a portion of the paratype series of $A$. janeae were collected between 21:00 and 23:00 h in the Rio Iriri, just upriver of a sandbank that separated the tributary from the mainstream Rio Xingu. In this area the river had a pronounced
current that was faster towards the centre of the channel, with the substrate in the sampled areas formed by pebbly sand. The depth of the fishing effort varied between $<10 \mathrm{~cm}$ and about 120 cm , and water parameters at that location were: $\mathrm{pH}=5.5$; $\mathrm{T}=31.1^{\circ} \mathrm{C}$ at night and $31.9^{\circ} \mathrm{C}$ during the day; dissolved oxygen (D.O.) $=5.9 \mathrm{mg} \mathrm{L}{ }^{-1} ; \quad$ conductivity $32.9 \mathrm{uScm}^{-1}$; and Secchi depth $=187 \mathrm{~cm}$ (H. LópezFernández, pers. comm.). Some non-type specimens of $A$. janeae that originated in the Rio Xingu were captured in the main stream of shallow rapids and small waterfalls, always over a rocky bottom (J.L.O. Birindelli, pers. comm.).

## Material examined

Holotype: BRAZIL. Pará: Rio Iriri, just upriver of its mouth into Rio Xingu, Município de Altamira ( $03^{\circ} 48^{\prime} 54^{\prime \prime} \mathrm{S}, \quad 052^{\circ} 37^{\prime} 09^{\prime \prime} \mathrm{W}$ ), collected by H. LópezFernández, G. Ortí, N. Meliciana, and C. Röepke, 15 August 2008; INPA 30832 (160).

Paratypes: BRAZIL. Pará: collected with holotype, INPA 36380, 16 (136-225; 2 CS); MPEG 21684, 1 (141); USNM 404355, 1 (191). Rio Xingu near cachoeira, collected by M. Goulding, 14 October 1983; MZUSP 31491, 2 (115-206).

Non-type specimens: BRAZIL. Pará: Rio Xingu, Cachoeira de Kaituká, Altamira ( $03^{\circ} 33^{\prime} 47$ "S, $51^{\circ} 51^{\prime} 20^{\prime} W$ ); MZUSP 3930, 13 (113-340). Rio Xingu, Furo da Mirandolina (approximately $03^{\circ} 12^{\prime} 43^{\prime \prime} \mathrm{S}$, $\left.052^{\circ} 12^{\prime} 12^{\prime \prime} \mathrm{W}\right)$; INPA 4131, 1 (221). Rio Jamanxim, Rio Tapajós basin, Novo Progresso near Vila Mil ( $07^{\circ} 43^{\prime} 51^{\prime \prime} \mathrm{S}, 055^{\circ} 16^{\prime} 36^{\prime \prime} \mathrm{W}$ ); MZUSP 97383, 10 (197402). Rio Jamanxim, Rio Tapajós basin, small beach near Novo Progresso ( $09^{\circ} 17^{\prime} 59^{\prime} \mathrm{S}, 054^{\circ} 50^{\prime} 00^{\prime} \mathrm{W}$ ); MZUSP 97514, 9 (213-222). Rio Iriri ( $03^{\circ} 48^{\prime} 49^{\prime \prime} \mathrm{S}$, $\left.052^{\circ} 38^{\prime} 06^{\prime \prime} \mathrm{W}\right)$, MZUSP 105843, 1 (300). Rio Xingu, above Belo Monte (approximately $03^{\circ} 17^{\prime} \mathrm{S}, 052^{\circ} 12^{\prime} \mathrm{W}$ ); MZUSP 106054, 1 (341). Rio Xingu at Altamira, near Kaituká ( $\left.03^{\circ} 33^{\prime} 48^{\prime \prime} \mathrm{S}, 051^{\circ} 51^{\prime} 49^{\prime \prime} \mathrm{W}\right)$; MZUSP 106223, 1 (372). Rio Xingu, Boa Esperança ( $03^{\circ} 33^{\prime} 44^{\prime \prime} \mathrm{S}$, $052^{\circ} 20^{\prime} 59^{\prime \prime} \mathrm{W}$ ); MZUSP 106287, 1 (only anterior half of specimen). Rio Xingu, Arroz Crú, Altamira ( $03^{\circ} 22^{\prime} 05^{\prime} \mathrm{S}, 051^{\circ} 58^{\prime} 02^{\prime} \mathrm{W}$ ); MZUSP 106290, 1 (209); MZUSP 106049, 1 (230). Mato Grosso: Rio Xingu basin, Rio Culuene, Paranatinga, cachoeira at site of Pequenia Centrais Hidreléctica Paranatinga II ( $13^{\circ} 51^{\prime} 08^{\prime} \mathrm{S}, ~ 053^{\circ} 15^{\prime} 22^{\prime \prime} \mathrm{W}$ ); MZUSP 89741, 3 (191232). Rio Xingu basin, Rio Culuene, Paranantinga, cofferdam of Pequena Centrais Hidreléctica 2 ( $13^{\circ} 49^{\prime} 00^{\prime \prime} \mathrm{S}, 053^{\circ} 15^{\prime} 00^{\prime \prime} \mathrm{W}$ ); MZUSP 94909, 6 (253368).

## Archolaemus luciae sp. nov. (Figs 6, 7; Table 2)

Archolaemus blax, Schwassmann \& Carvalho 1985: 233 (in part, specimens from Rio Tapajós and

Rio Cupixi; not species redescription or biological information). Ferreira, 1995: 51 (Brazil, Rio Trombetas basin, Cachoeira Porteira, Cachoeira Vira-Mundo, inhabitant of swift-flowing waters).

Diagnosis: Archolaemus luciae is diagnosed from A. blax by the depth of the caudal filament (3.3$4.8 \%$ CL versus $5.0-6.5 \% \mathrm{CL}$ ), the length of the coronomeckelian bone (50\% or more the length of Meckel's cartilage versus less than $20 \%$ the length of Meckel's cartilage), and the length of the posterior ceratohyal (approximately the same length as the ventral hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus luciae is diagnosed from A. ferreirai in the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris versus falling short of that line) and the distribution of teeth on the internal surface of the endopterygoid (between seven and ten irregularly distributed teeth versus five or six teeth arranged in a single row). Archolaemus luciae is diagnosed from A. janeae in the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris versus falling short of that line), the length of the caudal filament (28.6-46.1\% LEA versus 16.0-25.7\% LEA), the depth of the caudal filament (3.3-4.8\% CL versus $5.6-11.2 \% \mathrm{CL}$ ), the form of the premaxilla (the anteroposterior length greater than the transverse width versus the anteroposterior length equal to the transverse width), the distribution of the teeth on the dentary (dentition restricted to the anterior one-half or slightly more of the dorsal margin versus occupying most of the dorsal margin), the length of the coronomeckelian bone ( $50 \%$ or more of the length of Meckel's cartilage versus less than 20\% of the length of Meckel's cartilage), and the length of the posterior ceratohyal (approximately the same length as the ventral hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus luciae is diagnosed from $A$. orientalis by the pigmentation pattern (the presence of a narrow dark stripe along the lateral line and a broad band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin, versus the lack of such dark pigmentation), the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris versus falling short of that line), the total number of pectoral-fin rays (19-21 versus 16), the total number of anal-fin rays (192-213 versus 164 186), the form of the premaxilla (the anteroposterior length greater than the transverse width versus the anteroposterior length equal to the transverse width), and the length of the coronomeckelian bone ( $50 \%$ or more the length of Meckel's cartilage versus less than $20 \%$ the length of Meckel's cartilage). Archolaemus luciae is diagnosed from A. santosi by the pigmenta-


Figure 7. A, Archolaemus luciae sp. nov., holotype, female, 267 mm total length, INPA 20960; Brazil, Pará, Rio Trombetas, Cachoeira Porteira, below Furo nas Pedras (approximately $01^{\circ} 05^{\prime} \mathrm{S}, 057^{\circ} 02^{\prime} \mathrm{W}$ ); B, Archolaemus luciae sp. nov., male, 497 mm total length, INPA 6767; Brazil, Pará, Rio Jamanxim, Rio Tapajós basin, Ilha Terra Preta ( $05^{\circ} 27^{\prime} 04^{\prime \prime} \mathrm{S}, 055^{\circ} 52^{\prime} 40^{\prime \prime} \mathrm{W}$ ).
tion pattern (the presence of a narrow dark stripe along the lateral line and a broad band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin versus the lack of such dark pigmentation), the length of the gape (the rictus extending posteriorly beyond the vertical through the posterior naris versus falling short of that line), the length of the caudal filament (28.6-46.1\% of LEA versus 14.4$27.0 \%$ ), the depth of the caudal filament (3.3-4.8\% of CL versus $6.7-9.1 \%$ ), and the length of the coro-
nomeckelian bone ( $50 \%$ or more of the length of Meckel's cartilage versus less than $20 \%$ the length of Meckel's cartilage).

Description: Morphometric data for the examined specimens of A. luciae are presented in in Table 2.

Body elongate and laterally compressed. Greatest body depth located along abdominal cavity or slightly posterior to that region. Body profile gently convex from rear of head for approximately anterior one-third
of body, and then gradually angled posteroventrally along progressively tapering body and caudal filament. Ventral profile of body distinctly convex anteriorly along anterior half of abdominal cavity, and then gradually angled posterodorsally along base of anal fin and caudal filament. Anteriormost perforated lateral line scale located approximately at vertical through origin of pectoral fin. Lateral line continuous, extending from immediately posterior of vertical through origin of pectoral fin to end of caudal filament.

Head laterally compressed, widest at opercular region and deepest at nape. Dorsal profile of head varying from nearly straight to slightly concave along antorbital region, and then from straight to slightly convex from that point to rear of head. Ventral profile of head straight and posteroventrally aligned. Snout subconical, anteroventrally directed, and moderately developed to elongate with elongation particularly pronounced in largest examined specimen (INPA 6767; 497 mm TL). Eye small and located laterally on dorsal half of head. Orbital rim covered by membrane and attached to surface tissues of adjoining region of head in six specimens of approximately $97-135 \mathrm{~mm}$ TL, partially free on one side in two specimens of approximately $96-97 \mathrm{~mm}$ TL, and without membrane and unattached in individuals larger than 135 mm TL. Anterior naris located at end of short tube and positioned short distance posterior of tip of snout, but distant from anterior margin of eye. Posterior naris elliptical, without tube, and located nearer tip of snout than anterior margin of eye. Mouth distinctly inferior, with upper jaw clearly longer than, and overlapping, lower jaw. Gape relatively long and extending posteriorly beyond vertical through posterior margin of posterior nares. Branchial opening moderately elongate, located along posterior margin of opercle and immediately anterior to anterior margin of pectoral-fin origin. Branchial membranes joined at isthmus, and extending posteriorly to under pectoral-fin base. Anus situated proximate to elongate urogenital papilla, and both structures positioned approximately ventral of orbit. Position of anus and urogenital papilla shifting ontogenetically from posterior of vertical through posterior margin of orbit to under or slightly anterior of orbit.
Scales small, cycloid, and present from immediately posterior of head to end of caudal filament. Between 127 and 148 (127) scales along lateral line ( $N=15$ ). Between 13 and 16 scales above lateral line at midbody (14) ( $N=29$ ). Scales absent on head.

Pectoral fin long, approximately two-thirds of head length, broad and distally pointed, with rounded margin, and lateral rays longer overall. Pectoral-fin rays between ii, 17 and ii, 19 (19) ( $N=29$ ). Anal-fin elongate, extending from vertical through base of
pectoral-fin base along most of length of body. Fin margin gently convex anteriorly, and then straight for length of fin, but with rays becoming progressively shorter posteriorly. Anal fin with 22-32 (24) unbranched rays $(N=29)$ and 192 to 213 (192) total fin rays ( $N=29$ ). Caudal filament long, with elongation particularly pronounced in largest examined specimen (INPA 6767; 497 mm TL).

Coloration in alcohol: Available specimens somewhat faded compared with obvious coloration in live specimens (see Coloration in life). Overall ground head and body coloration tan. Body with obvious but not dark, narrow stripe along lateral line. Extent of stripe variable anteriorly, even in specimens of comparable size, with anterior limit ranging from above centre of pectoral fin to point slightly posterior to vertical through tip of pectoral fin. Body pigmentation darker than overall ground pigmentation along dorsal portion of body. Area overlying basal anal-fin pterygiophores with pterygiophores outlined by very narrow dark surface bars that cumulatively form dusky broad band running along ventral portion of body, with depth of band gradually decreasing posteriorly. Dusky stripe along lateral line and dark ventral band above anal fin separated by more lightly pigmented, broad midlateral stripe. Head lightly coloured ventrally, and somewhat darker on opercle and above horizontal between tip of snout and areas somewhat below orbit. Pectoral and anal fins hyaline.

Coloration in life: (Based on recently collected specimens from Rio Teles Pires, Rio Tapajós basin.) Head dark overall, more so dorsal of horizontal between mouth and base of pectoral fin. Body dark from lateral line dorsally, with lateral line more intensely pigmented. Basal portions of anal-fin basal pterygiophores outlined by dark bars on body surface; pigmentation cumulatively forming dark band above base of fin, with height of band gradually decreasing posteriorly. Pigmentation overlying distal portions of basal pterygiophores less intense than that near base of fin, resulting in faint dusky band variably separate, other than anteriorly, from dark band along ventral margin of body. Pectoral fin dusky. Anal fin with rays outlined by dark pigmentation and dusky overall.

Distribution: Archolaemus luciae is known within the Amazon basin from the Rio Trombetas basin at Cachoeira Porteira, Cachoeira Vira-Mundo, and the Rio Mapuera, all in the state of Pará, as well as the Rio Tapajós basin, where it has been collected in the Rio Jamanxim and Rio Teles Pires in the state of Mato Grosso. In the state of Amapá, north of the lower Amazon, it has been collected in the Rio Jari that
empties into the Amazon and the Rio Araguari that drains independently into the Atlantic Ocean (Fig. 6).

Secondary sexual dimorphism: Archolaemus luciae demonstrates pronounced sexual dimorphism in the form of the snout in mature individuals (Fig. 7). Whereas the snout in mature females of the species is overall comparable with that in mature males and females of various congeners, the snout in the large male of A. luciae is distinctly lengthened and more massive anteriorly.

Habitat: Ferreira (1995:51) reported A. luciae (as A. blax) from swift-flowing water habitats around Cachoeira Porteira and Cachoeira Vira-Mundo in the Rio Trombetas basin. Some of the non-type specimens of the species that originated in the Rio Tapajós were captured in the main stream of shallow rapids and small waterfalls, always over a rocky bottom (J. L. O. Birindelli, pers. comm.).

Etymology: The species name, Archolaemus luciae, is in honor of Lucia Py-Daniel of the Instituto Nacional de Pesquisas da Amazônia, in recognition of her many contributions to the knowledge of the fishes of the Amazon and her assistance to the authors over the years.

## Material examined

Holotype: BRAZIL. Pará: Rio Trombetas, Cachoeira Porteira, below Furo nas Pedras (approximately $01^{\circ} 05^{\prime} \mathrm{S}, 057^{\circ} 02^{\prime} \mathrm{W}$ ), collected by Equipe de Ictiologia do INPA, 19 April 1985; INPA 20960 (267).

Paratypes: BRAZIL. Collected with holotype: INPA 36381, 3 (137-276); Rio Trombetas, Cachoeira Porteira (approximately $01^{\circ} 05^{\prime} \mathrm{S}, 057^{\circ} 02^{\prime} \mathrm{W}$ ), collected by Equipe de Ictiologia do INPA, 19 April 1985; INPA 20962, 23 (40-202); INPA 20964, 12 (110-210; 4 CS); MPEG 21685, 1 (138); MZUSP 110184 (135); USNM 404356 (158). Rio Mapuera, Cachoeira Porteira (approximately $01^{\circ} 05^{\prime} \mathrm{S}, 057^{\circ} 02^{\prime} \mathrm{W}$ ); collected by E. Ferreira \& M. Jegú; INPA 20961, 2 (130-180).
Non-type specimens: BRAZIL. Amapá: Rio Cupixi, Rio Araguari basin, bridge on road to Serra do Navio, in channel of river (approximately $00^{\circ} 53^{\prime} 45^{\prime \prime} \mathrm{N}$, $052^{\circ} 00^{\prime} 07^{\prime} \mathrm{W}$ ); MZUSP 34920, 1 (238). Rio Iratapuru, left bank tributary of Rio Jari, Laranjal do Jari, at Cachoeira de São Raimundo, above Cachoeira de Santo Antonio ( $\left.00^{\circ} 33^{\prime} 59^{\prime \prime} \mathrm{S}, \quad 052^{\circ} 34^{\prime} 40^{\prime \prime} \mathrm{W}\right)$; MZUSP 103446, 1 (206). Pará: Rio Trombetas, Oriximiná, above Cachoeira Porteira (approximately $01^{\circ} 03^{\prime} \mathrm{S}, 057^{\circ} 01^{\prime} \mathrm{W}$ ); INPA 3289, 1 (210). Rio Trombetas, Oriximiná, near igarapé Caxipacoré (approximately $01^{\circ} 03^{\prime} \mathrm{S}, \quad 057^{\circ} 01^{\prime} \mathrm{W}$ ); INPA 3354, 1 (173); INPA 3386, 1 (160). Rio Trombetas, Oriximiná, above Cachoeira Vira-Mundo (approximately $01^{\circ} 03^{\prime} \mathrm{S}$,
$\left.057^{\circ} 01^{\prime} \mathrm{W}\right)$; INPA 3356, 3 (230-269); INPA 3378, 1 (250). Rio Mapuera, Cachoeira Porteira (approximately $01^{\circ} 05^{\prime} \mathrm{S}, 057^{\circ} 02^{\prime} \mathrm{W}$ ); INPA 4371, 10 (140-235); INPA 20957, 2 (130-141). Rio Trombetas, Cachoeira Porteira (approximately $01^{\circ} 05^{\prime} \mathrm{S}, \quad 057^{\circ} 02^{\prime} \mathrm{W}$ ); INPA 4372, 1 (102); INPA 19989, 1 (275); INPA 19990, 1 (181); INPA 20965, 1 (60). Trombetas, Oriximiná, Cachoeira Vira-Mundo (approximately $01^{\circ} 05^{\prime} \mathrm{S}, 057^{\circ} 02^{\prime} \mathrm{W}$ ); INPA 5081, 1 (220); INPA 5082, 1 (180). Rio Jamanxim, Rio Tapajós basin, Ilha Terra Preta ( $\left.05^{\circ} 27^{\prime} 04^{\prime \prime} \mathrm{S}, \quad 055^{\circ} 52^{\prime} 40^{\prime \prime} \mathrm{W}\right)$; INPA 6767, 1 (497); INPA 6812, 6 (92-130). Rio Tapajós, São Luis do Tapajós (approximately $04^{\circ} 25^{\prime} \mathrm{S}, \quad 056^{\circ} 13^{\prime} \mathrm{W}$ ); MZUSP 24268, 1 (400). Rio Tapajós basin, Jacareacanga, rapids along Rio Teles Pires, below Sete Quedas (approximately $06^{\circ} 13^{\prime} 20^{\prime} \mathrm{S}, \quad 057^{\circ} 45^{\prime} 10^{\prime \prime} \mathrm{W}$ ); MZUSP 96130, 1 (229). Mato Grosso: Rio Tapajós, Paranaíta, Rio Teles Pires, near ferry crossing of rodovia MT-416; MZUSP 95630, 4 (148-186). Rio Peixoto de Azevedo, Município do Peixoto de Azevedo, tributary of Rio Teles Pires near Peixoto de Azevedo (approximately $10^{\circ} 10^{\prime} \mathrm{S}, 054^{\circ} 50^{\prime} \mathrm{W}$ ); MZUSP 96618, 1 (203). Rio Teles Pires, Itaúba, below mouth of Rio Renato (approximately $11^{\circ} 03^{\prime} 43^{\prime \prime} \mathrm{S}, \quad 055^{\circ} 16^{\prime} 33^{\prime \prime} \mathrm{W}$ ); MZUSP 99362, 2 (363-445). Rio Tapajós, Itaúba, Rio Teles Pires (approximately $11^{\circ} 39^{\prime} 11^{\prime \prime} \mathrm{S}, 055^{\circ} 42^{\prime} 14^{\prime \prime} \mathrm{W}$ ); MZUSP 95909, 1 (409). Rio Braço Norte, Novo Mundo, above Pequena Centrais Hidreléctica Braço Norte IV (approximately $09^{\circ} 47^{\prime} \mathrm{S}, 054^{\circ} 54^{\prime} \mathrm{W}$ ); MZUSP 106461, 1 (292). Rio Teles Pires, Município da Paranatinga, Fazenda do Antônio Bezerra (approximately $\left.10^{\circ} 20^{\prime} \mathrm{S}, 054^{\circ} 45^{\prime} \mathrm{W}\right)$; MZUSP 107923, 1 (447).

## Archolaemus orientalis sp. nov. Stewart, Vari, de Santana \& Wosiacki (Figs 8, 9; TABLE 2)

Diagnosis: Archolaemus orientalis is diagnosed from all congeners by the pigmentation pattern (the lack of a narrow dark stripe along the lateral line and absence of a broad band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin, versus the presence of such dark pigmentation). Archolaemus orientalis is further distinguished from A. blax by the length of the gape (the rictus falling short of the vertical through the posterior border of the posterior nares versus extending beyond that line), the total number of pectoral-fin rays ( 16 versus $19-22$ ), the total number of anal-fin rays (164-186 versus 193-222), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), and the length of the posterior ceratohyal (approximately the same length as the ventral hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus orien-


Figure 8. Archolaemus orientalis sp. nov., holotype, 156 mm total length, MPEG 21508; Brazil, Minas Gerais, Rio São Francisco system, Rio Piracutu basin, Município Buritizerio, Rio do Sono ( $17^{\circ} 18^{\prime} \mathrm{S}, 045^{\circ} 20^{\prime} 59^{\prime \prime} \mathrm{W}$ ).


Figure 9. Map of upper portions of Rio São Francisco basin and adjoining areas showing geographical distribution of Archolaemus orientalis sp. nov. (1, holotype locality; some symbols represent more than one locality and/or lot of specimens).
talis is additionally diagnosed from A. ferreirai in the total number of pectoral-fin rays (16 versus 18-21), the total number of anal-fin rays (164-186 versus 190-222), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), and the length of the coronomeckelian bone (less than $20 \%$ the length of Meckel's cartilage versus $50 \%$ or more the length of Meckel's cartilage).

Archolaemus orientalis is further diagnosed from $A$. janeae in the total number of pectoral-fin rays (16 versus 19-21), the total number of anal-fin rays (164186 versus 205-228), the distribution of the teeth on the dentary (dentition restricted to the anterior onehalf or slightly more of the dorsal margin of the dentary versus occupying most of the dorsal margin of the dentary), and the length of the posterior ceratohyal (approximately the same length as the ventral
hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus orientalis is additionally diagnosed from A. luciae in the length of the gape (the rictus falling short of the vertical through the posterior border of the posterior nares versus extending beyond that line), the total number of pectoral-fin rays ( 16 versus $19-21$ ), the total number of anal-fin rays ( $164-186$ versus $192-213$ ), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), and the length of the coronomeckelian bone (less than $20 \%$ of the length of Meckel's cartilage versus $50 \%$ or more of the length of Meckel's cartilage). Archolaemus orientalis is further diagnosed from A. santosi in the total number of pectoral-fin rays (16 versus 19-21) and the total number of anal-fin rays (164-186 versus 192 213).

Description: Morphometric data for the examined specimens of $A$. orientalis are presented in Table 2.

Body elongate and laterally compressed. Greatest body depth located along abdominal cavity or slightly posterior to that region. Body profile very gently convex from rear of head for approximately anterior one-third of body, and then gradually angled posteroventrally along progressively tapering body and caudal filament. Ventral profile of body straight to slightly convex along anteriormost portion of base of anal fin, and then gradually angled posterodorsally along base of anal fin and caudal filament. Anteriormost perforated lateral line scale located at vertical approximately at origin of pectoral fin. Lateral line continuous, extending from immediately posterior of vertical through origin of pectoral fin to end of caudal filament.
Head laterally compressed, widest at opercular region and deepest at nape. Head profile convex anteriorly proximate to upper lip, and then straight from that point to rear of head. Ventral profile straight and posteroventrally aligned. Snout subconical and anteroventrally directed; length comparatively short. Eye small and located laterally on dorsal half of head. Orbital rim free, without membrane or attachment to surface tissues of adjoining region of head. Anterior naris located at end of short tube and positioned short distance posterior of tip of snout, but distant from anterior margin of eye. Posterior naris elliptical, without tube, and located approximately same distance from tip of snout and anterior margin of eye in adults. Mouth distinctly inferior, with upper jaw clearly longer than, and overlapping, lower jaw. Gape short in specimens of all sizes, extending posteriorly to point short of vertical through posterior naris. Branchial opening moderately elongate, located along posterior margin of opercle and immediately anterior
to anterior margin of pectoral-fin origin. Branchial membranes joined at isthmus and extending posteriorly to under pectoral-fin base. Anus proximate to elongate urogenital papilla, and both structures positioned approximately ventral of orbit. Position of anus and urogenital papilla shifting ontogenetically from posterior of vertical, through posterior margin of orbit, to under orbit.

Scales small, cycloid, and present from immediately posterior of head to end of caudal filament. Scales along lateral line 120 to $143(143)(N=2)$. Fifteen scales above lateral line at midbody (15) $(N=2)$. Scales absent on head.

Pectoral fin long, approximately two-thirds of head length, broad, and distally pointed with lateral rays longer. Pectoral-fin rays ii, $16(16)(N=4)$. Analfin elongate, extending from vertical through base of pectoral-fin base along most of length of body. Fin margin gently convex anteriorly, and then straight for much of length of fin, but with rays becoming progressively shorter posteriorly. Anal fin with 27-32 (27) unbranched rays $(N=2$ ) and 164-186 (164) total fin rays $(N=3)$. Caudal filament of moderate length.

Coloration in alcohol: Overall body coloration of head and body tan, with scattered small dark spots somewhat more concentrated dorsally, but not forming any discrete pigmentation pattern. Subset of specimens with series of small dark spots aligned on body surface between basal pterygiophores of anal fin, but with that region of body otherwise hyaline. Line of contact between hypaxial and epaxial muscle sometimes gives appearance of narrow dark line in vicinity of lateral line. Pectoral and anal fins hyaline.

Distribution: Archolaemus orientalis sp. nov. is only known from a relatively limited portion of the Rio São Francisco in the state of Minas Gerais, Brazil (Fig. 9).

Habitat: The holotype and paratype from the Rio do Sono in the Rio São Francisco system were collected in a portion of the river characterized by large rapids and a major waterfall, the Cachoeira das Almas, and rocky and sandy substrates. Those specimens were captured approximately 200 m downstream from the Cachoeira das Almas under a partially submerged grassy bank in swiftly flowing, approximately $1.5-\mathrm{m}$ deep water over a sand substrate (T. C. Pessali, pers. comm.)

Remarks: The description of $A$. orientalis from the Rio São Francisco basin represents the first record of the genus from outside the drainages within the Amazon catchment or the Rio Tocantins and Rio Araguari, which empty into or near the lower reaches of that massive drainage system. The holotype
(MPEG 21508) and the paratypes originating with it (MPEG 21509) were collected and originally intermingled with a large series of Eigenmannia microstoma (MPEG 20010). The two species are similar to a pronounced degree in their overall external appearances, albeit differing in the form of the mouth and the position of the eye.

Etymology: The species name, Archolaemus orientalis, from the Latin for 'of the east' refers to the presence of the species in the Rio São Francisco, the easternmost known occurrence of a species of Archolaemus.

## Material examined

Holotype: BRAZIL. Minas Gerais: Rio São Francisco system, Rio Piracutu basin, Município Buritizerio, Rio do Sono $\left(17^{\circ} 18^{\prime} \mathrm{S}, \quad 045^{\circ} 20^{\prime} 59^{\prime \prime} \mathrm{W}\right)$, collected by T. C. Pessali, 1 November 2010, MPEG 21508 (156).

Paratypes: BRAZIL. Minas Gerais: collected with holotype; MPEG 21509, 1 (150). Pirarapora, Rio São Francisco ( $\left.17^{\circ} 15^{\prime} \mathrm{S}, 045^{\circ} 00^{\prime} \mathrm{W}\right)$, collected by J. D. Haseman, 15 July 1907; FMNH 94418, 3 (120-181; 1 CS).

## ARCHOLAEMUS SANTOSI SP. NOV.

(Figs 10, 11; TABLE 2)
Diagnosis: Archolaemus santosi is diagnosed from A. blax in the length of the gape (the rictus falling short of the vertical through the posterior border of the posterior naris versus extending beyond that line), the depth of the caudal filament (6.7-9.1\% CL versus $5.0-6.5 \% \mathrm{CL}$ ), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), and the length of the posterior ceratohyal (approximately the same length as the ventral hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus santosi is diagnosed
from $A$. ferreirai in the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width) and the length of the coronomeckelian bone (less than $20 \%$ the length of Meckel's cartilage versus $50 \%$ or more the length of Meckel's cartilage). Archolaemus santosi is diagnosed from $A$. janeae by the head length to the rear of the opercle (15.0-16.3\% LEA versus 13.5-14.7\% LEA), the mouth width ( $7.0-10.3 \% \mathrm{HL}$ versus $11.0-19.1 \% \mathrm{HL}$ ), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), the distribution of the teeth on the dentary (dentition restricted to the anterior one-half or slightly more of the dorsal margin versus occupying most of the dorsal margin), and the length of the posterior ceratohyal (approximately the same length as the ventral hypohyal versus 1.5 times the length of the ventral hypohyal). Archolaemus santosi is diagnosed from A. luciae in the length of the gape (the rictus falling short of the vertical through the posterior border of the posterior nares versus extending beyond that line), the length of the caudal filament (14.4$27.0 \%$ LEA versus $28.6-46.1 \%$ LEA), the depth of the caudal filament (6.7-9.1\% CL versus $3.3-4.8 \% \mathrm{CL}$ ), the form of the premaxilla (the anteroposterior length equal to the transverse width versus the anteroposterior length greater than the transverse width), and the length of the coronomeckelian bone (less than $20 \%$ the length of Meckel's cartilage versus $50 \%$ or more of the length of Meckel's cartilage). Archolaemus santosi is diagnosed from $A$. orientalis by the pigmentation pattern (the presence of a narrow dark stripe along the lateral line and a broad band of dusky to dark pigmentation overlying the basal pterygiophores of the anal fin versus the lack of such dark pigmentation), the total number of pectoral-fin rays (19-21


Figure 10. Archolaemus santosi sp. nov., holotype, female, 197 mm total length, INPA 20966; Brazil, Rondônia, Rio Jamari, above site of future Usina Hidroelétrica Samuel (approximately $08^{\circ} 27^{\prime} \mathrm{S}, 063^{\circ} 30^{\prime} \mathrm{W}$ ).


Figure 11. Map of middle portions of Rio Madeira basin and adjoining areas, showing the geographical distribution of Archolaemus santosi sp. nov. (the dot indicates the holotype locality and more than one lot of specimens).
versus 16), and the total number of anal-fin rays (204-221 versus 164-186).

Description: Morphometric data for the examined specimens of $A$. santosi are presented in Table 2.

Body elongate and laterally compressed. Greatest body depth located along abdominal cavity in specimens of all sizes. Body profile gently convex from rear of head for approximately anterior one-third of body, and then gradually angled posteroventrally along progressively tapering body and caudal filament. Ventral profile of body straight and posteroventrally aligned in juveniles and convex anteriorly along anterior half of abdominal cavity in larger individuals, and then gradually angled posterodorsally along base of anal fin and caudal filament. Anteriormost perforated lateralline scale located at vertical situated approximately at origin of pectoral fin. Lateral line continuous, extending from immediately posterior of vertical through origin of pectoral fin to end of caudal filament.
Head laterally compressed, widest at opercular region and deepest at nape. Head profile convex anteriorly from margin of upper lip in specimens of all sizes, then convex to varying degrees along antorbital region, with concavity more pronounced in larger specimens, and then straight to slightly convex from that point to rear of head in specimens of all sizes. Ventral profile straight and posteroventrally aligned. Snout subconical and anteroventrally directed, mod-
erately elongate in smaller specimens to elongate in larger individuals. Eye small and located laterally on dorsal half of head. Orbital rim covered by membrane and attached to surface tissues of adjoining region of head in two specimens of approximately $73-104 \mathrm{~mm}$ TL, partially free on one side in an individual of 103 mm TL, and completely free in larger examined specimens. Anterior naris located at end of short tube and positioned short distance posterior of tip of snout, but distant from anterior margin of eye. Posterior naris elliptical, without tube, and located nearer to tip of snout than to anterior margin of eye in adults. Mouth distinctly inferior, with upper jaw clearly longer than, and overlapping, lower jaw. Gape short in adults, not extending posteriorly to vertical through anterior margin of posterior naris. Branchial opening moderately elongate, located along posterior margin of opercle and immediately anterior to anterior margin of pectoral-fin origin. Branchial membranes joined at isthmus and extending posteriorly to under pectoral-fin base. Anus situated proximate to elongate urogenital papilla, and both structures positioned approximately ventral of orbit. Position of anus and urogenital papilla shifting ontogenetically from distinctly posterior of vertical through posterior margin of orbit in juveniles, to along vertical through eye in largest specimens.

Scales small, cycloid, and present from immediately posterior of head to end of caudal filament. Scales
along lateral line 132 to 147 (147) ( $N=10$ ). Scales above lateral line at midbody 14 to $17(17)(N=12)$. Scales absent on head.

Pectoral fin long, approximately two-thirds of head length, broad, and distally pointed, with lateral rays longer. Pectoral-fin rays between ii, 19 and ii, 21 (21) ( $N=12$ ). Anal-fin elongate, extending from vertical through base of pectoral fin along most of length of body. Fin convex anteriorly, and then straight for length of fin, but with rays becoming progressively shorter posteriorly. Anal fin with 26-35 unbranched rays (26) ( $N=12$ ) and $204-221$ total fin rays (208) ( $N=12$ ). Caudal filament of moderate length.

Coloration in alcohol: Overall ground body coloration ranging from tan to brown. Head more lightly pigmented ventrally in specimens across all intensities of dark coloration, with dorsal two-thirds of head together with most of opercle distinctly darker in largest examined specimen. Narrow, dark stripe overlying lateral line for varying distances. Dark pigmentation along lateral line starts posterior of vertical through tip of pectoral fin in very small individuals, but extends anteriorly to beginning of lateral line in larger specimens. Basal anal-fin pterygiophores outlined on body surface by very narrow dark bars that cumulatively form dusky broad band running along ventral portion of body, with height of band gradually decreasing posteriorly. Surface bars along pterygiophore margins expand ontogenetically and coalesce to varying degrees to form solid dark band in larger specimens. Dark stripe along lateral line and dark ventral band separated by more lightly pigmented, broad midlateral stripe. Some specimens with ventral surface of the head tan, but most often with head dark over all surfaces. Pectoral fin hyaline overall, with scattered dark chromatophores. Anal fin hyaline.

Distribution: Archolaemus santosi is only known from the type locality at the Rio Jamari in Rondônia, Brazil (Fig. 11).

Habitat: The area of the type locality of $A$. santosi was reported by Santos (1996: 249) as being rapids (= corredeira), indicative of the rapid water flow conditions at the site.

Remarks: Archolaemus santosi is only known from juveniles and mature females.

Etymology: The species name, Archolaemus santosi, is in honor of Geraldo Mendes dos Santos of the Instituto Nacional de Pesquisas da Amazônia, the collector of the type series, in recognition of his many contributions to our knowledge of the Amazonian fish fauna.

Material examined
Holotype: BRAZIL. Rondônia, Rio Jamari, above site of Usina Hidroelétrica Samuel (approximately $08^{\circ} 27^{\prime} \mathrm{S}, 063^{\circ} 30^{\prime} \mathrm{W}$ ), station Polo 666, collected by G. M. Santos, 4 April 1985; INPA 20966 (197).

Paratypes: BRAZIL. Collected with holotype, INPA 36382, 9 (73-212; 3 CS); MPEG 21686, 1 (100); MZUSP 110185, 1 (132); USNM 404357, 1 (100).

## COMPARATIVE CLEARED AND STAINED MATERIAL EXAMINED

Apteronotidae. Apteronotus cf. albifrons, MPEG 2434, 1 (114), Brazil, Rio Goiapi. Adontosternarchus sachsi, INPA 18277, 2 (119-1220), Brazil, Rio Japurá. Compsaraia compsus, INPA 28916, 2 (116120), Brazil, Rio Negro. Sternarchogiton preto, INPA 289061 (69), Brazil, Rio Negro. Sternarchorhynchus mesensis, MNRJ 11611 (2), 130-140 mm, Brazil, Rio Tocantins.

Sternopygidae. Distocyclus conirostris, INPA 28879, 2 (142-239); INPA 28915, 2 (135-158), Brazil, Rio Negro. Eigenmannia limbata, INPA 18288, 2 (137-176), Brazil, Rio Japura. Eigenmannia cf. macrops, INPA 22687, 2 (65-153), Brazil, Rio Uatumã. Eigenmannia macrops, INPA 33259, 2 (67.7-94.1), Brazil, Rio Solimões. Eigenmannia gr. trilineata, MPEG 6887, 1 (167), Brazil, Rio Capim. Japigny kirschbaum, FMNH 50185, 3 (100-130), Guyana, New River. Rhabdolichops eastwardi, INPA 12361, 2 (60-172), Brazil, Rio Negro. Rhabdolichops troscheli, INPA 12363, 3 (70-174), Brazil, Rio Negro. Sternopygus astrabes, INPA 30502, 2 (112156), Brazil, Rio Negro. Sternopygus macrurus, INPA 4869, 4 (31.9-84.1), Brazil, Rio Solimões, Lago Castanho; INPA 16001, 1 (27), Brazil, Rio Urubu.

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## REFERENCES

Albert JS. 2001. Species diversity and phylogenetic systematics of American knifefishes (Gymnotiformes, Teleostei). Miscellaneous Publication, Museum of Zoology, University of Michigan 190: 1-127.
Albert JS. 2003. Family Sternopygidae (Glass knifefishes, Rattail knifefishes). In: Reis RE, Kullander SO, Ferraris CJ Jr, eds. Check list of the freshwater fishes of South and Central America. Porto Alegre: EDIPUCRS, 487-491.
Albert JS, Campos-da-Paz R. 1998. Phylogenetic systematics of Gymnotiformes with diagnoses of 58 clades: a review of available data. In: Malabarba LR, Reis RE, Vari RP, Lucena ZM, Lucena C, eds. Phylogeny and classification of Neotropical fishes. Porto Alegre: EDIPUCRS, 419-446.
Albert JS, Crampton WGR. 2005. Diversity and phylogeny of Neotropical electric fishes (Gymnotiformes). In: Bullock H, Hopkins CD, Popper AN, Fay RR, eds. Electroreception. 21. New York: Springer Handbook of Auditory Research, 360-409.
Albert JS, Fink WL. 1996. Sternopygus xingu, a new species of electric fish from Brazil (Teleostei: Gymnotoidei), with comments on the phylogenetic position of Sternopygus. Copeia 1996: 85-102.
Alves-Gomes JA. 1998. The phylogenetic position of the South American electric fish genera Sternopygus and Archolaemus (Ostariophysi: Gymnotiformes) according to 12 S and 16 S mitochondrial DNA sequences. In: Malabarba LR, Reis RE, Vari RP, Lucena ZM, Lucena C, eds. Phylogeny and classification of Neotropical fishes. Porto Alegre: EDIPUCRS, 446-459.
Alves-Gomes JA. 1999. Systematic biology of gymnotiform and mormyriform electric fishes: phylogenetic relationships, molecular clocks, and rates of evolution in the mitochondrial rRNA genes. Journal of Experimental Biology 202: 11671183.

Arratia G. 1992. Development and variation of the suspensorium of primitive catfishes (Teleostei: Ostariophysi) and their phylogenetic relationships. Bonner Zoologische Monographien 92: 1-149.
Bichuette ME, Trajano E. 2003. Epigean and subterranean ichthyofauna from the São Domingos karst area, upper Tocantins River basin, central Brazil. Journal of Fish Biology 63: 1100-1121.
Bichuette ME, Trajano E. 2006. Morphology and distribution of the cave knifefish Eigenmannia vicentespelaea Triques, 1996 (Gymnotiformes: Sternopygidae) from central Brazil, with an expanded diagnosis and comments on subterranean evolution. Neotropical Ichthyology 4: 99-106.
Bloom DD, Lovejoy NR. 2011. The biogeography of marine incursions in South America. In: Albert JS, Reis RE, eds. Historical biogeography of Neotropical freshwater fishes. Berkeley, CA: University of California Press, 137-144.
Camargo M, Giarrizzo T, Isaac V. 2004. Review of the
geographic distribution of fish fauna of the Xingu River basin, Brazil. Ecotropica 10: 123-147.
Campos JEG, Dardenne MA. 1997. Origem e evolução tectônica da bacia Sanfranscicana. Revista Brasileira de Geociêcias 27: 283-294.
Campos-da-Paz R. 1999. New species of Megadontognathus from the Amazon basin, with phylogenetic and taxonomic discussions on the genus (Gymnotiformes: Apteronotidae). Copeia 1999: 1041-1049.
Campos-da-Paz R. 2005. Previously undescribed dental arrangement amongst electric knifefishes, with comments on the taxonomic and conservation status of Tembeaussu marauna Triques (Otophysi: Gymnotiformes: Apteronotidae). Neotropical Ichthyology 3: 395-400.
Campos-da-Paz R. 2007. Família Sternopygidae. In: Buckup PA, Menezes NA, Ghazzi MS, eds. Catálogo das espécies de peixes de água doce do Brasil. Série livros 23. Rio de Janeiro: Museu Nacional, Universidade Federal do Rio de Janeiro, 121-122.
Correia SB, Crampton WGR, Albert JS. 2006. Three new species of the Neotropical electric fish Rhabdolichops (Gymnotiformes: Sternopygidae) from the Central Amazon, with a new diagnosis of the genus. Copeia 2006: 27-42.
Costa WJEM. 2003. The Simpsonichthys flavicaudatus species group (Cyprinodontiformes: Rivulidae: Cynolebiatinae): phylogenetic relationships, taxonomic revision and biogeography. Ichthyological Exploration of Freshwaters 14: 31-60.
Costa WJEM. 2010. Rivulus jalapensis, a new killifish from the Tocantins River basin, central Brazil (Cyprinodontiformes: Rivulidae). Ichthyological Exploration of Freshwaters 21: 193-198.
Crampton WGR. 2011. An ecology perspective on diversity and distributions. In: Albert JS, Reis RE, eds. Historical biogeography of Neotropical freshwater fishes. Berkeley, CA: University of California Press, 165-189.
Crampton WGR, Albert JS. 2006. Evolution of electric signal diversity in gymnotiform fishes. Part A. Phylogenetic systematics, ecology, and biogeography. In: Ladish R, Collin SP, Moller P, Kapoor BG, eds. Communication in fishes. Enfield: Science Publishers, 647-696.
Fearnside PM. 2006. Dams in the Amazon: Belo Monte and Brazil's hydroelectric development of the Xingu River basin. Environmental Management 38: 16-27.
Ferreira E, dos Santos GM, Jégu M. 1988. Aspectos ecológicos da ictiofauna do rio Mucajaí, na ilha da Paredão, Roraima. Amazoniana 3: 339-352.
Ferreira E, Zuanon J, Fosberg B, Goulding M, Briglia-Ferreira R. 2007. Rio Branco. Peixes, Ecologia e Conservação de Roraima. Manaus: Amazon Conservation Association, Instituto Nacional de Pesquisas da Amazônia, Sociedade Civil de Mamirauá.
Ferreira EFG. 1995. (issue for 1993). Composição, distribuição e aspectos ecológicos da ictiofauna de um trecho do rio Trombetas, na área de influência da future UHE cachoeira Porteira, Estado do Pará, Brasil. Acta Amazonica 23 (suplemento 1-4): 1-89.
Gayet M, Meunier F. 1991. Première découverte de Gym-
notiformes fossiles (Pisces, Ostariphysi) dans el Miocéne supériur de Bolivie. Comptes rendus hebdomadaires de l'Academie de Sciences de Paris 313: 471-476.
Hulen KG, Crampton WGR, Albert JS. 2005. Phylogenetic systematics and historical biogeography of the Neotropical electric fish Sternopygus (Teleostei: Gymnotiformes). Systematics and Biodiversity 3: 407-432.
Kirschbaum F. 1995. Discharge types of gymnotiform fishes. In: Moller P, ed. Electric fishes. History and behavior. London: Chapman \& Hall, 172-180.
Korringa M. 1970. A new gymnotoid fish from the Rio Tocantins, Brazil. Festschrift for George Sprague Myers. Proceedings of the California Academy of Sciences, Fourth Series 38: 265-271.
Kullander SO. 1988. Teleocichla, a new genus of South American rheophlic cichlid fishes with six new species (Teleostei: Cichlidae). Copeia 1988: 196-230.
Lima FCT, Caires RA. 2011. Peixes da Estação Ecológica Serra Geral do Tocantins, bacias dos Rios Tocantins e São Francisco, com observações sobre as implicações biogeográficas das ‘águas emendadas’ dos Rios Sapão e Galheiros. Biota Neotropica 11: 1-20.
Lima FCT, Ribeiro AC. 2011. Continental-scale tectonic controls of biogeography and ecology. In: Albert JS, Reis RE, eds. Historical biogeography of Neotropical freshwater fishes. Berkeley, CA: University of California Press, 145-164.
Lima FCT, Zuanon J. 2004. A new species of Astyanax (Characiformes: Characidae) from the rapids of the lower rio Xingu, Brazil. Neotropical Ichthyology 2: 117-122.
Lovejoy NR, Lester K, Crampton WGR, Marques FPL, Albert JS. 2010. Phylogeny, biogeography, and electric signal evolution of Neotropical electric fishes of the genus Gymnotus (Osteichthyes, Gymnotidae). Molecular Phylogenetics and Evolution 54: 278-290.
Lucinda PHF, Freitas IS, Soares AB, Marques EE, Agostinho CS, de Oliveira RJ. 2007. Fish, Lajeado reservoir, Rio Tocantins drainage, state of Tocantins, Brazil. Check List 3: 70-83.
Lujan NK, Armbruster JW. 2011. The Guiana Shield. In: Albert JS, Reis RE, eds. Historical biogeography of Neotropical freshwater fishes. Berkeley, CA: University of California Press, 211-224.
Lundberg JG, Cox-Fernandes C, Albert JS, Garcia M. 1996. Magosternarchus, a new genus with two new species of electric fishes (Gymnotiformes: Apteronotidae) from the Amazon River basin, South America. Copeia 1996: 657670.

Lundberg JG, Mago-Leccia F. 1986. A review of Rhabdolichops (Gymnotiformes, Sternopygidae), a genus of South American freshwater fishes, with description of four new species. Proceedings of the Academy of Natural Sciences of Philadelphia 138: 53-85.
Lundberg JG, Marshall LG, Guerrero J, Horton B, Malabarba MCSL, Wesselingh F. 1998. The stage for Neotropical fish diversification: a history of tropical South American rivers. In: Malabarba LR, Reis RE, Vari RP, Lucena ZM, Lucena C, eds. Phylogeny and classification of Neotropical fishes. Porto Alegre: EDIPUCRS, 13-48.

Mago-Leccia F. 1978. Los peces de la familia Sternopygidae de Venezuela. Acta Científica Venezolana 29: 1-91.
Mago-Leccia F. 1994. Electric fishes of the continental waters of America. Classification and catalogue of the electric fishes of the order Gymnotiformes (Teleostei: Ostariophysi) with descriptions of new genera and species. Caracas: Biblioteca de la academia de ciencias fisicas, matematics y naturales.
Mago-Leccia F, Lundberg JG, Baskin JN. 1985. Systematics of the South American freshwater fish genus Adontosternarchus (Gymnotiformes, Apteronotidae). Contributions in Sciences, Natural History Museum of Los Angeles County 358: 1-19.
Meunier FJ, Jégu M, Keith P. 2011. A new genus and species of Neotropical electric fish, Japigny kirschbaum (Gymnotiformes: Sternopygidae) from French Guiana. Cybium 35: 47-53.
Moller P. 1995. Electric fishes, history and behavior. New York: Chapman and Hall.
Montag LF, Freitas TG, Wosiacki WB, Barthem RB. 2008. Os peixes da Floresta Nacional de Caxiuanã (municípios de Melgaço e Portel, Pará - Brasil). The fishes of National Forest of Caxiuanã. (Melaka and Portal municipality, Pará State - Brazil). Boletim Museu Paraense Emílio Goeldi, Ciências Naturais 3: 1-34.
Nijssen H, Isbrücker IJH. 1972. On Hyopygus lepturus, a little know dwarf gymnotid fish from South America (Pisces, Cypriniformes, Gymnotoidei). Zoologische Mededelingen 47: 160-179.
de Santana CD, Vari RP. 2009. The South American electric fish genus Platyurosternarchus (Gymnotiformes: Apteronotidae). Copeia 2009: 233-244.
de Santana CD, Vari RP. 2010a. Electric fishes of the genus Sternarchorhynchus (Teleostei, Ostariophysi, Gymnotiformes); phylogenetic and revisionary studies. Zoological Journal of the Linnean Society 159: 223-371.
de Santana CD, Vari RP. 2010b. New rheophilic species of electric knifefish from the rapids and waterfalls of the lower Rio Xingu, Brazil (Gymnotiformes: Apteronotidae). Copeia 2010: 160-165.
Santos GM. 1996. Impactos da Hidelétrica Samuel sobre as communidades de peixes do rio Jamari (Rondônia, Brasil). Acta Amazonica 25: 247-280. (for 1995).
Schwassmann HO, Carvalho ML. 1985. Archolaemus blax Korringa (Pisces, Gymnotiformes, Sternopygidae): a redescription with note on ecology. Spixiana 8: 231240.

Taylor WR, Van Dyke GC. 1985. Revised procedures for staining and clearing small fishes and other vertebrates for bone and cartilage study. Cybium 9: 107-119.
Vari RP, Malabarba L. 1998. Neotropical Ichthyology: an overview. In: Malabarba LR, Reis RE, Vari RP, Lucena ZMS, Lucena CA, eds. Phylogeny and classification of Neotropical fishes. Porto Alegre: EDIPUCRS, 1-11.
Winemiller KO, Willis SC. 2011. The Vaupes Arch and Casiquiare Canal, barriers and passages. In: Albert JS, Reis RE, eds. Historical biogeography of Neotropical freshwater fishes. Berkeley, CA: University of California Press, 225242.


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[^1]:    $N$, number of specimens; H, holotype; range includes holotype of species.

